

Evaluation of Antibacterial Activity, Phytochemical Constituents, and Cytotoxicity Effects of Thai Household Ancient Remedies

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Abstract

Aim: Household ancient remedies reported here are described in the National List of Essential Medicines and have traditionally been used in Thailand to treat infection-related ailments. However, the safety and effectiveness of these remedies have been poorly evaluated. The aim of this study was to evaluate the antibacterial properties of these remedies against seven gram-positive and gram-negative multidrug-resistant bacteria species. Phytochemical constituents and cytotoxicity of these remedies were also determined.

Methods: Seven remedies, consisting of Um-Ma-Luk-Ka-Wa-Tee, Chan-Ta-Lee-La, Kheaw-Hom, Learng-Pid-Sa-Mud, Pra-Sa-Chan-Dang, Dhart-Ban-Chob, and Tree-Hom, were prepared by a licensed traditional medical doctor using a mixture of medicinal plants. Antibacterial activity of ethanol extracts of the remedies was determined by using a broth microdilution method. Qualitative phytochemical screening analysis was carried out to identify the presence of major components. Cytotoxicity activities of the extracts against Vero cells were assessed by green fluorescent protein-based assay.

Results: With the exception of Dhart-Ban-Chob extract, significant minimum inhibitory concentrations (MICs) of < 16 to 32 $\mu\text{g}/\text{mL}$ were observed for the remedy extracts depending on the bacterial strains. The Um-Ma-Luk-Ka-Wa-Tee extract was noncytotoxic against Vero cells and possessed the highest activity, with MICs of < 16 to 31 $\mu\text{g}/\text{mL}$ against all methicillin-resistant *Staphylococcus aureus* isolates.

Conclusions: Remarkable antibacterial activities against multidrug-resistant pathogens, as well as low toxicity on Vero cells, of Um-Ma-Luk-Ka-Wa-Tee support the use of this remedy in traditional medicine. Further investigation on other biological activities related to traditional applications, appropriate biomarkers, and treatment mechanisms of the household remedy are required.

Introduction

ANTIMICROBIAL DRUG RESISTANCE IS RECOGNIZED as one of the greatest problems in both developing and developed countries, in hospitals as well as in the community. Many studies have estimated the excess mortality and morbidity, and economic impact of infections caused by antibiotic-resistant pathogens.^{1,2} Even though enormous efforts have been made to discover new drugs, to implement a more appropriate prescribing antibiotic strategy, and to seek new ways of using older antibiotics, the prevalence of bacterial resistance has been increasing worldwide. In Europe and the United States, the excess hospital costs associated with the

infections were as high as \$30 billion.^{1–3} Infections with the multidrug-resistant bacteria are estimated to cause around 25,000–99,000 deaths per year.^{2,4}

Gram-positive cocci have emerged as significant pathogens associated with nosocomial infections in the past two decades. On the basis of results from the SENTRY Antimicrobial Surveillance Program, *Staphylococcus aureus* and coagulase-negative staphylococci are the microorganisms most frequently isolated from bloodstream infections, and about one third of the infections are caused by these pathogens.⁵ Methicillin-resistant *S. aureus* (MRSA) accounts for approximately 30% of *S. aureus* isolates and is also resistant to clindamycin, ciprofloxacin, and levofloxacin.⁵

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TABLE 1. MEDICAL APPLICATIONS AND INGREDIENTS OF SELECTED THAI HOUSEHOLD ANCIENT REMEDIES

Household remedies (net weight of remedy powder)	Herbal components (parts used)	Weight (g per the net weight)	Ailments/uses	Dosage ^a
Chan-Ta-Lee-La (33 g)	<i>Angelica dahurica</i> Benth. (root)	4	For relief of fever	Adults: 3–5 T×3 Children: 1–2 T×3
	<i>Atractylodes lancea</i> (Thunb.) DC. (rhizome)	4		
	<i>Artemisia annua</i> L. (whole plant)	4		
	<i>Myristica fragrans</i> Houtt. (mace)	4		
	<i>Dracaena loureiri</i> Gagnep. (heartwood)	4		
	<i>Gymnopetalum chinensis</i> (Lour.) Merr. (fruit)	4		
	<i>Tinospora crispa</i> (L.) Miers ex Hook.f. & Thomson (climber)	4		
	<i>Eurycoma longifolia</i> Jack. (root)	4		
	<i>Pogostemon cablin</i> (Blanco) Benth. (leaf)	1		
	Pra-Sa-Chan-Dang (64 g)	<i>Symplocos racemosa</i> Roxb. (root)		
<i>Bouea macrophylla</i> Griff (root)		4		
<i>Citrus aurantifolia</i> Swingle (root)		4		
<i>Kaempferia galanga</i> L. (rhizome)		4		
<i>Ligusticum sinense</i> Oliv. (rhizome)		4		
<i>Myristica fragrans</i> Houtt. (mace)		4		
<i>Caesalpinia sappan</i> Linn. (heartwood)		4		
<i>Nelumbo nucifera</i> Gaertn. (stamen)		1		
<i>Mesua ferrea</i> L. (pollen)		1		
<i>Mammea siamensis</i> (flower)		1		
<i>Jasminum sambac</i> (flower)		1		
<i>Dracaena loureiri</i> Gagnep. (heartwood)		32		
Kheaw-Hom (18 g)		<i>Pogostemon cablin</i> (Blanco) Benth. (leaf)	1	For relief of fever from chickenpox and measles and for thirst relief
	<i>Linnophila rugosa</i> (Roth.) Merr (leaf)	1		
	<i>Cordyline fruticosa</i> (L.) Goeppert (leaf)	1		
	<i>Eupatorium stoechadosmum</i> Hance (leaf)	1		
	<i>Vetiveria zizanioides</i> (L.) Nash ex Small. (root)	1		
	<i>Kaempferia galanga</i> L. (rhizome)	1		
	<i>Myristica fragrans</i> Houtt. (heartwood)	1		
	<i>Dracaena loureiri</i> Gagnep. (heartwood)	1		
	<i>Angiopteris evecta</i> Hoffm. (ND)	1		
	<i>Globba malaccensis</i> Ridl. (ND)	1		
	<i>Tacca chantrieri</i> Andre. (ND)	1		
	<i>Sophora exigua</i> Craib. (ND)	1		
	<i>Cyathea borneensis</i> Copel.(ND)	1		
	<i>Aristolochia</i> spp. (root) ^b	1		
	<i>Mimusops elengi</i> L. (flower)	1		
	<i>Mesua ferrea</i> L. (pollen)	1		
	<i>Mammea siamensis</i> Kosterm. (pollen)	1		
	<i>Nelumbo nucifera</i> Gaertn. (stamen)	1		
Tree-Hom (64 g)	<i>Terminalia</i> sp. (fruit)	4	For relief of fever Laxatives for children	Children: at 1–2 mo: 1 T×1 at 3–5 mo: 2–3 T×1 at 6–12 mo: 5–7 T×1
	<i>Terminalia bellirica</i> (Gaertn.) Roxb (fruit)	4		
	<i>Phyllanthus emblica</i> (fruit)	4		
	<i>Coriandrum sativum</i> L. (seed)	4		
	<i>Aristolochia</i> spp. (root)	1		
	<i>Angelica dahurica</i> Benth. (root)	1		
	<i>Glycyrrhiza glabra</i> (root)	1		
	<i>Trigonella foenum-graecum</i> L. (seed)	1		
	<i>Terminalia chebula</i> Retz. (fruitfruit)	22		
	<i>Rheum palmatum</i> L. (steamed root)	22		
Learng-Pid-Sa-Mud (18 g)	<i>Cyperus rotundus</i> L. (rhizome)	1	For treatment of diarrhea and dysentery	Adults: 5–7 T×3 Children: at 1–2 mo: 1 T×3 at 3–5 mo: 2 T×3 at 6–12 mo: 3–5 T×3 > 1 y: 5–7 T×3
	<i>Curcuma zedoaria</i> Rose (rhizome)	1		
	<i>Oroxylum indicum</i> (L.) Kurz (bark)	1		
	<i>Musa paradisiaca</i> L.; ABB Group (root)	1		
	<i>Allium sativum</i> Linn. (bulk)	1		
	<i>Piper retrofractum</i> Vahl (fruit)	1		

(continued)

TABLE 1. (CONTINUED)

Household remedies (net weight of remedy powder)	Herbal components (parts used)	Weight (g per the net weight)	Ailments/uses	Dosage ^a
Learng-Pid-Sa-Mud (Continued)	<i>Dipterocarpus alatus</i> Roxb. Ex G.Don (gum)	1		
	<i>Tachardia lacca</i> Kerr. (lac)	1		
	<i>Uncaria gambir</i> (Hunter) Roxb. (leaf and limb)	1		
	<i>Acacia catechu</i> (L.f.) Willd. (heartwood)	1		
	<i>Impatiens balsamina</i> L. (leaf)	1		
	<i>Punica granatum</i> L. (leaf)	1		
	<i>Curcuma longa</i> L. (rhizome)	6		
Dhart-Ban-Chob (112 g)	<i>Zingiber officinale</i> Roscoe (rhizome)	4	For treatment of diarrhea and bloated stomach	Adults: 3–5 T×3 Children: 2–3 T×3
	<i>Atractylodes lancea</i> (Thunb.) DC (rhizome)	4		
	<i>Dischidia rafflesiana</i> Wall. (leaf)	4		
	<i>Angelica sinensis</i> (Oliv) Diels (root)	4		
	<i>Angelica dahurica</i> Benth. (root)	4		
	<i>Abroma augusta</i> (L.) L. f. (seed)	4		
	<i>Lawsonia inermis</i> L.(seed)	4		
	<i>Pimpinella anisum</i> L. (seed)	4		
	<i>Trachyspermum ammi</i> (L.) Sprague (seed)	4		
	<i>Lepidium sativum</i> Linn. (seed)	4		
	<i>Diospyros decandra</i> Lour. (fruit)	4		
	<i>Myristica fragrans</i> Houtt. (mace)	4		
	<i>Syzygium aromaticum</i> (L.) Merr. & L.M. Perry (flower)	4		
	<i>Cinnamomum camphora</i> Th Fries (leaf)	4		
	<i>Cinnamomum bejolghota</i> (Ham.) Sweet (bark)	4		
	<i>Amomum krervanh</i> Pierre (fruit)	4		
	<i>Coriandrum sativum</i> L.(seed)	4		
	<i>Coriandrum sativum</i> L. (seed)	4		
	<i>Pogostemon cablin</i> (Blanco) Benth. (leaf)	4		
	<i>Aristolochia</i> spp. (root)	4		
	<i>Piper retrofractum</i> Vahl (fruit)	4		
	<i>Kaempferia galanga</i> L. (rhizome)	4		
	<i>Picrorrhiza kurroa</i> Benth. (rhizome)	8		
<i>Terminalia chebula</i> Retz. var. <i>chebula</i> (fruit)	16			
Um-Ma-Luk Ka-Wa-Tee (85 g)	<i>Aristolochia</i> spp. (root)	7	Used to relieve cough and reduce phlegm	Adults: 1–2 T×3 Children: 0.5–1 T×3
	<i>Dischidia rafflesiana</i> Wall. (leaf)	7		
	<i>Lawsonia inermis</i> L. (root)	7		
	<i>Coriandrum sativum</i> L. (root)	7		
	<i>Phyllanthus emblica</i> (fruit)	7		
	<i>Terminalia bellirica</i> (Gaertn.) Roxb. (fruit)	7		
	<i>Glycyrrhiza glabra</i> (root)	43		

^aDosage: tsp, teaspoon; T, 0.5-g tablet; ×1, once a day; ×2, twice a day; ×3, three times a day; ×4, four times a day.

^b*Aristolochia* spp. (root) was removed from Thai traditional herbal recipes in 2011.

ND, no data available.

Multidrug-resistant (MDR) gram-negative pathogens are less prevalent than gram-positive bacteria. However, infections due to pan-drug-resistant strains are sometimes untreatable. *Escherichia coli* is one of the most common causes of bloodstream infections, and data from Europe revealed that 9.3% (13,950 cases) of the isolates were resistant to a third-generation cephalosporin, a regularly used antibiotic.² The 30-day mortality rate among patients with third-generation

cephalosporin-resistant *E. coli* bloodstream infections was 2.5 times higher than that of the susceptible group.

Acinetobacter baumannii and *Pseudomonas aeruginosa* can cause a variety of infections and quickly become resistant to commonly prescribed antimicrobials. In a study from Karachi, Pakistan, nearly 50% of babies born with the infections caused by *Acinetobacter* spp. died, and approximately 70% of isolates were resistant to all antibiotics except polymyxin.⁶ Despite the

introduction of antipseudomonal antibiotics, *P. aeruginosa* continues to be a serious cause of infection associated with high rates of morbidity and mortality. Most studies have found that mortality rates varied from 30% to 60% among all patients with *P. aeruginosa* bloodstream infections and have remained high during the past few decades.^{7,8} In view of all this, the need to discover alternative and effective antibacterial agents is urgent.

Traditional Thai medicine (TTM) is a system of traditional medicine of the Kingdom of Thailand originated during the Sukhothai period (1238–1377) and was formally accepted as a primary health care resource in the late 19th century.⁹ Thai household ancient remedies have been noted in the National List of Essential Medicines, List of Herbal Medicinal Products D, 2006. Fifty ancient remedies and 21 herbal products were approved by the Ministry of Public Health for their effectiveness in treating ailments and are described in the National List of Essential Medicines, List of Herbal Medicinal Products A.D. 2011.¹⁰ However, scientific evidence of the biological effects of Thai household ancient remedies is limited.

Therefore, this study evaluated the *in vitro* antibacterial potential of seven Thai household ancient remedies that are traditionally used to treat infection-related ailments against seven gram-positive and gram-negative multidrug-resistant bacteria species. Moreover, the phytochemical constituents and cytotoxicity of these remedies were additionally determined. Results obtained from this study provide scientific information to support the use of herbal-based remedies for treatments of infectious diseases, particularly those caused by multidrug-resistant pathogens.

Materials and Methods

Thai household ancient remedies

Information on household ancient remedy uses in this paper was based on the National List of Essential Medicines, List of Herbal Medicinal Products A.D. 2006. The selection of seven Thai household ancient remedies—Um-Ma-Luk-Ka-Wa-Tee, Chan-Ta-Lee-La, Kheaw-Hom, Learng-Pid-Sa-Mud, Pra-Sa-Chan-Dang, Dhart-Ban-Chob, and Tree-Hom—screened in this study was based on their traditional claims for treatment or relief of fever, diarrhea, or sore throat. The list of medicinal components, medical applications, and dosages is summarized in Table 1 from the National List of Essential Medicines, List of

Herbal Medicinal Products, 2nd edition, B.E. 2554 (2011).¹⁰ The powdered remedies, prepared by a licensed Thai traditional medical doctor, Mr. Somchai Ontong (T.M.P. no. 22907), were purchased from Traditional Thai Medicine Hospital, Prince of Songkla University, Hat Yai, Thailand.

Remedies extracts

Five hundreds grams of each remedy was mixed with 1,000 mL of 95% ethanol. The mixtures were left for 7 days at 30°C and then filtered through Whatman no. 1 filter paper. The collected filtrates were further concentrated to dryness using a rotary evaporator and kept at 55°C. Yields (%; w/w) of each extracts were calculated as the ratio of the weight of the extract to the weight of the recipe powder (Table 2). The sample were stored at –40°C and further used for antibacterial tests.

Stock solutions of the extracts were prepared in advance at a concentration of 80 mg/mL in dimethylsulfoxide (DMSO; Merck, Darmstadt, Germany). Before the bioassay, working solutions were prepared by diluting the stock solutions to two times the maximum desired final testing concentrations in Mueller-Hinton broth (MHB; Difco, Strasbourg, France). The final concentration of DMSO in all assays was 1.25% or less, which is nontoxic for the tested pathogens.

Phytochemical screening tests

Qualitative phytochemical screening analysis of Thai household ancient remedy extracts was carried out to identify the presence of secondary metabolites, including alkaloids, terpenoids, condensed tannins, and hydrolysable tannins using Dragendorff reagent, Liebermann-Burchard reagent, and ferric chloride reagent, respectively.¹¹

Antibacterial properties

Tested pathogens. Microorganisms used to determine antibacterial activities of different extracts were as follows: (1) gram-positive bacteria: *S. aureus* ATCC 25923, clinically isolated methicillin resistant *S. aureus* (MRSA NPRC R003-R005), *S. epidermidis* ATCC 35984, coagulase-negative staphylococci (NPRC 301 and 308), and coagulase-positive staphylococci (NPRC 506–507) isolated from acne lesions; (2) gram-negative bacteria: *P. aeruginosa* ATCC 10145, clinically isolated MDR *P. aeruginosa* (2097 and 5351), MDR *A. baumannii* (NPRC AB002, 004, 005, and 034), *E. coli* ATCC

TABLE 2. EXTRACTION YIELDS, CYTOTOXICITY EFFECTS, AND PHYTOCHEMICAL CONSTITUENTS OF SELECTED THAI HOUSEHOLD ANCIENT REMEDIES

Household remedies	Extraction yield (%; w/w)	Cytotoxicity ^a (IC ₅₀ ; µg/mL)	Phytochemical constituents ^b			
			1	2	3	4
Chan-Ta-Lee-La	2.5	19.5	+	–	+	+
Pra-Sa-Chan-Dang	4.2	> 50	+	+	+	+
Kheaw-Hom	3.7	> 50	+	–	+	+
Tree-Hom	8.7	> 50	–	–	+	–
Learng-Pid-Sa-Mud	7.3	19.7	+	+	+	+
Dhart-Ban-Chob	7.9	> 50	+	+	+	+
Um-Ma-Luk-Ka-Wa-Tee	3.9	> 50	+	+	+	–

^a50% inhibitory concentration of ellipticine (a positive control) was 0.8 µg/mL.

^bThe results showed the presence (+) or absence (–) of phytochemical compounds including alkaloid, triterpenoid, condensed tannin, and hydrolysable tannin.

25922, MDR *E. coli* (2746-08 and 2809-08), and *E. coli* O15:H7 RIMD 05091078. All bacterial strains were obtained from Natural Products Research Center and Department of Microbiology, Faculty of Science, Prince of Songkla University. The strains were maintained on a Trypticase soy agar (Difco) slant at 4°C and activated at 37°C for 24 hours with a Trypticase soy agar plate prior to any antimicrobial tests.

Bacterial cultures were prepared by transferring three to five well-isolated colonies into a tube containing 2 mL of MHB (Difco) and grown overnight at 37°C. The turbidity of the culture was adjusted with sterile MHB to match 0.5 McFarland standard.

Assessment of minimum inhibitory concentration. A modified broth microdilution method using 96-well microplate according to the Clinical and Laboratory Standard Institute¹² was used to obtain minimum inhibitory concentrations (MICs) of the crude extracts against the tested pathogens. Each plant extract (80 mg/mL) was serially diluted two-fold to obtain 1 mg/mL final testing concentration in the first well. The range of final testing concentration of each extract was 16–1000 µg/mL. An equal volume of 100 µL fresh bacterial culture concentration corresponding to 10⁶ colony-forming units/mL was added to the wells. The plate was covered with lids and incubated for 24 hours at 37°C. The bacterial growth was measured by recording the absorbance at 620 nm, using a microplate reader (Sunrise, Tecan, Switzerland). The MIC was defined as the lowest concentration of the test agent that had restricted growth to a level less than 0.05 at 620 nm after the incubation period.

A growth control and a blank control were taken using the inoculated broth added into 1.25% of DMSO and fresh MHB added into each concentration of the extract, respectively. The MICs of antibiotics, including vancomycin and rifampicin, for all the reference strains were simultaneously determined. Each experiment was performed at least twice.

As previously proposed by Rios and Recio¹³ and Kuet,¹⁴ extracts were classified as significantly active when MICs were less than 100 µg/mL, moderately active when the values were 100–625 µg/mL, and weakly active when the values were greater than 625 µg/mL.

Kinetic growth inhibition assay. Kinetic growth inhibition assay was used to observe the growth inhibitory effects of effective remedies (Learng-Pid-Sa-Mud, Kheaw-Hom, and Um-Ma-Luk-Ka-Wa-Tee) on representative gram-positive pathogen (MRSA NPRC R003, *S. epidermidis* ATCC 35984, coagulase-positive staphylococci NPRC 308) and gram-negative pathogens (MDR *A. baumannii* NPRC AB034, MDR *E. coli* 2746-08, and MDR *P. aeruginosa* 5351). Bacterial inoculum prepared as described above (1 mL) was mixed with 1 mL of MHB containing each remedy extract at final concentrations of 16, 32, 64, 125, 250, 500, and 1000 µg/mL. The tubes were incubated at 37°C, and samples (100 µL) were taken at 0, 6, 12, and 18 hours. Bacterial growth of each sample was measured by recording the absorbance at 620 nm.

Cytotoxicity assay

Cytotoxicity activities of the formula extracts against Vero cells were determined by green fluorescent protein–

based assay at the National Center for Genetic Engineering and Biotechnology, National Science and Technology Development Agency, Pathum Thani, Thailand. Ellipticine used as a positive control exhibited cytotoxicity against Vero cell line with 50% inhibitor concentration (IC₅₀) of 0.8 µg/mL.¹⁵

Results

Table 1 provides the ethno-botanical information of the ancient household remedies, including scientific name, plant part used, weight ratio of medicinal components, usages, and dosage. The remedies were prepared using a mixture of plants as follows: Dhart-Ban-Chob, Kheaw-Hom, Learng-Pid-Sa-Mud, Pra-Sa-Chan-Dang, Tree-Hom, Chan-Ta-Lee-La, and Um-Ma-Luk-Ka-Wa-Tee containing 24, 17, 13, 12, 10, 9, and 7 medicinal plants, respectively. In total, this study reports seven plant-based formulas containing 68 species that belonged to 48 families; most of these medicinal plants belong to the Zingiberaceae and Umbelliferae families. Different plant parts, such as root, rhizome, wood, fruit, mace, leaf, and flower, were used to prepare the remedies; of these, root, rhizome, and leaf were the most commonly used plant material. The most frequently mentioned medicinal plants used as household remedies were *Myristica fragrans* (found in four remedies: Chan-Ta-Lee-La, Pra-Sa-Chan-Dang, Kheaw-Hom, and Dhart-Ban-Chob), *Aristolochia pierrei* (found in four remedies: Tree-Hom, Um-Ma-Luk-Ka-Wa-Tee, Kheaw-Hom, and Dhart-Ban-Chob), *Dracaena loureiri* (found in three remedies: Chan-Ta-Lee-La, Pra-Sa-Chan-Dang, and Kheaw-Hom), and *Angelica dahurica* (found in three remedies: Chan-Ta-Lee-La, Tree-Hom, and Dhart-Ban-Chob). Our preliminary phytochemical test revealed that condensed tannins were common principals found in all tested remedies. Other compounds, including alkaloids, triterpenoids, and hydrolysable tannins, were present in most of the selected remedies (Table 2).

To assess antibacterial activity, the ethanol extracts were tested against five species (20 isolates) of gram-positive and gram-negative pathogens. The evaluated MICs are reported in Table 3. The rationale for using these 16 clinical isolates was that their antibiotic susceptibility profile is fairly representative of most antibiotic-susceptible and antibiotic-resistant isolates. The activity of these remedy extracts was interpreted as inhibition at any MICs of 1000 µg/mL or less. The extracts that did not exhibit inhibition up to the concentration limit of 1000 µg/mL were considered inactive. The antibacterial activity of the remedies was further classified as significant if the MIC was less than 100 µg/mL, moderate if the MIC was greater than 100 and less than or equal to 625 µg/mL, or weak if the MIC was greater than 625 µg/mL. The MICs obtained in this study for all the tested extracts ranged from less than 16 to 1000 µg/mL against gram-positive pathogens, while MICs varied from 125 to greater than 1000 µg/mL against gram-negative bacteria. With the exception of Dhart-Ban-Chob extract, the notable MICs of less than 16 to 32 µg/mL was observed for the remedy extracts depending on the bacterial strains. MRSA isolates used in this investigation were the most susceptible bacterial strain and were found to be susceptible to the extracts of Pra-Sa-Chan-Dang, Kheaw-Hom, Learng-Pid-Sa-Mud, and Um-Ma-Luk-Ka-Wa-Tee.

TABLE 3. MINIMUM INHIBITORY CONCENTRATION OF ETHANOL EXTRACTS OF THAI HOUSEHOLD ANCIENT REMEDIES AGAINST HUMAN PATHOGENS

Tested pathogens	MIC of the remedy extracts ^{a,b} ($\mu\text{g/mL}$)						
	1	2	3	4	5	6	7
<i>Staphylococcus aureus</i> ATCC 25923	250	62 ^c	31 ^c	250	250	500	125
Methicillin resistant <i>S. aureus</i> (MRSA) NPRC R003 125	125	31 ^c	125	31 ^c	250	31 ^c	
MRSA NPRC R004	125	31 ^c	31 ^c	250	1000	250	31 ^c
MRSA NPRC R005	250	62 ^c	1000	125	31 ^c	500	<16 ^c
<i>S. epidermidis</i> ATCC 35984	250	125	31 ^c	125	<16 ^c	250	<16 ^c
Coagulase-positive staphylococci NPRC 301	1000	1000	125	62 ^c	250	1000	31 ^c
Coagulase-positive staphylococci NPRC 308 125	125	125	125	1000	500	125	
Coagulase-negative staphylococci NPRC 506	31 ^c	250	250	250	125	125	125
Coagulase-negative staphylococci NPRC 507 62 ^c	125	125	125	1000	125	125	
<i>Pseudomonas aeruginosa</i> ATCC 10145	500	250	>1000	500	250	250	250
MDR <i>P. aeruginosa</i> 2097	500	250	125	500	125	500	250
MDR <i>P. aeruginosa</i> 5351	1000	1000	1000	250	500	1000	>1000
MDR <i>Acinetobacter baumannii</i> NPRC AB002	>1000	1000	>1000	1000	1000	>1000	>1000
MDR <i>A. baumannii</i> NPRC AB004	1000	1000	1000	500	500	500	500
MRD <i>A. baumannii</i> NPRC AB005	>1000	1000	>1000	>1000	1000	500	1000
MRD <i>A. baumannii</i> NPRC AB034	1000	>1000	>1000	500	1000	500	500
<i>Escherichia coli</i> ATCC 25922	>1000	>1000	>1000	>1000	1000	1000	>1000
MDR <i>E. coli</i> 2746-08	1000	1000	1000	1000	1000	1000	1000
MDR <i>E. coli</i> 2809-08	>1000	1000	>1000	>1000	1000	>1000	>1000
<i>E. coli</i> O15:H7 RIMD 05091078	250	>1000	>1000	>1000	1000	1000	>1000

^aThe tested household remedies were Chan-Ta-Lee-La, Pra-Sa-Chan-Dang, Kheaw-Hom, Tree-Hom, Learng-Pid-Sa-Mud, Dhart-Ban-Chob, and Um-Ma-Luk-Ka-Wa-Tee.

^bMICs of vancomycin on *S. aureus* ATCC 25923 and rifampicin on *P. aeruginosa* ATCC 10145/*E. coli* ATCC 25922 were 0.5 and 40/5 $\mu\text{g/mL}$, respectively.

^cMICs considered as representing noteworthy antimicrobial activity. MIC, minimum inhibitory concentration; MDR, multidrug-resistant.

The extract of Um-Ma-Luk-Ka-Wa-Tee possessed the highest and most significant antibacterial activity, with MICs less than 16–31 $\mu\text{g/mL}$ against all MRSA isolates. In addition, this extract exhibited remarkable activity, with MICs of <16 and 31 $\mu\text{g/mL}$, respectively, against a biofilm-producing isolate, *S. epidermidis* ATCC 35984 and acne lesion-isolated coagulase-positive staphylococci NPRC 301. The growth studies, as presented in Figure 1A–C, confirmed these results. This remedy showed moderate activity on some isolates of *P. aeruginosa* and *A. baumannii* but had weak activity or even total inactivity against *E. coli* isolates. Although the extract was generally considered inactive against gram-negative pathogens, subinhibitory concentrations of the extract partially inhibited the growth of these bacteria (Figure 1D–F). A certain activity against gram-positive pathogens was also shown by the extract of Kheaw-Hom and Learng-Pid-Sa-Mud, which have an MIC corresponding to 31 $\mu\text{g/mL}$. On the other hand, these extracts showed no activity with MIC values higher than 1000 $\mu\text{g/mL}$ on tested gram-negative bacteria. Similarly, with Um-Ma-Luk-Ka-Wa-Tee extract, subinhibitory concentration of Kheaw-Hom and Learng-Pid-Sa-Mud affected the growth of *E. coli* and *A. baumannii* and *A. baumannii* and *P. aeruginosa*, respectively (data not shown).

Cytotoxicity effects of the remedies were additionally investigated on Vero cells for their potentially safe uses (Table 2). The tested remedies had no cytotoxic effects except for Chan-Ta-Lee-La and Learng-Pid-Sa-Mud, which had IC₅₀ values of 19.5 and 19.7 $\mu\text{g/mL}$, respectively, on Vero cells. The IC₅₀ value of the positive control ellipticine used in this

study was 0.8 $\mu\text{g/mL}$, which is approximately 24 times higher than those of Chan-Ta-Lee-La and Learng-Pid-Sa-Mud and more than 60 times higher than those of other remedies.

Discussion

Traditional plant-based formulas have been commonly used to treat skin and soft tissue infections, diarrhea, gastritis, fever, and peptic ulcer diseases in Asian countries such as China,¹⁶ India,¹⁷ Japan,¹⁸ Korea,¹⁹ and Thailand.^{20–23} Although previous studies have revealed that folk healers believe that combining more than one plant increases effectiveness of medicines and many diseases were treated using a combination of more than one plant, few pharmacologic studies support this information.²⁴ In the present study, Thai household ancient remedies widely used to treat infection-related ailments were selected from the List of Herbal Medicinal Products A.D. 2006 approved by the Ministry of Public Health of Thailand. On the basis of criteria proposed by Rios and Recio¹³ and Kuete,¹⁴ Um-Ma-Luk-Ka-Wa-Tee (which consists of *Aristolochia* spp., *Dischidia rafflesiana*, *Lawsonia inermis*, *Coriandrum sativum*, *Phyllanthus emblica*, *Terminalia bellirica*, and *Glycyrrhiza glabra*) possessed significant anti-MRSA activity. The remedy is more effective than the previously reported Thai traditional remedies (Pikutbenjakul,²¹ Prasaproyai,²² and Benchalokawichian²³), as well as other traditional Thai medicinal plants.

Krai-Krue, a dried root of *Aristolochia pierrei* Lecomte or *Aristolochia tagala* Cham, is commonly used in Thai traditional herbal remedies.²⁵ Aristolochic acid and its

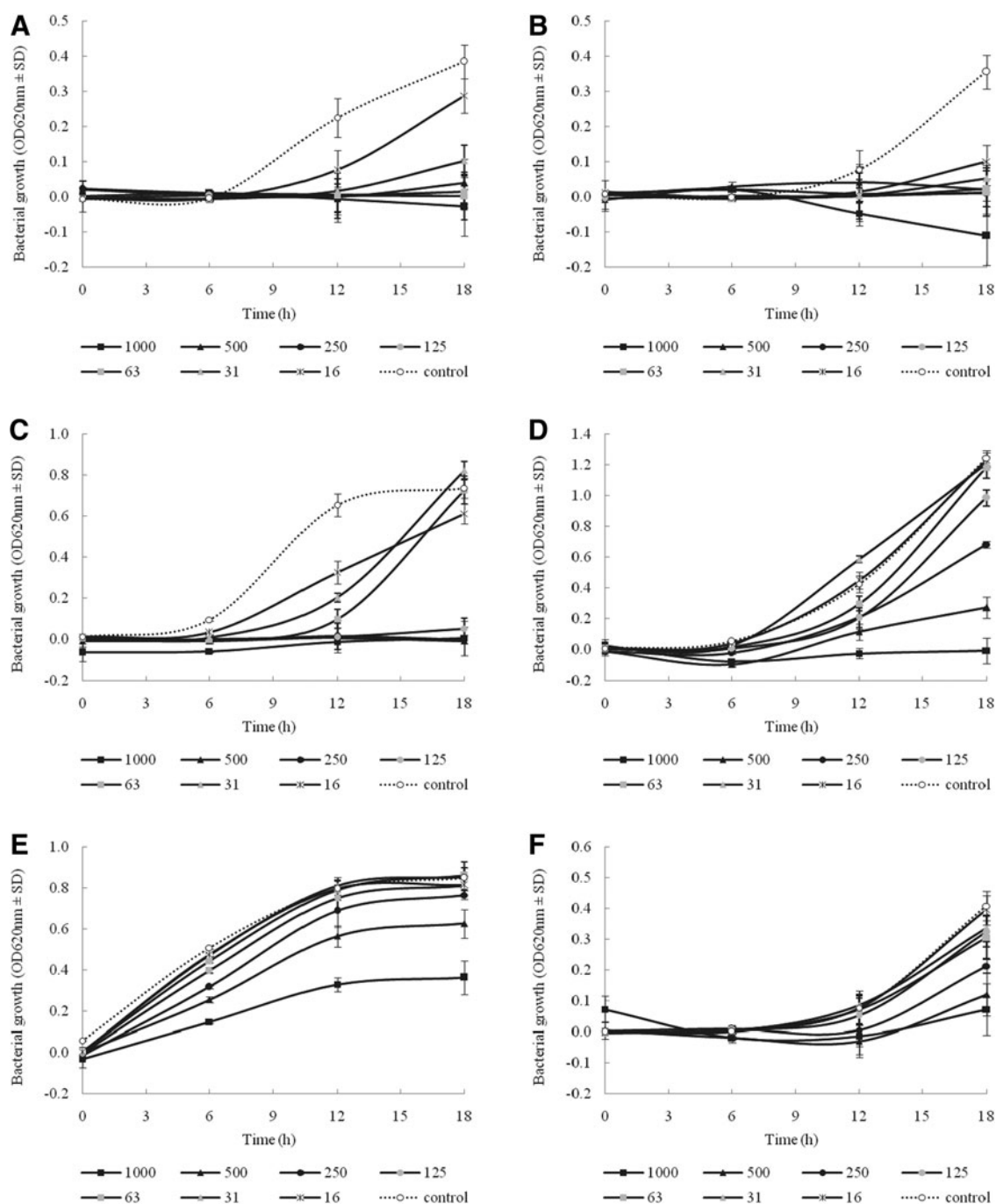


FIG. 1. Growth inhibition effects of Um-Ma-Luk-Ka-Wa-Tee ethanol extract on methicillin-resistant *Staphylococcus aureus* NPRC R003 (A), *S. epidermidis* ATCC 35984 (B), coagulase-positive staphylococci NPRC 308 (C), multidrug-resistant *Acinetobacter baumannii* NPRC AB034 (D), multidrug-resistant *Escherichia coli* 2746-08 (E), and multidrug-resistant *Pseudomonas aeruginosa* 5351 (F). For all graphs, continuous lines represent time points for test (with different concentrations of the extract) cultures and dashed lines represent control (with 1.25% dimethylsulfoxide) cultures. Each symbol indicates the mean \pm standard deviation for three independent experiments performed in duplicate. OD, optical density.

derivatives, found primarily in *Aristolochia* spp., are well-documented nephrotoxic and carcinogenic agents.²⁶ According to information from the U.S. Food and Drug Administration, Health Canada, and the Medicine Controls Agency (the United Kingdom), use of botanical products containing the compounds is no longer permitted.²⁷ In 2011,

therefore, the National Drug Committee removed Krai-Krue from Thai traditional herbal recipes.²⁰ For this reason, the remedies used in this investigation, including Um-Ma-Luk-Ka-Wa-Tee, did not contain *Aristolochia* spp. However, neither a Thai traditional preparation containing Krai-Krue, Homnawakod, nor a new version of the formula without this

medicinal plants caused any acute nephrotoxicity *in vivo*. In addition, the quality of the formulas in terms of chemical profile does not differ.²⁰

With the exception of *Dischidia rafflesiana*, antibacterial activities of *L. inermis*, *C. sativum*, *P. emblica*, *T. bellirica*, and *G. glabra* have been documented. Dried powdered Um-Ma-Luk-Ka-Wa-Tee contains approximately 55% (w/w) of *G. glabra*, traditionally used for treating upper respiratory tract ailments, including cough, hoarseness, sore throat, and bronchitis.²⁸ Various extracts from *G. glabra* showed antioxidant, anti-inflammatory, and antibacterial activities against human pathogens, such as *Bacillus coagulans*, *E. coli*, *Salmonella typhimurium*, and *S. aureus*. In previous investigations, MICs (0.3–40 mg/mL) of ethanol and water from root and leaves of this plant against *S. aureus*, *Bacillus subtilis*, *E. faecalis*, *Candida albicans*, and *Mycobacterium tuberculosis* were much higher than that of the recipe extract.^{28–31} Glycyrrhizin generally is considered the major biologically active principal and has been used industrially, but most of the antibacterial activity from this plant is due to glabridin, which was similar to the activities of Um-Ma-Luk-Ka-Wa-Tee (3.9–250 µg/mL).³⁰

Previous studies on *L. inermis*, traditionally used as antiseptic for burns and wounds in Yemen³² and Nigeria³³ have revealed that ethanol, water, and ethyl acetate extracts of the plant exhibited broad-spectrum antibacterial activity on *S. aureus*, *Streptococcus* sp., *E. faecalis*, *Proteus mirabilis*, *Klebsiella pneumoniae*, *P. aeruginosa*, and *E. coli* at certain concentrations. Lawsonia, which was known as the major bioactive constituent, had a wide spectrum of antimicrobial activity, including antiviral, antimycotic, and antiparasitic activities.³⁴ Um-Ma-Luk-Ka-Wa-Tee extract (MIC, 31 to <16 µg/mL) had potent anti-MRSA activity greater than that of *L. inermis* extract (5–80 mg/mL).^{33,35} In addition, *T. bellirica* chloroform extract—as well as the active constituents of this plant, termilignan, thannilignan, and anolignan—show antiviral, antifungal, and antimalarial activities.³⁶ Methanol extract of this plant had inhibitory activities on *Bacillus cereus*, *Listeria monocytogenes*, *S. aureus*, *E. coli*, and *Salmonella anatum*, when tested by agar well diffusion method (100 mg/mL; 60 µL/well).³⁷ Air-dried fruits of *Phyllanthus emblica* have been widely used as a source of natural antioxidants in traditional Chinese and Indian medicine.³⁸ Although anti-MRSA activity of *P. emblica* has never been reported, extract of *P. emblica* has been reported to have the ability to prevent the colonization of *C. albicans*.³⁹

In addition to *Aristolochia* spp., our results showed that *M. fragrans*, *D. loureiri*, and *A. daturica* were frequently mentioned medicinal plants used in the household remedies. *M. fragrans* (seed, seed kernel, and mace), *D. loureiri* (stem wood), and *A. daturica* (fruit and root) are widely used in Asian medicinal ingredients. This finding agrees with values reported in the literature; extracts and chemical constituents of the medicinal plant have been found to possess various biological activities. *M. fragrans* and its active principals, such as alkyl benzene derivatives (myristicin and elemicin), terpenes, α -pinene, β -pinene, and myristic acid, have been extensively examined for antibacterial, antiviral, antioxidant, anti-inflammatory, anticancer, and anticariogenic activities.^{40–42} Ethyl acetate, ethanol, and methanol extract possessed significant antibiofilm and antibacterial activities

against various oral primary colonizers, such as *Streptococcus mutans*, *S. sanguis*, *S. sobrinus*, *S. salivarius*, *Lactobacillus acidophilus*, *L. casei*, and *Actinomyces viscosus*.⁴³ *D. loureiri* (known in Thai as Chan-Dang) has been found to possess antinociceptive, antipyretic,⁴⁴ antioxidant, and anti-inflammatory activities.⁴⁵ Flavonoid derivatives of this plant, such as loureirin D and (2*S*)-pinocembrin, showed moderate antimicrobial activity against *S. aureus*, *B. subtilis*, *Botrytis cinerea*, and *Cladosporium herbarum*.⁴⁶ Root of *A. daturica* has been stated to have useful properties, including anti-inflammatory, antiasthmatic, antipyretic, and antiacne effects. Studies conducted by Nam⁴⁷ and Lechner⁴⁸ and colleagues additionally reported antibacterial activities on *S. aureus*, MRSA, *Mycobacterium fortuitum*, and *Propionibacterium acnes* of extracts from root and its active constituents, faltarindiol.

It would appear that this paper represents the first work exploring *in vitro* activity against MDR pathogens of the following Thai traditional remedies: Dhart-Ban-Chob, Kheaw-Hom, Learng-Pid-Sa-Mud, Pra-Sa-Chan-Dang, Tree-Hom, Chan-Ta-Lee-La, and Um-Ma-Luk-Ka-Wa-Tee. Additionally, it reports for the first time that the ethanol extract of Um-Ma-Luk-Ka-Wa-Tee possessed significant anti-MRSA activity with low cytotoxic effect. The information obtained from this study suggested that glabridin may possibly be used as a biomarker for antibacterial activity of this household remedy. However, it is necessary to further investigate and understand the relationship between antibacterial activity and glabridin content in the recipe. Further studies on other biological activities related to their traditional application are required to elucidate the appropriate biomarkers as well as to explain the treatment mechanisms of household remedies.

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Author Disclosure Statement

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References

1. Roberts RR, Hota B, Ahmad I, et al. Hospital and societal costs of antimicrobial-resistant infections in a Chicago teaching hospital: implications for antibiotic stewardship. *Clin Infect Dis* 2009;49:1175–1184.
2. de Kraker ME, Davey PG, Grundmann H. Mortality and hospital stay associated with resistant *Staphylococcus aureus* and *Escherichia coli* bacteremia: estimating the burden of antibiotic resistance in Europe. *PLoS Med* 2011;8:e1001104.
3. de Kraker ME, Wolkewitz M, Davey PG, et al. Burden of antimicrobial resistance in European hospitals: excess mortality and length of hospital stay associated with bloodstream infections due to *Escherichia coli* resistant to third-generation cephalosporins. *J Antimicrob Chemother* 2011;66:398–407.
4. Klevens RM, Edwards JR, Richards CL, et al. Estimating health care-associated infections and deaths in U.S. hospitals, 2002. *Public Health Rep* 2007;122:160–166.

5. Gales AC, Sader HS, Ribeiro J, Zoccoli C, Barth A, Pignatari AC. Antimicrobial susceptibility of gram-positive bacteria isolated in Brazilian hospitals participating in the SENTRY Program (2005–2008). *Braz J Infect Dis* 2009;13:90–98.
6. Saleem AF, Ahmed I, Mir F, Ali SR, Zaidi AK. Pan-resistant *Acinetobacter* infection in neonates in Karachi, Pakistan. *J Infect Dev Ctries* 2012;4:30–37.
7. Tuon FF, Gortz LW, Rocha JL. Risk factors for pan-resistant *Pseudomonas aeruginosa* bacteremia and the adequacy of antibiotic therapy. *Braz J Infect Dis* 2012;16:351–356.
8. Cattaneo C, Antoniazzi F, Casari S, et al. *P. aeruginosa* bloodstream infections among hematological patients: an old or new question? *Ann Hematol* 2012;91:1299–304.
9. Chokevivat V, Chuthaputti A. The role of Thai traditional medicine in health promotion. 6th Global Conference on Health Promotion (6GCHP). Department for the Development of Thai Traditional and Alternative Medicine, Ministry of Public Health, Thailand; 2005, pp. 1–25.
10. National Essential Drug List Committee. The National List of Essential Drugs: Last update 25th May 2011 (List of Herbal Medicinal Products). Bangkok: War Veterans Administration Printing; 2011.
11. Kaur GJ, Arora DS. Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare* and *Trachyspermum ammi*. *BMC Complement Altern Med* 2009;9:30.
12. Clinical and Laboratory Standards Institute. M07-A8: Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria That Grow Aerobically. Approved Standard-Eighth Edition. Wayne, PA: Clinical and Laboratory Standards Institute; 2009.
13. Rios JL, Recio MC. Medicinal plants and antimicrobial activity. *J Ethnopharmacol* 2005;100:80–84.
14. Kuete V. Potential of Cameroonian plants and derived products against microbial infections: a review. *Planta Med* 2010;76:1479–1491.
15. Chusri S, Settharaksa S, Chokpaisarn J, Limsuwan S, Voravuthikunchai SP. Thai Herbal formulas used for wound treatment: a study of their antibacterial potency, anti-inflammatory, antioxidant, and cytotoxicity effects. *J Altern Complement Med* 2013;19:671–676.
16. Li S, Zhao J, Liu J, et al. Prospective randomized controlled study of a Chinese herbal medicine compound Tangzu Yuyang Ointment for chronic diabetic foot ulcers: a preliminary report. *J Ethnopharmacol* 2011;133:543–550.
17. Shyni GL, Ratheesh M, Sindhu G, Helen A. Anti-inflammatory and antioxidant effects of Jeevaneeya Rasayana: an ayurvedic polyherbal formulation on acute and chronic models of inflammation. *Immunopharmacol Immunotoxicol* 2010;32:569–575.
18. Ohno T, Inoue M, Ogihara Y. Suppressive effect of Shichimotsu-koka-to (Kampo medicine) on pulmonary metastasis of B16 melanoma cells. *Biol Pharm Bull* 2002;25:880–884.
19. Hong HT, Kim HJ, Lee TK, et al. Inhibitory effect of a Korean traditional medicine, Honghwain-Jahage (water extracts of *Carthamus tinctorius* L. seed and *Hominis placenta*) on interleukin-1-mediated bone resorption. *J Ethnopharmacol* 2002;79:143–148.
20. Tripatara P, Onlamul W, Booranasubkajorn S, et al. The safety of Homnawakod herbal formula containing *Aristolochia tagala* Cham in Wistar rats. *BMC Complement Altern Med* 2012;12:170.
21. Kondo S, Sattaponpan C, Phongpaichit S, Srijan A, Itharat A. Antibacterial activity of Thai medicinal plants *Pikubtenjakul*. *J Med Assoc Thai* 2010;93:S131–135.
22. Sattaponpan C, Kondo S. Antibacterial activity of crude extracts of prasaproyhai formula and its components against pathogenic bacteria. *J Med Assoc Thai* 2011;94:S153–161.
23. Nuaeissara S, Kondo S, Itharat A. Antimicrobial activity of the extracts from Benchalokawichian remedy and its components. *J Med Assoc Thai* 2011;94:S172–177.
24. Flatie T, Gedif T, Asres K, Gebre-Mariam T. Ethnomedical survey of Berta ethnic group Assosa Zone, Benishangul-Gumuz regional state, mid-west Ethiopia. *J Ethnobiol Ethnomed* 2009;5:14.
25. Sathornviriyapong S, Picheansoonthon C, Tiasakul R, Tiyaworanant S, Reutrakul V. Botanical origin and identification of Krai-Krue herbal plant. *Kasetsart J (Nat Sci)* 2007;41:420–32.
26. Arlt VM, Stiborova M, Schmeiser HH. Aristolochic acid as a probable human cancer hazard in herbal remedies: a review. *Mutagenesis* 2002;17:265–77.
27. Debelle FD, Vanherweghem JL, Nortier JL. Aristolochic acid nephropathy: a worldwide problem. *Kidney Int* 2008;74:158–169.
28. Nitalikar MM, Munde KC, Dhore BV, Shikalgar SN. Studies of antibacterial activities of *Glycyrrhiza glabra* root extract. *Int J PharmTech Res* 2010;2:899–901.
29. Nirmala P, Selvaraj T. Anti-inflammatory and anti-bacterial activities of *Glycyrrhiza glabra* L. *J Agricult Technol* 2011;7:815–823.
30. Gupta VK, Fatima A, Faridi U, et al. Antimicrobial potential of *Glycyrrhiza glabra* roots. *J Ethnopharmacol* 2008;116:377–380.
31. Li W, Asada Y, Yoshikawa T. Antimicrobial flavonoids from *Glycyrrhiza glabra* hairy root cultures. *Planta Med* 1998;64:746–747.
32. Ali NA, Julich WD, Kusnick C, Lindequist U. Screening of Yemeni medicinal plants for antibacterial and cytotoxic activities. *J Ethnopharmacol* 2001;74:173–179.
33. Muhammad HS, Muhammad S. The use of *Lawsonia inermis* linn. (henna) in the management of burn wound infections. *Afr J Biotechnol* 2005;4:934–937.
34. Babu PD, Subhasree RS. Antimicrobial activities of *Lawsonia inermis*: a review. *Academic J Plant Sci* 2009;2:231–232.
35. Chomnawang MT, Surassmo S, Wongsariya K, Bunyapraphatsara N. Antibacterial activity of Thai medicinal plants against methicillin-resistant *Staphylococcus aureus*. *Fitoterapia* 2009;80:102–104.
36. Valsaraj R, Pushpangadan P, Smitt UW, et al. New anti-HIV-1, antimalarial, and antifungal compounds from *Terminalia bellerica*. *J Nat Prod* 1997;60:739–742.
37. Shan B, Cai YZ, Brooks JD, Corke H. The *in vitro* antibacterial activity of dietary spice and medicinal herb extracts. *Int J Food Microbiol* 2007;117:112–119.
38. Liu XL, Cui C, Zhao MM, et al. Identification of phenolics in the fruit of emblica (*Phyllanthus emblica* L.) and their antioxidant activities. *Food Chem* 2008;109:909–915.
39. Thaweboon B, Thaweboon S. Effect of *Phyllanthus emblica* Linn. on candida adhesion to oral epithelium and denture acrylic. *Asian Pac J Trop Med* 2011;4:41–45.
40. Ozaki Y, Soedigdo S, Wattimena YR, Suganda AG. Anti-inflammatory effect of mace, aril of *Myristica fragrans* Houtt., and its active principles. *Jpn J Pharmacol* 1989;49:155–163.

41. Akinboro A, Mohamed KB, Asmawi MZ, Othman AS, Ying TH, Maidin SM. Mutagenic and antimutagenic assessment of methanol leaf extract of *Myristica fragrans* (Houtt.) using *in vitro* and *in vivo* genetic assays. *Drug Chem Toxicol* 2011;35:412–422.
42. Chirathaworn C, Kongcharoensuntorn W, Dechdougchan T, Lowanitchapat A, Sa-nguanmoo P, Poovorawan Y. *Myristica fragrans* Houtt. methanolic extract induces apoptosis in a human leukemia cell line through SIRT1 mRNA downregulation. *J Med Assoc Thai* 2007;90: 2422–2428.
43. Shafiei Z, Shuhairi NN, Md Fazly Shah Yap N, Harry Sibungkil CA, Latip J. Antibacterial activity of *Myristica fragrans* against oral pathogens. *Evid Based Complement Alternat Med* 2012;2012:825362.
44. Reanmongkol W, Subhadhirasakul S, Bouking P. Antinociceptive and antipyretic activities of extracts and fractions from *Dracaena loureiri* in experimental animals. *Songklanakarin J Sci Technol* 2003;25:467–476.
45. Likhitwitayawuid K, Sawasdee K, Kirtikara K. Flavonoids and stilbenoids with COX-1 and COX-2 inhibitory activity from *Dracaena loureiri*. *Planta Med* 2002;68: 841–843.
46. Ichikawa K, Kitaoka M, Taki M, et al. Retrodihydrochalcones and homoisoflavones isolated from Thai medicinal plant *Dracaena loureiri* and their estrogen agonist activity. *Planta Med* 1997;63:540–543.
47. Lechner D, Stavri M, Oluwatuyi M, Pereda-Miranda R, Gibbons S. The anti-staphylococcal activity of *Angelica dahurica* (Bai Zhi). *Phytochemistry* 2004;65:331–335.
48. Nam C, Kim S, Sim Y, Chang I. Anti-acne effects of Oriental herb extracts: a novel screening method to select anti-acne agents. *Skin Pharmacol Appl Skin Physiol* 2003;16:84–90.

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