

# A novel nanobiotherapeutic poly-[hemoglobin-superoxide dismutase-catalase-carbonic anhydrase] with no cardiac toxicity for the resuscitation of a rat model with 90 minutes of sustained severe hemorrhagic shock with loss of 2/3 blood volume

Yuzhu Bian & Thomas Ming Swi Chang

*Artificial Cells and Organs Research Centre, Departments of Physiology, Medicine and Biomedical Engineering, Faculty of Medicine, McGill University, Montreal, QC, Canada*

## Abstract

We crosslink hemoglobin (Hb), superoxide dismutase (SOD), catalase (CAT), and carbonic anhydrase (CA) to form a soluble polyHb-SOD-CAT-CA nanobiotechnological complex. The obtained product is a soluble complex with three enhanced red blood cell (RBC) functions and without blood group antigens. In the present study, 2/3 of blood volume was removed to result in 90-min hemorrhagic shock at mean arterial blood pressure (MAP) of 30 mmHg. This was followed by the reinfusion of different resuscitation fluids, then followed for another 60 min. PolyHb-SOD-CAT-CA maintained the MAP at  $87.5 \pm 5$  mmHg as compared with 3 volumes of lactated Ringer's solution,  $43.3 \pm 2.8$  mmHg; blood,  $91.3 \pm 3.6$  mmHg; polyHb-SOD-CAT,  $86.0 \pm 4.6$  mmHg; poly stroma-free hemolysate (polySFHb),  $85.0 \pm 2.5$  mmHg; and polyHb,  $82.6 \pm 3.5$  mmHg. PolyHb-SOD-CAT-CA was superior to the blood and other fluids based on the following criteria. PolyHb-SOD-CAT-CA reduced tissue  $pCO_2$  from  $98 \pm 4.5$  mmHg to  $68.6 \pm 3$  mmHg. This was significantly ( $p < 0.05$ ) more effective than lactated Ringer's solution ( $98 \pm 4.5$  mmHg), polyHb ( $90.1 \pm 4.0$  mmHg), polyHb-SOD-CAT ( $90.9 \pm 1.4$  mmHg), blood ( $79.1 \pm 4.7$  mmHg), and polySFHb ( $77 \pm 5$  mmHg). PolyHb-SOD-CAT-CA reduced the elevated ST level to  $21.7 \pm 6.7\%$  and is significantly ( $< 0.05$ ) better than polyHb ( $57.7 \pm 8.7\%$ ), blood ( $39.1 \pm 1.5\%$ ), polySFHb ( $38.3\% \pm 2.1\%$ ), polyHb-SOD-CAT ( $27.8 \pm 5.6\%$ ), and lactated Ringer's solution ( $106 \pm 3.1\%$ ). The plasma cardiac troponin T (cTnT) level of polyHb-SOD-CAT-CA group was significantly ( $P < 0.05$ ) lower than that of all the other groups. PolyHb-SOD-CAT-CA reduced plasma lactate level from  $18 \pm 2.3$  mM/L to  $6.9 \pm 0.3$  mM/L. It was significantly more effective ( $P < 0.05$ ) than lactated Ringer's solution ( $12.4 \pm 0.6$  mM/L), polyHb ( $9.6 \pm 0.7$  mM/L), blood ( $8.1 \pm 0.2$  mM/L), polySFHb ( $8.4 \pm 0.1$  mM/L), and polyHb-SOD-CAT ( $7.6 \pm 0.3$  mM/L). PolyHb-SOD-CAT-CA can be stored for 320 days at room temperature. Lyophilized poly-Hb-SOD-CAT-CA can be heat pasteurized at 68F for 2 h. This can be important

if there is a need to inactivate human immunodeficiency virus, Ebola virus, and other infectious organisms.

**Keywords:** artificial cells, antioxidant, biotherapeutic, catalase, carbon dioxide carrier, carbonic anhydrase, cardiac toxicity, hemorrhagic shock, inactive infectious agents, ischemia–reperfusion, oxygen carrier, oxygen radicals, pasteurization, resuscitation, superoxide dismutase, storage stability

## Introduction

The blood cannot be pasteurized to inactivate potential infectious agents, and this had been a major problem during the human immunodeficiency virus (HIV) crisis in the year 1989. Other epidemics like Ebola may well come at any time. In addition, accidents, natural or man-made disasters, can result in severe injuries with extensive blood loss requiring urgent blood transfusion (Hess et al. 2011, Kim and Greenburg 2014). However, much time can be lost because of the need to transport the injured to the hospital for blood group test before transfusion can be given. There is a safety window of delay for blood replacement in severe hemorrhagic shock (Hess et al. 2011, Kim and Greenburg 2014, Chang 1964). Beyond this safety window, there can be problems related to ischemia–reperfusion injuries and irreversible shock. There are also problems related to the transport of donor blood, since it can only be stored for 1 day at room temperature and requires refrigeration at 4°C but even then only for about 42 days. Seven-micron-diameter red blood cells (RBCs) cannot easily perfuse into constricted microcirculation in severe hemorrhagic shock or partially obstructed vessels as in heart attack and stroke.

Polyhemoglobin (polyHb) is a water-soluble oxygen carrier based on the 1964 basic principle of nanobiotechno-

Copyright © 2014 Informa Healthcare USA, Inc. This is an open-access article distributed under the terms of the CC-BY-NC-ND 3.0 License which permits users to download and share the article for non-commercial purposes, so long as the article is reproduced in the whole without changes, and provided the original source is credited.

Correspondence: Professor Thomas Ming Swi Chang, Director, Artificial Cells and Organs Research Centre, Room 1004, Faculty of Medicine, McGill University, 3655 Promenade Sir William Osler, Montreal, QC H3G 1Y6, Canada. E-mail: artcell.med@mcgill.ca. website: www.medicine.mcgill.ca/artcell

(Received 15 July 2014; revised 1 September 2014; accepted 10 September 2014)

logical assembly of Hb molecules (Chang 1964, Chang 1971). There was no initial interest in this until the 1989 crisis of HIV-contaminated blood. It took 20 years to develop this and other modified Hb oxygen carriers for clinical trials (Gould et al. 1995, Chang 1997, Klein 2000, Winslow 2006, Chang 2005, Chang 2007, Jahr et al. 2008, Chang 2013, Burhop and Estep 2001, Natanson et al. 2008, Moore et al. 2009, Biro et al. 1995). It was too late for many patients infected with HIV from contaminated donor blood. We need to learn from this lesson and be ready in case there is another unexpected epidemic like Ebola.

Large-scale U.S. clinical trials on bovine polyHb have been carried out in surgical patients (Jahr et al. 2008). PolyHb has been approved for routine use in those countries, where these side effects are not as serious as the problem with insufficient supply of donor blood or HIV-contaminated donor blood (Kim and Greenburg 2014). A U.S. randomized clinical trial (Moore et al. 2009) in more than 700 prehospital ambulance patients for hemorrhagic shock shows that human polyHb can be given on the spot without the need for crossmatching. It can delay the need for blood transfusion for 12 h, whereas the control group that was given saline required blood transfusion in 30 min. There are nonfatal cardiac side effects in 3% of the polyHb group as compared with 0.6% in the control group.

Some other types of modified Hb have more severe side effects (Natanson et al. 2008). In conditions with endothelial dysfunction as in diabetes, there could be vasoconstriction (Yu et al. 2010). In severe ischemia as in sustained severe hemorrhagic shock, myocardial ischemia or cerebral ischemia, there is the potential for ischemia-reperfusion injuries (Biro et al. 1995, Alayash 2004, D'Agnillo and Chang 1998, Powanda and Chang 2002, Ma and Hsia 2013).

It is important to resolve the adverse effect for more widespread use. Especially, this is because now there is again the potential of infectious agents that can lead to epidemics and contaminated donor blood in the same way as the disastrous HIV crisis. In addition, large-scale wars, terrorisms, and major accidents that require on the spot use of blood substitutes can also happen any time. We are wiser now and know that we cannot wait for the needs to come before carrying out serious research to resolve the side effects.

PolyHb is only an oxygen carrier but in addition to the transport of oxygen, RBCs have other two important functions, removal of oxygen radicals and transport of carbon dioxide (CO<sub>2</sub>). We have shown earlier that polyHb needs antioxidant enzymes for the removal of oxygen radicals especially in ischemia-reperfusion (D'Agnillo and Chang 1998, Powanda and Chang 2002). This results in the removal of oxygen radicals and prevention of ischemia-reperfusion injuries (Powanda and Chang 2002, Chang et al. 2004, Nadithe and Bae 2011). Superoxide dismutase (SOD) can also remove superoxide that would otherwise remove nitrate oxide to form peroxyxynitrate. This may help in preventing the removal of nitrate oxide that can lead to vasoconstriction in conditions with endothelial dysfunction (Yu et al. 2010). An extension of this is oxygen carrier with synthetic antioxidant enzymes (Ma and Hsia 2013).

Do we need the third function of RBC—CO<sub>2</sub> transport (Geers and Gros 2000)—for the treatment of sustained severe hemorrhagic shock? The use of a novel intracellular pCO<sub>2</sub> microelectrode shows that in hemorrhagic shock, increase in intracellular pCO<sub>2</sub> is related to increase in mortality (Sims et al. 2001). In addition, increase in tissue pCO<sub>2</sub> is related to the severity of myocardial ischemia (Tronstad et al. 2010). Thus, we have added the CO<sub>2</sub> transport function of RBC by using carbonic anhydrase (CA) to form a soluble biotechnological complex of polyHb-SOD-catalase (CAT)-CA (Bian et al. 2011). With adjustments of the enzyme concentrations, this complex can have these RBC functions at enhanced levels.

Our earlier *in vivo* studies show that polyHb-SOD-CAT-CA could reduce the elevated tissue pCO<sub>2</sub> level in a rat model of hemorrhagic shock within the 60-min safety window (Bian et al. 2013). We now present our detailed research data in a 90-min severe hemorrhagic shock rat model with 2/3 blood volume loss, which is beyond the 60-min safety window for rats.

## Methods

### Materials and methods

The following are the places of purchase and the materials used in this experiment: Sigma: potassium phosphate, toluene, sodium phosphate, 99% lysine monohydrochloride, 25% glutaraldehyde, Sephacryl S-300HR, 37% hydrochloric acid, CAT, SOD, CA, 30% Brij 35 solution, Drabkin's solution, and 3% (w/w) hydrogen peroxide; Fisher: 1.5 × 98 cm chromatography column and Millipore 100 kDa centrifugal filter; B. Braun Medical Inc.: lactated Ringer's solution; McGill University (MacDonald campus): fresh bovine blood; and Biopure Cooperation: ultrapure bovine Hb.

### Preparation of SFHb

Stroma-free hemolysate (SFHb) is the content of RBCs with their membrane stroma removed. For preparing this, we used the method described in detail elsewhere (Chang 2007). Fresh bovine blood with heparin was centrifuged at 4000 g for 60 min at 4°C to remove the plasma supernatant and upper layer of cell pellet. The RBCs were washed four times with sterile, ice-cold saline and then suspended in twice the volume of potassium phosphate (12.5 mM; pH, 7.4) for 30 min. Then 2 volumes of ice-cold reagent-grade toluene was used to remove stromal lipids. The sample was centrifuged at 16,000 g for 2 h at 4°C to remove cellular debris. The concentration of Hb was tested using Drabkin's solution. Our analysis shows that SFHb contains enzymes (SOD, CAT, and CA) that are normally present in RBCs (Table I).

### Preparation of polyHb-SOD-CAT-CA, polySFHb, polyHb, and polyHb-SOD-CAT

#### PolyHb-SOD-CAT-CA

PolyHb-SOD-CAT-CA was prepared by the method described in detail elsewhere (Bian et al. 2011). SOD, CAT, and CA were added to SFHb with a final concentration as shown in Table I. These were crosslinked by glutaraldehyde to construct polyHb-SOD-CAT-CA. First, 1.3M lysine was added at

Table I. Composition of test liquids.

Test liquids	Type	Hb gm/dl	SOD Unit/dl	CAT Unit/dl	CA Unit/dl	P50 mmHg
Lactated Ringer's solution	solution	0	0	0	0	N/A
Blood	7 $\mu$ m	15	112,500	750,000	1,875,000	26.6
PolyHb	solution	10	0	0	0	18.2
PolySFHb	solution	10	70,000	470,000	950,000	19.15
PolyHb-SOD-CAT	solution	10	195,000	3,500,000	0	20.5
PolySFHb-SOD-CAT-CA	solution	10	195,000	3,500,000	1,360,000	19.92

a molar ratio of 7:1 lysine/Hb before the crosslinking reaction. The reaction vessel was flushed with nitrogen gas and kept at 4°C at all times to prevent the formation of metHb. Twenty milliliters of the mixture was placed on a shaker at 4°C for 1 h at 140 rpm. Without stopping the shaker, 0.7 ml of 5% glutaraldehyde was slowly added to it at a molar ratio of 16:1 glutaraldehyde/Hb at a rate of 0.15–0.20 mL every 5–10 min. The reaction mixture was allowed to crosslink in the shaker for 24 h at 4°C. The crosslinking was stopped by adding 2.0M lysine at a molar ratio of 200:1 lysine/Hb.

#### **PolySFHb**

PolySFHb was prepared using the same method, but using the stroma-free content of RBCs (SFHb) with the normal blood cell concentration of SOD, CAT, and CA without the addition of more enzymes.

#### **PolyHb**

PolyHb was prepared by crosslinking ultrapure Hb that contained no RBC enzymes following the above-mentioned procedure but without adding enzymes. This represented the polyHb used in clinical trial.

#### **PolyHb-SOD-CAT**

PolyHb-SOD-CAT was prepared following the first procedure mentioned above, except that ultrapure Hb was used with the additional CAT and SOD to make up the final concentration as shown in Table I. This represented the sample with O<sub>2</sub> transport and antioxidant functions but without CO<sub>2</sub> transport function.

#### **Purification and concentration**

Purification and concentration were carried out as follows: polyHb-SOD-CAT-CA, polySFHb, polyHb-SOD-CAT, and polyHb were each filtered using a sterile 0.45  $\mu$ m filter and dialyzed against lactated Ringer's solution overnight at 4°C using a dialysis membrane. A Sephacryl S-300 column in a 4°C cabinet was used to remove only the fraction of free tetrameric Hb with molecular weight greater than 300 Ka. The samples were placed in a dialysis membrane on a bed of Sephacryl/Gel absorbent at 4°C and concentrated to a final Hb concentration of 10 g/dL.

#### **The measurement of Hb concentration and SOD, CAT, and CA enzyme activities**

##### **Hb concentration**

The Hb concentration was determined by colorimetry.

The samples were first reacted with Drabkin's solution (Sigma-Aldrich).

Then the concentration of the resulting cyanmetHb solution was measured by spectrophotometry at 540 nm.

##### **SOD**

SOD activity was determined by HT Superoxide Dismutase Assay Kit (R&D Systems), which followed the inhibition of nitroblue tetrazolium reduction using xanthine/xanthine oxidase as a superoxide generator.

##### **CAT**

CAT measurement was based on the rate of disappearance of H<sub>2</sub>O<sub>2</sub> as described elsewhere (Chang 2007).

##### **CA**

For CA measurement, one Wilbur–Anderson (W-A) unit of CA activity was defined as the amount of enzyme that caused the pH of a 0.02 M Tris buffer to drop from 8.3 to 6.3. The reaction was initiated by the addition of substrate, and the time (T) needed for the pH of the reaction mixture to drop from 8.3 to 6.3 was recorded. A Fisher Accumet Basic pH Meter (Fisher Scientific, Pittsburgh, PA) with MI-407 (P) Needle pH Electrode (Microelectrodes Inc., Bedford, NH) was used to measure the change in pH caused by the hydration reaction of CO<sub>2</sub> catalyzed by CA. The control for the assay consisted of the same mixture without the test sample. The measurements in seconds were converted into W-A units according to the following formula:

$$1 \text{ W-A unit} = \frac{[2 \times (T_0 - T)]}{T}$$

The units were then plotted versus the Hb/CA concentration (Bian et al. 2011).

#### **Effects on tissue pCO<sub>2</sub>, mean arterial pressure, and electrocardiogram in a hemorrhagic shock rat model**

The Animal Ethics Committee of McGill University has approved this experimental protocol.

Sprague-Dawley rats, weighing 300  $\pm$  30 g, were randomly divided into 6 groups and each group contained 4 rats. Each of them was anesthetized with pentobarbital of 30 mg/g body weight. The animals were freely breathing and no ventilation support or oxygen was given during the protocol. This was to simulate hemorrhagic shock, where no ventilation support is available. Heparinized catheters were inserted into the right femoral vein and right femoral artery. Mean arterial pressure (MAP) was reduced by slowly withdrawing up to 2/3 volume of blood from the right femoral artery and maintained at 30 mmHg for 90 min. After this, each rat received one of

the following resuscitation fluids intravenously through the femoral vein in 15 min: (The detailed composition of each fluid is shown in Table I).

- (1) Sham control group: This group received the same general anesthesia, cannulation, and continuous monitoring for blood pressure, tissue  $p\text{CO}_2$ , and electrocardiogram. No blood was removed and no fluid was infused.
- (2) Whole blood shed by rats.
- (3) Lactated Ringer's solution, three volumes of shed blood.
- (4) PolyHb-SOD-CAT-CA in lactated Ringer's solution equivalent to the volume of shed blood.
- (5) PolySFHb in lactated Ringer's solution equivalent to the volume of shed blood.
- (6) PolyHb in lactated Ringer's solution equivalent to the volume of shed blood.
- (7) PolyHb-SOD-CAT in lactated Ringer's solution equivalent to the volume of shed blood.

MAP and electrocardiogram (ECG) were recorded using a Biopac Lab system (BIOPAC Systems Inc., Goleta, CA). Tissue  $p\text{CO}_2$  was measured by placing a  $p\text{CO}_2$  microelectrode (Lazar Research Labs, USA) on the surface of vastus medialis muscle at the incision site used for cannulation. MAP, tissue  $p\text{CO}_2$ , and ECG were followed throughout the experiment and continued for 60 min after the completion of the above reinfusions. After this, the rats were given IP Nembutal to terminate these acute studies. The blood was collected at the end of the resuscitation and centrifuged at 5000 g for 10 min at  $4^\circ\text{C}$  to collect the plasma. The plasma was stored at  $-80^\circ\text{C}$  before the assay. Cardiac troponin T (cTnT) was analyzed using a cTnT assay kit. Lactate assay kit was purchase from Biosource.

## Statistics

Primary endpoint is 60 min after the completion of the reinfusions of the resuscitation fluids. Data were expressed as mean  $\pm$  standard deviation (SD). Analysis of variance and Student's t-test were used to assess the difference between treatment groups. Statistical significance was defined as p value of less than 0.05 to reject a null hypothesis.

## Results

### Effects on mean arterial blood pressure

In the groups with 2/3 volume of the blood removed, the MAP fell to 30 mmHg. In each group after 90 min at MAP of 30 mmHg, reinfusions of resuscitation fluids with different compositions (Table I) had the effects that are shown in Figure 1. The results of these individual groups were pooled together for comparison (Figure 2). Here, a sham control group was also included. The sham control group showed no significant changes in MAP (Figure 2). When transfused with 3 volumes of lactated Ringer's solution, the MAP increased transiently from 30 mmHg to  $43.3 \pm 2.8$  mmHg after 1 h (Figures 1 and 2). Reinfusion of the lost blood (with Hb of 15 g/dl) maintained the MAP at  $91.3 \pm 3.6$  mmHg. When transfused with polyHb-SOD-CAT-CA (with Hb at 10 g/dL), the MAP was maintained at  $87.5 \pm 5$  mmHg when followed for 1 h after the infusion. PolySFHb, polyHb-SOD-CAT, and polyHb maintained the MAP at  $85.0 \pm 2.5$  mmHg,  $86.0 \pm 4.6$  mmHg, and  $82.6 \pm 3.5$  mmHg, respectively. However, there were other important criteria besides maintaining the MAP, which included changes in ST elevation, tissue  $p\text{CO}_2$ , plasma lactate, and the integrity of the intestine and other organs.

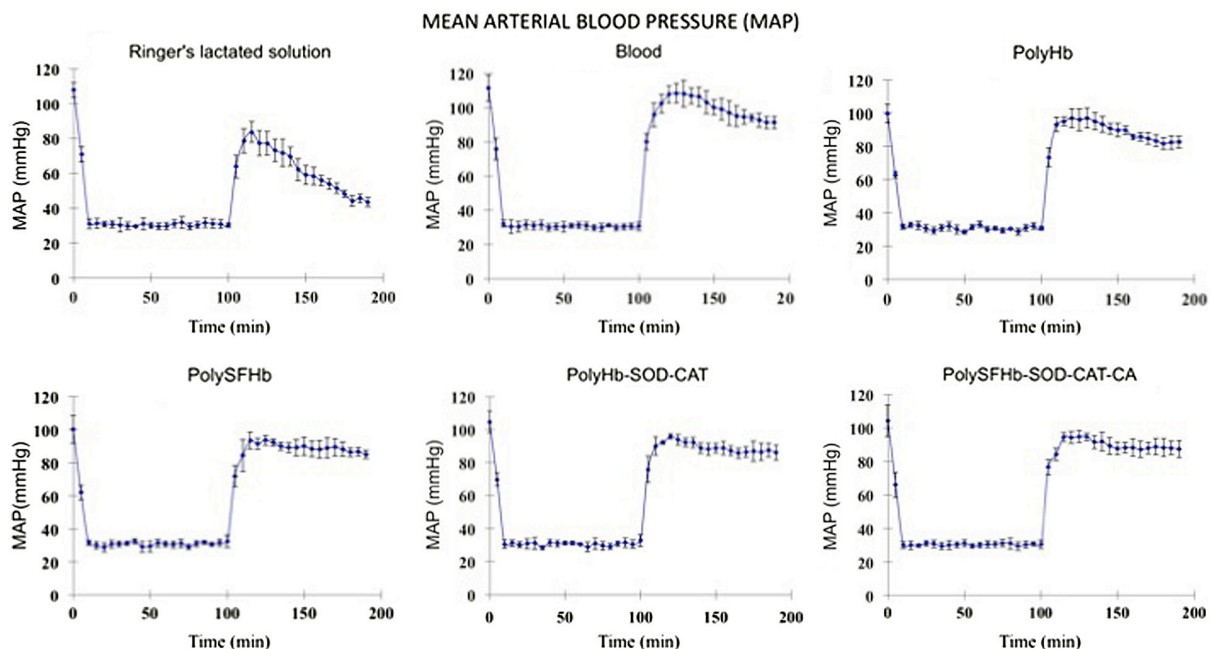


Figure 1. Mean arterial blood pressure. 2/3 volume of blood was removed to result in 90 min of hemorrhagic shock at MAP of 30 mmHg. After this, different fluids were reinfused. Reinfusion with 3 volumes of lactated Ringer's solution increased MAP transiently and fell to  $43.3 \pm 2.8$  mmHg. Blood (Hb 15 g/dl) maintained the MAP at  $91.3 \pm 3.6$  mmHg, while polyHb-SOD-CAT-CA (Hb 10 g/dL) maintained it at  $87.5 \pm 5$  mmHg, polyHb-SOD-CAT at  $86.0 \pm 4.6$  mmHg, polySFHb at  $85.0 \pm 2.5$  mmHg, and polyHb at  $82.6 \pm 3.5$  mmHg.

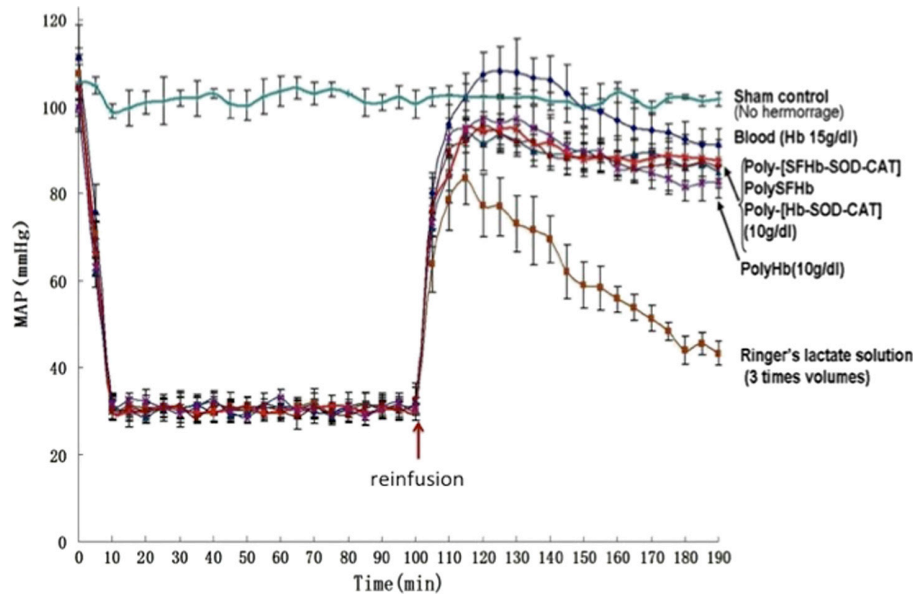


Figure 2. Comparison of mean blood pressures. The above individual measurements were pooled together for comparison. Sham control measurement, where no blood loss occurred, was added as control.

**Effects on elevated tissue pCO<sub>2</sub>**

The mean and standard deviation of each group were calculated and compared with the other groups. In the test groups with 2/3 volume of the blood removed, the baseline tissue pCO<sub>2</sub> of 54.7 ± 2.5 mmHg increased steadily to 98 ± 4.5 mmHg by 90 min (Figure 3). The effects on tissue pCO<sub>2</sub> during the 60-min period after the reinfusions of different resuscitation fluids are shown in Figures 3 and 4. The sham control group showed no significant changes in tissue pCO<sub>2</sub>

(Figure 4). The results of the comparison with other groups are as follows:

- (1) Reinfusion of lactated Ringer’s solution did not significantly ( $P > 0.05$ ) reduce the tissue pCO<sub>2</sub> level that remains at 98 ± 4.5 mmHg.
- (2) Rats that received their shed whole blood showed significant ( $P < 0.05$ ) reduction in the tissue pCO<sub>2</sub> levels, 79.1 ± 4.7 mmHg.

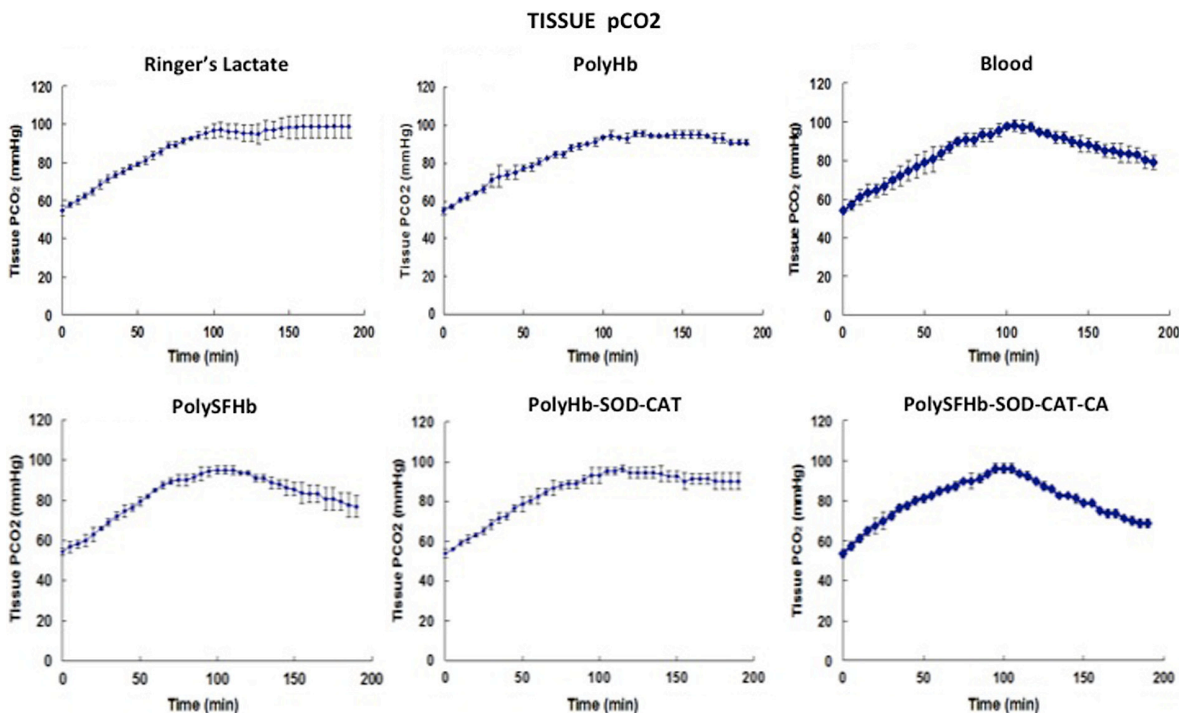


Figure 3. Tissue pCO<sub>2</sub> and plasma lactate. Following the 2/3 volume of blood loss, the tissue pCO<sub>2</sub> level increased steadily from the baseline tissue pCO<sub>2</sub> of 54.7 ± 2.5 mmHg to 98 ± 4.5 mmHg by 90 min after the hemorrhagic shock. Reinfusion with polyHb-SOD-CAT-CA reduced this to 68.6 ± 3 mmHg in 1 h. The effect that the other fluids had on tissue pCO<sub>2</sub> were lactated Ringer’s solution, 98 ± 4.5 mmHg; polyHb, 90.1 ± 4.0 mmHg; polyHb-SOD-CAT, 90.9 ± 1.4 mmHg; blood, 79.1 ± 4.7 mmHg; and polySFHb, 77 ± 5 mmHg.

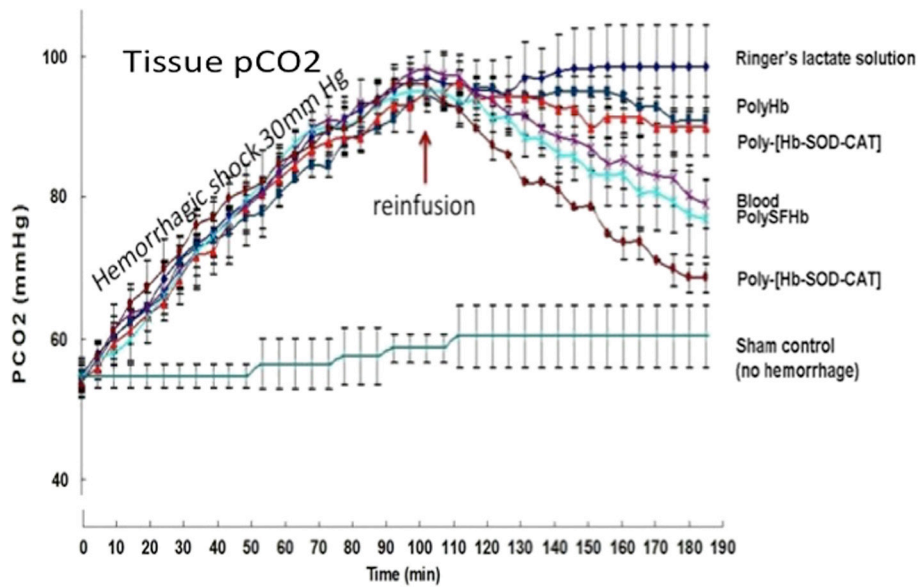


Figure 4. Comparison of  $p\text{CO}_2$ . The above measurements of each group were pooled together for comparison. Sham control measurement, where no blood loss occurred, was added as control. The results showed that reinfusion with polyHb-SOD-CAT-CA reduced this to  $68.6 \pm 3$  mmHg in 1 h. This was significantly ( $p < 0.05$ ) more effective than lactated Ringer's solution ( $98 \pm 4.5$  mmHg), polyHb ( $90.1 \pm 4.0$  mmHg), polyHb-SOD-CAT ( $90.9 \pm 1.4$  mmHg), blood ( $79.1 \pm 4.7$  mmHg), and polySFHb ( $77 \pm 5$  mmHg).

- (3) Reinfusion of polyHb (crosslinked ultrapure Hb with no RBC enzymes) did not significantly ( $P > 0.05$ ) reduce the elevated tissue  $p\text{CO}_2$  levels that remained at  $90.1 \pm 4.0$  mmHg.
- (4) Reinfusion of polyHb-SOD-CAT (crosslinked ultrapure Hb plus the addition of antioxidant enzymes—SOD and CAT but not CA) did not significantly ( $P > 0.05$ ) reduce the elevated tissue  $p\text{CO}_2$  levels that remain at  $90.9 \pm 1.4$  mmHg.
- (5) Reinfusion of polySFHb (crosslinking Hb and the content of RBC with the enzymes SOD, CAT, and CA at the concentration normally present in the RBC) was as efficient as blood in reducing the elevated tissue  $p\text{CO}_2$  levels to  $77 \pm 5$  mmHg.
- (6) PolyHb-SOD-CAT-CA was formed as in (5) above, but more SOD, CAT, and CA enzymes were added to enhance the normal RBC enzyme activities. Reinfusion reduced the elevated tissue  $p\text{CO}_2$  to  $68.6 \pm 3$  mmHg toward the normal levels after 60 min. This was significantly more effective than blood ( $p < 0.05$ ) that only reduced the tissue  $p\text{CO}_2$  to  $79.1 \pm 4.7$  mmHg.

### Effects on ST elevation

ST elevation in the ECG could be caused by ischemia, ischemia-reperfusion injury, or abnormal tissue metabolism. The results of reinfusion of different fluids are shown in Figure 5. Infusion of lactated Ringer's solution did not reduce the ST as it remained at  $106 \pm 3.1\%$  of the 90-min hemorrhagic shock ST level (Figure 5). For polyHb, an oxygen carrier without any enzyme activities, the ST only recovered to  $57.7 \pm 8.7\%$  of the elevated level. Blood reduced the ST to  $39.1 \pm 1.5\%$ , while polySFHb to  $38.3\% \pm 2.1\%$ . PolyHb-SOD-CAT with its enhanced antioxidant enzymes reduced the ST to  $27.8 \pm 5.6\%$ . PolyHb-SOD-CAT-CA reinfusion reduced the ST to  $21.7 \pm 6.7\%$  of the hemorrhagic

shock ST level, which is significantly better than the others ( $P < 0.05$ ). This is likely because polyHb-SOD-CAT-CA has all three RBC functions at an enhanced level, that is, oxygen and  $\text{CO}_2$  transport plus antioxidant functions.

### Effects on plasma lactate levels

In the case of plasma lactate level, after 90 min of sustained severe hemorrhagic shock it increased to  $18 \pm 2.3$  mM/L. PolyHb-SOD-CAT-CA reduced this to  $6.9 \pm 0.3$  mM/L in 60 min (Figure 6). It is significantly more effective ( $P < 0.05$ ) than lactated Ringer's solution ( $12.4 \pm 0.6$  mM/L), polyHb ( $9.6 \pm 0.7$  mM/L), blood ( $8.1 \pm 0.2$  mM/L), polySFHb ( $8.4 \pm 0.1$  mM/L), and polyHb-SOD-CAT ( $7.6 \pm 0.3$  mM/L).

### Histology

Detailed results of the histological studies will be finalized and reported elsewhere. The most striking changes were in the intestine. Lactated Ringer's solution group showed

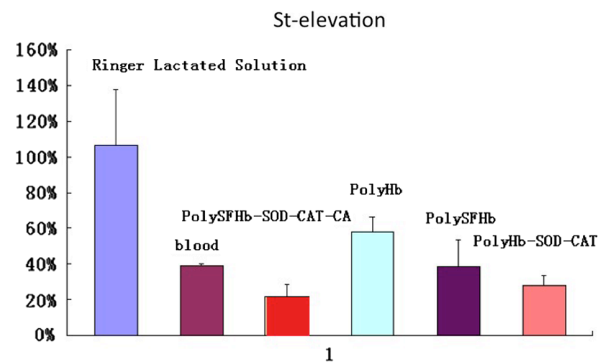


Figure 5. ST elevation. PolyHb-SOD-CAT-CA reduced the ST to  $21.7 \pm 6.7\%$ . This was significantly ( $p < 0.05$ ) better than polyHb ( $57.7 \pm 8.7\%$ ), blood ( $39.1 \pm 1.5\%$ ), polySFHb ( $38.3\% \pm 2.1\%$ ), and polyHb-SOD-CAT ( $27.8 \pm 5.6\%$ ). Lactated Ringer's solution exhibited no significant ( $p > 0.05$ ) effects ( $106 \pm 3.1\%$ ).

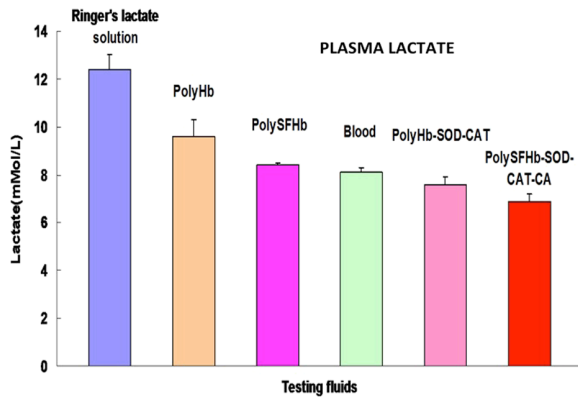


Figure 6. Plasma lactate. PolyHb-SOD-CAT-CA reduced the plasma lactate level from  $18 \pm 2.3$  mM/L to  $6.9 \pm 0.3$  mM/L. It was significantly ( $p < 0.05$ ) more effective than lactated Ringer's solution ( $12.4 \pm 0.6$  mM/L), polyHb ( $9.6 \pm 0.7$  mM/L), blood ( $8.1 \pm 0.2$  mM/L), polySFHb ( $8.4 \pm 0.1$  mM/L), and polyHb-SOD-CAT ( $7.6 \pm 0.3$  mM/L).

damaged villi and glands. The polyHb group showed damaged villi and mild damage of glands. The blood reinfusion group showed damaged villi but intact glands. There were no signs of injury in the polySFHb, polyHb-SOD-CAT, and polyHb-SOD-CAT-CA groups. The histological results suggest that the presence of antioxidant enzymes, SOD and CAT, in the polySFHb, polyHb-SOD-CAT, and polyHb-SOD-CAT-CA could protect the intestine from ischemia-reperfusion injuries in severe 90-min hemorrhagic shock beyond the 60-min safety window. A summary of other findings can be found in Table II.

**Stability at different temperatures including pasteurization**

PolyHb can be heat sterilized and stored for 1 year even at room temperature (Kim and Greenburg 2014, Gould et al. 1995). However, polyHb-SOD-CAT-CA contains both Hb and enzymes, and enzymes are particularly sensitive to storage and heat. We thus carried out studies to analyze its heat sterilization stability and storage stability. The detailed methods and results will be finalized and reported elsewhere. A summary of the result is as follows (Table II):

RBCs cannot be heat sterilized. On the other hand, after heat sterilization at 68°C for 2 h, lyophilized polyHb-SOD-CAT-CA retained good enzyme activities of SOD,  $100 \pm 2.5\%$ ; CAT,  $63.8 \pm 4\%$ ; and CA,  $97 \pm 4\%$ . More CAT can be added during the crosslinking process to maintain the same enzyme ratio after heat sterilization. It is important to note that heat sterilization at 68°C for 2 h is possible only for the lyophilized

form of polyHb-SOD-CAT-CA and not for the solution. After sterilization, the lyophilized form can be stored as such and can be easily reconstituted by dissolving in suitable solutions. Allowable storage time for RBC is about 1 day at room temperature and 42 days at 4C. The lyophilized form of polyHb-SOD-CAT-CA can retain sufficient enzyme activities when stored at room temperature for 320 days (Table III).

**Discussions**

In ischemia-reperfusion study of hemorrhagic shock, stroke or myocardial infarction in animals, researchers use an ischemic period ranging from 30 min to 1 h with variable results. Our earlier studies on ischemia-reperfusion injuries in a rat model of combined hemorrhagic shock-brain ischemia show that a minimum of 60 min is needed (Powanda and Chang 2002). However, our more recent results show that 60 min is the safety window for the treatment of hemorrhagic shock in rats (Bian et al. 2013). Thus, in order to avoid the pitfall of a very short ischemic period, and also to carry out a more stringent test, we are using 90 min for the present study.

We show in the present study that in severe hemorrhagic shock lasting for 90 min beyond the 60-min safety window, the novel nanobiotherapeutic polyHb-SOD-CAT-CA is superior to blood and other resuscitation fluids. It is more effective than RBC in reducing tissue pCO<sub>2</sub>. This is important since increased tissue pCO<sub>2</sub> has been related to increased mortality in hemorrhagic shock (Sims et al. 2001) and severity of cardiac ischemia (Tronstad et al. 2010). It is also more effective than blood and the other resuscitation fluids for the recovery of ST and reducing the elevated plasma lactate level. It is also more effective in protecting the intestine in the reperfusion and resuscitation process. Being a solution, it can also better perfuse into the constricted microcirculation than RBC in hemorrhagic shock or partially obstructed vessels in heart attack and stroke. The enhanced antioxidant enzymes, CAT and SOD, can remove damaging oxygen radicals. SOD also removes superoxide and prevents it from reacting and removing nitric oxide to cause vasoconstriction. The lyophilized form can be heat sterilized to remove infectious agents like HIV or future infectious agents like ebola to avoid contaminated donor blood crisis in the future. It is not specific to blood groups and the lyophilized powder can be stored at room temperature for 320 days (Table III). The lyophilized powder can be quickly reconstituted and dissolved for use. Since it does not need refrigeration or blood group

Table II. Histology of organs 60 min after reinfusion.

	Heart	Intestine	Kidney	Liver
Ringers lactated solution	Sarcoplasmic condensation; Contraction bands	Damaged villi as well as gland	Cytoplasmic vacuoles; tubular dilatation; necrosis	Sinusoidal dilatation; hepatocyte necrosis and congestion
PolyHb	Band contraction	Injured villi and mild injured gland	Disorder in the structure of tubular and cell necrosis	Eosinophilic change and nuclear loss
Blood	Sarcoplasmic condensation and eosinophilic changes	Injured villi but intact gland	Congestion and necrotic cells	Mild hepatocyte necrosis and congestion
PolySFHb	Eosinophilic changes	No damage	Tubular epithelial cell swelling	Mild congestion
PolyHb-SOD-CAT	No injury signals	No damage	Tubular epithelial cell swelling	Mild congestion
PolyHb-SOD-CAT-CA	No injury signals	No damage	Tubular epithelial cell swelling	Mild congestion

Table III. Effects of storage time and temperature on stability.

Stability at different temperatures		
Donor red blood cells		
Maximal time of storage allowed	20–25C 1 day?	4C 42 days
Poly-[Hb-SOD-CAT-CA] solution		
T1/2 of enzyme activities:	20–25C	4C
CAT	172 days	380 days
SOD	92 days	198 days
CA	51 days	231 days
Poly-[Hb-SOD-CAT-CA] lyophilized:		
Day 320 % enzyme activity	20–25C	4C
CAT	73% (day 320)	85% (day 320)
SOD	67% (day 320)	76% (day 320)
CA	73% (day 320)	85% (day 320)

testing, medics or ambulance can easily carry it and use it on the spot. This would avoid the life-threatening delays in emergency situations like mass disasters, war, and remote regions. However, polyHb-CAT-SOD-CA is just a potential nanobiotherapeutic with enhanced functions for specific applications. It is not meant to replace donor RBC with its much longer circulation time, which is essential for other long-term elective nonemergency uses.

PolyHb-SOD-CAT-CA requires the use of enzymes that can be expensive if purchased in purified form from commercial sources. Thus, we have successfully extracted enzymes from RBC and used them in the preparation of polyHb-SOD-CAT (Gu and Chang 2009). We have more recently successfully extracted all three enzymes, SOD, CAT, and CA, for the preparation of polyHb-SOD-CAT-CA (Chen, Glynn and Chang, personal communication). This way, Hb and RBC enzymes can be extracted from outdated donor blood, placental cord blood (Li et al. 2006), and from other sources, thus further utilizing RBC that would otherwise be discarded.

Our earlier study shows that polySFHb (contains enzymes at the same concentration as those in the RBCs) does not involve in immunological reactions and is not activated by any complement (Zhu et al. 2010). If the higher amounts of enzymes in the new polyHb-CAT-SOD-CA resulted in immunological reaction, then we can use human enzymes extracted from outdated human donor RBC or human placenta cord RBC (Li et al. 2006), or recombinant human enzymes.

Massive fluid and RBC replacement in severe injuries can lead to dilution of platelets and clotting factors resulting in bleeding (Hess et al. 2011, Kim and Greenburg 2014, Moore et al. 2009). Our earlier study in rats also shows that the clotting time is markedly increased when more than 80% of blood is exchanged with polyHb (Wong and Chang 2009). We have designed a soluble polyHb–fibrinogen complex that does not cause any increase in clotting time even when 98% of the blood has been exchanged with polyHb–fibrinogen (Wong and Chang 2007). If needed, we can apply this to the polyHb-SOD-CAT-CA to obtain a nanobiotherapeutic that has not only enhanced RBC functions, but also a platelet-like function.

One theory regarding vasoconstriction in endothelial dysfunction is the removal of nitrate oxide by modified Hb (Yu et al. 2010). Superoxide may contribute to this problem since it removes nitric oxide to form peroxyxynitrate. The SOD

Table IV. Comparison of polyHb-SOD-CAT-CA with RBC.

Properties	RBCs	PolyHb-SOD-CAT-CA
Oxygen transport	7 microns	Solution: better perfusion
Carbon dioxide transport	7 microns, enzymes in RBC	Solution + Enhanced enzymes
Oxygen radicals removal	7 microns, enzymes in RBC	Solution + Enhanced enzymes
Storage	1 day, unless refrigerated: 42 days	> 320 days room temperature
Blood groups	Yes, unless O	None
Heat sterilization	Cannot	Yes for lyophilized form
Platelet-like function	None	Possible
Availability	Limited	No limit
Circulation time	Donor RBC, 30 days	1–3 days
Potential applications	Better for elective long-term RBC replacement	As nanobiotherapeutic for emergency uses

component in polyHb-CAT-SOD-CA is effective in removing superoxide. This would prevent the removal of nitrate oxide to avoid vasoconstriction. In the unlikely event that polyHb-CAT-SOD-CA still causes vasoconstriction in endothelial dysfunction, we can use one of the available Hbs that has been modified to prevent vasoconstriction (Lui and Kluger 2011). Other ways to prevent adverse vasoconstriction of modified Hb are also available (Yu et al. 2010, Jia and Alayash 2013).

Stem cell culture for producing blood cells (Mazurier et al. 2011) has much promise for platelets and leucocytes that require only small volume. However, scaling up the liters of RBC needed for clinical use is still a major problem. When scale-up becomes practical, universal donor—group O—RBC can be produced. This will be very important for many important nonurgent uses of RBC. However, we shall still need polyHb-CAT-SOD-CA for emergency uses like sustained severe hemorrhagic shock for a number of reasons (Table IV).

## Declaration of interest

The authors have an agreement with McGill University for a patent application on this invention to allow for future development. They do not have conflicts of interest related to any connections with commercial companies in this.

The support for this research was provided by operating grants from the Canadian Institutes of Health Research (CIHR) and the Canadian Blood Service/CIHR joint award to Professor TMS Chang.

## References

- Alayash AI. 2004. Oxygen therapeutics: can we tame Hb. *Nat Rev Drug Discov.* 3:152–159.
- Bian Y, Rong Z, Chang TMS. 2011. PolyHb-superoxide dismutase-catalase-carbonic anhydrase: a novel biotechnology-based blood substitute that transports both oxygen and carbon dioxide and also acts as an antioxidant. *Artif Cells Blood Substit Biotechnol.* 39:127–136.
- Bian YZ, Wei G, Chang TMS. 2013. Lowering of elevated tissue PCO<sub>2</sub> in a hemorrhagic shock rat model after reinfusion of a novel nanobiotechnological PolyHb-superoxide dismutase-catalase-carbonic anhydrase that is an oxygen and carbon dioxide carrier with enhanced antioxidant properties. *Artif Cells Nanomed Biotechnol.* 41:60–68.



- Biro GP, Ou C, Ryan-MacFarlane C, Anderson PJ. 1995. Oxyradical generation after resuscitation of hemorrhagic shock with blood or stroma-free Hb. *Artif Cells Blood Substit Immobiliz Biotechnol.* 23:631-645.
- Burhop KE, Estep TE. 2001. Hb induced myocardial lesions. *Artif Cells Blood Substit Immobil Biotechnol.* 29:101-106.
- Chang EJ, Lee TH, Mun KC, Kim HC, Suh SI, Bae JH, et al. 2004. Effects of polyHb-antioxidant enzyme complex on ischemia-reperfusion in kidney. *Transplant Proc.* 36:1952-1954.
- Chang TMS (Ed.). 2013. *Selected Topics in Nanomedicine.* Singapore, London: World Science Publisher/Imperial College Press, 590 pages.
- Chang TMS. 1964. Semipermeable microcapsules. *Science.* 146:524-525.
- Chang TMS. 1971. Stabilisation of enzymes by microencapsulation with a concentrated protein solution or by microencapsulation followed by cross-linking with glutaraldehyde. *Biochem Biophys Res Commun.* 44:1531-1536.
- Chang TMS. 1997. *Red Blood Cell Substitutes: Principles, Methods, Products and Clinical Trials, Vol. I (Monograph).* Karger/Landes Systems, Basel, Switzerland (available for free online viewing at [www.medicine.mcgill.ca/artcell](http://www.medicine.mcgill.ca/artcell)).
- Chang TMS. 2005. Therapeutic applications of polymeric artificial cells. *Nat Rev Drug Discov.* 4:221-235.
- Chang TMS. 2007. Monograph on "ARTIFICIAL CELLS: Biotechnology, Nanotechnology, Blood Substitutes, Regenerative Medicine, bioencapsulation, cell/stem cell therapy". Singapore, London: World Scientific Publisher/Imperial College Press, 454 pages. (available for free online viewing/download at [www.medicine.mcgill.ca/artcell](http://www.medicine.mcgill.ca/artcell)).
- D'Agnillo F, Chang TMS. 1998. PolyHb-superoxide dismutase-catalase as a blood substitute with antioxidant properties. *Nat Biotechnol.* 16:667-671.
- Geers C, Gros G. 2000. Carbon dioxide transport and carbonic anhydrase in blood and muscle. *Physiol Rev.* 80:681-715.
- Gould, SA, Sehgal LR, Sehgal HL, GS Moss. 1995. The development of Hb solutions as red cell substitutes:Hb solutions. *Transfu Sci.* 16:5-17.
- Gu G, Chang TMS. 2009. Extraction of erythrocyte enzymes for the preparation of PolyHb-catalase-superoxide dismutase. *Artif Cells Blood Substit Biotechnol.* 37:69-77.
- Hess JR, Johansson PI, Holcomb JB, et al. 2011. Massive transfusion and transfusion therapy in trauma In: Mintz PD, Ed. *Transfusion Therapy: Clinical Principles and Practice.* 3rd ed. Bethesda, Maryland: AABB Press, pp. 305-322.
- Jahr JS, Mackenzie C, Pearce LB, Pitman A, Greenburg AG. 2008. HBOC-201 as an alternative to blood transfusion: efficacy and safety evaluation in a multicenter phase III trial in elective orthopaedic surgery. *J Trauma.* 64:1484-1497.
- Jia YP, Alayash AI. 2013. *Molecular Basis of Haptoglobin and Hb Complex Formation and Protection against Oxidative Stress and Damage in "Selected Topics in Nanomedicine"* Editor TMS Chang, World Science Publisher/Imperial College Press, Singapore London 149-168.
- Kim HW, Greenburg AG (Eds.). 2014. *Hemoglobin-Based Oxygen Carriers as Red Cell Substitutes and Oxygen Therapeutics.* New York, London: Springer, 738 pages
- Klein HG. 2000. The prospects for red-cell substitutes. *N Engl J Med.* 342:1666-1668.
- Li T, Yu R, Zhang HH, Liang WG, Yang XM, Yang CM. 2006. A method for purification and viral inactivation of human placenta Hb. *Artif Cells Blood Substit Biotechnol.* 34:175-188.
- Lui FE, Kluger R. 2011. Reviving artificial blood: meeting the challenge of NO scavenging by hemoglobin. *Chem Biochem.* 11:1816-1824.
- Ma L, Hsia CJC. 2013. Polynitroxylated Hb as a multifunctional therapeutic for critical care and transfusion medicine. In: Chang TMS (Ed.). *Selected Topics in Nanomedicine.* Singapore, London: World Science Publisher/Imperial College Press, pp. 169-194.
- Mazurier C, Douay L, Lapillonne H. 2011. RBC from induced pluripotent stem cells: hurdles and developments. *Curr Opin Hematol.* 18:249-253.
- Moore E, Moore FA, Fabian TC, Bernard AC, Fulda GJ, Hoyt DB, et al. 2009. Human polymerized hemoglobin for the treatment of hemorrhagic shock when blood is unavailable: the USA multicenter trial. *J Am Coll Surg.* 208:1-13.
- Nadithe V, Bae YH. 2011. Hb conjugates with antioxidant enzymes(Hb-superoxide dismutase-catalase) via poly(ethylene glycol) cross-linker for protection of pancreatic beta RINm5F cells in hypoxia. *Tissue Eng.* 17:2453-2462.
- Natanson C, Kern SJ, Lurie P, Banks SM, Wolfe SM. 2008. Cell-free hemoglobin-based blood substitutes and risk of myocardial infarction and death: a meta-analysis. *JAMA.* 299:2304-2312.
- Powanda D, Chang TMS. 2002. Cross-linked polyHb-superoxide dismutase-catalase supplies oxygen without causing blood brain barrier disruption or brain edema in a rat model of transient global brain ischemia-reperfusion. *Artif Cells Blood Substit Immobiliz Biotechnol.* 30:25-42.
- Sims C, Seigne P, Menconi M, Monarca J, Barlow C, Pettit J, Puyana JC. 2001. Skeletal muscle acidosis correlates with the severity of blood volume loss during shock and resuscitation. *J Trauma.* 51:1137-1146.
- Tronstad C, Pischke SE, Holhjem L, Tonnessen TI, Martinsen OG, Grimnes S. 2010. Early detection of cardiac ischemia using a conductometric PCO<sub>2</sub> sensor: real-time drift correction and parameterization. *Physiol Meas.* 31:1241-1255.
- Winslow RM (Ed.). 2006. *Blood Substitutes.* Amsterdam: Academic Press.
- Wong N, Chang TMS. 2007. PolyHb-fibrinogen: a novel blood substitutes with platelet-like activity for extreme hemodilution. *Artif Cells Blood Substit Biotechnol.* 35:481-489.
- Yu B, Shahid M, Egorina EM, Sovershaev MA, Raher MJ, Lei C, et al. 2010. Endothelial dysfunction enhances vasoconstriction due to scavenging of nitric oxide by a hemoglobin-based oxygen carrier. *Anesthesiology.* 112:586-594.
- Zhu HL, Du QQ, Chen C, Chang TMS. 2010. The immunological properties of stroma-free polyhemolysate containing catalase and superoxide dismutase activities prepared by polymerized bovine stroma-free hemolysate. *Artif Cells Blood Substit Biotechnol.* 38:57-63.