

ORIGINAL ARTICLE

Omega-3 polyunsaturated fatty acid and ursodeoxycholic acid have an additive effect in attenuating diet-induced nonalcoholic steatohepatitis in mice

Ja Kyung Kim¹, Kwan Sik Lee¹, Dong Ki Lee¹, Su Yeon Lee², Hye Young Chang², Junjeong Choi³ and Jung Il Lee¹

Nonalcoholic steatohepatitis (NASH) can progress into liver cirrhosis; however, no definite treatment is available. Omega-3 polyunsaturated fatty acid (omega-3) has been reported to alleviate experimental NASH, although its beneficial effect was not evident when tested clinically. Thus, this study aimed to investigate the additive effect of omega-3 and ursodeoxycholic acid (UDCA) on diet-induced NASH in mice. C57BL/6 mice were given a high-fat diet (HFD) for 24 weeks, at which point the mice were divided into three groups and fed HFD alone, HFD with omega-3 or HFD with omega-3 in combination with UDCA for another 24 weeks. Feeding mice an HFD and administering omega-3 improved histologically assessed liver fibrosis, and UDCA in combination with omega-3 further attenuated this disease. The assessment of *collagen α1(I)* expression agreed with the histological evaluation. Omega-3 in combination with UDCA resulted in a significant attenuation of inflammation whereas administering omega-3 alone failed to improve histologically assessed liver inflammation. Quantitative analysis of *tumor necrosis factor α* showed an additive effect of omega-3 and UDCA on liver inflammation. HFD-induced hepatic triglyceride accumulation was attenuated by omega-3 and adding UDCA accentuated this effect. In accordance with this result, the expression of *sterol regulatory binding protein-1c* decreased after omega-3 administration and adding UDCA further diminished *SREBP-1c* expression. The expression of inducible *nitric oxide synthase (iNOS)*, which may reflect oxidative stress-induced tissue damage, was suppressed by omega-3 administration and adding UDCA further attenuated *iNOS* expression. These results demonstrated an additive effect of omega-3 and UDCA for alleviating fibrosis, inflammation and steatosis in diet-induced NASH.

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INTRODUCTION

Nonalcoholic fatty liver disease, which is one of the most prevalent chronic liver diseases worldwide, consists of a spectrum of conditions that includes nonalcoholic steatohepatitis (NASH).^{1–3} Although simple steatosis may undergo an indolent natural course, when it is accompanied by advanced liver fibrosis it often takes a similar clinical course to that of chronic hepatitis C virus infection with advanced fibrosis.⁴ However, with all of these possible liver-associated morbidities and mortalities, a definitive treatment that would stop or reverse the progression of the disease has yet to be discovered.

Omega-3 polyunsaturated fatty acid (omega-3) is clinically used for treating hypertriglyceridemia.⁵ There is evidence that omega-3 suppresses hepatic triglyceride (TG) accumulation and reduces inflammation, thereby alleviating NASH in animal models.^{6–9} However, solid clinical evidence of the benefit of omega-3 in NASH is lacking, even though the absence of significant adverse effects of omega-3 makes it convenient to use as a supplement.^{10–12}

Ursodeoxycholic acid (UDCA) is reported to have a beneficial role in experimental NASH because of its anti-inflammatory, hepato-protective and antioxidant effects.^{13–16} However, clinical trials have failed to show meaningful

¹Department of Internal Medicine, Yonsei University College of Medicine, Seoul, Korea; ²Medical Research Center, Gangnam Severance Hospital, Yonsei University College of Medicine, Seoul, Korea and ³Department of Pathology, Yonsei University Wonju College of Medicine, Wonju, Korea
Correspondence: Professor JI Lee or Professor DK Lee, Department of Internal Medicine, Gangnam Severance Hospital, Yonsei University College of Medicine, 211 Eonju-ro Gangnam-gu, Seoul 135-720, Korea.

E-mail: mdflorence@yuhs.ac or DKLEE@yuhs.ac

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improvement in liver inflammation or fibrosis in NASH patients treated with UDCA.^{17,18} Although negative results may be attributable to improperly constructed studies, including suboptimal drug dosage and study duration, it has been suggested that a combination of UDCA with other agents, such as vitamin E, might result in clinically meaningful outcomes;¹⁹ however, a study showed that long-term consumption of high doses of vitamin E (daily dose of 400 IU) increased mortality.²⁰ Safety should be a concern when combination therapy is tested.

Agents that demonstrate beneficial effects on experimental NASH often fail to show significant improvement in human studies. Therefore, a combination therapy that is safe and more effective than either agent alone might have a better outcome in clinical settings. Thus, this study aimed to investigate the effect of omega-3 in combination with UDCA on diet-induced NASH in mice and to compare the results with those of omega-3 alone to assess the possibility of this combination as a treatment for NASH.

MATERIALS AND METHODS

Ethics

The animal experimental procedures and protocols were approved by the Institutional Animal Care and Use Committee of Gangnam Severance Hospital, Yonsei University College of Medicine (Permit Number: 0011). The study was carried out in accordance with the recommendations and restrictions of the Institutional Animal Care and Use Committee.

Animals

Male C57BL/6 mice (12 weeks of age) were obtained from the Central Lab Animal (Seoul, Korea) and housed under a 12-h light–dark cycle. Mice were fed with a diet that consisted of 15% anhydrous milk fat, 1.0% cholesterol and 50% sucrose (a high-fat diet (HFD)) that was purchased from Picolab (Bethlehem, PA, USA). These mice were fed with the HFD for 24 weeks, at which point they were divided into three groups and fed with HFD alone (HFD), HFD and 70 mg kg⁻¹ omega-3 (Pronova Biocare, Sandefjord, Norway; HFD/Omega-3) or HFD and omega-3 in combination with 20 mg kg⁻¹ UDCA (Dae-woong Pharm., Seoul, Korea; HFD/Omega-3+UDCA) for another 24 weeks. Omega-3 and UDCA were diluted in 0.75% Tween-80 and administered orally through sonde for 24 weeks. After 48 weeks of HFD feeding with or without administering experimental agents, the mice were sacrificed and their livers were harvested. A sample of fresh liver tissue was fixed in 10% buffered formalin, and the remaining tissue was snap-frozen in liquid nitrogen and stored at -80 °C. Blood samples were collected by cardiac puncture after the mice were anesthetized and were stored at -80 °C.

Histological evaluation

Sections of liver tissue specimens, fixed in 10% formalin and embedded in paraffin wax, were stained with H&E and Sirius Red for histological evaluation. A blinded pathologist evaluated the slides for inflammation as described in previous studies with minor modifications.^{21–23} Inflammation was histologically quantified by counting inflammatory foci in 20 consecutive high-power fields (×40 objective; average histological grade, grade 0: no foci, grade 1: <2 foci per high-power field, grade 2: ≥2 foci per high-power field). Liver fibrosis was evaluated by calculating the fibrosis ratio using an

image analysis system as described in a previous study with some modifications.²⁴ Briefly, images of Sirius Red-stained sections were captured under ×10 magnification. In the stained section, red indicated fibrosis and gray indicated parenchyma. After interactive thresholding, the image was converted into a binary image. The two-dimensional patterns were measured by direct pixel counting on the binary images. The total area was the sum of the areas of the microscopic fields that included parenchyma and fibrosis. For each slide, the area of fibrosis was evaluated in 20 consecutive high-power fields and was then averaged.

RNA extraction and gene expression analysis by quantitative real-time PCR

Total RNA was extracted from frozen whole liver using Trizol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's protocol. RNA samples were quantified by spectrophotometry. RNA integrity was assessed using agarose gel electrophoresis and ethidium bromide staining. The RNA samples were then diluted in RNase-free water and stored at -70 °C until use. Five micrograms of RNA was reverse transcribed using an RNA PCR kit (version 1.2; Takara Bio Inc., Japan) according to the manufacturer's recommendations. Oligonucleotide primers and TaqMan probes for *collagen α1(I)*, *transforming growth factor (TGF) β*, *tumor necrosis factor (TNF) α*, *sterol regulatory binding protein (SREBP)-1c*, *fatty acid synthase*, *stearoyl-carnitine palmitoyl-coenzyme A desaturase*, *carnitine palmitoyl-coenzyme A transferase* and *inducible nitric oxide synthase (iNOS)* were used with 18S as an internal control. The probes were obtained from Applied Biosystems (Perkin-Elmer/PE Applied Biosystems, Foster City, CA, USA) and purchased in a ready-for-use form in Assays-on-Demand Gene Expression Products. The TaqMan probes were labeled at the 5' end with the reporter dye fluorescein and at the 3' end with minor groove binder nonfluorescent quencher. Quantitative PCR was performed in triplicate for each sample on a Step One Plus Real Time System (Applied Biosystems). Each 20-μl reaction contained 10 μl of TaqMan Fast Universal Master Mix (Applied Biosystems, Darmstadt, Germany), 1 μl of Gene Expression Mix (Perkin-Elmer/PE Applied Biosystem) and 2 μl of cDNA diluted in 7 μl RNase-free water. The thermal cycler conditions were 20 s at 95 °C, 40 cycles of 5 s at 95 °C and then 20 s at 60°. Fold changes in target gene messengerRNA (mRNA) relative to the endogenous 18S control were calculated as described in previous studies.²⁵

Measurement of TG content

Hepatic TG content was quantified using a commercial kit (ab65336, Abcam, Cambridge, MA, USA) according to the manufacturer's recommendations. Briefly, lipid extracts were prepared by the homogenization of 50 mg of frozen tissue in a solution of 5% NP-40. The homogenized tissue was heated to 80–100 °C in a water bath for 2–5 min, then cooled down to room temperature. The samples were purified and diluted (1:10) in distilled H₂O to be measured at OD_{570 nm} according to the manufacturer's instructions.

Statistical analysis

All results are shown as the mean ± (s.e.m.). Data were analyzed by nonparametric analysis (Kruskal–Wallis or Mann–Whitney test) or one-way analysis of variance with Tukey's *post-hoc* analysis. *P* < 0.05 was considered statistically significant. All calculations were performed with SPSS version 15.0 software for Windows (SPSS Inc., Chicago, IL, USA).

RESULTS

Characteristics of mice fed an HFD with or without experimental materials

The changes in body weight during the 48 weeks of the experiment are shown in Figure 1 and serum biochemical findings are summarized in Table 1. Administering omega-3 did not induce significant changes in body weight independent of UDCA combinations. However, serum TG levels decreased significantly as a result of administering omega-3 alone ($P=0.012$) or the concomitant administration of omega-3 and UDCA ($P<0.011$). There was no difference in TG levels between the omega-3 group and omega-3 with UDCA group ($P=0.995$, when compared with the omega-3 only group). Furthermore, serum alanine aminotransferase decreased when mice were fed omega-3 alone ($P<0.001$) or in combination with UDCA ($P<0.001$). There was no difference in alanine aminotransferase levels between the omega-3 group and omega-3 with UDCA group ($P=0.999$, when compared with the omega-3 only group).

Omega-3-attenuated NASH-associated liver fibrosis and adding UDCA further enhanced its anti-fibrotic effect

Histological assessment of liver fibrosis was performed using an image analysis system after Sirius Red staining. Administering

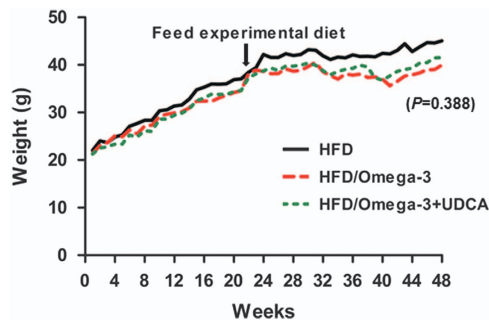


Figure 1 Changes in bodyweight during 48 weeks of the experiment. All the mice were fed a high fat diet (HFD) for 24 weeks and were then divided into three groups. One group ($n=8$) was fed an HFD only (HFD), another group ($n=8$) was fed an HFD and administered omega-3 (HFD/Omega-3) and the other group ($n=8$) was fed an HFD and administered omega-3 in combination with UDCA (HFD/Omega-3+UDCA) for another 24 weeks.

omega-3 at a daily dose of 70 mg kg^{-1} with an HFD-improved histologically evaluated liver fibrosis ($P<0.001$; Figures 2a and b). Administration of UDCA at a daily dose of 20 mg kg^{-1} in combination with omega-3 further attenuated HFD-induced liver fibrosis ($P=0.026$, compared with the omega-3 only group; Figure 2b).

The effect of omega-3 and UDCA on NASH-associated liver fibrosis was also investigated by quantitative analyses of *collagen $\alpha 1(I)$* and *TGF β* mRNA expression. Omega-3 alleviated HFD-induced collagen deposition ($P=0.002$) and administering UDCA in combination with omega-3 further decreased *Col $\alpha 1(I)$* expression ($P=0.021$, compared with the omega-3 only group; Figure 3a). In addition, administering omega-3 as a supplement to the HFD diminished the mRNA expression of the pro-fibrotic gene *TGF β* ($P=0.012$), and administering UDCA in combination with omega-3 further accentuated this effect ($P=0.029$, compared with the omega-3 only group; Figure 3a).

Omega-3 and UDCA-attenuated NASH-associated liver inflammation

Histological analysis of liver inflammation was carried out after H&E staining. Administering UDCA in combination with omega-3 ameliorated histologically assessed liver inflammation when compared with that of mice fed an HFD only ($P=0.036$; Figures 2a and c). In addition, when liver expression of *TNF α* , an inflammatory cytokine, was assessed, administering omega-3 attenuated the *TNF α* mRNA increase ($P<0.001$) and adding UDCA further decreased *TNF α* expression ($P=0.008$, compared with the omega-3 only group; Figure 3b).

Omega-3 and UDCA-attenuated hepatic TG accumulation

The effect of omega-3 and UDCA on HFD-induced hepatic TG accumulation was evaluated by measuring the tissue TG content. Administering omega-3 with the HFD resulted in decreased hepatic TG content compared with that of mice fed the HFD only ($P=0.029$). In addition, the combination of UDCA and omega-3 further attenuated hepatic TG accumulation ($P=0.021$; Figure 4).

Table 1 Characterization of mice fed an HFD and administered omega-3 with or without UDCA

Diet	HFD Vehicle ($n=8$)	HFD Omega-3 ($n=8$)	HFD Omega-3+UDCA ($n=8$)
Serum glucose (mg dl^{-1})	116.00 ± 28.00	120.25 ± 12.45	146.66 ± 12.13
Serum TG (mg dl^{-1})	124.00 ± 7.23	79.33 ± 5.24^a	78.33 ± 9.13^a
Serum Chol (mg dl^{-1})	158.00 ± 5.19	145.25 ± 9.07	153.00 ± 13.40
Serum ALT (IU l^{-1})	393.00 ± 26.57	88.25 ± 11.32^a	87.25 ± 6.67^a

Abbreviations: ALT, alanine aminotransferase; Chol, cholesterol; HFD, high fat diet; Omega-3, omega-3 polyunsaturated fatty acid; TG, triglyceride; UDCA, ursodeoxycholic acid.

Data are expressed as the mean \pm s.e.

^a $P<0.05$, when compared with mice fed HFD alone.

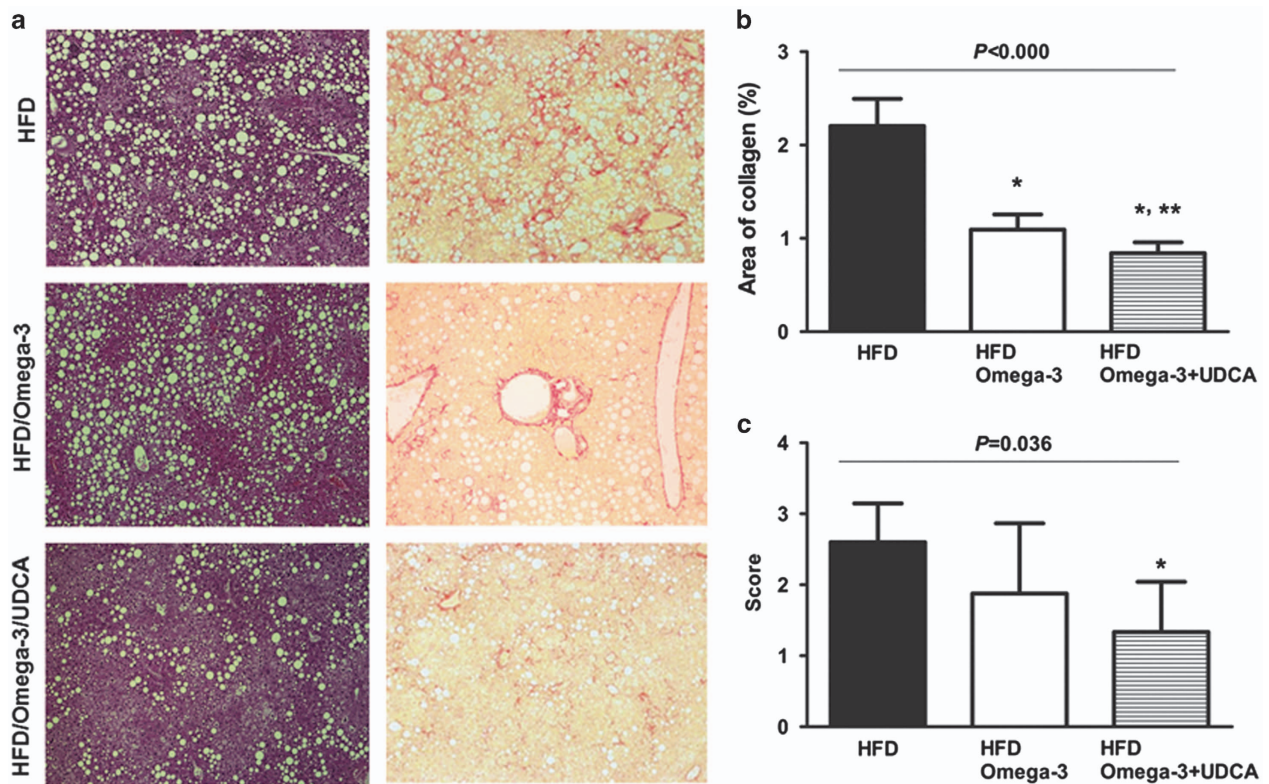


Figure 2 Histological assessment of fibrosis and inflammation in diet-induced NASH. Histological analysis of the liver to evaluate inflammation and fibrosis was performed after H&E and Sirius Red staining. Mice were fed with a high fat diet (HFD) for 24 weeks and were then divided into three groups. One group ($n=8$) was fed an HFD only (HFD), another group ($n=8$) was fed an HFD and administered omega-3 (HFD/Omega-3) and the other group ($n=8$) was fed an HFD and administered omega-3 in combination with UDCA (HFD/Omega-3+UDCA) for 24 weeks. (a) Representative histological evaluation using H&E ($\times 100$) and Sirius Red ($\times 100$). (b) Percent region of fibrosis. Liver fibrosis was evaluated by calculating the fibrosis ratio using an image analysis system. The total area was the sum of the area of microscopic fields including parenchyma and fibrosis. (c) Inflammation was histologically quantified by counting inflammatory foci in 20 consecutive high-power fields ($\times 40$ objective). The averaged histological grades are expressed as the following; grade 0: no foci, grade 1: < 2 foci per high-power field, grade 2: ≥ 2 foci per high-power field. $*P < 0.05$ when compared with that of the HFD group. $**P < 0.05$ when compared with that of HFD/Omega-3 group.

Omega-3 suppressed the expression of lipogenesis-associated genes and adding UDCA augmented this effect

The mRNA expression of *SREBP-1c*, which is known to regulate lipogenic enzymes,²⁶ was decreased after omega-3 administration in HFD-induced NASH ($P=0.014$). Administering UDCA in combination with omega-3 further diminished *SREBP-1c* expression ($P=0.001$, when compared with the omega-3 only group; Figure 3c). The mRNA expression of *fatty acid synthase*, a gene involved in *de novo* lipogenesis, decreased when omega-3 was administered ($P=0.035$), but UDCA did not show an additive effect ($P=0.343$, when compared with the omega-3 only group). The mRNA expression of *stearoyl-carnitine palmitoyl-coenzyme A desaturase*, another gene involved in *de novo* lipogenesis, was also reduced when omega-3 was administered ($P=0.022$), but there was no significant difference when UDCA was administered in combination with omega-3 ($P=0.083$ when compared with the omega-3 only group). The mRNA expression of *carnitine palmitoyl-coenzyme A transferase*, a mitochondrial β -oxidation enzyme, decreased with omega-3 supplement ingestion ($P=0.045$), but adding UDCA did not

produce a further reduction in *carnitine palmitoyl-coenzyme A transferase* mRNA expression ($P=0.686$, when compared with the omega-3 only group).

The effect of UDCA on NASH was associated with decreased *iNOS* mRNA expression

The upregulation of *iNOS* is known to be associated with liver inflammation and tissue damage,^{27,28} and UDCA has been reported to reduce *iNOS* in several inflammatory diseases.^{29,30} We evaluated whether the beneficial effect of UDCA on NASH in our study was associated with decreased *iNOS* mRNA expression. Administration of omega-3 with the HFD resulted in diminished *iNOS* mRNA ($P=0.012$), and adding UDCA in combination with omega-3 further suppressed *iNOS* mRNA expression ($P=0.029$, when compared with the omega-3 only group; Figure 3d).

DISCUSSION

Although several previous studies have reported that omega-3 attenuates diet-induced NASH,^{31–33} to our knowledge, this

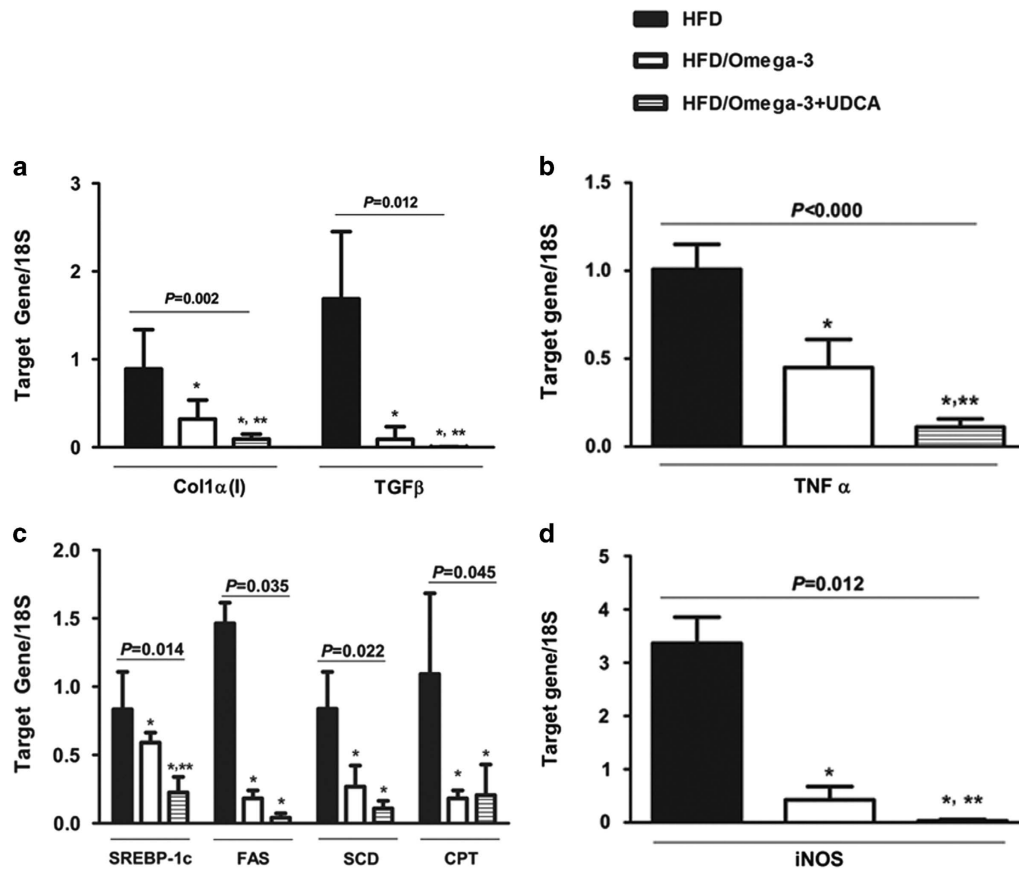


Figure 3 Changes in gene expression related to fibrosis, inflammation, steatosis and oxidative stress. Mice were fed a high fat diet (HFD) for 24 weeks and were then divided into three groups. One group ($n=8$) was fed an HFD only (HFD), another group ($n=8$) was fed an HFD and administered omega-3 (HFD/Omega-3) and the other group ($n=8$) was fed an HFD and administered omega-3 in combination with UDCA (HFD/Omega-3+UDCA) for 24 weeks. (a) Expression of fibrosis related genes in the liver. Quantitative analysis of liver *Col1α(I)* and *TGFβ* mRNA, markers of fibrosis, decreased when omega-3 was added to HFD. The HFD/Omega-3+UDCA group exhibited decreased *Col1α(I)* and *TGFβ* expression when compared with that of the HFD/Omega-3 group. (b) Expression of inflammation-related *TNFα* in the liver. Quantitative analysis of liver *TNFα* mRNA was decreased in the HFD/Omega-3 and HFD/Omega-3+UDCA groups when compared with the HFD group. The HFD/Omega-3+UDCA group had decreased *TNFα* expression when compared with that of the HFD/Omega-3 group. (c) Expression of lipogenic genes. mRNA expression of *SREBP-1c*, the lipogenic gene regulator, was decreased in the HFD/Omega-3 and HFD/Omega-3+UDCA groups when compared with the HFD group. The HFD/Omega-3+UDCA group had decreased *SREBP-1c* expression when compared with that of the HFD/Omega-3 group. mRNA expression of *fatty acid synthase* and *stearyl-carnitine palmitoyl-coenzyme A desaturase*, genes involved in *de novo* lipogenesis, were significantly decreased when omega-3 was administered. However, no additive effect was found with UDCA. *CPA*, a fatty acid oxidation related gene, was diminished by administering omega-3 with an HFD, but no further reduction was noticed by administering UDCA in combination with omega-3. (d) Expression of an oxidative stress-related gene. Decreased liver *iNOS* mRNA expression was observed with omega-3 as a supplement to HFD and adding UDCA further augmented this effect. * $P<0.05$ when compared with that of the HFD group. ** $P<0.05$ when compared with that of HFD/Omega-3 group.

is the first study to report an additive effect of omega-3 and UDCA on NASH. In accordance with other studies, our study also demonstrated that omega-3 alone could alleviate liver fibrosis identified by decreased *collagen α1(I)* mRNA expression. However, administering UDCA in combination with omega-3 further reduced fibrosis at the mRNA level and also further attenuated histologically assessed liver fibrosis when compared with the group to which omega-3 was administered alone. It has been repeatedly suggested that liver fibrosis is one of the most important factors in forecasting the progression of NASH,^{34–36} and an additive effect of omega-3

and UDCA in diminishing NASH-associated fibrosis makes this combination a possible candidate for the treatment of NASH.

TGFβ has been identified as a key regulator of liver fibrosis that exerts its profibrogenic effect through hepatic stellate cells.³⁷ The role of *TGFβ* signaling in the progression of NASH-associated fibrosis has also been reported in several studies, making it an attractive target for treatment.^{38,39} Our study demonstrated that omega-3 and UDCA had an additive effect in suppressing *TGFβ* mRNA expression, although *TGFβ* is not suggested as a target for this combination therapy, according to our results.

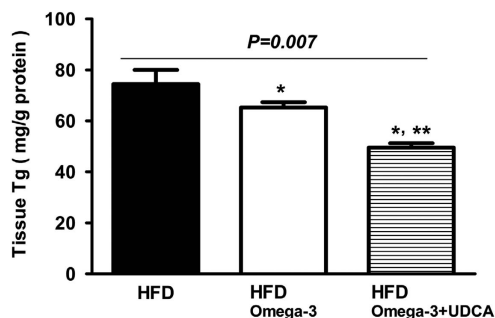


Figure 4 Hepatic triglyceride content. Hepatic triglyceride content was measured in frozen liver tissue using a commercial kit. The mice were fed a high fat diet (HFD) for 24 weeks and were then divided into three groups. One group ($n=8$) was fed an HFD only (HFD), another group ($n=8$) was fed an HFD and administered omega-3 (HFD/Omega-3) and the other group ($n=8$) was fed an HFD and administered omega-3 in combination with UDCA (HFD/Omega-3+UDCA) for another 24 weeks. * $P<0.05$ when compared with that of the HFD group. ** $P<0.05$ when compared with that of the HFD/Omega-3 group.

Even with the promising results of either omega-3 or UDCA on experimentally induced NASH, neither omega-3 nor UDCA has succeeded in showing a significant therapeutic effect when tested in NASH patients. Randomized controlled clinical trials investigating the effect of either omega-3 or UDCA on NASH in patients have failed to show significant improvement when compared with a placebo.^{17,18} However, a systemic review has suggested that negative results on the beneficial role of these agents might be attributed to improperly constructed study designs including suboptimal drug dosage and study duration.¹⁹ Meanwhile, several studies testing UDCA-based combination regimens, such as UDCA with vitamin E, resulted in biochemical or histological improvements in NASH patients.^{40,41} However, as described earlier, there has been a report showing the adverse effects of long-term vitamin E consumption,²⁰ and safety should be the utmost concern in medical treatments of NASH. No serious side-effects have been recorded with UDCA treatment, and a human study using 25 mg kg⁻¹ of UDCA, which was higher than the dose used in our study, has been carried out without reporting any serious adverse events.⁴² As for omega-3, the currently recommended maximal dose is 4000 mg per day, which is equivalent to the dose used in this study.

In addition to liver fibrosis, inflammation is also known to have a major role in the development and progression of NASH.^{23,43} Although administering omega-3 alone failed to show significant histologic changes, administering UDCA in combination with omega-3 resulted in histologically improved inflammation in diet-induced NASH, suggesting an additive effect. A previous study also demonstrated decreased expression of the proinflammatory cytokine TNF α after omega-3 supplementation,⁴⁴ which is in accordance with our study results.

TG accumulation in the liver is known to be controlled by SREBP-1c, a transcriptional factor that regulates lipogenic

enzymes.^{45,46} Several studies have reported that omega-3 supplementation downregulated *SREBP-1c* gene expression,⁶⁻⁹ and a recent study demonstrated that UDCA mediated lipogenesis by repressing *SREBP-1c* expression.⁴⁷ Our study suggested an additive effect of omega-3 and UDCA for suppressing *SREBP-1c*, although this study could not reveal an additive effect of UDCA for regulating genes related to *de novo* lipogenesis or fatty acid oxidation.

Although our study does not delineate the mechanistic role of omega-3 and UDCA-combined treatment for improving NASH, suppressed *iNOS* expression suggests that reduced liver damage from oxidative stress may be a possible explanation. Increased *iNOS* production has been demonstrated in fructose-induced hepatic steatosis,⁴⁸ and suppressed *iNOS* production has been associated with the use of anti-inflammatory drugs.⁴⁹ Studies investigating more detailed mechanisms are needed in the future.

In conclusion, our study demonstrated an additive effect of omega-3 and UDCA for alleviating fibrosis and inflammation in diet-induced NASH. Our study also demonstrated the additive effect of omega-3 and UDCA for suppressing the lipogenic gene *SREBP-1c* and the oxidative stress related gene *iNOS*. These results suggest that the combination of omega-3 and UDCA may be a possible treatment for NASH, although validation using different NASH models and human studies are necessary.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

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- Angulo P. GI Epidemiology: nonalcoholic fatty liver disease. *Aliment Pharmacol Ther* 2007; **25**: 883-889.
- Argo CK, Caldwell SH. Epidemiology and Natural History of Non-Alcoholic Steatohepatitis. *Clin Liver Dis* 2009; **13**: 511-531.
- Angulo P. Long-term mortality in nonalcoholic fatty liver disease: is liver histology of any prognostic significance? *Hepatology* 2010; **51**: 373-375.
- Bhala N, Angulo P, van der Poorten D, Lee E, Hui JM, Saracco G *et al*. The natural history of nonalcoholic fatty liver disease with advanced fibrosis or cirrhosis: an International Collaborative Study. *Hepatology* 2011; **54**: 1208-1216.
- Yokoyama M, Origasa H, Matsuzaki M, Matsuzawa Y, Saito Y, Ishikawa Y *et al*. Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): a randomised openlabel, blinded endpoint analysis. *Lancet* 2007; **369**: 1090-1098.
- Kim HJ, Takahashi M, Ezaki O. Fish oil feeding decreases mature sterol regulatory element-binding protein 1 (SREBP-1) by down-regulation of SREBP-1c mRNA in mouse liver. A possible mechanism for down-regulation of lipogenic enzyme mRNAs. *J Biol Chem* 1999; **274**: 25892-25898.
- Yahagi N, Shimano H, Hasty AH, Amemiya-Kudo M, Okazaki H, Tamura Y *et al*. A crucial role of sterol regulatory element-binding protein-1 in the regulation of lipogenic gene expression by polyunsaturated fatty acids. *J Biol Chem* 1999; **274**: 35840-35844.
- Xu J, Nakamura MT, Cho HP, Clarke SD. Sterol regulatory element binding protein-1 expression is suppressed by dietary polyunsaturated fatty acids. A mechanism for the coordinate suppression of lipogenic genes by polyunsaturated fats. *J Biol Chem* 1999; **274**: 23577-23583.

- 9 Botolin D, Wang Y, Christian B, Jump DB. Docosahexaenoic acid (22:6, n-3) regulates rat hepatocyte SREBP-1 nuclear abundance by Erk- and 26S proteasome-dependent pathways. *J Lipid Res* 2006; **47**: 181–192.
- 10 Zhu FS, Liu S, Chen XM, Huang ZG, Zhang DW. Effects of n-3 polyunsaturated fatty acids from seal oils on nonalcoholic fatty liver disease associated with hyperlipidemia. *World J Gastroenterol* 2008; **14**: 6395–6400.
- 11 Spadaro L, Magliocco O, Spampinato D, Piro S, Oliveri C, Alagona C *et al*. Effects of n-3 polyunsaturated fatty acids in subjects with nonalcoholic fatty liver disease. *Dig Liver Dis* 2008; **40**: 194–199.
- 12 Parker HM, Johnson NA, Burdon CA, Cohn JS, O'Connor HT, George J. Omega-3 supplementation and non-alcoholic fatty liver disease: a systematic review and meta-analysis. *J Hepatol* 2012; **56**: 944–951.
- 13 Chamulitrat W, Burhenne J, Rehlen T, Pathil A, Stremmel W. Bile salt-phospholipid conjugate ursodeoxycholy lysophosphatidylethanolamide as a hepatoprotective agent. *Hepatology* 2009; **50**: 143–154.
- 14 Pathil A, Warth A, Chamulitrat W, Stremmel W. The synthetic bile acid-phospholipid conjugate ursodeoxycholy lysophosphatidylethanolamide suppresses TNF α -induced liver injury. *J Hepatol* 2011; **54**: 674–684.
- 15 Mitsuyoshi H, Nakashima T, Sumida Y, Yoh T, Nakajima Y, Ishikawa H *et al*. Ursodeoxycholic acid protects hepatocytes against oxidative injury via induction of antioxidants. *Biochem Biophys Res Commun* 1999; **263**: 537–542.
- 16 Uzun MA, Koksal N, Aktas S, Gunerhan Y, Kadioglu H, Dursun N *et al*. The effect of ursodeoxycholic acid on liver regeneration after partial hepatectomy in rats with non-alcoholic fatty liver disease. *Hepatol Res* 2009; **39**: 814–821.
- 17 Leuschner UFH, Lindenthal B, Herrmann G, Arnold JC, Rossle M, Cordes HJ *et al*. High-dose ursodeoxycholic acid therapy for nonalcoholic steatohepatitis: a double-blind, randomized, placebo-controlled trial. *Hepatology* 2010; **52**: 472–479.
- 18 Dasarathy S, Dasarathy J, Khiyami A, Yerian LM, Sargent R, Hawkins CA *et al*. Randomized controlled trial of omega 3 fatty acids in the treatment of non alcoholic steatohepatitis in type 2 diabetes mellitus. *Hepatology* 2013; **58**: 518a–518a.
- 19 Xiang Z, Chen YP, Ma KF, Ye YF, Zheng L, Yang YD *et al*. The role of ursodeoxycholic acid in non-alcoholic steatohepatitis: a systematic review. *BMC Gastroenterol* 2013; **13**: 140.
- 20 Miller ER, Pastor-Barriuso R, Dalal D, Riemersma RA, Appel LJ, Guallar E. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Ann Intern Med* 2005; **142**: 37–46.
- 21 Brunt EM. Nonalcoholic steatohepatitis: definition and pathology. *Semin Liver Dis* 2001; **21**: 3–16.
- 22 Kirsch R, Clarkson V, Shephard EG, Marais DA, Jaffer MA, Woodburne VE *et al*. Rodent nutritional model of non-alcoholic steatohepatitis: species, strain and sex difference studies. *J Gastroenterol Hepatol* 2003; **18**: 1272–1282.
- 23 Kleiner DE, Brunt EM, Van Natta M, Behling C, Contos MJ, Cummings OW *et al*. Design and validation of a histological scoring system for nonalcoholic fatty liver disease. *Hepatology* 2005; **41**: 1313–1321.
- 24 O'Brien MJ, Keating NM, Elderiny S, Cerda S, Keaveny AP, Afhdal NH *et al*. An assessment of digital image analysis to measure fibrosis in liver biopsy specimens of patients with chronic hepatitis C. *Am J Clin Pathol* 2000; **114**: 712–718.
- 25 Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(T) $-(\Delta\Delta C)$ method. *Methods* 2001; **25**: 402–408.
- 26 Nagaya T, Tanaka N, Suzuki T, Sano K, Horiuchi A, Komatsu M *et al*. Down-regulation of SREBP-1c is associated with the development of burned-out NASH. *J Hepatol* 2010; **53**: 724–731.
- 27 Droge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002; **82**: 47–95.
- 28 Brenner C, Galluzzi L, Kepp O, Kroemer G. Decoding cell death signals in liver inflammation. *J Hepatol* 2013; **59**: 583–594.
- 29 Carotti S, Guarino MP, Cicala M, Perrone G, Alloni R, Segreto F *et al*. Effect of ursodeoxycholic acid on inflammatory infiltrate in gallbladder muscle of cholesterol gallstone patients. *Neurogastroenterol Motil* 2010; **22**: e232.
- 30 Moon SJ, Jeong JH, Jhun JY, Yang EJ, Min JK, Choi JY *et al*. Ursodeoxycholic acid ameliorates pain severity and cartilage degeneration in monosodium iodoacetate-induced osteoarthritis in rats. *Immune Netw* 2014; **14**: 45–53.
- 31 Ishii H, Horie Y, Ohshima S, Anezaki Y, Kinoshita N, Dohmen T *et al*. Eicosapentaenoic acid ameliorates steatohepatitis and hepatocellular carcinoma in hepatocyte-specific Pten-deficient mice. *J Hepatol* 2009; **50**: 562–571.
- 32 Kajikawa S, Imada K, Takeuchi T, Shimizu Y, Kawashima A, Harada T *et al*. Eicosapentaenoic acid attenuates progression of hepatic fibrosis with inhibition of reactive oxygen species production in rats fed methionine- and choline-deficient diet. *Dig Dis Sci* 2011; **56**: 1065–1074.
- 33 Depner CM, Philbrick KA, Jump DB. Docosahexaenoic acid attenuates hepatic inflammation, oxidative stress, and fibrosis without decreasing hepatosteatosis in a Ldlr(-/-) mouse model of western diet-induced nonalcoholic steatohepatitis. *J Nutr* 2013; **143**: 315–323.
- 34 Harrison SA, Torgerson S, Hayashi PH. The natural history of nonalcoholic fatty liver disease: a clinical histopathological study. *Am J Gastroenterol* 2003; **98**: 2042–2047.
- 35 Fassio E, Alvarez E, Dominguez N, Landeira G, Longo C. Natural history of nonalcoholic steatohepatitis: a longitudinal study of repeat liver biopsies. *Hepatology* 2004; **40**: 820–826.
- 36 Adams LA, Sanderson S, Lindor KD, Angulo P. The histological course of nonalcoholic fatty liver disease: a longitudinal study of 103 patients with sequential liver biopsies. *J Hepatol* 2005; **42**: 132–138.
- 37 Friedman SL. Mechanisms of hepatic fibrogenesis. *Gastroenterology* 2008; **134**: 1655–1669.
- 38 Cayon A, Crespo J, Mayorga M, Guerra A, Pons-Romero F. Increased expression of Ob-Rb and its relationship with the overexpression of TGF- β 1 and the stage of fibrosis in patients with nonalcoholic steatohepatitis. *Liver Int* 2006; **26**: 1065–1071.
- 39 Klein S, Mittendorfer B, Eagon JC, Patterson B, Grant L, Feirt N *et al*. Gastric bypass surgery improves metabolic and hepatic abnormalities associated with nonalcoholic fatty liver disease. *Gastroenterology* 2006; **130**: 1564–1572.
- 40 Dufour JF, Oneta CM, Gonvers JJ, Bihl F, Cerny A, Cereda JM *et al*. Randomized placebo-controlled trial of ursodeoxycholic acid with vitamin E in nonalcoholic steatohepatitis. *Clin Gastroenterol Hepatol* 2006; **4**: 1537–1543.
- 41 Pietu F, Guillaud O, Walter T, Vallin M, Hervieu V, Scazec JY *et al*. Ursodeoxycholic acid with vitamin E in patients with nonalcoholic steatohepatitis: long-term results. *Clin Res Hepatol Gastroenterol* 2012; **36**: 146–155.
- 42 Azzaroli F, Raspani ME, Simoni P, Montagnani M, Lisotti A, Cecinato P *et al*. High doses of ursodeoxycholic acid up-regulate the expression of placental breast cancer resistance protein in patients affected by intrahepatic cholestasis of pregnancy. *PLoS ONE* 2013; **8**: e64101.
- 43 Tilg H, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010; **52**: 1836–1846.
- 44 Stienstra R, Mandard S, Patsouris D, Maass C, Kersten S, Muller M. Peroxisome proliferator-activated receptor alpha protects against obesity-induced hepatic inflammation. *Endocrinology* 2007; **148**: 2753–2763.
- 45 Browning JD, Horton JD. Molecular mediators of hepatic steatosis and liver injury. *J Clin Invest* 2004; **114**: 147–152.
- 46 Ferre P, Foufelle F. SREBP-1c transcription factor and lipid homeostasis: clinical perspective. *Horm Res* 2007; **68**: 72–82.
- 47 Lee JM, Gang GT, Kim DK, Kim YD, Koo SH, Lee CH *et al*. Ursodeoxycholic acid inhibits liver X receptor alpha-mediated hepatic lipogenesis via induction of the nuclear corepressor SMILE. *J Biol Chem* 2014; **289**: 1079–1091.
- 48 Spruss A, Kanuri G, Wagnerberger S, Haub S, Bischoff SC, Bergheim I. Toll-Like receptor 4 is involved in the development of fructose-induced hepatic steatosis in mice. *Hepatology* 2009; **50**: 1094–1104.
- 49 Moeslinger T, Friedl R, Spieckermann PG. Inhibition of inducible nitric oxide synthesis by azathioprine in a macrophage cell line. *Life Sci* 2006; **79**: 374–381.



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