

Recent Advances in *Toxoplasma gondii* Immunotherapeutics

Sherene Swee-Yin Lim^{1,*}, Rofina Yasmin Othman^{1,2}

¹Genetics and Molecular Biology Department, Institute of Biological Sciences, Faculty of Science, University of Malaya, 50603, Kuala Lumpur, Malaysia; ²Centre for Research in Biotechnology for Agriculture, University of Malaya, 50603 Kuala Lumpur, Malaysia

Abstract: Toxoplasmosis is an opportunistic infection caused by the protozoan parasite *Toxoplasma gondii*. *T. gondii* is widespread globally and causes severe diseases in individuals with impaired immune defences as well as congenitally infected infants. The high prevalence rate in some parts of the world such as South America and Africa, coupled with the current drug treatments that trigger hypersensitivity reactions, makes the development of immunotherapeutics intervention a highly important research priority. Immunotherapeutics strategies could either be a vaccine which would confer a pre-emptive immunity to infection, or passive immunization in cases of disease recrudescence or recurrent clinical diseases. As the severity of clinical manifestations is often greater in developing nations, the development of well-tolerated and safe immunotherapeutics becomes not only a scientific pursuit, but a humanitarian enterprise. In the last few years, much progress has been made in vaccine research with new antigens, novel adjuvants, and innovative vaccine delivery such as nanoparticles and antigen encapsulations. A literature search over the past 5 years showed that most experimental studies were focused on DNA vaccination at 52%, followed by protein vaccination which formed 36% of the studies, live attenuated vaccinations at 9%, and heterologous vaccination at 3%; while there were few on passive immunization. Recent progress in studies on vaccination, passive immunization, as well as insights gained from these immunotherapeutics is highlighted in this review.

Key words: *Toxoplasma gondii*, immunotherapeutics, vaccine, passive immunization

INTRODUCTION

Toxoplasmosis is a zoonotic disease caused by the obligate intracellular parasite *Toxoplasma gondii*. Toxoplasmosis can cause severe neurologic, ocular, and systemic diseases in neonates and individuals with weakened immune system [1]. In immunocompetent adults, *T. gondii* infection is usually asymptomatic and forms dormant tissue cysts containing bradyzoites at immune-privileged sites such as the brain and muscles [1]. When immunity is impaired, latent infection is then reactivated causing widespread tissue destruction and severe pathology, and is among the major causes of mortality in AIDS patients. Disease severity is exacerbated in underdeveloped countries with limited financial resources and infrastructure to acquire

and distribute the anti-retroviral drugs [2]. The sub-Saharan African continent is a dire example of this. A high rate of HIV infection combined with high seroprevalence of *T. gondii* co-infection would result in an estimated 2.5 to 10 million people in this region who are at risk of dying from toxoplasmic encephalitis [3]. Atypical strains, predominantly found in South and Central America, have caused debilitating diseases, especially in the form of recurrent ocular disease that can lead to blindness in otherwise healthy adults [4,5]. This is an especially critical problem in Brazil where the current risk of ocular toxoplasmosis is at a high rate of 18% [6]. Thus, this pathogen remains as a severe public health problem in developing countries.

The development of a vaccine against toxoplasmosis is highly important to arrest the spread of the disease because in theory, a single treatment confers life-long protective immunity. Significant strides have been made in the past 15 years in antigen isolation and characterization, gene-cloning, generation of parasite mutants, and immunological methods [7]. Vaccine studies demonstrating robust disease control are antigens that could elicit a protective Th1-biased immune re-

•Received 3 April 2014, revised 21 August 2014, accepted 22 August 2014.

*Corresponding author (sheranelim@um.edu.my)

© 2014, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

sponse and generation of long-lived cytotoxic CD8⁺ T cells producing IFN- γ [8]. Vaccinations have thus far proven effective at reducing disease mortality and parasite cyst burden in mice, but do not achieve complete protection. Thus, sterile immunity from the disease remains an elusive target.

As toxoplasmosis is an opportunistic infection causing illness in immunocompromised patients with reduced cellular immunity, therapeutic intervention to reduce pathogenesis is also needed. Current anti-parasitic drug treatments cause toxic hypersensitivity reactions and are teratogenic, presenting a need for safer and well-tolerated treatment options. As the major cause of toxoplasmosis mortality is due to reactivation of latent infection, while considerable morbidity can result from the tendency of congenital infections to relapse with retinal damage, studies on passive immunization to decrease parasitemia burden is necessary for limiting the disease pathology. The aim of this review is to focus on recent advances in both the active and passive immunization studies against toxoplasmosis in murine models.

VACCINES

Based on observations that chronic infection is able to confer immunological memory to pregnant mothers, normally preventing disease transmission to fetuses even if re-exposed during gestation, a toxoplasmosis vaccine seems highly feasible. Various vaccination strategies against toxoplasmosis in rodent models have been employed in the past decades, ranging from inactivated vaccines, protein vaccines in subunit or multi-antigenic cocktails, and DNA vaccines. However, efficacious vaccines remains a challenge as vaccine experiments have not been able to confer sterile immunity against infection. Sterile immunity is a protection endpoint that is assessed not only by survival rate, but also by the lack of tissue cysts after challenge infection, which is a hallmark of chronic infection. Only a few live, attenuated strains of *T. gondii* have been able to elicit a potent Th1 immune response for effective disease resistance, but live vaccines such as the commercially-licensed ToxoVax[®] (Intervet B.V.) used for sheep immunization is considered unsafe for human use and is restricted in its applications due to safety concerns as well as a short shelf-life.

It has been established that the IFN- γ -secreting CD8⁺ T lymphocyte is the major T cell subset involved in crucial long-term protective immunity in toxoplasmosis [9,10]. Thus, an ideal vaccine to treat toxoplasmosis should be sufficiently immuno-

genic to induce a Th1-type immune response to control disease pathogenesis, as well as to prevent the development of chronic tissue cysts. There are several challenging features in toxoplasmosis vaccine development. First is the stage-specific expression of parasite proteins which may render vaccines targeted to the acute stage ineffective at the chronic stage and vice versa. Second is the presence of varied *T. gondii* genotype strains means that vaccinations against a particular strain may not be protective against infections with other strains, especially with growing concern over the virulent, atypical strains from South America [11]. Third is the parasite's innate immune evasion mechanisms that enable the sequestration of dormant tissue cysts as a life-long chronic infection has been recalcitrant to both drug treatments and vaccination attempts to eliminate infection.

PROTEIN VACCINES

Significant advances made in the past decade in understanding the cell and molecular biology of the *T. gondii* parasite has provided the impetus to the current focus of vaccines based on defined subcellular components of the parasite. Among the parasite antigens that have been researched extensively in vaccination experiments are the surface antigen glycoproteins, or SAGs. Notably, investigations on highly immunogenic SAG1 expressed on tachyzoites have repeatedly yielded high survival rates and significant reduction in brain tissue cysts load [reviewed in 7]. SAG1 immunization studies by Khan et al. [12] and Debard et al. [13] demonstrated 100% and 85% reduction in brain cysts load when adjuvanted with QuilA and cholera toxin (CT) respectively. In a move to use less toxic adjuvants, a SAG1 vaccine co-administered with non-toxic heat-labile enterotoxin (LT) also provided significant but slightly lowered protection and a 78% reduction in brain cyst loads following intranasal delivery [14]. However, this highlights the need to develop adjuvants to simultaneously maximize immunostimulatory efficacy and safety. A recent development was the use of saponins from *Panax ginseng* roots as a non-toxic adjuvant in a ROP18 vaccine formulation, which improved protective efficacy of the vaccine [15]. Thus, these ginseng roots that have been used as traditional medicine hold potential in enhancement of the anti-toxoplasmosis immune responses and could be investigated further in vaccine formulations.

Immunity against *T. gondii* is highly complex and studies have shown how differences in host genetic background can influence vastly divergent disease outcome. This was apparent

in a SAG1 vaccination which proved inhibitory to the mother-fetal transmission of toxoplasmosis in BALB/c mice by 50%, but not with CBA/J mice, which in fact exhibited increased disease transmission to fetuses [16]. As congenital toxoplasmosis can cause severe damage to a baby's brain, eyes, and nervous system, and current drug treatments are teratogenic, immunization efficacy in limiting congenital transmission should be also considered an important endpoint in more vaccine research.

In recent years, vaccination with several protein antigen combinations resulted in increased survival rates in mice following lethal challenge with type I parasite. The most significant gains in survival rates were reported for vaccine antigen SAG1 (80%) [17], SAG1+SAG2 (83%) [18], TgACT (50%) [19], and TgRACK-1 (45%) [20]. Among these findings, the unique study by Chuang et al. [17] used poly (lactide-co-glycolide) (PLG) microparticle encapsulation of the SAG1 and SAG2 protein antigens as a sustained-release vaccine formulation, which is an attractive concept for inducing prolonged disease immunity. Overall, with proper optimization to prevent antigens from denaturing during the encapsulation process, this is a strategy which could be also applied in various other immunotherapeutics formats to maintain therapeutic levels in blood or tissues for extended periods of time.

An important endpoint in the evaluation of toxoplasmosis vaccines is the provision of sterile immunity from the cystic bradyzoite stage, which is recalcitrant to current drug treatments and the source of disease reactivation in chronically-infected individuals. Thus, scoring the effects of immunization on brain tissue cyst loads in mice model gives crucial insights on vaccine utility. In lieu of this, the parasite antigens that have been included in numerous multi-antigen vaccines with good success are the ROP2 and GRA4 proteins. ROP2 is an abundant rhoptry protein essential in parasite multiplication and host invasion [21]. GRA4 is a dense granule protein localized to the intravacuolar network of the parasitophorous vacuole (PV) and involved in nutrient acquisition [22]. Both antigens have shown Th1-type immunogenicity, and its protective efficacy in various mouse strains [23] indicates these antigens as vaccine candidates that warrant further studies. The highest reductions in brain tissue cyst loads conferred were through the immunization with protein antigen combinations of ROP2+ROP4+SAG1 (90%) [24], ROP2+ROP4+GRA4 (84%) [25], and GRA7 nanoparticles (72%) [26]. In particular, the GRA7 nanoparticle is a novel vaccine based on monomeric T cell

epitopes of GRA7 protein that self assembles into nanoparticles similar to a viral capsid. This nanoparticle vaccine is self-adjuncting and demonstrates partial protective immunity to both type I and type II challenge infections.

Another interesting development was the vaccination with a recombinant plant-produced GRA4 antigen [27]. While a plant-produced toxoplasmosis antigen is not novel in itself [28], this work done by del L. Yácono et al. [27] involved the oral immunization of each mouse with leaf extracts equivalent to approximately 0.5 µg of GRA4 antigen. The substantial reduction in tissue cysts (59%) achieved by a low-dose vaccine followed by challenge with the cyst-forming type II strain potentiates the GRA4 antigen as a strong vaccine candidate. As *T. gondii* parasites mainly gain host entry by the intestinal mucosa route, an oral vaccine that can be potentially produced at lowered cost in plants shows good prospects. A summary of some of the most recent studies in protein vaccines against toxoplasmosis is provided in Table 1.

DNA VACCINES

DNA vaccination has the distinct advantage of being capable of generating an effective immune response with substantial cost benefits. DNA vaccinations against toxoplasmosis would generally induce an enhanced Th1 immune response with an increased production of inflammatory cytokines IFN-γ and IL-2 to limit infection. An analysis across *T. gondii* DNA vaccination studies reveals that the inducement of a Th1-type cellular immune response with significantly elevated levels of IFN-γ and IL-2 does not correlate with the degree of protective outcomes in mice. This means that the level of immunogenicity is not necessarily predictive of survival rates, as can be appreciated in several recent examples [33-36]. However, DNA vaccinations in general has shown to be effective at inducing the activation and proliferation of CD4⁺ and CD8⁺ T cells, along with eliciting specific antibodies essential for control of chronic infection.

The drawback of DNA vaccines is that the immune responses it generates are usually too weak to be significantly protective when administered in higher primates and humans [37]. Improvement in its potency can be gained by strategies such as codon optimization for increased protein expression, co-expression of cytokines, and molecular adjuvants, and using a heterologous prime-boost strategy [7,38]. In addition, DNA vaccination strategies using a combination of multiple antigens

Table 1. Examples of immunization with protein vaccines against *Toxoplasma gondii*

Antigen	Antigen delivery	Mouse strain	Challenge	Results	Study
ROP5	Protein (100 µg) +FCA+2 boosters in IFA, sc	BALB/c	100 tachyzoites, RH strain	Increased survival time relative to controls, but no complete protection, complete mortality by day 12	[29]
ROP5+SAG1	Proteins (100 µg each) +FCA+2 boosters in IFA, sc	BALB/c	100 tachyzoites, RH strain	Increased survival time (12.1 ± 3.4 days). No complete protection, complete mortality by day 17. Mixed Th1/Th2 immune response.	[29]
ROP2+GRA4 +SAG1	Proteins (total amount: 10 µg)+IFA, sc	C3H/HeJ (H-2 ^k) and C57BL/6 (H-2 ^b)	5 tissue cysts, DX (Type II) strain	C3H/HeJ: No significant reduction in brain tissue cysts C57BL/6: Reduced brain tissue cysts (55%)	[24]
ROP2+ROP4+ SAG1	Proteins (total amount: 10 µg)+IFA, sc	C3H/HeJ (H-2 ^k) and C57BL/6 (H-2 ^b)	5 tissue cysts, DX (Type II) strain	C3H/HeJ: Reduced brain tissue cysts (71%) C57BL/6: Reduced brain tissue cysts (90%)	[24]
ROP2+ROP4+ GRA4	Proteins (total amount: 10 µg)+IFA, sc	C3H/HeJ (H-2 ^k) and C57BL/6 (H-2 ^b)	5 tissue cysts, DX (Type II) strain	C3H/HeJ: Reduced brain tissue cysts (59%) C57BL/6: Reduced brain tissue cysts (41%) Reduced parasitaemia to 71% (C3H/HeJ) and 90% (C57BL/6)	[24]
ROP2+ROP4+ SAG1	Proteins (total amount: 10 µg)+IFA, sc	BALB/c (H-2 ^d)	5 tissue cysts, DX (Type II) strain	Reduced brain tissue cysts (77%)	[25]
ROP2+ROP4+ GRA4	Proteins (total amount: 10 µg)+IFA, sc	BALB/c (H-2 ^d)	5 tissue cysts, DX (Type II) strain	Reduced brain tissue cysts (84%)	[25]
ROP2	Protein (10 µg)+CpG (10 µg), im	C3H/HeN (H-2 ^k)	20 tissue cysts, Me49 (Type II) strain	Reduced brain tissue cysts (63%)	[30]
ROP2+GRA4	Proteins (10 µg each) +CpG (10 µg), im	C3H/HeN (H-2 ^k)	20 tissue cysts, Me49 (Type II) strain	Reduced brain tissue cysts (66%)	[30]
SAG1	Protein (10 µg)+PLG encapsulation ^a , ip	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	Increased survival (endpoint: 28 days) of 80% Control immunized group survival was 20%	[17]
SAG1+SAG2	Protein (10 µg)+PLG encapsulation, ip	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	Increased survival (endpoint: 28 days) of 83%	[18]
SAG1+GRA1+ GRA4 (T- and B-cell epitopes)	Protein (50 µg)+FCA +2 boosters in IFA, im	BALB/c and Kunming	10 tachyzoites, GJS (Type I) strain	BALB/c: Increased survival time (18.3 ± 0.9 days) Kunming: Increased survival time (16.0 ± 0.8 days)	[31]
GRA4	Protein from leaf extracts of GRA4- recombinant plants (0.5 µg) ^b , oral	C57BL/6	20 tissue cysts, Me49 (Type II) strain	Reduced brain tissue cysts relative to control (59%) Increased Th1/Th2 cytokine	[27]
TgACT ^c	Protein (30 µg), in	BALB/c	Acute infection: 4 × 10 ⁴ tachyzoites, RH strain Chronic infection: 1 × 10 ⁴ tachyzoites, RH strain	Increased survival rate of 50% relative to control Reduced tachyzoites load in liver (60.05%) and brain (49.75%)	[19]
TgPDI ^d	Protein (30 µg), in	BALB/c	Acute infection: 4 × 10 ⁴ tachyzoites, RH strain Chronic infection: 1 × 10 ⁴ tachyzoites, RH strain	Increased survival rate of 31% relative to control Reduced tachyzoites load in liver and brain (statistically significant, <i>P</i> < 0.05)	[32]
TgRACK-1 ^e	Protein (35 µg), in	BALB/c	4 × 10 ⁴ tachyzoites, RH strain	Increased survival rate of 45% relative to control Decreased parasite burden in liver and brain (50%)	[20]

SAG, surface antigen; ROP, rhoptry antigen; GRA, dense granule antigen; IFA, incomplete Freund's adjuvant; FCA, Freund's complete adjuvant; CpG, oligodeoxynucleotides CpG motif adjuvant; sc, subcutaneous; im, intramuscular; ip, intraperitoneal; in, intranasal.

^aAntigen encapsulation in poly (lactide-co-glycolide) microparticles as adjuvant and sustained-release of antigen.

^bMice were fed with 83 mg of leaf extracts, containing an equivalent of 1 µg of rGRA4 in 5 doses at 1 week intervals.

^c*T. gondii* actin.

^d*T. gondii* protein disulfide isomerase (PDI), localized mainly in the ER, involved in cell signaling and homeostasis; also found on tachyzoite membrane surface and modulates host-parasite cell interactions.

^e*T. gondii* receptor for activated C kinase-1.

have been shown to be more effective in enhancing protective immunity than single subunit antigens [reviewed in 11]. A case in point is the antigenic combination of micronemal proteins

(MIC2, MIC3, MIC4, M2AP, and AMA1) which achieved significant protection of 84% reduction in the brain tissue cyst burden and longer survival durations than single subunit anti-

gen vaccination [39]. To ensure immunogenicity of the vaccines, the authors formulated a DNA vaccine based on known T-cell epitopes instead of using full-length antigens. This highlights that the identification of immunogenic epitopes capable of eliciting humoral and cellular immune responses is another important parameter to consider in vaccine design. The *in silico* identification of these functional epitopes seems increasingly possible in recent times via predictive binding algorithms for humans' MHC I molecules [8, reviewed in 40].

Co-administration of adjuvants has also shown to be a useful approach in boosting the immune response in DNA vaccinations to therapeutic levels. A recent example of this is the MIC8 vaccination studies. The MIC8 gene previously garnered a lot of interest due to the completely defective invasion phenotype of MIC8-parasite knockout mutant [41]. However, DNA vaccination with MIC8 alone generated only a modest increase in survival time [42]. In a strategy to enhance the protective immunity from MIC8 vaccination, co-administration of mIL-21 and mIL-15 cytokines showed improved survival time from 10.3 ± 0.9 days (without cytokines) to 16.2 ± 1.3 days (with cytokines) [43].

The highest protection achieved with DNA vaccination against toxoplasmosis was obtained by immunization with a plasmid encoding SAG, which conferred 100% survival in CH3 mice and 80% survival in BALB/c mice for at least 6 months post-lethal challenge [44]. The potency of this protective immunity exceeded a similar experiment using SAG1 protein vaccine which prolonged mouse survival, but couldn't prevent complete mortality [45]. However, several more recent studies using either the SAG1 antigen alone or in combination with other antigens showed less robust protective immunity, where the highest survival rate was 40%, even with the use of the same strain of parasite and mouse host [46-48]. This ambiguity may be attributed to the use of different SAG1 epitopes in each study. Another immunization study which also demonstrated a significant survival rate of 100% and 40% in BALB/c and C57BL/6 mice respectively following a lethal challenge, used a multi-epitope DNA vaccination [49]. This multi-epitope vaccine comprising 6 antigenic epitopes from SAG1, GRA1, GRA2 and GRA4 antigens also showed effective stimulation of a protective Th1 immune response; and forms part of an accumulating body of evidence on the efficacy of multi-antigen vaccinations against a complex parasite such *T. gondii* [49-51].

Over the past 4 years, toxoplasmosis DNA vaccination studies expanded to the use of many new parasite antigens, several

of which yielded promising protective outcomes (Table 2). Among these latest developments, several single-antigen vaccines were successful in producing survivors from a lethal infection challenge, such as ROP8, TgCyP, and GRA6 with a range of survival rate between 37.5-50.0% [52-54]. Another subset of these DNA vaccine studies produced a significant increase in survival time (≥ 3 weeks) but no survivors following a lethal challenge, which was the ROP13, ROP16, ROP18, MIC13, and eIF4A [34,36,55-57]. There was a less significant survival outcome (≤ 2 weeks) from yet another subset of antigens, such as TgROM1, TgPLP1, TgCDPK3, and TgCDPK5 [33,58-60]. For effective comparisons, these recent studies in DNA vaccinations against toxoplasmosis are summarized in Table 2. Taken together, the next generation of vaccine design could include the empirical evaluation of functional antigenic epitope combinations in conjunction with immunological adjuvants for maximal efficacy.

LIVE, ATTENUATED VECTORS

Another vaccination strategy against toxoplasmosis involves the use of live, attenuated vectors, such as viruses and bacteria as vehicles to express recombinant *T. gondii* antigens in the host. The use of a live, attenuated vector to deliver *T. gondii* antigens provides the benefits of mimicry of the intracellular niche of the parasite by using innocuous recombinant virus or bacteria, and has been shown to be able to induce complete protection in some instances due to its intrinsic adjuvant properties. An example of this was shown in a DNA/viral vector heterologous prime-boost vaccination experiment. The GRA4 antigen delivered via recombinant attenuated vaccinia virus afforded 100% survival from infection with type II PLK/GFP tachyzoites, no detectable brain tissue cysts on 45 days post-infection, and significantly reduced brain parasite load [67]. In contrast, the plasmid-plasmid DNA vaccination group showed a lower survival rate of 70%. Some of the live vectors used in *T. gondii* vaccination studies include *Salmonella typhimurium*, pseudorabies virus, adenovirus, and modified vaccinia Ankara (MVA) (Table 3). The major advantages of live vectors as vaccine vehicles are its prospect as oral vaccines leading to inducement of mucosal immunity, as well as the potent immune response it elicits. These advantages has led to the development of these live vaccine formats for diseases ranging from cholera [68], to HIV infection [69] and cancer [70]. However, the use of recombinant, live vaccines faces the inevitable obstacles of

Table 2. Examples of immunization with DNA vaccines against *Toxoplasma gondii*

Antigen	Antigen delivery	Mouse strain	Challenge	Results	Study
TgPLP1 ^a	Plasmid, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Increased survival time (11.3 ± 0.9 days). Unvaccinated cohort succumbed by day 6	[59]
TgPLP1	Plasmid+IL-18, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Improved survival time with IL18 co-administration (12.7 ± 1.2 days)	[59]
ROP9	Plasmids, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Increased survival time (12.9 ± 2.9 days)	[35]
ROP8	Plasmid, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Increased survival rate (50%) ^b	[52]
ROP13	Plasmid+IL-18, im	Kunming	Acute: 1 × 10 ³ tachyzoites (RH) Chronic: 10 tissue cysts, PRU (Type II) strain	RH strain challenge: Increased survival time (32.3 ± 2.7 days) PRU strain challenge: Reduction in brain tissue cysts load (66%)	[55]
ROP16	Plasmid, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Improved survival time (21.6 ± 9.9 days)	[56]
ROP18	Plasmid, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Increased survival time (27.9 ± 15.1 days). Unvaccinated cohort succumbed by day 7	[57]
SAG1	Plasmid, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	No complete protection, 100% mortality by day 8	[46]
SAG1+ SAG3	Plasmids, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Increased survival rate (20%)	[46]
SAG1+ SAG3	Plasmids+CT, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Increased survival rate with CT adjuvant (40%). Increased secretion of IFN-γ and higher lymphocyte proliferation	[46]
SAG1	Plasmid, im	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	Increased survival time (7.7 ± 2.5 days)	[47]
SAG1+ 14-3-3 ^c	Plasmids, im	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	Increased survival time (11.5 ± 3.5 days)	[47]
SAG2C+ SAG2D+ SAG2X ^d	Plasmids, im	BALB/c	20 tissue cysts, PRU (Type II) strain	Significant reduction in brain tissue cysts: SAG2C (72%), SAG2X (69%) and combined SAG2CDX (77%) No significant reduction in brain cysts for SAG2D-immunized group	[61]
SAG1+ GRA2	Plasmids, im	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	All mice succumbed by day 16 (n=12)	[48]
SAG1+ GRA2	Plasmids+ pSPreS2 ^e , im	BALB/c	1 × 10 ⁴ tachyzoites, RH strain	All mice succumbed by day 17 (n=12) No significant difference between immunization with or without pSPreS2 adjuvant	[48]
AMA1	Plasmid+recombinant adenovirus, ip	C57BL/6	1 × 10 ³ tachyzoites, PLK-GFP (Type II) strain	Heterologous prime-boost strategy using plasmid and virus vector bearing AMA1 antigen showed 50% survival rate, 23% reduction in brain tissue cysts	[62]
GRA4	Plasmid+peptide, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Heterologous prime-boost strategy using plasmid and GRA4 peptide antigen showed increased survival time (16.5 ± 5.4 days) and 40% survival rate	[63]
GRA6 ^f	Plasmid+LMS ^g , im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Partial protection. Higher survival rate with LMS (53.3%). Survival rate without adjuvant was lower (40%). Control group had 0% survival No detectable parasites in brain, liver and spleen of immunized groups (with and without LMS)	[54]
MIC13	Plasmid, im	Kunming	Acute: 1 × 10 ³ tachyzoites, RH strain Chronic: 10 tissue cysts, PRU (Type II) strain	RH strain challenge: Increased survival time (21.3 ± 11.3 days). PRU strain challenge: Reduction in brain tissue cysts load (57%)	[34]
MIC8	Plasmid+IL-12, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Increased survival time (15 days). No complete protection, all mice succumbed to illness	[64]
MIC8	Plasmid+mIL-21 +mIL-15, im	Kunming	Acute: 1 × 10 ³ tachyzoites, RH strain Chronic: 20 tissue cysts, PRU (Type II) strain	RH strain challenge: Increased survival time (16.2 ± 1.3 days) PRU strain challenge: Reduction in brain tissue cysts load (63.8%)	[43]
TgCDPK5 ^h	Plasmid, im	Kunming	Acute: 1 × 10 ³ tachyzoites, RH strain Chronic: 10 tissue cysts, PRU strain	Increased survival time (8.7 ± 4.3 days) Reduced brain tissue cysts (40%)	[33]

(Continued to the next page)

Table 2. Continued

Antigen	Antigen delivery	Mouse strain	Challenge	Results	Study
TgCDPK3 ⁱ	Plasmid, im	Kunming	Acute: 1 × 10 ³ tachyzoites, RH strain Chronic: 10 tissue cysts, PRU strain	Increased survival time (13.5 ± 4.9 days) Reduced brain tissue cysts (54%)	[60]
TgCyP ^j	Plasmid, im	BALB/c	500 tachyzoites, RH strain	Increased survival rate (37.5%) No reduction in brain parasite load	[53]
eIF4A ^k	Plasmid, im	Kunming	1 × 10 ³ tachyzoites, RH strain	Increased survival time (23.0 ± 5.5 days)	[36]
eIF2α	Plasmid, im	Kunming	Acute: 1 × 10 ³ tachyzoites, RH strain Chronic: 10 tissue cysts, PRU (Type II) strain	Increased survival time (15.9 ± 4.6 days) Reduced brain tissue cysts (44%)	[65]
TgROM1 ^l	Plasmid, im	BALB/c	1 × 10 ³ tachyzoites, RH strain	Increased survival time (12.5 ± 0.7 days)	[58]
TgIMP-1 ^m	Plasmid, im	BALB/c	500 tachyzoites, RH strain	Increased survival time (15.8 ± 6 days)	[66]

SAG, surface antigen; ROP, rhoptry antigen; GRA, dense granule antigen; AMA, apical membrane antigen; MIC, microneme antigen; CT, cholera toxin; im, intramuscular; ip, intraperitoneal; in, intranasal.

^a*T. gondii* perforin-like protein 1.

^bAuthors did not state the survival rate at endpoint of day 29, therefore the rate was inferred from the survival curve. The study showed a 100% survival at day 9 while control group had total mortality.

^cAntigen 14-3-3 is a protein found on parasitophorous vacuoles (PV) of tachyzoites and is a member of excretory secretory antigen (ESA) fraction.

^dSAG2C/D/X are bradyzoite-specific antigens, important in chronic infection.

^ePSPreS2 is a novel genetic adjuvant encoding for hepatitis B virus surface antigen S and PreS2 - a highly immunogenic region within native HbsAg.

^fDNA vaccine encodes for the immunodominant GRA6 epitope HF-10 from a Type II *T. gondii* strain which binds the H-2L^D MHC class 1 molecule.

^gLevamisole (LMS), a chemical adjuvant.

^h*T. gondii* calcium-dependent protein kinase 5.

ⁱ*T. gondii* calcium-dependent protein kinase 3.

^j*T. gondii* cyclophilin.

^kEukaryotic translation initiation factor, an RNA helicase.

^lRhomboid protein 1, an intramembrane serine protease.

^m*T. gondii* immune-mapped protein-1, a highly conserved antigen in apicomplexan parasites.

Table 3. Examples of immunization with live, attenuated vectors expressing *Toxoplasma gondii* antigens

Antigen	Antigen delivery	Mouse strain	Challenge	Results	Study
GRA4	DNA plasmid, gg+Vaccinia	C57BL/6	2 × 10 ⁴ tachyzoites, PLK/GFP (Type II) strain	100% survival rate. No detectable brain tissue cysts in vaccinated cohort, while controls showed 400-600 tissue cysts per brain. Significantly reduced brain tachyzoites load	[67]
SAG1	Heterologous vaccination: Adenovirus, sc+MVA ^a , im	C57BL/6	10 tissue cysts, Me49 (Type II) strain	80% survival rate, which is higher than homologous vaccination protocol (60%). Significantly reduced brain tissue cysts compared to homologous vaccination	[72]
SAG1+ SAG2	<i>Salmonella typhimurium</i> + CT, ig	BALB/c	1 × 10 ³ tachyzoites, RH strain	40% survival rate	[73]
SAG1	Recombinant Pseudorabies virus (rPRV), im	BALB/c	50 tachyzoites, RH strain	60% survival rate with a single dose immunization Survival rate was reduced to 20% when 3 doses of rPRV were administered in 15 days intervals	[74]
SAG1+ MIC3	Recombinant Pseudorabies virus (rPRV), im	BALB/c	100 tachyzoites, RH strain	66.7% survival rate	[75]

SAG, surface antigen; GRA, dense granule antigen; MIC, microneme antigen; CT, cholera toxin (as an adjuvant); gg, gene gun; sc, subcutaneous; im, intramuscular; ig, intragastric.

^aModified vaccinia virus ankara.

safety risks and genetically-modified organisms (GMO) regulatory issues. The safety concerns that need to be addressed other than the more obvious ablation of virulence, is the lowering of risk of autoimmune responses and non-administration to immunocompromised individuals [reviewed in 71]. The vaccination studies using live, attenuated vectors against

toxoplasmosis are summarized in Table 3.

Protective immunity to toxoplasmosis is highly complex as the interplay between the parasite's strain type and the host's genetic background influences disease pathogenesis [76,77]. The host's genetic influence over the selective processing of antigenic peptide, with the subsequent ability to present certain

peptides determines disease severity in each individual. Therefore, the challenge of vaccine design is to determine protective epitopes across the antigenic diversity in *T. gondii* strains, capable of engaging with the diverse MHCs encompassing a majority of the human population to produce protective immunity [40]. Vaccine researchers now have an additional resource for testing vaccines by referring to the toxoplasmosis infection and pathogenesis model developed by Dubey et al. [78]. This in-depth study details infection responses across different mouse strains and various *T. gondii* strains, including the atypical strains. Thus, this model can be a highly useful tool in testing vaccine efficacy by providing a foundational standard for comparison.

In considering the platforms for vaccine delivery from the myriad of strategies ranging from DNA to live vaccines, it is worth noting that each platform presents its own benefits and disadvantages that are beyond the scope of this article to delve into. The interested reader is referred to an excellent review by Bruna-Romero et al. [79] on this topic. The development of pre-emptive vaccines remains a highly important pursuit in light of the persistent nature of chronic toxoplasmosis that is refractory to current drug treatments, and the increasing mortality associated with reactivated infections.

PASSIVE IMMUNIZATION

In the acute stage of toxoplasmosis, the host's innate immune response plays a key role in the control of infection [80], while long-term protective immunity is mediated by the adaptive immune response that drives it into a quiescent chronic stage but does not completely eradicate the parasite [81,82]. The persistent chronic stage can be lethal in reactivated infections affecting immunocompromised hosts. Thus, passive immunization could be a useful means of limiting clinical disease in hosts with reduced cellular immunity.

A major cell subset critical in disease control is the CD8⁺ T cells which produces the cytokine IFN- γ [9,10]. It was revealed that the adoptive transfer of immune CD8⁺ T cells to naïve animals could confer protective immunity and prolong survival in acute infections [82]. Although these are encouraging findings, translation of the application of syngeneic cellular immunotherapeutics to the masses could be challenging. Nonetheless, studies on passive cellular immunization could still provide invaluable insights on the factors affecting long-term immune control of toxoplasmosis. The inbred C57BL/6 mouse strain is

mostly used in such studies due to its susceptibility to succumb to chronic toxoplasmosis, even with avirulent strains. This condition mimics reactivation of latent infections arising from immune compromise. One of the recent and interesting findings of this is that adoptive transfer of immune CD8⁺ T cells to pre-crudescent chronically-infected C57BL/6 mice could provide transient protection from disease reactivation for up to 4 weeks post-treatment [83]. However, this same treatment could not rescue endogenous CD8⁺ T cells from functional exhaustion leading to apoptosis and the impairment of memory T cells development. Thus, long-term protective immunity was not achieved. These results seem to suggest that adoptive cellular immunotherapy provides significant protection in acute toxoplasmosis, but not so in a model of disease reactivation in chronic toxoplasmosis.

While CD8⁺ T cells and IFN- γ are important mediators of the host immune responses to resist *T. gondii* infection, antibodies produced by B cells are essential for long-term resistance to the disease by the production of specific antibodies [84,85]. The absence of antibody production in the host exacerbates infection and results in chronic stage mortality. In a study using CD4⁺ depleted mice to parallel CD4⁺ deficiency in AIDS patients and other immunocompromised individuals, passive immunization with immune serum has shown to be able to prolong survival and transiently overcome impaired disease resistance [86]. This suggests that the production of specific antibodies together with cellular immune responses is an important feature in the generation of adaptive immunity to *T. gondii*. Therefore, the potential development of antibodies as immunotherapeutic agents to arrest disease progression would prove highly beneficial to individuals suffering from impaired immunity.

In an experiment using Fab antibody fragments specific to *T. gondii* SAG1 antigen, it was shown that the antibodies blocked parasites attachment to host cells in vitro by 52% at an antibody concentration of 200 $\mu\text{g/ml}$ [87]. Passive immunization of mice with 10 mg of this antibody provided improved survival rates of up to 50% following lethal challenge infection [87]. This survival rate improvement is a significant finding considering that the Fab fragments used in this study is lacking the Fc region of full antibodies which mediates Fc-receptor-dependent phagocytosis of opsonized tachyzoites, cellular cytotoxicity and complement activation. Therefore, it would seem that the antibodies mediate its protective function by binding to the parasites' surface and directly blocking infection of host

cells, which is consistent with an observation by Sayles and co-workers in their study [85].

Passive immunization with anti-GRA2 and anti-GRA6 monoclonal antibodies have also resulted in inhibition of parasite invasion and enhanced survival times in mice. However, the anti-SAG1 antibody yielded a more significant protective immunity than the antibodies targeted to the dense granules antigens [88]. Another passive immunization experiment using monoclonal antibodies against nucleoside triphosphate hydrolase-II (NT-Pase-II) showed no inhibition of parasite invasion, but a significant reduction in *T. gondii* replication and promoted prolonged survival in mice after a lethal challenge [89]. Taken together, the development of anti-toxoplasmosis antibodies providing greater protection from host infection could benefit from a combinatorial antibody formulation that blocks parasite attachment, invasion and proliferation; and thus warrants further investigations.

Recently, polyclonal antibodies against *T. gondii* was produced in egg yolks from immunized chicken and showed a comparable avidity and reactivity profile to mammalian-derived antibodies [90]. The main advantage of using a chicken platform is its' very high yield of antibodies (4 mg/ml purified IgY) which can be purified inexpensively and obviates the need to bleed animals. Because of the phylogenetic distance between avian and mammalian species, chicken IgY antibodies has the inherent biochemical advantage of not activating the mammalian complement system, and does not react with rheumatoid factors and anti-human IgG antibodies [91]. However, the therapeutic effect of these avian antibodies remains uncertain and requires further study. In conclusion, regardless of the production platform for anti-toxoplasmosis antibodies, therapeutic antibodies to ameliorate the significant disease morbidity and mortality associated with reactivated infections is obviously needed and evidence from past studies shows that this can be a feasible toxoplasmosis treatment for immunocompromised patients.

CONCLUDING REMARKS

The immune evasive nature and complex life cycle of *T. gondii* has made the design of immunotherapeutics aimed at providing complete protection a daunting challenge. Generally, vaccine studies thus far have shown that multi-antigenic formulations confer better protection than single-subunit vaccines. At this juncture, research on adjuvants for vaccine co-ad-

ministration is also crucial as alum has been the only one out of five licensed human adjuvants tested with toxoplasmosis vaccine formulations. In context of toxoplasmosis vaccination, this isn't an ideal option as alum augments the Th2 immune response, instead of the predominantly Th1 responses usually seen in protective immunity. Therefore, studies on multiple combinations of antigens with co-delivery of adjuvants or cytokines may yield better protective outcomes and progressively narrow the gap towards achieving sterile immunity. Vaccines and immunotherapeutic antibodies targeted to the parasite's surface membrane antigens present highly promising prospects considering the high immunogenicity of these antigens and the resulting significant parasitaemia inhibition from multiple studies reviewed thus far. Therefore, immunotherapeutics targeting either a single or multiple parasite surface antigens, in combination with invasion-related antigens such as the MIC, AMA1 and ROP proteins can be a potential strategy for generating a neutralizing immune response against toxoplasmosis. The discovery of atypical strains in the South American region which are more virulent than the classic European types I, II and III strains, and causes severe illness; would mean that next generation research efforts should also consider designing assays for cross-protection evaluation. These research efforts could be aided by a presently available model of toxoplasmosis infection strains in mouse [78]. A universal or multi-type anti-*T. gondii* immunotherapeutics is a highly valuable goal for global disease control. To this end, the completion of the genomes of predominant strains of *T. gondii* together with bioinformatics algorithms to predict MHC I-binding peptides could be a valuable tool to inform the design of broadly protective immunotherapeutics.

ACKNOWLEDGMENTS

Work related to the preparation of this review was supported by University of Malaya, Malaysia (grant: UM.C/625/1/HIR/MOHE/SCI/18).

CONFLICT OF INTEREST

We have no conflict of interest related to this review article.

REFERENCES

1. Chowdhury M. *Toxoplasmosis*: a review. *J Med* 1986; 17: 373-396.

2. Brindle R, Holliman R, Gilks C, Waiyaki P. *Toxoplasma* antibodies in HIV-positive patients from Nairobi. *Trans R Soc Trop Med Hyg* 1991; 85: 750-751.
3. Mui EJ, Schiehsler GA, Milhous WK, Hsu H, Roberts CW, Kirisits M, Muench S, Rice D, Dubey JP, Fowble JW, Rathod PK, Queenier SF, Liu SR, Jacobus DP, McLeod R. Novel triazine JPC-2067-B inhibits *Toxoplasma gondii* in vitro and in vivo. *PLoS Negl Trop Dis* 2008; 2: e190.
4. Khan A, Jordan C, Muccioli C, Vallochi AL, Rizzo LV, Belfort R Jr, Vitor RWA, Silveira C, Sibley LD. Genetic Divergence of *Toxoplasma gondii* Strains Associated with Ocular Toxoplasmosis, Brazil. *Emerg Infect Dis* 2006; 12: 942-949.
5. Bottos J, Miller RH, Belfort RN, Macedo AC, UNIFESP Toxoplasmosis Group, Belfort R Jr, Grigg ME. Bilateral retinochoroiditis caused by an atypical strain of *Toxoplasma gondii*. *Br J Ophthalmol* 2009; 93: 1546-1550.
6. Garcia J, Innes EA, Katzer F. Current progress toward vaccines against *Toxoplasma gondii*. *Vaccine Devel Ther* 2014; 4: 23-37.
7. Kur J, Holec-Gasior L, Hiszczynska-Sawicka E. Current status of toxoplasmosis vaccine development. *Expert Rev Vaccines* 2009; 8: 791-808.
8. Tan TG, Mui E, Cong H, Witola W, Montpetit A, Muench SP, Sidney J, Alexander J, Sette A, Grigg M, Maewal A, McLeod R. Identification of *T. gondii* epitopes, adjuvants, and host genetic factors that influence protection of mice and humans. *Vaccine* 2010; 28: 3977-3989.
9. Gazzinelli RT, Hakim FT, Hieny S, Shearer GM, Sher A. Synergistic role of CD4⁺ and CD8⁺ T lymphocytes in IFN-gamma production and protective immunity induced by an attenuated *Toxoplasma gondii* vaccine. *J Immunol* 1991; 146: 286-292.
10. Brown CR, McLeod R. Class I MHC genes and CD8⁺ T cells determine cyst number in *Toxoplasma gondii* infection. *J Immunol* 1990; 145: 3438-3441.
11. Jongert E, Roberts CW, Gargano N, Förster-Waldl E, Petersen E. Vaccines against *Toxoplasma gondii*: challenges and opportunities. *Mem Inst Oswaldo Cruz* 2009; 104: 252-266.
12. Debard N, Buzoni-Gatel D, Bout D. Intranasal immunization with SAG1 protein of *Toxoplasma gondii* in association with cholera toxin dramatically reduces development of cerebral cysts after oral infection. *Infect Immun* 1996; 64: 2158-2166.
13. Khan IA, Ely KH, Kasper LH. A purified parasite antigen (p30) mediates CD8⁺ T cell immunity against fatal *Toxoplasma gondii* infection in mice. *J Immunol* 1991; 147: 3501-3506.
14. Bonenfant C, Dimier-Poisson I, Velge-Roussel F, Buzoni-Gatel D, Del Giudice G, Rappuoli R, Bout D. Intranasal immunization with SAG1 and nontoxic mutant heat-labile enterotoxins protects mice against *Toxoplasma gondii*. *Infect Immun* 2001; 69: 1605-1612.
15. Qu D, Han J, Du A. Enhancement of protective immune response to recombinant *Toxoplasma gondii* ROP18 antigen by ginsenoside Re. *Exp Parasitol* 2013; 135: 234-239.
16. Letscher-Bru V, Pfaff AW, Abou-Bacar A, Filisetti D, Antoni E, Villard O, Klein JP, Candolfi E. Vaccination with *Toxoplasma gondii* SAG-1 protein is protective against congenital toxoplasmosis in BALB/c mice but not in CBA/J mice. *Infect Immun* 2003; 71: 6615-6619.
17. Chuang SC, Ko JC, Chen CP, Du JT, Yang CD. Induction of long-lasting protective immunity against *Toxoplasma gondii* in BALB/c mice by recombinant surface antigen 1 protein encapsulated in poly (lactide-co-glycolide) microparticles. *Parasit Vectors* 2013; 6: 34.
18. Chuang SC, Ko JC, Chen CP, Du JT, Yang CD. Encapsulation of chimeric protein rSAG1/2 into poly (lactide-co-glycolide) microparticles induces long-term protective immunity against *Toxoplasma gondii* in mice. *Exp Parasitol* 2013; 134: 430-437.
19. Yin LT, Hao HX, Wang HL, Zhang JH, Meng XL, Yin GR. Intranasal immunisation with recombinant *Toxoplasma gondii* actin partly protects mice against Toxoplasmosis. *PLoS ONE* 2013; 8: e82765.
20. Wang HL, Pang M, Yin LT, Zhang JH, Meng XL, Yu BF, Guo R, Bai JZ, Zheng GP, Yin GR. Intranasal immunisation of the recombinant *Toxoplasma gondii* receptor for activated C kinase 1 partly protects mice against *T. gondii* infection. *Acta Trop* 2014; 137: 58-66.
21. Nakaar V, Ngô HM, Aaronson EP, Coppens I, Stedman TT, Joiner KA. Pleiotropic effect due to targeted depletion of secretory rhoptry protein ROP2 in *Toxoplasma gondii*. *J Cell Sci* 2003; 116: 2311-2320.
22. Nam HW. GRA Proteins of *Toxoplasma gondii*: maintenance of host-parasite interactions across the parasitophorous vacuolar membrane. *Korean J Parasitol* 2009; 47 (suppl): S29-S37.
23. Martin V, Supanitsky A, Echeverria PC, Litwin S, Tanos T, Roodt ARD, Guamera EA, Angel SO. Recombinant GRA4 or ROP2 protein combined with alum or the gra4 gene provides partial protection in chronic murine models of toxoplasmosis. *Clin Diagn Lab Immunol* 2004; 11: 704-710.
24. Dziadek B, Gatkowska J, Brzostek A, Dziadek J, Dzitko K, Grzybowski M, Dlugonska H. Evaluation of three recombinant multi-antigenic vaccines composed of surface and secretory antigens of *Toxoplasma gondii* in murine models of experimental toxoplasmosis. *Vaccine* 2011; 29: 821-830.
25. Dziadek B, Gatkowska J, Grzybowski M, Dziadek J, Dzitko K, Dlugonska H. *Toxoplasma gondii*: the vaccine potential of three trivalent antigen-cocktails composed of recombinant ROP2, ROP4, GRA4 and SAG1 proteins against chronic toxoplasmosis in BALB/c mice. *Exp Parasitol* 2012; 131: 133-138.
26. El Bissati K, Zhou Y, Dasgupta D, Cobb D, Dubey JP, Burkhard P, Lanar DE, McLeod R. Effectiveness of a novel immunogenic nanoparticle platform for *Toxoplasma* peptide vaccine in HLA transgenic mice. *Vaccine* 2014; 32: 3243-3248.
27. del L. Yácono M, Farran I, Becher ML, Sander V, Sánchez VR, Martín V, Veramendi J, Clemente M. A chloroplast-derived *Toxoplasma gondii* GRA4 antigen used as an oral vaccine protects against toxoplasmosis in mice. *Plant Biotechnol J* 2012; 10: 1136-1144.
28. Laguia-Becher M, Martin V, Kraemer M, Corigliano M, Yacono

- ML, Goldman A, Clemente M. Effect of codon optimization and subcellular targeting on *Toxoplasma gondii* antigen SAG1 expression in tobacco leaves to use in subcutaneous and oral immunization in mice. *BMC Biotechnol* 2010; 10: 52.
29. Zheng B, Lu S, Tong Q, Kong Q, Lou D. The virulence-related rhopty protein 5 (ROP5) of *Toxoplasma gondii* is a novel vaccine candidate against toxoplasmosis in mice. *Vaccine* 2013; 31: 4578-4584.
 30. Sanchez VR, Pitkowski MN, Fernandez Cuppari AV, Rodriguez FM, Fenoy IM, Frank FM, Goldman A, Corral RS, Martin V. Combination of CpG-oligodeoxynucleotides with recombinant ROP2 or GRA4 proteins induces protective immunity against *Toxoplasma gondii* infection. *Exp Parasitol* 2011; 128: 448-453.
 31. Wang Y, Wang M, Wang G, Pang A, Fu B, Yin H, Zhang D. Increased survival time in mice vaccinated with a branched lysine multiple antigenic peptide containing B- and T-cell epitopes from *T. gondii* antigens. *Vaccine* 2011; 29: 8619-8623.
 32. Wang HL, Li YQ, Yin LT, Meng XL, Guo M, Zhang JH, Liu HL, Liu JJ, Yin GR. *Toxoplasma gondii* Protein Disulfide Isomerase (TgPDI) is a novel vaccine candidate against toxoplasmosis. *PLoS ONE* 2013; 8: e70884.
 33. Zhang NZ, Huang SY, Xu Y, Chen J, Wang JL, Tian WP, Zhu XQ. Evaluation of immune responses in mice after DNA immunization with putative *Toxoplasma gondii* Calcium-Dependent Protein Kinase 5 (TgCDPK5). *Clin Vaccine Immunol*. 2014; 21: 924-929.
 34. Yuan ZG, Ren D, Zhou DH, Zhang XX, Petersen E, Li XZ, Zhou Y, Yang GL, Zhu XQ. Evaluation of protective effect of pVAX-TgMIC13 plasmid against acute and chronic *Toxoplasma gondii* infection in a murine model. *Vaccine* 2013; 31: 3135-3139.
 35. Chen J, Zhou DH, Li ZY, Petersen E, Huang SY, Song HQ, Zhu XQ. *Toxoplasma gondii*: protective immunity induced by rhopty protein 9 (TgROP9) against acute toxoplasmosis. *Exp Parasitol* 2014; 139: 42-48.
 36. Chen J, Huang SY, Li ZY, Yuan ZG, Zhou DH, Petersen E, Zhang NZ, Zhu XQ. Protective immunity induced by a DNA vaccine expressing eIF4A of *Toxoplasma gondii* against acute toxoplasmosis in mice. *Vaccine* 2013; 31: 1734-1739.
 37. Liu Q, Singla LD, Zhou H. Vaccines against *Toxoplasma gondii*: status, challenges and future directions. *Hum Vaccines Immunother* 2012; 8: 1305-1308.
 38. Saade F, Petrovsky N. Technologies for enhanced efficacy of DNA vaccines. *Expert Rev Vaccines* 2012; 11: 189-209.
 39. Beghetto E, Nielsen HV, Porto PD, Buffolano W, Guglietta S, Felici F, Petersen E, Gargano N. A combination of antigenic regions of *Toxoplasma gondii* microneme proteins induces protective immunity against oral infection with parasite cysts. *J Infect Dis* 2005; 191: 637-645.
 40. Henriquez FL, Woods S, Cong H, McLeod R, Roberts CW. Immunogenetics of *Toxoplasma gondii* informs vaccine design. *Trends Parasitol* 2010; 26: 550-555.
 41. Kessler H, Herm-Gotz A, Hegge S, Rauch M, Soldati-Favre D, Frischknecht F, Meissner M. Microneme protein 8-a new essential invasion factor in *Toxoplasma gondii*. *J Cell Sci* 2008; 121: 947-956.
 42. Liu MM, Yuan ZG, Peng GH, Zhou DH, He XH, Yan C, Yin CC, He Y, Lin RQ, Song HQ, Zhu XQ. *Toxoplasma gondii* microneme protein 8 (MIC8) is a potential vaccine candidate against toxoplasmosis. *Parasitol Res* 2010; 106: 1079-1084.
 43. Li ZY, Chen J, Petersen E, Zhou DH, Huang SY, Song HQ, Zhu XQ. Synergy of mIL-21 and mIL-15 in enhancing DNA vaccine efficacy against acute and chronic *Toxoplasma gondii* infection in mice. *Vaccine* 2014; 32: 3058-3065.
 44. Nielsen HV, Lauemøller SL, Christiansen L, Buus S, Fomsgaard A, Petersen E. Complete protection against lethal *Toxoplasma gondii* infection in mice immunized with a plasmid encoding the SAG1 gene. *Infect Immun* 1999; 67: 6358-6363.
 45. Petersen E, Nielsen HV, Christiansen L, Spenter J. Immunization with *E. coli* produced recombinant *T. gondii* SAG1 with alum as adjuvant protect mice against lethal infection with *Toxoplasma gondii*. *Vaccine* 1998; 16: 1283-1289.
 46. Cong H, Zhang M, Xin Q, Wang Z, Li Y, Zhao Q, Zhou H, He S. Compound DNA vaccine encoding SAG1/SAG3 with A2/B subunit of cholera toxin as a genetic adjuvant protects BALB/c mice against *Toxoplasma gondii*. *Parasit Vectors* 2013; 6: 63.
 47. Meng M, He S, Zhao G, Bai Y, Zhou H, Cong H, Lu G, Zhao Q, Zhu XQ. Evaluation of protective immune responses induced by DNA vaccines encoding *Toxoplasma gondii* surface antigen 1 (SAG1) and 14-3-3 protein in BALB/c mice. *Parasite Vector* 2012; 5: 273.
 48. Zhou H, Min J, Zhao Q, Gu Q, Cong H, Li Y, He S. Protective immune response against *Toxoplasma gondii* elicited by a recombinant DNA vaccine with a novel genetic adjuvant. *Vaccine* 2012; 30: 1800-1806.
 49. Liu S, Shi L, Cheng YB, Fan GX, Ren HX, Yuan YK. Evaluation of protective effect of multi-epitope DNA vaccine encoding six antigen segments of *Toxoplasma gondii* in mice. *Parasitol Res* 2009; 105: 267-274.
 50. Fachado A, Rodriguez A, Angel SO, Pinto DC, Vila I, Acosta A, Amendoeira RR, Lannes-Vieira J. Protective effect of a naked DNA vaccine cocktail against lethal toxoplasmosis in mice. *Vaccine* 2003; 21: 1327-1335.
 51. Ivory C, Chadee K. DNA vaccines: designing strategies against parasitic infections. *Genet Vaccines Ther* 2004; 2: 17.
 52. Parthasarathy S, Fong MY, Ramaswamy K, Lau YL. Protective immune response in BALB/c mice induced by DNA vaccine of the ROP8 gene of *Toxoplasma gondii*. *Am J Trop Med Hyg* 2013; 88: 883-887.
 53. Gong P, Huang X, Yu Q, Li Y, Huang J, Li J, Yang J, Li H, Zhang G, Ren W, Zhang X. The protective effect of a DNA vaccine encoding the *Toxoplasma gondii* cyclophilin gene in BALB/c mice. *Parasite Immunol* 2013; 35: 140-146.
 54. Sun XM, Zou J, AA ES, Yan WC, Liu XY, Suo X, Wang H, Chen QJ. DNA vaccination with a gene encoding *Toxoplasma gondii* GRA6 induces partial protection against toxoplasmosis in BALB/c mice. *Parasit Vectors* 2011; 4: 213.
 55. Wang PY, Yuan ZG, Petersen E, Li J, Zhang XX, Li XZ, Li HX, Lv

- ZC, Cheng T, Ren D, Yang GL, Lin RQ, Zhu XQ. Protective efficacy of a *Toxoplasma gondii* Rhopty Protein 13 plasmid DNA vaccine in mice. *Clin Vaccine Immunol* 2012; 19: 1916-1920.
56. Yuan ZG, Zhang XX, He XH, Petersen E, Zhou DH, He Y, Lin RQ, Li XZ, Chen XL, Shi XR, Zhong XL, Zhang B, Zhu XQ. Protective immunity induced by *Toxoplasma gondii* rhopty protein 16 against toxoplasmosis in mice. *Clin Vaccine Immunol* 2011; 18: 119-124.
57. Yuan ZG, Zhang XX, Lin RQ, Petersen E, He S, Yu M, He XH, Zhou DH, He Y, Li HX, Liao M, Zhu XQ. Protective effect against toxoplasmosis in mice induced by DNA immunization with gene encoding *Toxoplasma gondii* ROP18. *Vaccine* 2011; 29: 6614-6619.
58. Li J, Han Q, Gong P, Yang T, Ren B, Li S, Zhang X. *Toxoplasma gondii* rhomboid protein 1 (TgROM1) is a potential vaccine candidate against toxoplasmosis. *Vet Parasitol* 2012; 184: 154-160.
59. Yan HK, Yuan ZG, Petersen E, Zhang XX, Zhou DH, Liu Q, He Y, Lin RQ, Xu MJ, Chen XL, Zhong XL, Zhu XQ. *Toxoplasma gondii*: protective immunity against experimental toxoplasmosis induced by a DNA vaccine encoding the perforin-like protein 1. *Exp Parasitol* 2011; 128: 38-43.
60. Zhang NZ, Huang SY, Zhou DH, Chen J, Xu Y, Tian WP, Lu J, Zhu XQ. Protective immunity against *Toxoplasma gondii* induced by DNA immunization with the gene encoding a novel vaccine candidate: calcium-dependent protein kinase 3. *BMC Infect Dis* 2013; 13: 512.
61. Zhang M, Zhao L, Song J, Li Y, Zhao Q, He S, Cong H. DNA vaccine encoding the *Toxoplasma gondii* bradyzoite-specific surface antigens SAG2CDX protect BALB/c mice against type II parasite infection. *Vaccine* 2013; 31: 4536-4540.
62. Yu L, Yamagishi J, Zhang S, Jin C, Aboge GO, Zhang H, Zhang G, Tanaka T, Fujisaki K, Nishikawa Y, Xuan X. Protective effect of a prime-boost strategy with plasmid DNA followed by recombinant adenovirus expressing TgAMA1 as vaccines against *Toxoplasma gondii* infection in mice. *Parasitol Int* 2012; 61: 481-486.
63. Meng M, Zhou A, Lu G, Wang L, Zhao G, Han Y, Zhou H, Cong H, Zhao Q, Zhu XQ, He S. DNA prime and peptide boost immunization protocol encoding the *Toxoplasma gondii* GRA4 induces strong protective immunity in BALB/c mice. *BMC Infect Dis* 2013; 13: 494.
64. Zhao H, Huang FY, Guo JL, Tan GH. Evaluation on the immune response induced by DNA vaccine encoding MIC8 co-immunized with IL-12 genetic adjuvant against *Toxoplasma gondii* infection. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi (Chinese J Parasitol Parasit Dis)* 2013; 31: 284-289.
65. Chen J, Huang SY, Zhou DH, Li ZY, Petersen E, Song HQ, Zhu XQ. DNA immunization with eukaryotic initiation factor-2 α of *Toxoplasma gondii* induces protective immunity against acute and chronic toxoplasmosis in mice. *Vaccine* 2013; 31: 6225-6231.
66. Cui X, Lei T, Yang D, Hao P, Li B, Liu Q. *Toxoplasma gondii* immune mapped protein-1 (TgIMP1) is a novel vaccine candidate against toxoplasmosis. *Vaccine* 2012; 30: 2282-2287.
67. Zhang G, Huong VT, Battur B, Zhou J, Zhang H, Liao M, Kawase O, Lee EG, Dautu G, Igarashi M, Nishikawa Y, Xuan X. A heterologous prime-boost vaccination regime using DNA and a vaccinia virus, both expressing GRA4, induced protective immunity against *Toxoplasma gondii* infection in mice. *Parasitology* 2007; 134: 1339-1346.
68. Taylor DN, Tacket CO, Losonsky G, Castro O, Gutierrez J, Meza R, Nataro JP, Kaper JB, Wasserman SS, Edelman R, Levine MM, Cryz SJ. Evaluation of a bivalent (CVD 103-HgR/CVD 111) live oral cholera vaccine in adult volunteers from the United States and Peru. *Infect Immun* 1997; 65: 3852-3856.
69. Virnik K, Hockenbury M, Ni Y, Beren J, Pavlakis GN, Felber BK, Berkower I. Live attenuated rubella vectors expressing SIV and HIV vaccine antigens replicate and elicit durable immune responses in rhesus macaques. *Retrovirology* 2013; 10: 99.
70. Toussaint B, Chauchet X, Wang Y, Polack B, Le Gouvellec A. Live-attenuated bacteria as a cancer vaccine vector. *Expert Rev Vaccines* 2013; 12: 1139-1154.
71. Detmer A, Glenting J. Live bacterial vaccines-a review and identification of potential hazards. *Microb Cell Fact* 2006; 5: 23.
72. Mendes ÉA, Fonseca FG, Casério BM, Colina JP, Gazzinelli RT, Caetano BC. Recombinant vaccines against *T. gondii*: comparison between homologous and heterologous vaccination protocols using two viral vectors expressing SAG1. *PLoS One* 2013; 8: e63201.
73. Cong H, Gu QM, Jiang Y, He SY, Zhou HY, Yang TT, Li Y, Zhao QL. Oral immunization with a live recombinant attenuated *Salmonella typhimurium* protects mice against *Toxoplasma gondii*. *Parasite Immunol* 2005; 27: 29-35.
74. Liu Q, Gao S, Jiang L, Shang L, Men J, Wang Z, Zhai Y, Xia Z, Hu R, Zhang X, Zhu XQ. A recombinant pseudorabies virus expressing TgSAG1 protects against challenge with the virulent *Toxoplasma gondii* RH strain and pseudorabies in BALB/c mice. *Microbes Infect* 2008; 10: 1355-1362.
75. Nie H, Fang R, Xiong BQ, Wang LX, Hu M, Zhou YQ, Zhao JL. Immunogenicity and protective efficacy of two recombinant pseudorabies viruses expressing *Toxoplasma gondii* SAG1 and MIC3 proteins. *Vet Parasitol* 2011; 181: 215-221.
76. Mack DG, Johnson JJ, Roberts F, Roberts CW, Estes RG, David C, Grumet CF, McLeod R. HLA-class II genes modify outcome of *Toxoplasma gondii* infection. *Int J Parasitol* 1999; 29: 1351-1358.
77. Suzuki Y, Wong SY, Grumet FC, Fessel J, Montoya JG, Zolopa AR, Portmore A, Schumacher-Perdreau E, Schrappe M, Koppen S, Ruf B, Brown BW, Remington JS. Evidence for genetic regulation of susceptibility to toxoplasmic encephalitis in AIDS patients. *J Infect Dis* 1996; 173: 265-268.
78. Dubey JP, Ferreira LR, Martins J, McLeod R. Oral oocyst-induced mouse model of toxoplasmosis: effect of infection with *Toxoplasma gondii* strains of different genotypes, dose, and mouse strains (transgenic, out-bred, in-bred) on pathogenesis and mortality. *Parasitology* 2012; 139: 1-13.
79. Bruna-Romero O, de Oliveira DM, de Andrade-Neto VF. Toxoplasmosis: Advances and Vaccine Perspectives. In Rodriguez-Morales AJ, eds., *Current Topics in Tropical Medicine*. Rijeka, Croatia

- tia. InTech. 2012.
80. Yarovinsky F. Innate immunity to *Toxoplasma gondii* infection. *Nat Rev Immunol* 2014; 14: 109-121.
81. Beghetto E, Spadoni A, Buffolano W, Del Pezzo M, Minenkova O, Pavoni E, Pucci A, Cortese R, Felici F, Gargano N. Molecular dissection of the human B-cell response against *Toxoplasma gondii* infection by lambda display of cDNA libraries. *Int J Parasitol* 2003; 33: 163-173.
82. Bhadra R, Gigley JP, Khan IA. The CD8 T-cell road to immunotherapy of toxoplasmosis. *Immunotherapy* 2011; 3: 789-801.
83. Bhadra R, Cobb DA, Khan IA. Donor CD8⁺ T cells prevent *Toxoplasma gondii* de-encystation but fail to rescue the exhausted endogenous CD8⁺ T cell population. *Infect Immun* 2013; 81: 3414-3425.
84. Kang H, Remington JS, Suzuki Y. Decreased resistance of B cell-deficient mice to infection with *Toxoplasma gondii* despite unimpaired expression of IFN- γ , TNF- α , and inducible nitric oxide synthase. *J Immunol* 2000; 164: 2629-2634.
85. Sayles PC, Gibson GW, Johnson LL. B cells are essential for vaccination-induced resistance to virulent *Toxoplasma gondii*. *Infect Immun* 2000; 68: 1026-1033.
86. Johnson LL, Sayles PC. Deficient humoral responses underlie susceptibility to *Toxoplasma gondii* in CD4-deficient mice. *Infect Immun* 2002; 70: 185-191.
87. Fu YF, Feng M, Ohnishi K, Kimura T, Itoh J, Cheng XJ, Tachibana H. Generation of a neutralizing human monoclonal antibody Fab fragment to surface antigen 1 of *Toxoplasma gondii* tachyzoites. *Infect Immun* 2011; 79: 512-517.
88. Cha DY, Song IK, Lee GS, Hwang OS, Noh HJ, Yeo SD, Shin DW, Lee YH. Effects of specific monoclonal antibodies to dense granular proteins on the invasion of *Toxoplasma gondii* in vitro and in vivo. *Korean J Parasitol* 2001; 39: 233-240.
89. Tan F, Hu X, Pan CW, Ding JQ, Chen XG. Monoclonal antibodies against nucleoside triphosphate hydrolase-II can reduce the replication of *Toxoplasma gondii*. *Parasitol Int* 2010; 59: 141-146.
90. Ferreira \tilde{A} Jr, Santiago FM, Silva MV, Ferreira FB, Macedo AG Jr, Mota CM, Faria MS, Silva Filho HH, Silva DA, Cunha-Júnior JP, Mineo JR, Mineo TW. Production, characterization and applications for *Toxoplasma gondii*-specific polyclonal chicken egg yolk immunoglobulins. *PLoS ONE* 2012; 7: e40391.
91. Larsson A, Balow RM, Lindahl TL, Forsberg PO. Chicken antibodies: taking advantage of evolution-a review. *Poult Sci* 1993; 72: 1807-1812.

