



CrossMark
click for updates

Review

Cite this article: Blaschuk OW. 2015

N-cadherin antagonists as oncology
therapeutics. *Phil. Trans. R. Soc. B* **370**:
20140039.

<http://dx.doi.org/10.1098/rstb.2014.0039>

One contribution of 19 to a discussion meeting
issue 'Cell adhesion century: culture
breakthrough'.

Subject Areas:

cellular biology

Keywords:

N-cadherin antagonists, ADH-1, endothelial
cells, angiogenesis, cancer

Author for correspondence:

Orest W. Blaschuk

e-mail: orest.blaschuk@mcgill.ca

N-cadherin antagonists as oncology therapeutics

Orest W. Blaschuk

Division of Urology, Department of Surgery, McGill University, Montreal, Quebec, Canada H3A 1A1

The cell adhesion molecule (CAM), N-cadherin, has emerged as an important oncology therapeutic target. N-cadherin is a transmembrane glycoprotein mediating the formation and structural integrity of blood vessels. Its expression has also been documented in numerous types of poorly differentiated tumours. This CAM is involved in regulating the proliferation, survival, invasiveness and metastasis of cancer cells. Disruption of N-cadherin homophilic intercellular interactions using peptide or small molecule antagonists is a promising novel strategy for anti-cancer therapies. This review discusses: the discovery of N-cadherin, the mechanism by which N-cadherin promotes cell adhesion, the role of N-cadherin in blood vessel formation and maintenance, participation of N-cadherin in cancer progression, the different types of N-cadherin antagonists and the use of N-cadherin antagonists as anti-cancer drugs.

1. Introductory comments concerning N-cadherin and calcium-dependent cell adhesion

Neural (N)-cadherin is an integral membrane, calcium-binding glycoprotein that mediates the intercellular adhesion of neuronal cells and many other types of non-neuronal cells [1–3]. The extracellular domain of N-cadherin is composed of five subdomains (EC1–EC5; figure 1) [1,2]. Three calcium ions bind between each successive duo of subdomains (i.e. EC1–EC2, EC2–EC3, EC3–EC4 and EC4–EC5). The binding of calcium ions stabilizes the elongated, curved structure of N-cadherin thus enabling the proper function of this cell adhesion molecule (CAM) [2].

Cells comprising vertebrate tissues have been shown to possess calcium-dependent and calcium-independent adhesion mechanisms [4,5]. The calcium-dependent adhesion mechanism is resistant to proteolysis in the presence of calcium, whereas the calcium-independent adhesion mechanism is not. Treatment of cells with proteases (e.g. trypsin) in the presence of ethylene glycol tetraacetic acid (EGTA, which results in the removal of calcium from the medium) abolishes calcium-dependent adhesion.

N-cadherin was identified in the 1980s by two groups: Volk & Geiger [6–8] and Hatta & Takeichi [9]. Volk and Geiger were seeking to elucidate the CAMs present in a particular intercellular junction, known as the adherens junction. They prepared an intercalated disc-enriched membrane fraction containing adherens junctions from chicken cardiac muscle and then extracted it with detergent [6]. This detergent extract was used to immunize mice and generate hybridomas. One of the hybridomas was found to secrete a monoclonal antibody (designated ID-7.2.3) that recognized a membrane glycoprotein component of adherens junctions present in intercalated discs, as judged by immunoelectron microscopy [6,7]. Volk and Geiger named this glycoprotein adherens junction-specific CAM (A-CAM) [7]. They proceeded to demonstrate that A-CAM was degraded by trypsin in the presence of EGTA. It is noteworthy that the formation of adherens junctions does not occur in the absence of calcium. Finally, monovalent Fab fragments of ID-7.2.3 were shown to be capable of blocking the formation of adherens junctions by cultured cells [8]. Collectively, these observations provided strong evidence that A-CAM was directly involved in mediating the calcium-dependent formation of adherens junctions. It was later shown that A-CAM was identical to N-cadherin [10].

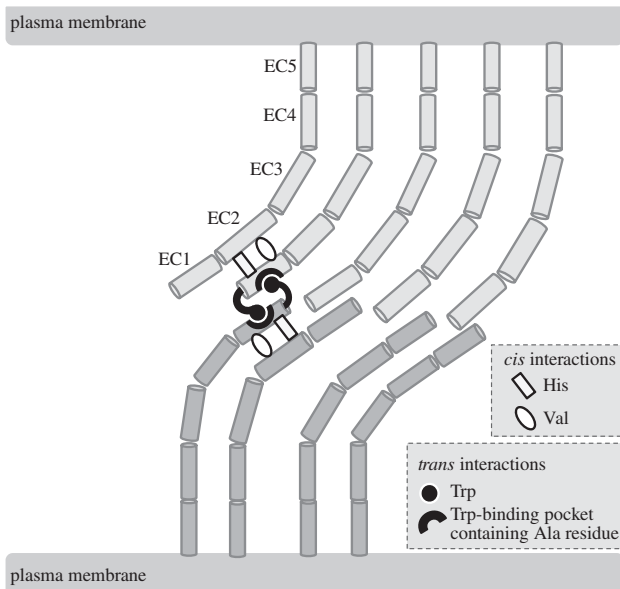


Figure 1. Diagram showing the extracellular domains of N-cadherin monomers simultaneously interacting within the plane of the cell's plasma membrane (referred to as *cis* interactions) and at adhesive contacts between apposed cells (referred to as *trans* interactions). His and Val residues within the first extracellular (EC1) subdomain of N-cadherin monomers mediate *cis* interactions with the second extracellular (EC2) subdomain of adjacent N-cadherin monomers. The Trp and Ala residues within the EC1 subdomain of N-cadherin monomers directly mediate *trans* interactions. The Trp residue at the N-terminus of each N-cadherin monomer inserts into an apposed hydrophobic pocket containing an Ala residue. See text for further details.

Hatta & Takeichi [9] speculated that surface proteins present on cells after treatment with trypsin in the presence of calcium were candidate CAMs responsible for mediating calcium-dependent adhesion. They immunized rats with cells obtained from the neural retina of 6-day-old chick embryos dissociated by trypsin in the presence of calcium and subsequently generated hybridomas from the splenocytes of these animals. The monoclonal antibodies secreted by the hybridomas were then screened against western blots containing protein extracts of either cells treated with trypsin in the presence of calcium or cells treated with trypsin in the presence of EGTA. A hybridoma was identified that secreted a monoclonal antibody (designated NCD-2) which recognized an antigen (molecular mass 127 kDa) on western blots prepared from the protein extracts of cells treated with trypsin in the presence of calcium, but not from cells treated with trypsin in the presence of EGTA. NCD-2 reacted with the surface of cells dissociated from neural retina of 6-day-old chick embryos by trypsin in the presence of calcium. Furthermore, NCD-2 blocked the aggregation of these cells. Finally, NCD-2 recognized the antigen in all neuronal tissues examined, as well as the heart, as judged by probing western blots of protein extracts from these tissues with this monoclonal antibody. The antigen was not detected in skin, liver, kidney and lung protein extracts. This distribution was similar to that observed in mouse embryos [11]. Based on these observations, the antigen was named N-cadherin.

The nucleotide sequence of cDNA encoding chicken N-cadherin was subsequently determined and the amino acid sequence was deduced from this information [1]. The data suggested that N-cadherin had extracellular, transmembrane and cytoplasmic domains. An expression vector containing N-cadherin cDNA was transfected into mouse L cells that do

not express this CAM or display calcium-dependent cell adhesion. The transfected cells exhibited calcium-dependent aggregation in suspension. NCD-2 prevented the aggregation of these cells. These observations substantiate the contention that N-cadherin is a calcium-dependent CAM.

2. Mechanism by which N-cadherin promotes cell adhesion

N-cadherin monomers exist in many different states within the plane of the cell's plasma membrane and at intercellular adhesive contacts, where they function as homophilic CAMs [2,12,13]. In order to form stable intercellular adhesive junctions, the monomers apparently must interact within the plane of the plasma membrane (referred to as *cis* interactions) and also with monomers on the surface of apposing cells (referred to as *trans* interactions) (figure 1). These *cis* and *trans* interactions are centred around the cell adhesion recognition (CAR) sequence Histidine–Alanine–Valine (His79–Ala80–Val81) which is found towards the end of the first extracellular (EC1) subdomain of N-cadherin [1,2,13,14]. This sequence was predicted to be directly involved in mediating cell adhesion over two decades ago [14,15]. Recent studies suggest that the His79 and Val81 residues promote *cis* interactions, whereas the Ala80 residue facilitates *trans* interactions [2,12,13]. The Ala80 residue is a constituent of a hydrophobic pocket into which docks a tryptophan (Trp2) residue at the N-terminus of an apposed N-cadherin EC1 domain, thus promoting intercellular adhesion (figure 1).

3. Role of N-cadherin in the vasculature

N-cadherin is intimately involved in the formation of blood vessels (a process known as angiogenesis) and the maintenance of their integrity [3,16–18]. Blood microvessels are composed of endothelial and mural cells (pericytes). One of the fundamental functions of pericytes is to stabilize mature microvessels [17,18]. N-cadherin is found at adhesive complexes between endothelial cells and the pericytes that ensheath them [3,16–18]. Inhibition of N-cadherin function destabilizes microvessels [16,18]. For example, antibodies directed against N-cadherin disrupt endothelial cell–pericyte adhesive complexes and cause microvessels to haemorrhage [16]. These observations underline the importance of N-cadherin-mediated endothelial cell–pericyte adhesive interactions to microvessel stability.

4. Role of N-cadherin in tumour growth and progression

Increased tumour growth is dependent on an adequate blood supply [19,20]. Angiogenesis therefore plays a central role in enabling tumour growth. As discussed in §3, N-cadherin is involved in angiogenesis and the maintenance of blood vessel stability. Tumour growth is consequently dependent on N-cadherin. Antagonists of this CAM could conceivably affect tumour growth.

N-cadherin is also expressed by many tumour types: poorly differentiated carcinomas [21], melanoma [22], neuroblastoma [23] and multiple myeloma [6].

Normal epithelial cells express epithelial (E)-cadherin, a CAM closely related to N-cadherin [3,21,24]. Poorly differentiated carcinoma cells cease to express E-cadherin and instead display N-cadherin. Numerous studies have shown that loss of E-cadherin and the aberrant expression of N-cadherin causes tumour cells to lose their polarity, resist apoptosis, become invasive and metastatic. N-cadherin antagonists therefore have the potential of causing tumour cell apoptosis and suppressing metastasis. The ability of N-cadherin to regulate the behaviour of tumour cells has been attributed in part to the interaction with, and activation of, fibroblast growth factor receptor [24–26]. *In vivo*, further complexities are apparent in the molecular mechanisms underlying the ability of N-cadherin to influence cancer cell metastasis which remain to be fully elucidated. In particular, the N-cadherin-mediated interactions between tumour cells and either endothelial or stromal cells during the metastatic process have not been thoroughly investigated. Studies are also needed to determine the role of N-cadherin in the establishment of metastases at secondary sites [27].

5. N-cadherin antagonists

Several types of N-cadherin antagonists have been discovered. Three types of antagonists are based on the CAR sequence His–Ala–Val (HAV): synthetic linear peptides, synthetic cyclic peptides and non-peptidyl peptidomimetics [28].

Synthetic linear peptides containing the HAV motif were the first peptides shown to be capable of inhibiting N-cadherin-dependent processes [14,29–31]. For example, the decapeptide N-Ac-LRAHAVDING-NH₂, whose sequence is identical to that found in the EC1 subdomain of human N-cadherin, was shown to block neurite outgrowth [29], myoblast fusion [30] and Schwann cell migration on astrocytes [31].

Synthetic cyclic peptides harbouring the HAV motif were subsequently shown to act as N-cadherin antagonists [32]. The most studied cyclic peptide is N-Ac-CHAVC-NH₂ (designated ADH-1) [3]. This cyclic peptide, like the linear peptide, is capable of disrupting a wide variety of N-cadherin-mediated processes. Importantly, ADH-1 has been shown to inhibit angiogenesis [33]. In addition, studies have demonstrated that ADH-1 can cause apoptosis of multiple myeloma [34], neuroblastoma [23] and pancreatic [35] cancer cells. Not all tumour cells displaying N-cadherin undergo apoptosis when exposed to ADH-1. For example, this cyclic peptide does not cause apoptosis in various melanoma cell lines that express N-cadherin [36]. Instead, ADH-1 appears to sensitize these cells to the cytotoxic agent melphalan. The mechanism by which ADH-1 increases the cytotoxic effects of melphalan remains to be elucidated. ADH-1 has been shown to inhibit pancreatic and melanoma tumour growth in animal models when used alone [35], or in combination with cytotoxic agents [36], respectively.

Non-peptidyl peptidomimetics of ADH-1 have also been identified [3,37]. Unlike peptides which are rapidly degraded by gastrointestinal enzymes, such small molecule inhibitors

might be suitable for oral administration. The biological properties of these peptidomimetics remain to be determined.

A different type of N-cadherin antagonist has recently been discovered [38]. This type of antagonist is a synthetic linear peptide that harbours a Trp residue in the second position from the N-terminus (similar to N-cadherin). The peptide H-SWTLYTPSGQSK-NH₂ inhibits endothelial cell tube formation *in vitro* indicating that it has anti-angiogenic properties [38]. The biological properties of this peptide have not been extensively investigated.

The final type of N-cadherin antagonist to be developed is monoclonal antibodies. Two such monoclonal antibodies directed against the N-cadherin extracellular domain are capable of inhibiting the invasiveness and proliferation of N-cadherin expressing PC3 human prostate carcinoma cells *in vitro* [39]. These antibodies also suppress PC3 tumour growth and lymph node metastases *in vivo*.

Collectively, these observations indicate the potential of N-cadherin antagonists for serving as inhibitors of tumour growth and metastasis.

6. N-cadherin antagonists as anti-cancer drugs

The use of N-cadherin antagonists in the clinic as oncology therapeutics is at an early stage of investigation. Only a few clinical trials have been conducted with ADH-1 and there have been no trials with other N-cadherin antagonists. ADH-1 was not toxic at the doses tested in animals [36] and humans [40] indicating that this N-cadherin antagonist does not affect the normal vasculature. There have been indications that ADH-1 might be useful in treating ovarian cancer [40]. In addition, clinical trials testing ADH-1 in combination with melphalan have shown promise in the treatment of melanoma [41].

7. Summary and future directions

The CAM N-cadherin is capable of regulating the proliferation, survival, invasiveness and metastasis of certain tumour cell types, as well as blood vessel formation and stability. Consequently, N-cadherin antagonists can potentially affect many aspects of tumour growth and progression.

The N-cadherin antagonist ADH-1 has been shown to be an anti-angiogenic agent and can cause tumour cell apoptosis. Limited clinical trials with ADH-1 have suggested that it may be useful as an anti-cancer drug. Clearly, more studies are needed to test this hypothesis. It also remains to be determined which tumour types are most responsive to ADH-1.

The identification of orally available, small molecule N-cadherin antagonists is encouraging. Studies are needed to determine the effects of these antagonists on tumours and their vasculature.

References

- Hatta K, Nose A, Nagafuchi A, Takeichi M. 1988 Cloning and expression of cDNA encoding a neural calcium-dependent cell adhesion molecule: its identity in the cadherin gene family. *J. Cell Biol.* **106**, 873–881. (doi:10.1083/jcb.106.3.873)
- Harrison OJ *et al.* 2011 The extracellular architecture of adherens junctions revealed by crystal structures of type I cadherins. *Structure* **19**, 244–256. (doi:10.1016/j.str.2010.11.016)
- Blaschuk OW, Devemy E. 2009 Cadherins as novel targets for anti-cancer therapy.

- Eur. J. Pharmacol.* **625**, 195–198. (doi:10.1016/j.ejphar.2009.05.033)
4. Takeichi M. 1988 The cadherins: cell–cell adhesion molecules controlling animal morphogenesis. *Development* **102**, 639–655.
 5. Magnani JL, Thomas WA, Steinberg MS. 1981 Two distinct adhesion mechanisms in embryonic neural retina cells. I. A kinetic analysis. *Dev. Biol.* **81**, 96–105. (doi:10.0012-1606(81)90351-1)
 6. Volk T, Geiger B. 1984 A 135-kd membrane protein of intercellular adherens junctions. *EMBO J.* **3**, 2249–2260.
 7. Volk T, Geiger B. 1986 A-CAM: a 135-kD receptor of intercellular adherens junctions. I. Immunoelectron microscopic localization and biochemical studies. *J. Cell Biol.* **103**, 1441–1450. (doi:10.1083/jcb.103.4.1441)
 8. Volk T, Geiger B. 1986 A-CAM: a 135-kD receptor of intercellular adherens junctions. II. Antibody-mediated modulation of junction formation. *J. Cell Biol.* **103**, 1451–1464. (doi:10.1083/jcb.103.4.1451)
 9. Hatta K, Takeichi M. 1986 Expression of N-cadherin adhesion molecules associated with early morphogenetic events in chick development. *Nature* **320**, 447–449. (doi:10.1038/320447a0)
 10. Volk T, Volberg T, Sabanay I, Geiger B. 1990 Cleavage of A-CAM by endogenous proteinases in cultured lens cells and in developing chick embryos. *Dev. Biol.* **139**, 314–326. (doi:10.1016/0012-1606(90)90301-X)
 11. Hatta K, Okada TS, Takeichi M. 1985 A monoclonal antibody disrupting calcium-dependent cell-cell adhesion of brain tissues: possible role of its target antigen in animal pattern formation. *Proc. Natl Acad. Sci. USA* **82**, 2789–2793. (doi:10.1073/pnas.82.9.2789)
 12. Bunse S, Garg S, June K, Vogel D, Ansari N, Stelzer EH, Schuman E. 2013 Role of N-cadherin *cis* and *trans* interfaces in the dynamics of adherens junctions in living cells. *PLoS ONE* **8**, e81517. (doi:10.1371/journal.pone.0081517)
 13. Brasch J, Harrison OJ, Honig B, Shapiro L. 2012 Thinking outside the cell: how cadherins drive adhesion. *Trends Cell Biol.* **22**, 299–310. (doi:10.1016/j.tcb.2012.03.004)
 14. Blaschuk OW, Sullivan R, David S, Pouliot Y. 1990 Identification of a cadherin cell adhesion recognition sequence. *Dev. Biol.* **139**, 227–229. (doi:10.1016/0012-1606(90)90290-Y)
 15. Blaschuk OW, Pouliot Y, Holland PC. 1990 Identification of a conserved region common to cadherins and influenza strain A hemagglutinins. *J. Mol. Biol.* **211**, 679–682. (doi:10.1016/0022-2836(90)90065-T)
 16. Gerhardt H, Wolburg H, Redies C. 2000 N-cadherin mediates pericyte–endothelial interaction during brain angiogenesis in the chicken. *Dev. Dyn.* **218**, 472–479. (doi:10.1002/1097-0177(200007)218:3<472::AID-DVDY1008>3.0.CO;2-#)
 17. Gaengel K, Genové G, Armulik A, Betsholtz C. 2009 Endothelial-mural cell signalling in vascular development and angiogenesis. *Arterioscler. Thromb. Vasc. Biol.* **29**, 630–638. (doi:10.1161/ATVBAHA.107.161521)
 18. Paik J-H, Skoura A, Chae S-S, Cowan AE, Han DK, Proia RL, Hla T. 2004 Sphingosine 1-phosphate receptor regulation of N-cadherin mediates vascular stabilization. *Genes Dev.* **18**, 2392–2403. (doi:10.1101/gad.1227804)
 19. Folkman J. 2007 Angiogenesis: an organizing principle for drug discovery? *Nat. Rev. Drug Discov.* **6**, 273–286. (doi:10.1038/nrd2115)
 20. Naumov GN, Folkman J, Straume O, Akslen LA. 2008 Tumor-vascular interactions and tumor dormancy. *APMIS* **116**, 569–585. (doi:10.1111/j.1600-0463.2008.01213.x)
 21. Bex G, van Roy F. 2009 Involvement of members of the cadherin superfamily in cancer. *Cold Spring Harbor Perspect. Biol.* **1**, a003129. (doi:10.1101/cshperspect.a003129)
 22. Beasley GM *et al.* 2011 Prospective multicenter phase II trial of systemic ADH-1 in combination with melphalan via isolated limb infusion in patients with advanced extremity melanoma. *J. Clin. Oncol.* **29**, 1210–1215. (doi:10.1200/JCO.2010.32.1224)
 23. Lammens T *et al.* 2012 N-Cadherin in neuroblastoma disease: expression and clinical significance. *PLoS ONE* **7**, e31206. (doi:10.1371/journal.pone.0031206)
 24. Wheelock MJ, Shintani Y, Maeda M, Fukumoto Y, Johnson KR. 2008 Cadherin switching. *J. Cell Sci.* **121**, 727–735. (doi:10.1242/jcs.000455)
 25. Suyama K, Shapiro I, Guttman M, Hazan RB. 2002 A signaling pathway leading to metastasis is controlled by N-cadherin and the FGF receptor. *Cancer Cell* **2**, 301–314. (doi:10.1016/S1535-6108(02)00150-2)
 26. Qian X *et al.* 2013 N-cadherin/FGFR promotes metastasis through epithelial-to-mesenchymal transition and stem/progenitor cell-like properties. *Oncogene* **33**, 3411–3421. (doi:10.1038/onc.2013.310)
 27. Grinberg-Rashi H *et al.* 2009 The expression of three genes in primary non-small cell lung cancer is associated with metastatic spread to the brain. *Clin. Cancer Res.* **15**, 1755–1761. (doi:10.1158/1078-0432.CCR-08-2124)
 28. Blaschuk OW. 2012 Discovery and development of N-cadherin antagonists. *Cell Tissue Res.* **348**, 309–313. (doi:10.1007/s00441-011-1320-5)
 29. Doherty P, Rowett LH, Moore SE, Mann SE, Walsh FS. 1991 Neurite outgrowth in response to transfected N-CAM and N-cadherin reveals fundamental differences in neuronal responsiveness to CAMs. *Neuron* **6**, 247–258. (doi:10.1016/0896-6273(91)90360-C)
 30. Mege RM, Goudou D, Diaz C, Nicolet M, Garcia L, Geraud G, Rieger F. 1992 N-cadherin and N-CAM in myoblast fusion: compared localisation and effect of blockade by peptides and antibodies. *J. Cell Sci.* **103**, 897–906.
 31. Wilby MJ, Muir EM, Fok-Seang J, Gour BJ, Blaschuk OW, Fawcett JW. 1999 N-cadherin inhibits Schwann cell migration on astrocytes. *Mol. Cell. Neurosci.* **14**, 66–84. (doi:10.1006/mcne.1999.0766)
 32. Blaschuk OW, Gour B. 2000 *Compounds and methods for modulating cell adhesion*. United States Patent Number 6,031,072.
 33. Blaschuk OW, Gour BJ, Farookhi R, Anmar A. 2003 *Compounds and methods for modulating endothelial cell adhesion*. United States Patent Number 6,610,821.
 34. Sadler NM, Harris BR, Metzger BA, Kirshner J. 2013 N-cadherin impedes proliferation of the multiple myeloma cancer stem cells. *Am. J. Blood Res.* **3**, 271–285.
 35. Shintani Y, Fukumoto Y, Chaika N, Grandgenett PM, Hollingsworth MA, Wheelock MJ, Johnson KR. 2008 ADH-1 suppresses N-cadherin-dependent pancreatic cancer progression. *Int. J. Cancer* **122**, 71–77. (doi:10.1002/ijc.23027)
 36. Augustine CK, Yoshimoto Y, Gupta M, Zipfel PA, Selim MA, Febbo P, Pendergast AM, Peters WP, Tyler DS. 2008 Targeting N-cadherin enhances antitumor activity of cytotoxic therapies in melanoma treatment. *Cancer Res.* **68**, 3777–3784. (doi:10.1158/0008-5472.CAN-07-5949)
 37. Gour BJ, Blaschuk OW, Ali A, Ni F, Chen Z, Michaud S, Wang S, Hu Z. 2007 *Peptidomimetic modulators of cell adhesion*. United States Patent Number 7,268,115.
 38. Devery E, Blaschuk OW. 2008 Identification of a novel N-cadherin antagonist. *Peptides* **29**, 1853–1861. (doi:10.1016/j.peptides.2008.06.025)
 39. Tanaka H *et al.* 2010 Monoclonal antibody targeting of N-cadherin inhibits prostate cancer growth, metastasis and castration resistance. *Nat. Med.* **16**, 1414–1420. (doi:10.1038/nm.2236)
 40. Perotti A *et al.* 2009 Clinical and pharmacological phase I evaluation of Exherin (ADH-1), a selective anti-N-cadherin peptide in patients with N-cadherin expressing solid tumours. *Ann. Oncol.* **20**, 741–745. (doi:10.1093/annonc/mdn695)
 41. Beasley GM *et al.* 2009 A phase 1 study of systemic ADH-1 in combination with melphalan via isolated limb infusion in patients with locally advanced in-transit malignant melanoma. *Cancer* **115**, 4766–4774. (doi:10.1002/cncr.24509)