

Serological and Genetic Diversity of Capsular Polysaccharides in *Enterococcus faecalis*

Markus Hufnagel,^{1,2} Lynn E. Hancock,^{3†} Stefanie Koch,¹ Christian Theilacker,¹
Michael S. Gilmore,^{3,4} and Johannes Huebner^{1,5*}

Channing Laboratory, Department of Medicine, Brigham & Women's Hospital, Harvard Medical School,¹ and Division of Infectious Diseases, Children's Hospital, Harvard Medical School,⁵ Boston, Massachusetts; University Children's Hospital, University Hospital Schleswig-Holstein, Campus Kiel, Kiel, Germany²; and Department of Microbiology and Immunology³ and Department of Ophthalmology,⁴ University of Oklahoma Health Sciences Center, Oklahoma City, Oklahoma

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Enterococci possess capsular carbohydrate antigens that are targets of opsonic antibodies. These antigens may be used to develop alternative options for the treatment and prevention of enterococcal infections. The present study was done to analyze the diversity of capsular polysaccharides in *Enterococcus faecalis*. Four type-specific sera were used in an enzyme-linked immunosorbent assay format to detect polysaccharide antigen extracted from bacterial cell walls. A total of 55% of a collection of 29 *E. faecalis* strains could be grouped into one of four serogroups. Additional analysis of the strains by opsonophagocytic assays revealed agreement between the results of the two methods for 72% of the isolates. An additional four strains could be assigned to a serogroup on the basis of opsonic killing by sera with antibodies against the four prototypes strains, provisionally named CPS-A to CPS-D. The results of the two methods disagreed for one strain (4%). When the results of both methods were combined, 66% of the strains could be classified. One strain had to be assigned to two serogroups. The assignments to the four serogroups were confirmed by analysis of the genetic organization of the biosynthetic capsular polysaccharide (*cps*) locus. All strains grouped into serotypes CPS-A and CPS-B possessed only the *cpsA* and *cpsB* genes, while all strains grouped into serogroups CPS-C and CPS-D possessed an additional eight or nine genes. Our results suggest the existence of a limited number of *E. faecalis* capsule serotypes, and we provisionally propose four serotypes, named CPS-A to CPS-D, and the respective prototype strains for these families.

Enterococci are one of the leading causes of infections in hospitalized patients and the third most common cause of nosocomial bloodstream infections in the United States (6–8, 20). The number of strains resistant to virtually all clinically available antibiotics is increasing (20). Recent studies demonstrate that up to 25% of enterococcal hospital isolates are resistant to glycopeptide antibiotics (1, 20), the mainstay of the therapeutic armory. Resistant enterococcal strains cause significant morbidity and mortality, especially among immunocompromised patients.

Huebner et al. (16) recently identified a capsular polysaccharide in an *Enterococcus faecalis* strain and a vancomycin-resistant *E. faecium* strain that is the target of opsonic antibodies. Compositional and structural analyses revealed a teichoic acid-like molecule. Antibodies raised against this antigen protected mice in an experimental model of systemic enterococcal infection (15), thus offering new options for treatment and prevention. In 2002, Hancock and colleagues (10, 12) identified a genetic locus involved in the biosynthesis of a cell wall polysaccharide of *E. faecalis* strain FA2-2. This strain reacts with a type-specific antiserum raised against Maekawa

E. faecalis strain type 2 (19). The corresponding *cps* locus consists of 11 open reading frames.

Some clues to the diversity of *E. faecalis* capsular polysaccharides can be gleaned from previous attempts to establish serotyping systems (19, 24), although it is not known at present how many structurally different capsular polysaccharides exist. Sharpe and Shattok (24) used HCl extracts of bacteria to raise rabbit sera with antibodies against surface polysaccharides. Their serotyping scheme is likely based on polysaccharide antigens, but considerable taxonomic changes subsequent to development of the scheme (19) limited its applicability. Maekawa et al. (19) used formaldehyde-killed bacteria to immunize rabbits against 21 prototype strains. Their serogroups are probably based not only on polysaccharide antigens but on a number of different surface antigens as well.

With an interest in the ultimate development of immunotherapeutics, we undertook a study to reexamine the diversity of *E. faecalis* capsules using strains identified by modern taxonomy and to compare the serological diversity observed with the organization of the genes in the recently identified *E. faecalis* capsule locus (10).

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MATERIALS AND METHODS

Strains, culture, and sera. The bacterial strains and sera used in these studies are described in Tables 1 and 2 (10, 15–17, 19, 21). *E. faecalis* 12030 is the only *E. faecalis* strain for which structural information on the capsular polysaccharide

* Corresponding author. Mailing address: Channing Laboratory, Brigham and Women's Hospital, 181 Longwood Ave., Boston, MA 02115. Phone: (617) 525-2645. Fax: (617) 731-1541. E-mail: jhuebner@channing.harvard.edu.

† Present address: Department of Molecular and Experimental Medicine, The Scripps Research Institute, La Jolla, Calif.

TABLE 1. *E. faecalis* strains used in the present study

Strain	Origin (reference or source)
12030	Clinical isolate (16)
12107	Clinical isolate (15)
type 1-21	Prototype Japanese strains from Maekawa et al. (19)
FA2-2	Laboratory strain
HG101	<i>cpsI</i> mutant of FA2-2 (10)
OG1-RF	Laboratory strain (ATCC 47077)
V583	Vancomycin-resistant clinical strain (ATCC 700802)
MMH 594	Epidemic strain (17)
R19.001	Epidemic strain from Children's Hospital, Boston, Mass. (21)
68114	Clinical isolate

exists (16, 27). Strain *E. faecalis* 12107 (15) is a clinical isolate from the United States, and the 21 prototype *E. faecalis* strains previously described by Maekawa et al. (19) are of *E. faecalis* types 2 and 5. HG101 is an insertion mutant of laboratory strain FA2-2 that is unable to produce the capsular polysaccharide (10). All *E. faecalis* strains were grown without agitation at 37°C either in Todd-Hewitt broth (Becton Dickinson, Sparks, Md.) or, for the capsular polysaccharide-specific enzyme-linked immunosorbent assay (CPS-ELISA), in Columbia broth (Difco Laboratories, Detroit, Mich.) with the addition of 0.5% glucose. Erythromycin (Sigma Chemical Co., St. Louis, Mo.) was added at a concentration of 10 µg/ml for mutant strain HG101.

Sera with antibodies against the purified capsular polysaccharides of strains *E. faecalis* 12030 (15) and *E. faecalis* 12107 (16) were prepared as described elsewhere (15). In addition, sera with antibodies against two of the prototype strains described by Maekawa et al. (19) (*E. faecalis* Maekawa type 2 and *E. faecalis* Maekawa type 5) were prepared by using heat-killed and proteinase K-treated whole bacterial cells. New Zealand White rabbits were injected with two doses of 100 µg of purified polysaccharide antigen subcutaneously and three doses of 10 µg of purified polysaccharide antigen intravenously as described previously (15). Immunization with whole bacterial cells (i.e., *E. faecalis* type 2 and type 5) was done by using doses of 3 × 10⁸ bacteria injected intravenously three times a week for 5 weeks. The bacterial cells were heat killed (70°C for 1 h) and digested with proteinase K (0.1 mg/ml at 56°C for 1 h; Invitrogen, Carlsbad, Calif.) prior to intravenous injection. Rabbits were bled 5 weeks after the last injection and periodically for 20 weeks without boosting. The resulting four groups of serum samples, representing four serologically distinct families, were used in a newly developed capsular polysaccharide-specific ELISA (CPS-ELISA) and in opsonophagocytic assays.

CPS-ELISA. Crude polysaccharide extracts were prepared from the enterococcal strains. Bacteria were grown overnight at 37°C to stationary phase in 50 ml of Columbia broth supplemented with 0.5% glucose. The cell pellet was suspended in 500 µl of 10% trichloroacetic acid (TCA) and incubated on a rotor rack at 4°C for 18 to 24 h (a modification of the procedure of Heckels and Virji [13]). The suspension was then centrifuged at 12,000 rpm for 10 min (Hettich Micro 20 instrument), and the supernatant was precipitated with 1.25 ml of ethanol overnight at -20°C. After centrifugation (10 min at 12,000 rpm) the supernatant was discarded and the pellet was dissolved in 1 ml of distilled H₂O. Microtiter plates (96 wells; Immulon 2HB; Dynex Technologies, Chantilly, Va.) were coated overnight with 80 µl of antigen solution along with 20 µl of 5× sensitizing buffer (180 mg of NaH₂PO₄ and 1.7 g of NaHPO₄ per 100 ml) at 4°C. The plates were washed three times with phosphate-buffered saline (PBS), blocked with 3% skim milk in PBS for 1 h, and again washed three times with PBS. The primary antibody was applied in dilutions of 1:100 and 1:500, and the plates were incubated for 60 min at 37°C. In order to eliminate cross-reactivity and to improve specificity, all four prototype groups of sera were absorbed with either one or a combination of three different heterologous *E. faecalis* strains (Table 2). All serum samples were absorbed at a 1:20 dilution with 2 × 10⁹ bacteria per ml at 4°C for 60 min on a rotor rack. This procedure was done three times for every serum sample. After an additional washing step, the secondary antibody (goat anti-rabbit alkaline phosphatase-conjugated immunoglobulin G antibody; ICN Biomedicals, Aurora, Ohio) was applied at a dilution of 1:1,000 and the mixture was incubated for 60 min at 37°C. After the plates were washed with PBS, the detection reagent *p*-nitrophenyl-phosphate (Sigma Chemical Co.) was added. The plates were read at 405 nm after 15, 30, 60, and 90 min in an EL 309 ELISA reader (Bio-Tek Instruments, Winooski, Vt.). Each strain was tested at least in duplicate. Both positive and negative controls were included on all plates.

Characterization of TCA extracts. For quantitative measurement of the protein contents of TCA extracts, a modified dye-binding assay of Bradford (9) was used. For quantitative estimation of the carbohydrate contents of TCA extracts, the phenol-sulfuric acid method was performed (9).

Opsonophagocytic assays. Opsonophagocytic assays with all enterococcal strains were performed as described elsewhere (16). Each bacterial strain was tested with the four groups of unabsorbed prototype sera. In addition, the sera were absorbed either with the homologous, immunizing strain or with single or multiple combinations of heterologous prototype strains (Table 2). Samples without serum and samples with normal rabbit serum (NRS) were used as controls, and each experiment was performed at least in duplicate. The percent killing was calculated by comparing the inoculum (viable counts at time zero [*T*₀]) with the colony counts obtained after incubation for 90 min at 37°C (*T*₉₀) on a rotor rack by use of the following formula: [(mean CFU at *T*₀ - mean CFU at *T*₉₀)/(mean CFU at *T*₀)] × 100.

Immunoelectron microscopy. Immunoelectron microscopy was performed with the four prototype strains, *E. faecalis* 12030, 12107, type 2, and type 5, as described elsewhere (16). The four groups of unabsorbed sera (data not shown) and all four groups of sera absorbed either with the homologous strain or with each of the three heterologous strains were used as primary antibodies.

PCR analysis of *cps* locus. The primers and PCR conditions used to amplify the 11 genes of the *cps* locus have been described elsewhere (10).

Restriction fragment length polymorphism (RFLP) analysis of the *cps* locus. Southern blot hybridization analysis of chromosomal DNA digested with different restriction enzymes (Invitrogen) was performed as described elsewhere (10).

RESULTS

Generation of capsule-specific antisera. Four groups of rabbit sera with antibodies against enterococcal surface polysaccharide antigens were prepared to establish a system for the serotyping of *E. faecalis*. Each of the four groups of sera reacted specifically with the homologous strain (Fig. 1), outlining a capsule-like structure in all cases. Absorption of the respective serum group with the corresponding homologous strain abolished the visualization of the capsule-like structure (Fig. 1, second column), whereas absorption of the four groups of sera with any of the three heterologous strains had no effect on the ability to visualize the capsule (Fig. 1, columns three to five). This indicates that all four groups of sera reacted with antigens present on the surface of the bacterial cells and that each type of serum bound specifically to its prototype strain. Only low levels of binding of gold-labeled protein A were observed when NRS was used as a negative control (data not shown).

Type-specific reactivity of antisera. Three of the groups of absorbed prototype sera (i.e., α12030, α12107, and αType 5; Table 2) reacted only with the corresponding prototype strain by CPS-ELISA (Table 3). The type 2-specific serum reacted with both *E. faecalis* type 2 and *E. faecalis* type 5 antigens, indicating cross-reactivity between the two strains (Table 3).

TABLE 2. Sera and serum absorptions used in the present study

Antigen against which immune rabbit serum was directed	<i>E. faecalis</i> strain(s) used for absorption in CPS-ELISA	Homologous and heterologous strains used for absorption in opsonophagocytic assay
Strain 12030 polysaccharide	Type 20	12030, ^a 12107 ^b
Strain 12107 polysaccharide	68114	12107, ^a 12030 ^b
Type 2 whole bacteria (heat and proteinase K treated)	12107	Type 2, ^a 12030 ^b and 12107 ^b
Type 5 whole bacteria (heat and proteinase K treated)	12030, 12107, type 2	Type 5, ^a 12030 ^b and 12107 ^b

^a Homologous strain.

^b Heterologous strain.

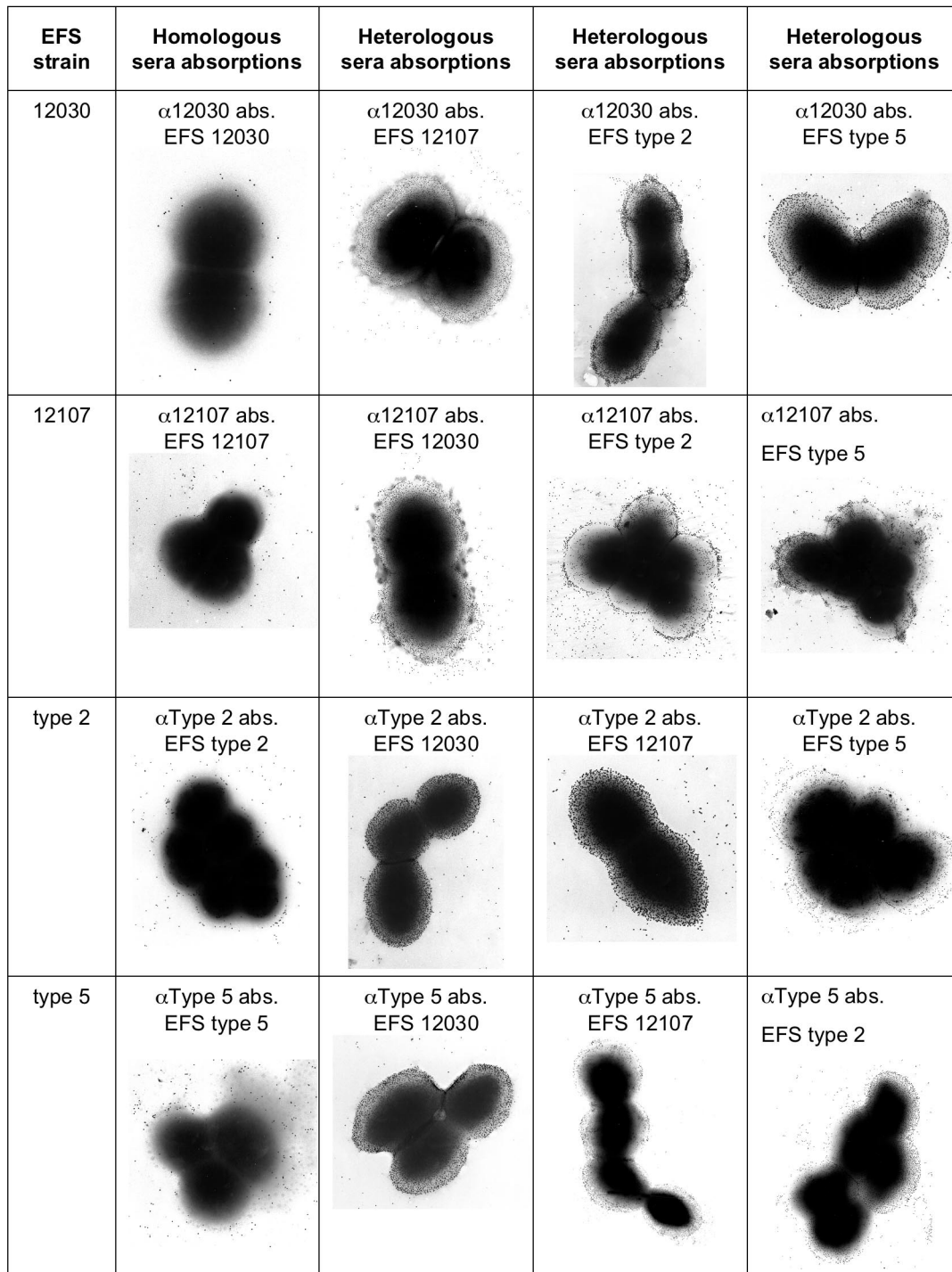


FIG. 1. Immunoelectron microscopy of the four prototype *E. faecalis* (EFS) strains labeled with the prototype serum absorbed with either the homologous strain or the three heterologous strains.

This cross-reactivity was further substantiated by the testing of the two strains in an absorption CPS-ELISA (Fig. 2). The two groups of sera were absorbed with the respective homologous strains as well as with each of the three heterologous strains. Binding of both groups of sera was reduced by absorption with the homologous strain (84% reduction for the type 5-specific serum, 91% reduction for the type 2-specific serum). Absorp-

tion of the two groups of sera with the respective heterologous strain, *E. faecalis* type 2 or *E. faecalis* type 5, led to a higher degree of reduction (57% reduction for the type 5-specific serum, 83% reduction for the type 2-specific serum) than absorption with the heterologous strains, *E. faecalis* 12030 or *E. faecalis* 12107 (levels of reduction, between 11 and 22%). Despite the cross-reactivity of the type 2-specific serum be-

TABLE 3. Results of CPS-ELISA with TCA extracts of 14 *E. faecalis* strains

Proposed serogroup	<i>E. faecalis</i> strain	OD ₄₀₅ ^a			
		α12030	α12107	αType 2	αType 5
CPS-A	12030	2.56	0.772	0.456	0.324
	Type 7	1.855	0.495	0.459	0.253
CPS-B	12107	0.232	2.529	0.501	0.356
	Type 1	0.275	2.232	0.843	0.306
	OG1-RF	0.319	2.263	0.489	0.285
CPS-C	Type 2	0.586	0.384	2.444	0.243
	FA2-2	0.469	0.546	1.86	0.818
	Type 12	0.254	0.306	2.029	0.218
	R19.001	0.288	0.642	2.402	0.33
	V583	0.42	0.718	2.476	0.514
CPS-D	Type 5	0.222	0.482	1.128	1.512
	Type 18	0.523	0.482	0.976	1.305
CPS-C/D	MMH 594	0.184	0.337	2.388	2.396
NT ^b	Type 9	0.347	0.775	1.4	0.515

^a The OD₄₀₅s of the four prototype groups of sera for the respective prototype strain are shown in boldface and italic. Boldface alone indicates the assignment of the strain to one of the four prototype strain types.

^b NT, nontypeable.

tween *E. faecalis* type 2 and *E. faecalis* type 5, these two strains were distinguished in the CPS-ELISA. Type 2-related strains showed high levels of reactivity (as determined by measurement of the optical density at 405 nm [OD₄₀₅]) with the type 2-specific serum but low levels of reactivity with the type 5-specific serum. In contrast, type 5-related strains reacted with both types of sera, with the strains having higher levels of reactivity with type 5-specific serum than type 2-specific serum (134 and 100%, respectively; Table 3).

Diversity of binding of antisera to capsular polysaccharides in the CPS-ELISA. The assignment of a test strain to a prototype strain was made if the OD₄₀₅ was at least 70% of the value obtained with the corresponding prototype strain (Table 3). This threshold was chosen on the basis of the assumption that the reactivity of a related strain with one of the prototype groups of sera should exceed the cross-reactivity of the other three heterologous prototype strains with the respective groups of sera. The following formula was used to obtain a threshold value: sum of the OD₄₀₅s of all four groups of sera reacting with all three heterologous strains divided by the sum of the OD₄₀₅s of all four groups of sera reacting with the respective homologous strains. A value of 62.9% was obtained by this

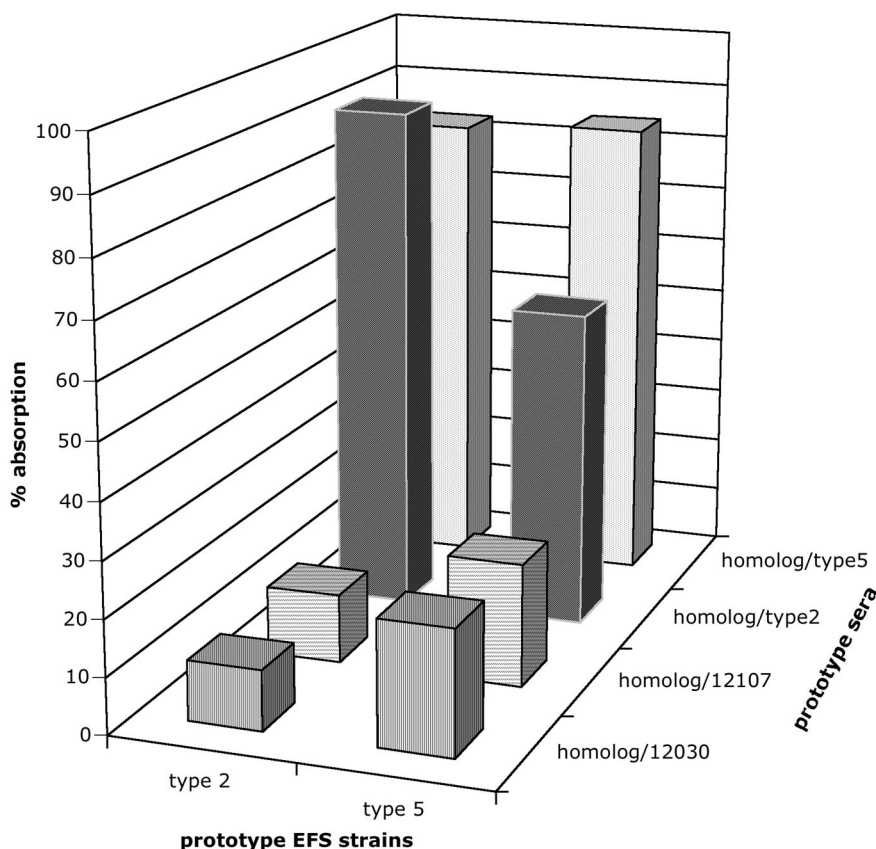


FIG. 2. Absorption CPS-ELISA of *E. faecalis* (EFS) type 2 and type 5 prototype strains with the two corresponding prototype sera. The two types of sera were absorbed either with the respective homologous strain or with each of the three heterologous strains. The percent absorption values reflect the ratio of the amount of absorbed sera to the amount of unabsorbed sera. Both types of sera could be absorbed with the homologous strain (84 and 91%, respectively). Absorption of both types of sera with heterologous strains *E. faecalis* type 2 and *E. faecalis* type 5, respectively, led to a higher degree of absorption (57 and 83%, respectively) than absorption with the heterologous strains *E. faecalis* 12030 and *E. faecalis* 12107 (range, 11 to 22%).

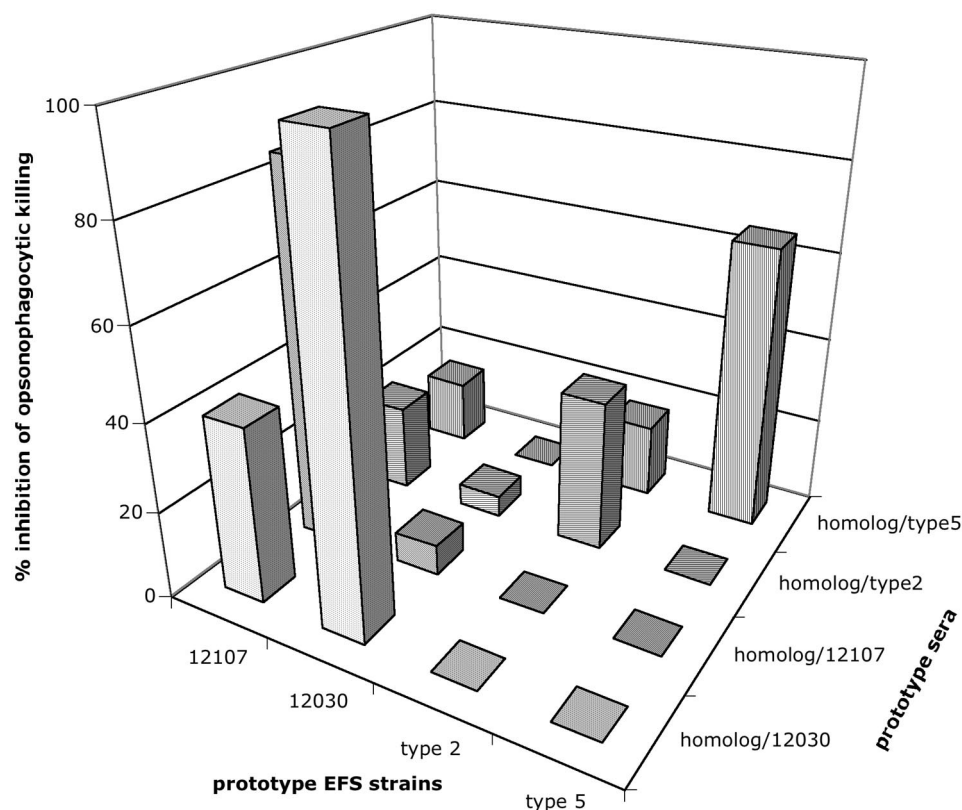


FIG. 3. Phagocytic inhibition assay of the four prototype groups of sera against the four prototype *E. faecalis* (EFS) strains. Each prototype serum was tested against its homologous prototype *E. faecalis* strain by absorption with the respective homologous strain and with the three heterologous strains. The percent inhibition values reflect the ratio of the amount of absorbed sera to the amount of unabsorbed sera. The killing activities of the four types of sera were most effectively inhibited by absorption with the homologous strain (100% for *E. faecalis* 12030, 84% for *E. faecalis* 12107, 66% for *E. faecalis* type 5). Type 2-specific sera could not be absorbed to the same degree (absorption, 34%). Inhibition of killing activity by heterologous strains was greatly reduced for all four prototype sera, with cross-reactivity between strains *E. faecalis* 12030 and *E. faecalis* 12107.

formula. To decrease the error margin, a 70% cutoff was finally chosen as the assignment criterion.

A panel of 29 different *E. faecalis* strains was tested by the CPS-ELISA. By using the criteria described above, 55% (i.e., 16 of 29) of the strains tested could be classified into one of the four serotypes. One strain (*E. faecalis* MMH 594) reacted with two groups of sera (i.e., type 2- and type 5-specific sera), and thus, 41% (12 of 29) of the strains could not be classified unambiguously by the CPS-ELISA.

Diversity of capsular polysaccharides in opsonophagocytic killing. To confirm the findings obtained by CPS-ELISA, all strains were analyzed by opsonophagocytic assays that tested for a clinically relevant biological function, i.e., the opsonic killing activity of the sera. To assess the specificity of the killing, the sera were absorbed with the respective homologous strain as well as with different heterologous strains (Table 2). Isolates were assigned to one of the four serotypes if (i) opsonic killing by a prototype serum was greater than 70% and (ii) the killing was abolished by more than 50% by absorption with the homologous strain but not to the same degree by absorption with the heterologous strains.

Again, the three prototypes strains, *E. faecalis* 12030, *E. faecalis* 12107, and *E. faecalis* type 5, were most effectively killed and, according to the assignment criterion, were also specifically killed by their homologous sera (Fig. 3). Type 2-specific

serum killed both *E. faecalis* type 2 and *E. faecalis* type 5, supporting the findings of cross-reactivity between these two strains. Ten of the 29 (34%) *E. faecalis* strains could be unambiguously assigned to one of the four serotypes. For a further 24% (7 of 29) of the strains, the opsonic killing activity was not able to distinguish between type 2- or type 5-related strains. One strain (Maekawa strain type 7) was killed to the same degree by both strain 12030- and strain 12107-specific sera. The overall agreement between the CPS-ELISA and the opsonophagocytic assay was 72% (21 of 29 strains). Seven (24%) strains that were nontypeable by one assay could be assigned to a serogroup by the other test. Only one strain (Maekawa strain type 6 strain) gave conflicting results by the two assays.

When the results of the two assays are taken together, 66% (19 of 29) of the *E. faecalis* strains could be assigned to one of the four serogroups. Two additional strains were grouped into two serogroups (types 2 and 5; Table 4).

Genetic diversity of capsular polysaccharides. The assignment of a given strain to a serogroup was correlated with the genetic organization of the *cps* locus, shown by Hancock and Gilmore (10) to be involved in the biosynthesis of capsular polysaccharides. Representative strains of the four serogroups were chosen, and PCR was used to test for the presence of the 11 *cps* genes (Table 5). Two different patterns could be shown: *E. faecalis* strains of the strain 12030- and 12107-related groups

TABLE 4. Proposal for a serotyping system for *E. faecalis* based on capsular polysaccharides and their relationship with the *cps* locus^a

Proposed serogroup	<i>E. faecalis</i> strain	Homology by:		RFLP pattern of CPS locus
		CPS-ELISA	Opsonophagocytic assay	
CPS-A	12030	12030	12030	I
	Type 3		12030	I
	Type 7	12030	12030 and 12107	I
CPS-B	12107	12107	12107	I
	OG1-RF	12107	12107	I
	Type 1	12107		I
	Type 10		12107	I
	Type 16		12107	I
	Type 20	12107		I
CPS-C	Type 2	Type 2	Type 2	III
	V583	Type 2	Type 2 and type 5	III
	FA2-2	Type 2	Type 2 and type 5	III
	R19.001	Type 2	Type 2	ND ^b
	Type 12	Type 2	Type 2 and type 5	II
	Type 14	Type 2	Type 2 and type 5	II
	Type 21		Type 2	II
CPS-D	Type 5	Type 5	Type 2 and type 5	II
	68114	Type 5	Type 2 and type 5	ND
	Type 18	Type 5	Type 2 and type 5	II
CPS-C and CPS-D	MMH 594	Type 2 and type 5		III
	Type 6	Type 5	Type 2	II
NT ^c	Type 4			I
	Type 8			III
	Type 9			I
	Type 11			III
	Type 13			I
	Type 15			I
	Type 17			I
	Type 19			I

^a *E. faecalis* strains were assigned to one of the four capsular polysaccharide serotypes (CPS-A to CPS-D) if the definition criteria for one of the two diagnostic assays were met or if one assay eliminated cross-reactivity between two serotypes in the second assay. Boldface indicates the prototype strain.

^b ND, not determined.

^c NT, nontypeable.

consistently possessed only the first two genes of the *cps* locus (i.e., *cpsA* and *cpsB*), while all strains of the type 2- and type 5-related groups had the *cpsA* to *cpsE* and the *cpsG* to *cpsK* genes. The *cpsF* gene was consistently missing from the type 5-related strains and was variably present in the type 2-related strains. No specific pattern could be observed for the nontypeable strains. The results of the PCR were verified by Southern blot hybridization analysis. Hybridization analysis of DNA digested with ClaI, using the 14.8-kb *cpsA* to *cpsK* fragment amplified from strain V583 as a probe, demonstrated various degrees of conservation among the isolates examined. Strains belonging to the *E. faecalis* 12030 and *E. faecalis* 12107 groups all showed an RFLP class I pattern, with only a single hybridization band, the size of which corresponds to those of the *cpsA* and *cpsB* genes, as well as a conserved region downstream of *cpsK* (Fig. 4). *E. faecalis* type 5 strains yielded an RFLP class II pattern and were more closely related to class III strains than to the other classes of strains, with the class II and class III strains appearing to differ by only a single ClaI restriction fragment.

The difference between RFLP class II and III strains is likely attributable to the absence of the *cpsF* gene in the class II strains. The 5.7-kb ClaI fragment, which spans the 3' end of *cpsE* through the 5' end of *cpsJ* and which is present in class III strains, is absent from class II strains. However, a conserved 4.7-kb ClaI fragment from class II strains hybridizes to the probe (Fig. 4). An apparent ClaI polymorphism was observed for class II strains. With the exception of the 2.4-kb ClaI fragment, *E. faecalis* type 5 and type 6 strains have ClaI restriction patterns identical to those of *E. faecalis* types 12, 14, and 18. In place of the 2.4-kb fragment, *E. faecalis* type 5 has a slightly smaller ClaI fragment of approximately 1.8 kb, whereas *E. faecalis* type 6 has a much larger ClaI fragment of 6.0 kb (Fig. 4). *E. faecalis* class III strains previously identified as being reactive with type 2-specific serum (10), namely, *E. faecalis* FA2-2, *E. faecalis* V583, and *E. faecalis* MMH 594, showed identical hybridization patterns with the *E. faecalis* type 2 prototype strain (Fig. 4).

TABLE 5. PCR analysis of the *cps* locus in a representative collection of *E. faecalis* strains

Proposed serogroup	<i>E. faecalis</i> strain	Result for gene:										
		<i>cpsA</i>	<i>cpsB</i>	<i>cpsC</i>	<i>cpsD</i>	<i>cpsE</i>	<i>cpsF</i>	<i>cpsG</i>	<i>cpsH</i>	<i>cpsI</i>	<i>cpsJ</i>	<i>cpsK</i>
CPS-A	12030	+	+	-	-	-	-	-	-	-	-	-
	Type 7	+	+	-	-	-	-	-	-	-	-	-
CPS-B	12107	+	+	-	-	-	-	-	-	-	-	-
	Type 1	+	+	-	-	-	-	-	-	-	-	-
	OG1-RF	+	+	-	-	-	-	-	-	-	-	-
CPS-C	Type 2	+	+	+	+	+	+	+	+	+	+	+
	FA2-2	+	+	+	+	+	+	+	+	+	+	+
	Type 12	+	+	+	+	+	-	+	+	+	+	+
	R19.001	+	+	+	+	+	-	+	+	+	+	+
CPS-D	Type 5	+	+	+	+	+	-	+	+	+	+	+
	Type 18	+	+	+	+	+	-	+	+	+	+	+
CPS-C and CPS-D	MMH 594	+	+	+	+	+	+	+	+	+	+	+
NT ^a	Type 9	+	+	-	-	-	-	-	-	-	-	-

^a NT, nontypeable.

^b +, present; -, absent.

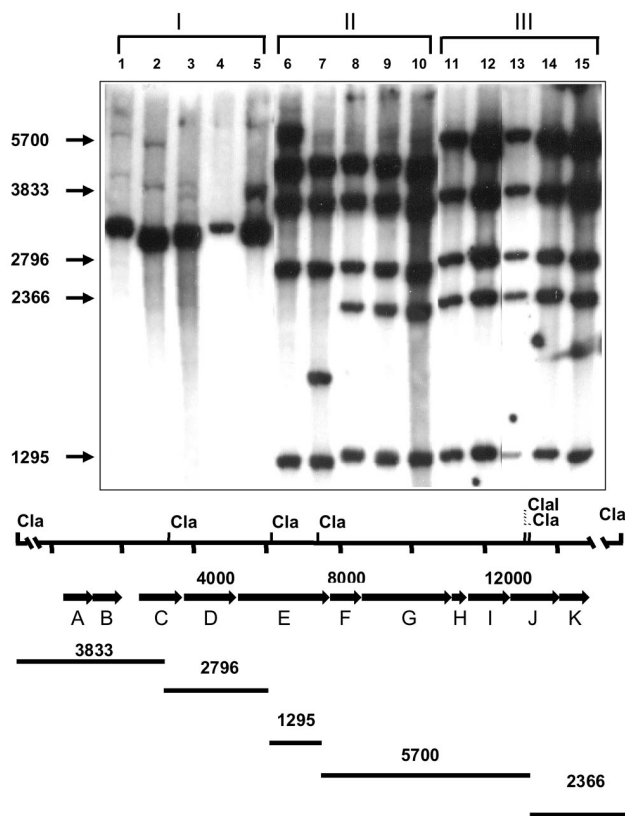


FIG. 4. Southern blot analysis of capsular polysaccharide determinants from various *E. faecalis* strains. DNA was restricted with *Cla*I and probed with the radiolabeled fragment of the *cpsA* to *cpsK* genes from *E. faecalis* strain V583. Lanes 1 to 5, strains with RFLP pattern I; lanes 6 to 10, strains with RFLP pattern II; lanes 11 to 15, strains with RFLP pattern III. Lane 1, *E. faecalis* 12107; lane 2, *E. faecalis* 12030; lane 3, *E. faecalis* type 4; lane 4, *E. faecalis* type 1; lane 5, *E. faecalis* type 7; lane 6, *E. faecalis* type 6; lane 7, *E. faecalis* type 5; lane 8, *E. faecalis* type 18; lane 9, *E. faecalis* type 14; lane 10, *E. faecalis* type 12; lane 11, *E. faecalis* type 8; lane 12, *E. faecalis* type 2; lane 13, *E. faecalis* MMH 594; lane 14, *E. faecalis* V583; lane 15, *E. faecalis* FA2-2. The sizes (in base pairs) of the *E. faecalis* V583 sequence fragments hybridizing to the probe are shown at the left. The *Cla*I restriction map for the *E. faecalis* V583 capsular polysaccharide determinant is shown below. The molecular sizes (in base pairs) of the regions spanned by the *Cla*I sites in *E. faecalis* V583 are listed below the map.

Capsule-specific reactivity of antisera. To confirm the hypothesis that the typing sera reacted with capsular antigens encoded by the *cps* locus, a CPS-ELISA (data not shown) and an opsonophagocytic assay were performed with the acapsular mutant HG101 and its wild-type (WT) capsular strain, *E. faecalis* FA2-2 (Fig. 5). *E. faecalis* FA2-2 reacts specifically with type 2-specific serum (10). NRS was used as a negative control. The WT strain was resistant to killing by NRS, whereas the capsule-negative mutant was highly susceptible to killing by NRS. Type 2-specific serum killed both strains efficiently. Absorption of the serum with the WT strain abolished the killing activity against the WT strain but not the mutant, and absorption of the serum with the mutant had little or no effect on the killing activity of the type 2-specific serum. These data support the hypothesis that the activity of the type 2-specific serum is directed against the capsule produced at least in part by enzymes encoded by the *cps* locus.

DISCUSSION

An enterococcal surface antigen that is the target of opsonic antibodies has previously been isolated from *E. faecalis* 12030 and characterized (16). Compositional and structural analyses of this antigen revealed a teichoic acid composed of a repeating unit of α -D-glucopyranosyl-(1-2)- α -D-glucopyranose (kojibiose) attached to a glycerol diphosphate (16, 27). Passive and active immunotherapies reduced the bacterial counts in a mouse model of bacteremia when the animals were challenged with the homologous strain. This serum also protected mice when they were challenged with a vancomycin-resistant *E. faecium* strain (15). However, it is not known how many structurally and immunologically different capsular polysaccharides that may be used as vaccine candidates exist in enterococci.

Earlier studies by Sharpe (23), Sharpe and Shattok (24), and Maekawa et al. (18, 19) attempted to establish systems for the serotyping of enterococci. The latter scheme was based on 6 of Sharpe's original prototype strains, 2 strains from the Centers for Disease Control and Prevention, and 13 clinical isolates from Japan (19). In order to relate our present findings to those of previous investigators, we included the strain collection of Maekawa et al. (19) in the present effort, since these isolates represent *E. faecalis* strains with a high degree of antigenic diversity (although the diversity is not necessarily based solely on carbohydrate antigens). Maekawa's collection also included the strains from the study of Sharpe (23) and that of Sharpe and Shattok (24).

A CPS-ELISA was developed on the basis of the preferential extraction of cell wall teichoic acids. In the assay, we used sera that were absorbed with different heterologous strains (Table 2). The four prototype strains used in this study were chosen for the following reasons: the capsular polysaccharide structure of *E. faecalis* 12030 has been elucidated (16, 27). The opsonic killing activities of antibodies against the purified capsular polysaccharide from this strain were tested against a collection of clinical isolates, and it was found that 33% of the strains were killed by this serum (16). One of these strains (*E. faecalis* 12107) was not killed by this serum, and preliminary structural analysis of capsular polysaccharide material from this strain indicated that its cell wall teichoic acid structure may be different from that of *E. faecalis* 12030 (unpublished data).

In studies performed by Maekawa and Habadera (18), 30% of urinary tract isolates were of the type 2 serotype. Studies performed contemporaneously by Hancock and Gilmore (10) indicated that the *cps* locus in the 21 Maekawa prototype strains showed three different patterns, with *E. faecalis* 12030 and *E. faecalis* 12107 being indistinguishable by this method. However, two additional patterns were observed, and *E. faecalis* type 2 and *E. faecalis* type 5 were arbitrarily chosen as representatives of the remaining two RFLP patterns, respectively.

The four types of sera used in this study were all directed against capsular polysaccharides and reacted specifically with the respective prototype strain, independently of the method used to raise the sera. Immunoelectron microscopy of the four prototype strains reacting with the respective sera clearly showed that the capsule is the target of the antibodies (Fig. 1). However, *E. faecalis* type 2 and *E. faecalis* type 5 showed cer-

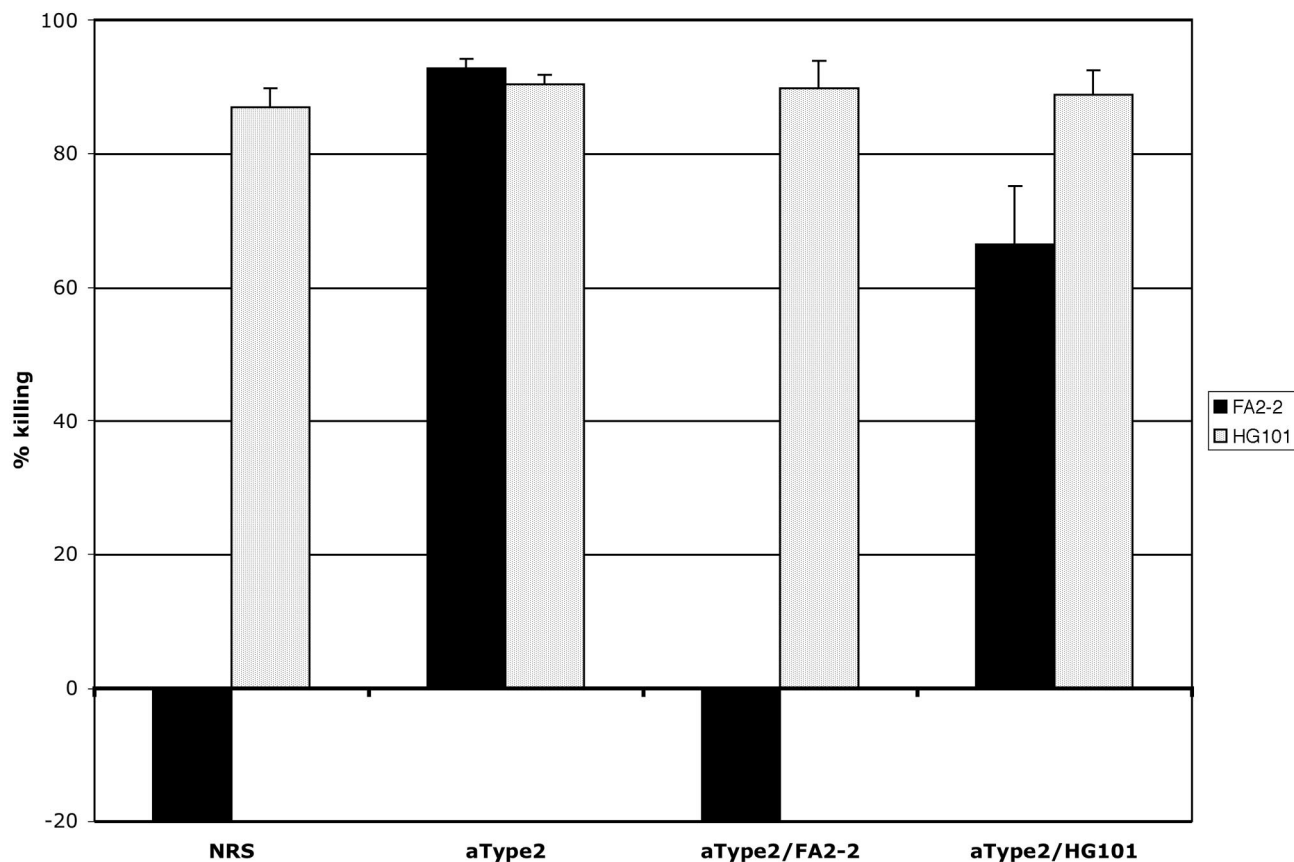


FIG. 5. Comparison of *E. faecalis* strains FA2-2 and HG101 in the opsonophagocytic assay with NRS and the type 2-specific sera. Negative killing reflects growth during the 90-min incubation period of the assay.

tain degrees of cross-reactivity. Heat inactivation and proteinase K treatment of whole bacterial cells elicited antibodies directed against capsular structures, a finding which is confirmed by the opsonophagocytic assay with WT *E. faecalis* strain FA2-2 and its capsule-negative mutant, HG101. The WT strain was killed only by the type 2-specific serum, indicating that the serum contained opsonic antibodies directed against the capsule. In contrast, the capsule-negative mutant was already sensitive to killing by preexisting natural antibodies in NRS, which were not necessarily directed against the capsule. A similar phenomenon has been observed previously (2, 3, 15, 16).

In order to increase the specificity of the reactivities of the four types of sera against capsular polysaccharides, we used TCA extraction to isolate crude polysaccharide antigens from the bacterial cells in the CPS-ELISA. We found that the protein levels in TCA extracts were below the detection limit of the modified Bradford assay used. Treatment of TCA extracts with sodium periodate reduced the carbohydrate content to 18% of that of pure TCA extracts and reduced the reactivity with the type-specific serum to 8% (data not shown), indicating that the majority of the antibody binding activity was to a carbohydrate antigen. In contrast, treatment of the TCA extract with proteinase K had no effect on the reactivities of the antisera (98% of that of pure TCA extracts). From these data we conclude that the TCA extracts primarily consisted of carbohydrate antigens, with the majority presumably being capsular polysaccharide antigens. This assumption was further

supported by the observation that type 2-specific serum revealed a strong reactivity with *E. faecalis* FA2-2 (OD_{405} , 1.860) but a strongly reduced reactivity with capsule-negative mutant HG101 (OD_{405} , 0.526).

Using the criteria mentioned above to assign an isolate to one of the four serogroups, we were unequivocally able to group 55% of the strains tested. However, one of the serum types (i.e., the type 2-specific serum) showed a high degree of cross-reactivity with a different prototype strain (i.e., *E. faecalis* type 5). This may be explained by chemical and/or structural similarities of the different polysaccharides, a situation not uncommon for polysaccharide antigens from other bacteria (5, 25). Alternatively, a given strain may be able to produce two different variant forms of carbohydrate antigen, depending on the conditions used.

To correlate the data obtained by CPS-ELISA with the biological response (i.e., opsonic killing) relevant to the intended use of the antigens as the basis of a vaccine, we compared the results obtained by CPS-ELISA with those obtained by an opsonophagocytic assay. While the majority of strains reacted identically in the opsonophagocytic assay, we also observed some differences. The high degree of cross-reactivity between *E. faecalis* type 2 and *E. faecalis* type 5 in the opsonophagocytic assay points toward the possibility that these two antigens have close structural similarity. Taking into account this cross-reactivity, as well as the number of strains that were not typeable by both methods, the agreement between the two assays was 72%.

Four strains could be grouped only by the opsonophagocytic assay, perhaps because the amount of specific polysaccharide on the surface of the bacterial cell was too small to be detected by the CPS-ELISA. This amount of antigen may still provide enough antigenic epitopes for opsonization, leading to killing in the opsonophagocytic assay. One of the two strains typeable only by CPS-ELISA (Maekawa strain type 20) was highly resistant to killing by any of the sera, and factors other than capsule production that contribute to serum resistance may be responsible for this effect. The other strain, *E. faecalis* type 1, was highly sensitive to killing by all four types of sera, including NRS (data not shown), suggesting that this strain was unable to resist non-antibody-mediated opsonic killing, even though it has a capsule.

To date there is no proven link between the structure of capsular polysaccharide and a genetic locus responsible for the biosynthesis of the capsular polysaccharide. *E. faecalis* 12030 is the only strain for which structural analysis of the capsular polysaccharide has been performed (16, 27). The *cps* locus is known to be involved in the biosynthesis of capsular polysaccharides (10), but so far only a compositional analysis of the respective antigen exists. The homology of one of the enzymes within this gene locus with a glycerophosphotransferase involved in the synthesis of teichoic acids in *Bacillus subtilis* and the presence of quantitative amounts of phosphate indicate that the locus described by Hancock and Gilmore (10) may be responsible for the production of the capsular polysaccharide described by Huebner and colleagues (16, 27). Three different organizations of the gene locus have been observed: *E. faecalis* 12030 and *E. faecalis* 12107 showed an RFLP class I pattern, with the presence of only two open reading frames, i.e., *cpsA* and *cpsB*. Both of these genes may be essential in *E. faecalis*, and they are present in all strains tested so far, independent of their serotype.

Hancock and Gilmore (11) and Xu et al. (28–30) have described a different locus involved in the synthesis of an integral enterococcal cell wall polysaccharide. While Huebner and colleagues (16, 27) were able to isolate and initially characterize a similar tetraheteroglycan material from the cell wall of our prototype strain, *E. faecalis* 12030, this antigen is distinct from the teichoic acid-like capsular polysaccharides, and the preparation of this tetraheteroglycan shows no immunoreactivity with serum raised against the capsular polysaccharides examined here (data not shown).

E. faecalis type 2- and *E. faecalis* type 5-related strains, which we refer to as the CPS-C and the CPS-D serogroups, respectively, possessed at least 10 of the 11 genes of the *cps* locus. While strains of the CPS-D serogroup did not contain the *cpsF* gene, its presence was variable in CPS-C strains. At present, it is unclear why RFLP class II strains represent both CPS-C and CPS-D strains. Some class II strains (i.e., *E. faecalis* types 5 and 18) react with type 5-specific serum in the CPS-ELISA, whereas others (i.e., *E. faecalis* types 12 and 14) react with type 2-specific serum. *E. faecalis* strain MMH 594, which was previously shown to react with type 2-specific serum (10), produced a hybridization pattern identical to that of RFLP class III strains; but it is now known that it cross-reacts with type 5-specific serum, which suggests that it may modulate expression of its *cps* genes, possibly leading to antigenic variation through the expression of multiple serotypes. Interestingly, *E.*

faecalis type 8 produced the same hybridization profile as *E. faecalis* type 2 (Fig. 4) but was nontypeable with type 2-specific serum. It remains unclear how *E. faecalis* types 2, 8, and 11 (data not shown) can yield identical restriction patterns for the *cps* determinant yet still be serologically distinct. It is possible that *E. faecalis* types 8 and 11 have lost the ability to produce the capsular serotype, thus exposing alternative typing antigens. Alterations in promoter sequences or the presence of regulatory elements could modulate the extent to which the polysaccharide is expressed. Alternatively, the product(s) of polysaccharide biosynthetic genes not found in the *cps* determinant could alter the capsular structure through the placement of additional carbohydrate residues on the capsular polysaccharide.

The presence of the genes in the *cps* locus does not completely explain the serologic specificity observed. Structural analysis of defined knockout mutants may be able to determine whether the difference between CPS-C and CPS-D can be explained by the expression of the *cpsF* gene. The additional genes that are probably necessary for the production of the CPS-A and CPS-B polysaccharides need to be identified and sequenced.

It appears that *E. faecalis* capsular polysaccharides have evolved from two distinct families. The first family comprises lineages containing all (RFLP class III) or most (RFLP class II) of the *cpsC* to *cpsK* determinant, whereas the second family consists of a third lineage that lacks this determinant. The importance of the capsular polysaccharide to the biology of the organism would suggest that at some time in the distant past the second family acquired the ability to make a capsule, independent of the *cps* determinant. A closer examination of the Southern blot analysis (Fig. 4) reveals additional faint bands for DNA from strains of the second family, suggesting that the evolution of capsular polysaccharide diversity in this family may have paralleled that in the first family to allow some cross hybridization with related genes located elsewhere on the chromosome. This observation is consistent with the evolution of capsular polysaccharide determinants in other organisms (22). In *Staphylococcus aureus*, the capsular polysaccharides of serotypes 5 and 8 are similar and their biosynthetic genes occur at the same point on the *S. aureus* genomic map, whereas the determinant for type 1 capsule biosynthesis occurs elsewhere on the chromosome. The limited homology between these genetic pathways is detectable only by hybridization under low-stringency conditions, even though the structures of the type 1, 5, and 8 capsular polysaccharides are similar (22).

From the present data we conclude that a limited number of enterococcal serogroups exist. The four serogroups described here represent a major portion (about 60%) of clinically relevant *E. faecalis* isolates. The methods described here can be used to identify strains with serologically different polysaccharides. Ongoing studies aimed at identifying additional serogroups have so far not given an indication of any serologically diverse polysaccharide capsules (data not shown). The presence of strains that do not react with our antisera may be due to the absence of a polysaccharide capsule in these strains.

In contrast to the studies of Sharpe and Shattok (24), we investigated the serologic diversity of enterococcal polysaccharides not for epidemiologic analysis but for better evaluation of the feasibility of immunotherapy regimens based on capsu-

lar polysaccharide antigens. Serotyping of bacteria has been widely used as a tool in bacterial epidemiology. Powerful new molecular typing methods have replaced this technique. Methods such as pulsed-field gel electrophoresis (26) have better discriminatory power, and randomly amplified polymorphic DNA analysis (4) and multilocus sequence typing (14) are also able to show phylogenetic and evolutionary relationships between strains. Studies are under way to elucidate the possible relationships between serogroups and the evolutionary lineages of enterococcal strains of different geographical origins.

Compositional and structural analyses of the polysaccharides are under way to confirm the serogroups described here. These data will facilitate the comprehensive establishment of a system for the serotyping of enterococci and will help to determine the relevant serotypes to be included in a broadly active enterococcal polysaccharide vaccine. The limited number of serotypes found so far suggests that the development of an enterococcal vaccine will be feasible.

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