CROSSTALK

CrossTalk opposing view: The dominant mechanism causing disuse muscle atrophy is proteolysis

Michael B. Reid¹, Andrew R. Judge² and Sue C. Bodine³

¹Department of Applied Physiology and Kinesiology, University of Florida, Gainesville, FL, USA ²Department of Physical Therapy, University of Florida, Gainesville, FL, USA ³Department of Neurobiology, Physiology and Behavior, University of California, Davis, Davis, CA, USA

Email: michael.reid@ufl.edu

Skeletal muscle adapts to a prolonged reduction in physical activity by decreasing muscle fibre size and protein content. This physiological response is commonly termed disuse atrophy. It is most clearly stimulated by conditions that reduce mechanical loading of postural or respiratory muscles in the absence of disease and neural deficits. Examples of such conditions include bed rest, joint immobilization, limb suspension, microgravity and mechanical ventilation. In muscle disuse, as in all atrophic processes, the rate of proteolysis exceeds the rate of protein synthesis causing net loss of protein. A dispassionate review of the literature shows that both aspects of muscle protein homeostasis are disrupted by prolonged disuse. Proteolysis increases and synthesis declines. These changes have additive effects, contributing jointly to protein loss and muscle atrophy. The relative magnitudes of these contributions have not been measured directly and are not known. Therefore, like most scientists, we are resolutely ecumenical on the issue of proteolysis *vs.* synthesis. In contrast, a few individuals fervently argue that disuse atrophy is caused by decreased protein synthesis and that proteolysis is unimportant. This viewpoint has created unnecessary controversy in the literature and heated discussion at scientific conferences. This controversy is the focus of our CrossTalk debate.

To be clear, the 'dominant mechanism' of disuse atrophy is not known. Rates of proteolysis vs. protein synthesis cannot be quantified using existing methods nor can differences in these rates be measured directly. Attempts to estimate these values must be based on first principles, must involve untested assumptions, and cannot be validated experimentally. This makes it impossible to assign 'dominance' with scientific rigor. Instead of speculating on this issue, we will present evidence for the biological importance of proteolysis in disuse atrophy. The extent of peer-reviewed research, the diversity of experimental approaches, and the level of mechanistic detail make a compelling case for our position that proteolysis is essential for disuse atrophy.

Disuse stimulates proteolysis

For almost four decades, we have known that disuse promotes atrophy of skeletal muscle by stimulating protein breakdown. This process involves the co-activation of multiple proteolytic mechanisms that have complementary roles in degrading muscle protein. The ubiquitin–proteasome pathway is the dominant mechanism of protein breakdown during muscle atrophy (Bodine, 2013). Disuse activates this pathway in muscle, as shown by three lines of evidence: First, muscle increases

the expression of key gene products that regulate pathway activity. Mechanical unloading stimulates a rise in mRNA for the polyubiquitin gene, E2 ubiquitin conjugating proteins, E3 ubiquitin ligases, and subunits of the proteasome complex (Taillandier et al. 1996; Ikemoto et al. 2001; Jones et al. 2004; Nikawa et al. 2004; Hussain et al. 2010). Gene targets of particular interest are MAFbx/atrogin1 and MuRF1, two muscle-specific E3 ligases that regulate muscle atrophy in a variety of experimental models (Bodine et al. 2001). Like virtually all conditions of muscle wasting, MAFbx/atrogin1 and MuRF1 mRNA are increased in disuse atrophy. This response has been documented in diaphragm of mechanically ventilated humans and rodents (DeRuisseau et al. 2005; Levine et al. 2008, 2011; Hussain et al. 2010), limb muscles of humans after bed rest, lower limb suspension, or knee immobilization (Jones et al. 2004; de Boer et al. 2007; Abadi et al. 2009; Gustafsson et al. 2010; Reich et al. 2010; Wall et al. 2014), and limb muscles of rodents after hindlimb unloading, limb immobilization, or spaceflight (Nikawa et al. 2004; Senf et al. 2008; Andrianjafiniony et al. 2010). Second, upregulation of these pathway elements has the predicted outcome: ubiquitin conjugation to muscle proteins is increased in both humans and rodents (Ikemoto et al. 2001; DeRuisseau et al. 2005; Abadi et al. 2009; Ferreira et al. 2009; Levine et al. 2011; Brocca et al. 2012). Third, unloading increases the proteolytic activities of the 20S- and 26S-proteasome complexes. This occurs in human and rodent muscles (Ikemoto et al. 2001; Shanely et al. 2002; Levine et al. 2011), facilitating breakdown of ubiquitin-conjugated proteins.

Michael Reid is Professor of Applied Physiology and Kinesiology, and Dean of the College of Health and Human Performance at the University of Florida. He is an established investigator whose research addresses the cellular and molecular mechanisms that underlie muscle weakness and fatigue. Andrew Judge is currently an Associate Professor of Physical Therapy at the University of Florida where he runs a muscle plasticity lab focused primarily on skeletal muscle wasting associated with disuse and cancer-induced cachexia. His research studies the regulation of intracellular signalling pathways which cause skeletal muscle wasting during these conditions. Sue Bodine is currently a Professor of Neurobiology, Physiology



and Behaviour at the University of California, Davis where she runs a Neuromuscular Physiology Lab focused on muscle plasticity and the regulation of skeletal muscle mass. Her research investigates the mechanisms underlying both skeletal muscle growth and atrophy, with particular focus on mechanisms of protein degradation.

Muscle proteases and autophagy are also activated during disuse atrophy. This is particularly well documented for the cysteine-aspartic protease family, i.e. the caspases. The original discovery that unloading increases caspase-3 mRNA in the human diaphragm (Levine et al. 2008) led to animal studies confirming that caspase-3 enzyme activity is increased in unloaded diaphragm and limb muscles (Ferreira et al. 2009; Nelson et al. 2012; Talbert et al. 2013). Other members of the caspase family that are upregulated by unloading include caspase-6, -8, -9 and -12 (Taillandier et al. 1996; Andrianjafiniony et al. 2010). Calpain, a calcium-dependent protease, exhibits a similar response. Mechanical unloading of either diaphragm or limb muscle causes an increase in calpain mRNA and calpain activity (Taillandier et al. 1996; Shanely et al. 2002; DeRuisseau et al. 2005; Ferreira et al. 2009; Andrianiafinionv et al. 2010; Nelson et al. 2012; Talbert et al. 2013). Finally, emerging evidence indicates that autophagy via the lysosomal system is also increased during disuse atrophy of human and rodent muscles (Taillandier et al. 1996; Ikemoto et al. 2001; Andrianjafiniony et al. 2010; Hussain et al. 2010; Brocca et al. 2012). In aggregate, unloading activates all major proteolytic mechanisms - the ubiquitin-proteasome pathway, protease activity, and autophagy-thereby promoting the degradation of large molecular weight myofibrillar proteins, cytosolic proteins and smaller polypeptide residues.

Upregulation of proteolytic mechanisms (above) increases protein breakdown in unloaded skeletal muscle. This fact has been demonstrated in rodents using limb immobilization, hindlimb unloading, and mechanical ventilation (Goldspink, 1977; Loughna *et al.* 1987; Shanely *et al.* 2002) and in humans using unilateral lower limb suspension (Tesch *et al.* 2008). The latter study employed an innovative microdialysis technique to measure inter-

A E2 conjugases mice atroginbedrest ubiquitin conjugates muscle in vivo autophagy Proteolysis electron microscopy mRNA isolates Immobilization protease activity quadriceps hindlimb unloading RT-PCR Tats vastus lateralis isolated muscles Northern blotprotein breakdown mechanical ventilation immunohistochemistry diaphragm microgravity Western blot proteasome humans 3-MH releaseprotein extracts calpains MuRF-1

Figure 1. Word clouds illustrating the experimental approaches used to assess proteolysis (A) and protein synthesis (B) during disuse atrophy 3-MH, 3-methylhistidine; ULLS, unilateral lower limb suspension.

CrossTalk

stitial levels of 3-methylhistidine, an endogenous biomarker of myofibrillar degradation. Unloading for 3 days increased 3-methylhistidine levels by over 40%: evidence of increased breakdown of myofibrillar protein. This finding is consistent with recent evidence that human muscle undergoes significant atrophy within 2–5 days of being unloaded (Grosu *et al.* 2012; Suetta *et al.* 2012; Wall *et al.* 2014) and shows that greater protein breakdown is fundamental to the disuse atrophy process across species.

Conclusion

It is clear that proteolysis plays an important role in disuse atrophy, perhaps *the most* important role. This conclusion is firmly rooted in modern interdisciplinary biology (Fig. 1*A*). It incorporates what we know about muscle cell signalling, gene expression at the mRNA and protein levels, protein biochemistry, proteasome and protease activities, and regulation of protein breakdown. Plus it is inclusive, applying equally to postural and respiratory muscles of rodents and humans.

How can our learned opponent and a handful of other skeptics reject the preponderance of evidence on this issue? Historically, the position that proteolysis is unimportant has derived from measurements of fractional synthesis rate (FSR) in human vastus lateralis muscle during disuse. FSR data have been used to estimate the importance of synthesis based on first principles and assumptions that are either unstated or untested. This narrow approach (Fig. 1B), based on one end-point from one muscle of one species, has been justified by arguments that rodent models of disuse are 'heuristically misleading', that 'dynamic' FSR data are inherently more valid than 'snapshot' (i.e. any other) measurements, and that approaches which yield different outcomes are flawed and unreliable (Rennie *et al.* 2010). Such a position leaves little room for open-minded inquiry or free exchange of ideas. We must move beyond an either/or approach in order to broaden our understanding and advance the field. The time has come to acknowledge the fact that proteolysis is an essential component of disuse atrophy.

Call for comments

Readers are invited to give their views on this and the accompanying CrossTalk articles in this issue by submitting a brief (250 word) comment. Comments may be submitted up to 6 weeks after publication of the article, at which point the discussion will close and the CrossTalk authors will be invited to submit a 'Last Word'. Please email your comment to journals@physoc.org.

References

- Abadi A, Glover EI, Isfort RJ, Raha S, Safdar A, Yasuda N, Kaczor JJ, Melov S, Hubbard A, Qu X, Phillips SM & Tarnopolsky M (2009). Limb immobilization induces a coordinate down-regulation of mitochondrial and other metabolic pathways in men and women. *PLoS One* **4**, e6518.
- Andrianjafiniony T, Dupre-Aucouturier S, Letexier D, Couchoux H & Desplanches D (2010). Oxidative stress, apoptosis, and proteolysis in skeletal muscle repair after unloading. *Am J Physiol Cell Physiol* **299**, C307–C315.
- Bodine SC (2013). Disuse-induced muscle wasting. *Int J Biochem Cell Biol* **45**, 2200–2208.
- Bodine SC, Latres E, Baumhueter S, Lai VK, Nunez L, Clarke BA, Poueymirou WT, Panaro FJ, Na E, Dharmarajan K, Pan ZQ, Valenzuela DM, DeChiara TM, Stitt TN, Yancopoulos GD & Glass DJ (2001). Identification of ubiquitin ligases required for skeletal muscle atrophy. Science 294, 1704–1708.
- Brocca L, Cannavino J, Coletto L, Biolo G, Sandri M, Bottinelli R & Pellegrino MA (2012). The time course of the adaptations of human muscle proteome to bed rest and the underlying mechanisms. *J Physiol* 590, 5211–5230.
- de Boer MD, Selby A, Atherton P, Smith K, Seynnes OR, Maganaris CN, Maffulli N, Movin T, Narici MV & Rennie MJ (2007). The temporal responses of protein synthesis, gene expression and cell signalling in human quadriceps muscle and patellar tendon to disuse. J Physiol 585, 241–251.
- DeRuisseau KC, Kavazis AN, Deering MA, Falk DJ, Van Gammeren D, Yimlamai T, Ordway GA & Powers SK (2005). Mechanical ventilation induces alterations of the ubiquitin-proteasome pathway in the diaphragm. *J Appl Physiol (1985)* **98**, 1314–1321.

J Physiol 592.24



- Ferreira R, Vitorino R, Neuparth MJ, Appell HJ, Duarte JA & Amado F (2009). Proteolysis activation and proteome alterations in murine skeletal muscle submitted to 1 week of hindlimb suspension. *Eur J Appl Physiol* **107**, 553–563.
- Goldspink DF (1977). The influence of immobilization and stretch on protein turnover of rat skeletal muscle. *J Physiol* **264**, 267–282.
- Grosu HB, Lee YI, Lee J, Eden E, Eikermann M & Rose KM (2012). Diaphragm muscle thinning in patients who are mechanically ventilated. *Chest* **142**, 1455–1460.
- Gustafsson T, Osterlund T, Flanagan JN, von Waldén F, Trappe TA, Linnehan RM & Tesch PA (2010). Effects of 3 days unloading on molecular regulators of muscle size in humans. J Appl Physiol (1985) 109, 721–727.
- Hussain SN, Mofarrahi M, Sigala I, Kim HC, Vassilakopoulos T, Maltais F, Bellenis I, Chaturvedi R, Gottfried SB, Metrakos P, Danialou G, Matecki S, Jaber S, Petrof BJ & Goldberg P (2010). Mechanical ventilation-induced diaphragm disuse in humans triggers autophagy. *Am J Respir Crit Care Med* **182**, 1377–1386.
- Ikemoto M, Nikawa T, Takeda S, Watanabe C, Kitano T, Baldwin KM, Izumi R, Nonaka I, Towatari T, Teshima S, Rokutan K & Kishi K (2001). Space shuttle flight (sts-90) enhances degradation of rat myosin heavy chain in association with activation of ubiquitin-proteasome pathway. *FASEB J* 15, 1279–1281.
- Jones SW, Hill RJ, Krasney PA, O'Conner B, Peirce N & Greenhaff PL (2004). Disuse atrophy and exercise rehabilitation in humans profoundly affects the expression of genes associated with the regulation of skeletal muscle mass. *FASEB J* **18**, 1025–1027.
- Levine S, Biswas C, Dierov J, Barsotti R, Shrager JB, Nguyen T, Sonnad S, Kucharchzuk JC, Kaiser LR, Singhal S & Budak MT (2011). Increased proteolysis, myosin depletion, and atrophic AKT-FOXO signaling in human diaphragm disuse. *Am J Respir Crit Care Med* **183**, 483–490.

CrossTalk

- Levine S, Nguyen T, Taylor N, Friscia ME, Budak MT, Rothenberg P, Zhu J, Sachdeva R, Sonnad S, Kaiser LR, Rubinstein NA, Powers SK & Shrager JB (2008). Rapid disuse atrophy of diaphragm fibers in mechanically ventilated humans. *N Engl J Med* **358**, 1327–1335.
- Loughna PT, Goldspink DF & Goldspink G (1987). Effects of hypokinesia and hypodynamia upon protein-turnover in hindlimb muscles of the rat. *Aviat Space Environ Med* **58**, A133–A138.
- Nelson WB, Smuder AJ, Hudson MB, Talbert EE & Powers SK (2012). Cross-talk between the calpain and caspase-3 proteolytic systems in the diaphragm during prolonged mechanical ventilation. *Crit Care Med* **40**, 1857–1863.
- Nikawa T, Ishidoh K, Hirasaka K, Ishihara I, Ikemoto M, Kano M, Kominami E, Nonaka I, Ogawa T, Adams GR, Baldwin KM, Yasui N, Kishi K & Takeda S (2004). Skeletal muscle gene expression in space-flown rats. *FASEB J* **18**, 522–524.
- Reich KA, Chen YW, Thompson PD, Hoffman EP & Clarkson PM (2010). Forty-eight hours of unloading and 24 h of reloading lead to changes in global gene expression patterns related to ubiquitination and oxidative stress in humans. *J Appl Physiol (1985)* **109**, 1404–1415.
- Rennie MJ, Selby A, Atherton P, Smith K, Kumar V, Glover EL & Philips SM (2010). Facts, noise and wishful thinking: Muscle protein turnover in aging and human disuse atrophy. *Scand J Med Sci Sports* **20**, 5–9.
- Senf SM, Dodd SL, McClung JM & Judge AR (2008). Hsp70 overexpression inhibits NF- κ B and Foxo3a transcriptional activities and prevents skeletal muscle atrophy. *FASEB J* **22**, 3836–3845.
- Shanely RA, Zergeroglu MA, Lennon SL, Sugiura T, Yimlamai T, Enns D, Belcastro A & Powers SK (2002). Mechanical ventilationinduced diaphragmatic atrophy is associated with oxidative injury and increased proteolytic activity. *Am J Respir Crit Care Med* **166**, 1369–1374.

- Suetta C, Frandsen U, Jensen L, Jensen MM,
 Jespersen JG, Hvid LG, Bayer M, Petersson SJ,
 Schroder HD, Andersen JL, Heinemeier KM,
 Aagaard P, Schjerling P & Kjaer M (2012).
 Aging affects the transcriptional regulation of
 human skeletal muscle disuse atrophy. *PLoS One* 7, e51238.
- Taillandier D, Aurousseau E, Meynial-Denis D, Bechet D, Ferrara M, Cottin P, Ducastaing A, Bigard X, Guezennec CY, Schmid HP & Attaix D (1996). Coordinate activation of lysosomal, Ca²⁺-activated and ATP-ubiquitin-dependent proteinases in the unweighted rat soleus muscle. *Biochem* J **316**, 65–72.
- Talbert EE, Smuder AJ, Min K, Kwon OS & Powers SK (2013). Calpain and caspase-3 play required roles in immobilization-induced limb muscle atrophy. *J Appl Physiol (1985)* 114, 1482–1489.
- Tesch PA, von Walden F, Gustafsson T, Linnehan RM & Trappe TA (2008). Skeletal muscle proteolysis in response to short-term unloading in humans. *J Appl Physiol (1985)* **105**, 902–906.
- Wall BT, Dirks ML, Snijders T, Senden JM, Dolmans J & van Loon LJ (2014). Substantial skeletal muscle loss occurs during only 5 days of disuse. *Acta Physiol* (*Oxf*) **210**, 600–611.

Additional information

Competing interests

None declared.

Funding

This work was supported by the U.S. National Institute of Arthritis, Musculoskeletal, and Skin Diseases grants R01AR060209 (to A.R.J.), R01AR062083 (to M.B.R.), U.S. National Institute of Diabetes and Digestive and Kidney Diseases grant DK75801 (to S.C.B.), and U.S. Veterans Administration Rehabilitation Research and Development grant E7766R (to S.C.B.).