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Dopamine receptors – IUPHAR Review 13

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The variety of physiological functions controlled by dopamine in the brain and periphery is mediated by the D₁, D₂, D₃, D₄ and D₅ dopamine GPCRs. Drugs acting on dopamine receptors are significant tools for the management of several neuropsychiatric disorders including schizophrenia, bipolar disorder, depression and Parkinson's disease. Recent investigations of dopamine receptor signalling have shown that dopamine receptors, apart from their canonical action on cAMP-mediated signalling, can regulate a myriad of cellular responses to fine-tune the expression of dopamine-associated behaviours and functions. Such signalling mechanisms may involve alternate G protein coupling or non-G protein mechanisms involving ion channels, receptor tyrosine kinases or proteins such as β -arrestins that are classically involved in GPCR desensitization. Another level of complexity is the growing appreciation of the physiological roles played by dopamine receptor heteromers. Applications of new *in vivo* techniques have significantly furthered the understanding of the physiological functions played by dopamine receptors. Here we provide an update of the current knowledge regarding the complex biology, signalling, physiology and pharmacology of dopamine receptors.

Abbreviations

β Arr2, β -arrestin 2; BDNF, brain-derived neurotrophic factor; CaMKII, Ca²⁺/calmodulin-dependent PK II; CDK5, cyclin-dependent kinase 5; DAT, dopamine transporter; DARPP-32, dopamine and cAMP-regulated phosphoprotein, 32 kDa; GluA1, glutamate receptor, ionotropic, AMPA 1 subunit; GluN2B, glutamate receptor, ionotropic, NMDA 2B subunit; GIRKs, G protein coupled inwardly rectifying potassium channels; GSK3, glycogen synthase kinase; HTT, huntingtin; KO, knockout; IP₃, inositol trisphosphate; PDK, phosphatidylinositol-dependent kinase; PP1, protein phosphatase 1; PP2A, protein phosphatase 2A; rpS6, ribosomal protein S6; RTK, receptor tyrosine kinases; TCS1/2, tuberous sclerosis proteins 1 and 2; TrkB, neurotrophic tyrosine kinase, receptor, type 2.

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This article, written by members of the International Union of Basic and Clinical Pharmacology Committee on Receptor Nomenclature and Drug Classification (NC-IUPHAR) subcommittee for the dopamine receptors, confirms the existing nomenclature for these receptors and reviews our current understanding of their structure, pharmacology and functions and their likely physiological roles in health and disease. More information on these receptor families can be found in the Concise Guide to PHARMACOLOGY (<http://onlinelibrary.wiley.com/doi/10.1111/bph.12445/abstract>) and for each member of the family in the corresponding database <http://www.guidetopharmacology.org/GRAC/FamilyDisplayForward?familyId=20&familyType=GPCR>.

Table of Links

TARGETS			LIGANDS		
GPCRs^a	Enzymes^b	Transporters^c	amphetamine	fluoxetine	quetiapine
5-HT receptors	adenylyl cyclases	DAT	apomorphine	GABA	quinpirole
Adenosine A ₁ receptor	Akt	Catalytic receptors^d	aripiprazole	haloperidol	risperidone
Adenosine A _{2A} receptor	calpain	ErbB (epidermal growth factor) receptor family	asenapine	huntingtin	SCH23390
α _{1B} -Adrenoceptor	CaMKII	ErbB-1	BDNF	IGF	SKF 38393
β ₁ -Adrenoceptor	CDK5	IGFR1	blonanserin	IGF-1	SKF 81297
Dopamine D ₁ receptor	ERK	PDGFRβ	brexpiprazole	iloperidone	thioridazine
Dopamine D ₂ receptor	Epac1	RTKs	bromocriptine	insulin	UNC0006
Dopamine D ₃ receptor	Epac2	TrkB	cAMP	L-DOPA	UNC9975
Dopamine D ₄ receptor	GRK2	Ion channels^e	cariprazine	lithium	UNC9994
Dopamine D ₅ receptor	GSK3	GIRKs	chlorpromazine	lurasidone	xanomeline
Ghrelin receptor	GSK3α	K _v 2 channels	clozapine	NMDA	ziprasidone
Muscarinic M ₄ receptor	GSK3β	IP ₃ receptor	cocaine	olanzapine	
TA ₁ receptor	MAPK	Ligand-gated ion channels^f	dopamine	perphenazine	
	PDK1	GluA1			
	PKA	GluN2B			
	PKC	Ionotropic glutamate receptors			
	PLC				
	PLCβ				

This table lists protein targets and ligands, which are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson *et al.*, 2014) and the Concise Guide to PHARMACOLOGY 2013/14 (^{a,b,c,d,e,f}Alexander *et al.*, 2013a,b,c,d,e,f).

Introduction

Five subtypes of dopamine receptors (D₁, D₂, D₃, D₄ and D₅ receptors, encoded in humans by genes *DRD1*, *DRD2*, *DRD3*, *DRD4* and *DRD5*, respectively) are known to mediate essentially all of the physiological functions of dopamine. These functions include, but are not limited to, the following: voluntary movement, reward, sleep regulation, feeding, affect, attention, cognitive function, olfaction, vision, hormonal regulation, sympathetic regulation and penile erection. Dopamine receptors are also known to influence the immune system as well as cardiovascular, renal and gastrointestinal functions. As members of the GPCR superfamily, dopamine receptors have a canonical seven-transmembrane structure and can signal through both G protein-dependent and -independent mechanisms. Based on coupling to either Gα_{s,olf} proteins or Gα_{i/o} proteins to stimulate or inhibit the production of the second messenger cAMP, respectively, dopamine receptors are classified as D₁-class receptors (D₁ and D₅) or D₂-class receptors (D₂, D₃ and D₄) (Kebabian, 1978; Spano *et al.*, 1978). The alternative splicing of D₂ results in the generation of two major D₂ dopamine receptor variants that differ in the presence of an additional 29 amino acids on the third intracellular loop with distinct physiological, signalling and pharmacological properties, and are classified as D_{2S} (D₂-short) and D_{2L} (D₂-long). Dopamine receptors are well-established targets in the clinical pharmacology of numerous disorders and conditions such as schizophrenia, Parkinson's

disease, bipolar disorder, depression, restless leg syndrome, hyperprolactinaemia, pituitary tumours, hypertension, gastroparesis, nausea and erectile dysfunction. The basic principles of dopamine receptor structure, signalling, function and pharmacology are covered in detail in several excellent reviews (Niznik and Van Tol, 1992; Sibley and Monsma, 1992; Sokoloff *et al.*, 1992; Civelli *et al.*, 1993; Missale *et al.*, 1998; Vallone *et al.*, 2000; Carlsson, 2001; Seeman, 2006). Recently, we have provided a comprehensive overview of the field in *Pharmacological Reviews* (Beaulieu and Gainetdinov, 2011). However, although the basic information regarding the structural, genetic and biochemical properties of dopamine receptors has remained essentially unchanged in the last 4 years, a significant amount of new information has emerged on dopamine receptor signalling, functional relevance and pharmacology that requires an update of the status of current knowledge. Here we will focus on newly emerging topics and trends in understanding dopamine receptor biology as well as topics that were not covered or only partially discussed in our previous review (Beaulieu and Gainetdinov, 2011).

Mechanisms of dopamine receptor signalling

The prevailing convention was that dopamine receptors were considered to signal exclusively through G protein-dependent cellular processes. The D₁-class receptors (D₁ and

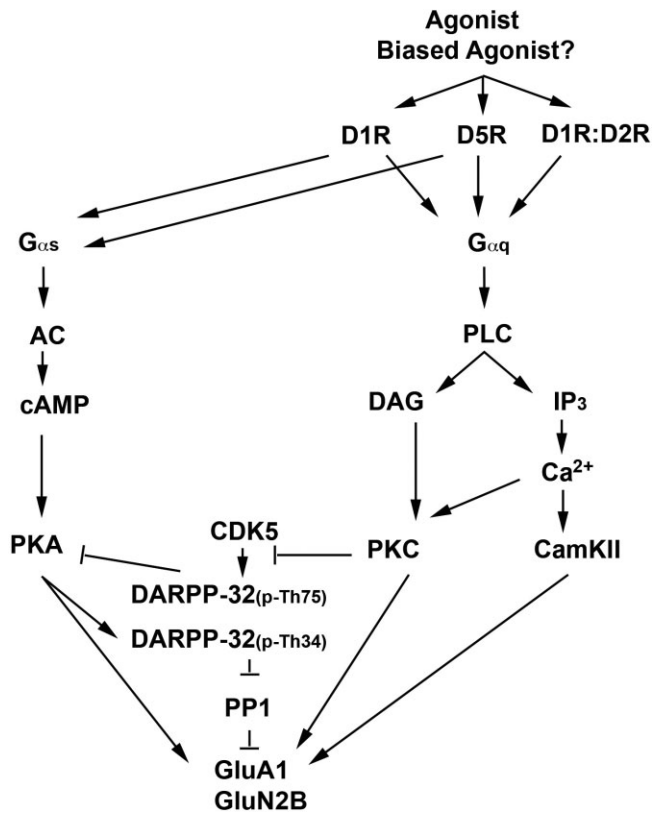


Figure 1

Schematic diagram representing the signalling cascades activated by the D₁ dopamine receptor (D1R). D5R, D₅ dopamine receptor; D1R:D2R, D₁-D₂ receptor heteromer.

D₅ receptors) are primarily coupled to G $\alpha_{s/oif}$ proteins and stimulate the activity of AC and the production of the second messenger cAMP (Figure 1). In contrast, the D₂ class receptors (D_{2s}, D_{2L}, D₃ and D₄ receptors) are associated with G $\alpha_{i/o}$ proteins to inhibit the production of cAMP (Kebabian, 1978; Spano *et al.*, 1978) (Figure 2).

Modulation of cAMP synthesis by dopamine receptors results in the regulation of PKA and potentially of other exchange proteins activated by cAMP (Epac1 and Epac2) (Svenningsson *et al.*, 2004; Beaulieu and Gainetdinov, 2011). Among PKA substrates, the multifunctional dopamine and cAMP-regulated phosphoprotein (DARPP-32/PPP1R1B) has been extensively studied over the last 30 years. When phosphorylated on Thr³⁴ by PKA, DARPP-32 is a negative regulator of protein phosphatase 1 (PP1). In contrast, phosphorylation of DARPP-32 on Thr⁷⁵ by cyclin-dependent kinase 5 (CDK5), in response to sustained D₁ receptor activation, results in PKA inhibition (Figures 1 and 2). The roles of PKA and DARPP-32 in dopamine receptor signalling are well characterized, and strong evidence supports their contribution to the physiological functions of dopamine receptors (Svenningsson *et al.*, 2004; Girault, 2012).

cAMP-mediated signalling and mRNA translation

An interesting development in the characterization of cAMP-mediated dopamine receptor signalling involves its recently

appreciated contribution to the regulation of mRNA translation mechanisms. Either D₁ receptor activation or D₂ receptor blockade by haloperidol has been shown to promote the phosphorylation of the ribosomal protein S6 (rpS6) on Ser^{235/236} and Ser^{240/244} (Santini *et al.*, 2009; Valjent *et al.*, 2011). Phosphorylation of rpS6 on these and adjacent residues results in enhanced CAP-dependent mRNA translation (Roux *et al.*, 2007; Hutchinson *et al.*, 2011). Increased phosphorylation of rpS6 via D₁ receptors would involve activation of PKA, subsequent inhibition of PP1 by DARPP-32 and activation of the mammalian target of rapamycin (mTOR) complex 1 (Santini *et al.*, 2009; 2012; Bonito-Oliva *et al.*, 2013). In medium spiny neurons expressing D₂ receptors, activation of PKA and DARPP-32 by the adenosine A_{2A} receptors also plays a role (Valjent *et al.*, 2001; 2011; Santini *et al.*, 2009), whereas activation of ERK signalling by a DARPP-32-dependent mechanisms are thought to be involved in D₁ receptor-expressing medium spiny neurons (Santini *et al.*, 2012). Interestingly, D₁ receptor stimulation also promotes rpS6 phosphorylation in the dentate gyrus, albeit through a different, mTOR-independent, pathway involving ERK activation (Gangarossa and Valjent, 2012). Understanding the overall importance of dopamine receptor-mediated regulation of rpS6 on mRNA translation and behaviour is still in its infancy. However, preliminary evidence supports its involvement in the development of L-DOPA-induced dyskinesia (Santini *et al.*, 2009; 2012; Subramaniam *et al.*, 2012) and cocaine sensitization, seeking and relapse behaviours (Wu *et al.*, 2011).

cAMP-independent dopamine receptor signalling

In addition to the regulation of cAMP, several studies have revealed that dopamine receptors can exert some of their biological effects through alternative signalling pathways (Beaulieu *et al.*, 2004; 2005; Hasbi *et al.*, 2009). For instance, there are indications that both D₁ and D₂ receptors can transactivate the brain-derived neurotrophic factor (BDNF) receptor in neurons (Swift *et al.*, 2011). These two dopamine receptors can also regulate calcium channels through a direct protein-protein interaction *in vivo* (Kisilevsky and Zamponi, 2008; Kisilevsky *et al.*, 2008). Direct interaction of D₁ and D₂ receptors and Na⁺-K⁺-ATPase has also been demonstrated (Hazelwood *et al.*, 2008; Blom *et al.*, 2012). Under certain circumstances, dopamine receptors can also regulate IP₃-mediated signalling (Medvedev *et al.*, 2013; Perreault *et al.*, 2014), and there is evidence for alternative coupling of D₁-class receptors to G α_q (Figure 1).

The D₂-class D₂ and D₃ receptors have been shown to signal through both G protein-dependent and G protein-independent mechanisms (Beaulieu and Gainetdinov, 2011). G protein-dependent mechanisms for D₂ dopamine receptors are represented by the well-known G $\alpha_{i/o}$ subunit-mediated cAMP-PKA-DARPP32 cascade (Svenningsson *et al.*, 2004) and the G $\beta\gamma$ -mediated activation of PLC, leading to increased cytoplasmic calcium and downstream signalling events (Hernandez-Lopez *et al.*, 2000; Beaulieu and Gainetdinov, 2011). Furthermore, G $\beta\gamma$ -mediated mechanisms are involved in the regulation of activity of the L- and N-type calcium channels (Yan *et al.*, 1997) as well as G protein coupled inwardly rectifying potassium channels (GIRKs) (Kuzhikandathil *et al.*, 1998; Beaulieu and Gainetdinov,

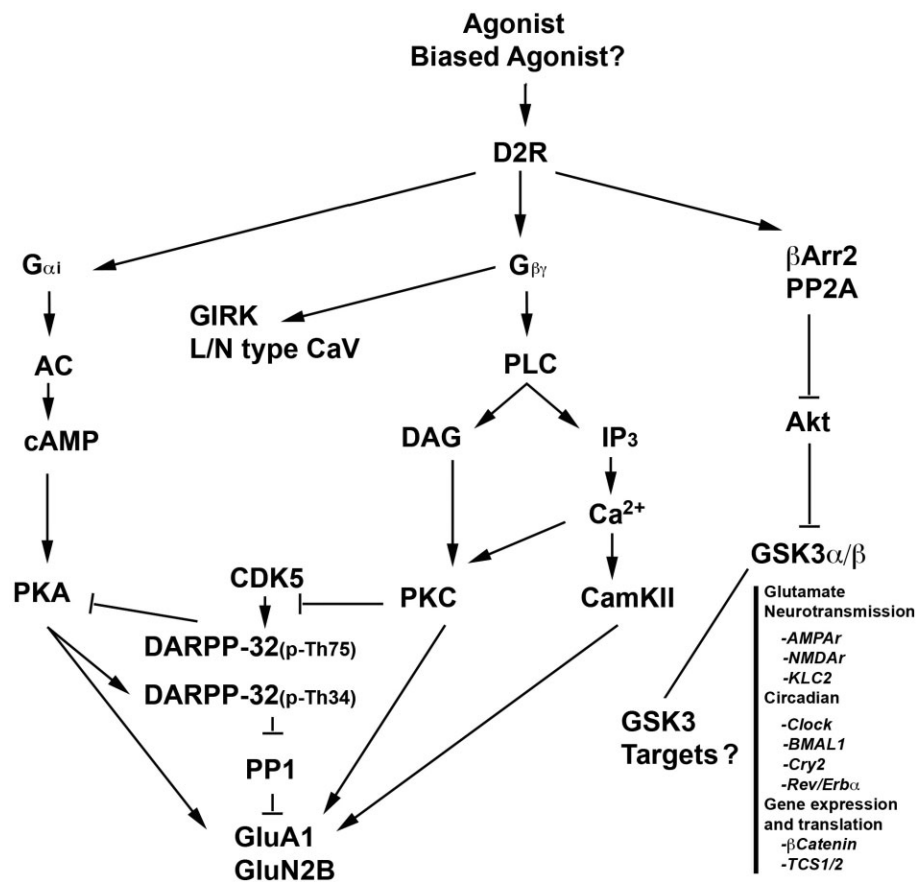


Figure 2

Schematic diagram representing the signalling cascades activated by the D₂ dopamine receptor (D2R). BMAL1, aryl hydrocarbon receptor nuclear translocator-like protein; *Clock*, circadian locomotor output cycles kaput gene; *Cry2*, cryptochrome 2; KLC2, kinesin light chain 2; Rev/Erb α , nuclear receptor subfamily 1, group D, member 1.

2011). Recent evidence indicates that all of these G protein-mediated signalling cascades converge on, among other targets, phosphorylation of two subunits of ionotropic glutamate receptors, GluA1 and GluN2B, which are critically involved in glutamatergic transmission (Jenkins and Traynelis, 2012; Dell’anno *et al.*, 2013; Hobson *et al.*, 2013; Jia *et al.*, 2013; Song *et al.*, 2013; Flores-Barrera *et al.*, 2014; Jenkins *et al.*, 2014; Murphy *et al.*, 2014) (Figure 2).

Finally, there is strong evidence that D₂ dopamine receptors can signal *in vivo* by activating cAMP-independent mechanisms involving the multifunctional adaptor protein β -arrestin 2 (β Arr2) (Beaulieu *et al.*, 2004; 2005; 2008b; Urs *et al.*, 2012) (Figure 2). In the remaining parts of this subsection, we will provide an overview of recent evidence underscoring the importance of cAMP-independent mechanisms in dopamine receptor function.

Coupling of dopamine receptors to G α_q

Several lines of evidence support the regulation of PLC and calcium signalling by dopamine receptors. As early as 1989, Felder *et al.* reported that the D₁ receptor agonist SKF 82526 stimulates PLC activity independently of cAMP in renal tubular membranes (Felder *et al.*, 1989). Activation of PLC

leads to the production of inositol trisphosphate (IP₃) and DAG. This results in the activation of PKC by DAG and an increased mobilization of intracellular calcium in response to IP₃ (Berridge, 2009). The increase of intracellular calcium in the cytoplasm leads to the activation of calcium-dependent PKC variants as well as calcium-regulated enzymes, such as the calcium/calmodulin-dependent PK II (CaMKII) and the protein phosphatase calcineurin/protein phosphatase 2B (PP2B).

The most common way for a GPCR to regulate PLC activity is by coupling to G α_q . Putative D₁–D₂ receptor heterodimers have been suggested to regulate DAG and IP₃ signalling by activating G $\alpha_{q/11}$ in transfected cells as well as in striatal membrane preparations (Lee *et al.*, 2004; Rashid *et al.*, 2007b). The physiological relevance of D₁–D₂ receptor heterodimers is supported by the co-expression of D₁ and D₂ receptors in small populations of medium spiny neurons of the nucleus accumbens in the mouse (Rashid *et al.*, 2007b) and in other regions of the basal ganglia (Perreault *et al.*, 2010). Notably, analysis of BAC transgenic mice that express fluorescent gene-reporter proteins driven by D₁ and D₂ receptor promoters showed that the majority of D₁ receptor-positive pyramidal neurons in the prefrontal cortex also express low

levels of D₂ receptors (Zhang *et al.*, 2010). In addition to co-expression studies, FRET studies conducted with fluorescent proteins in transfected cells and treatments of tissue sections with labelled antibodies have produced results that suggest the formation of receptor heterodimers (Rashid *et al.*, 2007b; Perreault *et al.*, 2013).

Despite accumulating evidence, the involvement of D₁–D₂ receptor heterodimers in the regulation of PLC-mediated signalling *in vivo* remains poorly understood. It should be noted that recent studies have questioned the selectivity (Chun *et al.*, 2013) and PLC activity (Lee *et al.*, 2014) of the putative D₁–D₂ receptor heteromer agonist SKF 83959 that was used to characterize the role of D₁–D₂ receptor heterodimers in the regulation of PLC *in vivo* (Rashid *et al.*, 2007b). One important aspect of D₁–D₂ heterodimer signalling in cells is the requirement of co-activation of both the D₁ and D₂ receptor moiety to activate G $\alpha_{q/11}$. Furthermore, the formation of the D₁–D₂ receptor heterodimers would prevent coupling of either receptors to G $\alpha_{s/olf}$ or G $\alpha_{i/o}$ (Perreault *et al.*, 2014). This theory, however, is in contrast with several *in vivo* observations supporting the regulation of PLC by D₁-class receptors without the need for D₂ receptor involvement. It was recently reported that acute systemic administration of cocaine, amphetamine, apomorphine or the D₁-class receptor agonist SKF 81297 to wild-type mice increases striatal IP₃ synthesis (Medvedev *et al.*, 2013). Co-treatments with selective antagonists as well as the use of D₁ and D₂ receptor knockout (KO) animals revealed that the production of IP₃ in response to these pharmacological treatments requires D₁, but not D₂ receptor activation. Importantly, PLC β inhibition suppressed spontaneous locomotor hyperactivity in hyperdopaminergic mice lacking the dopamine transporter (DAT) and antagonized the effects of amphetamine, cocaine, SKF 81297 and apomorphine on forward locomotion. Furthermore, the restoration of locomotion by L-DOPA in dopamine-depleted mice (Sotnikova *et al.*, 2005) is also reduced by inhibition of PLC β resulting in mostly vertical activity following these treatments (Medvedev *et al.*, 2013). These data strongly support a contribution of PLC in mediating the effects of dopamine on forward locomotion. However, further investigation is necessary to decipher the relative contribution of different modes of PLC regulation on the various aspects of dopamine-related behaviours.

At the same time, expression of D₁ receptors in transfected HEK293 cells does not affect intracellular calcium signalling. However, expression of D₅ receptors in the same cells induces extensive calcium mobilization after stimulation (So *et al.*, 2009), and the D₁-class receptor agonist SKF 38393 activates PLC-mediated signalling in D₁ receptor KO mice (Friedman *et al.*, 1997). Furthermore, this same agonist, as well as dopamine and SKF 83959, failed to increase IP₃ levels in brain slices prepared from mice lacking D₅ receptors (Sahu *et al.*, 2009). A similar lack of responsiveness of PLC-mediated signalling to SKF 83959 was also reported following systemic administration of this compound to D₅ receptor KO mice (Sahu *et al.*, 2009), suggesting that activation of D₅ receptors is sufficient to activate G $\alpha_{q/11}$ in response to selected doses of certain D₁-class receptor agonists.

Thus, several independent studies support the regulation of PLC-mediated signalling through dopamine receptors, however, these studies are in disagreement with regard to the

detailed mechanism of this regulation. Current evidence does not allow us to rule out the contributions of D₁ receptors, D₅ receptors or D₁–D₂ receptor heterodimers in this phenomenon (Figure 1). Discrepancies between the results of different research groups raise the possibility that several mechanisms may be involved, perhaps in different neuronal populations. It is also conceivable that different D₁-class receptor agonists may be functionally selective for PLC-mediated mechanisms when activating D₁ receptors, D₅ receptors or D₁–D₂ receptor heterodimers.

Beyond the question of its detailed mechanism, activation of PLC-mediated signalling by dopamine also raises the question of possible crosstalk between this modality of signalling and cAMP-mediated mechanisms. Among several possibilities, activation of PKC and CaMKII through calcium signalling could affect glutamate receptors concomitantly with PKA (Figure 1). Different mechanisms involving either positive or negative regulation of CDK5 by calcium may also be an important nexus for crosstalk. For instance, PKC has previously been shown to prevent the phosphorylation of DARPP-32 and other substrates by CDK5 (Sahin *et al.*, 2008). Because the global activity of DARPP-32 is modulated by an equilibrium between its phosphorylation by CDK5 and PKA (Bibb *et al.*, 1999) it is possible that G $\alpha_{q/11}$ -mediated dopaminergic signalling may reduce the phosphorylation of DARPP-32 by CDK5 and potentiate PKA-mediated signalling (Figure 1). In contrast, cleavage of the CDK5 co-activator p35 by the calcium-regulated protease calpain (Lee *et al.*, 2000; Beaulieu and Julien, 2003) may result in CDK5 hyperactivity and an inhibition of PKA signalling. Furthermore, changes in calcium concentration may also affect the activity of PP2B (calcineurin), which is involved in the dephosphorylation of DARPP-32 at Thr³⁴ (Halpain *et al.*, 1990). Overall, the full understanding of the regulation of PLC activity by dopamine remains incomplete yet holds promise for exciting future investigations.

Coupling of D₂-class receptors to β Arr2, Akt and glycogen synthase kinase (GSK3)

G protein-independent D₂ receptor signalling is represented by β Arr2-mediated mechanisms. Arrestins are a family of four molecular adaptor proteins that were originally characterized for their role in mediating GPCR desensitization and internalization (Lohse *et al.*, 1990; Ferguson *et al.*, 1996). In addition to these functions, the two ubiquitous arrestins, β Arr1 and β Arr2, have also been shown to act as molecular scaffolds for signalling molecules such as kinases and phosphatases (Luttrell *et al.*, 2001; Beaulieu *et al.*, 2005).

Several lines of evidence have pointed towards the contribution of a β Arr-mediated mechanism in the regulation of the serine/threonine kinases Akt and GSK3 by dopamine. Akt is involved in several cellular processes such as glucose metabolism, gene transcription, cell proliferation, migration and neurotrophin action through the stimulation of receptor tyrosine kinases (RTKs) (Cross *et al.*, 1995; Alessi *et al.*, 1996; Scheid and Woodgett, 2001). Activation of RTKs and some GPCRs regulates PI3K, which converts phosphatidylinositol-2-phosphate (PIP₂) to phosphatidylinositol-3-phosphate (PIP₃) (Martelli *et al.*, 2010). This newly formed PIP₃ interacts with the pleckstrin homology domain of Akt, inducing the recruitment of Akt to the plasma membrane. This, in turn,

results in the phosphorylation of Akt at the Thr³⁰⁸ and Ser⁴⁷³ residues by two phosphatidylinositol-dependent kinases, PDK1 and PDK2/ricor-mTOR respectively (Scheid and Woodgett, 2001; Jacinto *et al.*, 2006). Once activated, Akt phosphorylates several substrates including GSK3 (Rossig *et al.*, 2002). Mammalian cells express two isoforms of GSK3, GSK3 α and GSK3 β , which are constitutively active and can phosphorylate several cellular substrates (Woodgett, 1990; Kaidanovich-Belin and Woodgett, 2011). Phosphorylation by Akt inhibits both isoforms of GSK3 in response to growth factors and hormones, including insulin, IGF, and BDNF (Yamada *et al.*, 2002; Altar *et al.*, 2008). Specifically, Akt phosphorylates Ser²¹ on GSK3 α and Ser⁹ on GSK3 β , which are located on their respective N-terminal domains (Stambolic and Woodgett, 1994; Frame and Cohen, 2001).

Experiments using dopamine receptor agonists/antagonists, dopamine depletion and hyperdopaminergic DAT-KO mice have provided converging evidence for the negative regulation of Akt, resulting in the activation of both GSK3 isoforms by D₂-class receptors in mammals and other vertebrates (Beaulieu *et al.*, 2004; Bychkov *et al.*, 2007; Chen *et al.*, 2007; Souza *et al.*, 2011). Consequently, D₂-class receptor antagonists induce Akt activation and subsequent GSK3 inhibition (Beaulieu *et al.*, 2004; Emamian *et al.*, 2004). Additional investigations conducted using mice lacking various dopamine receptors have shown that a loss of D₂, but not D₁ receptors prevents the inactivation of striatal Akt by drugs acting on dopamine neurotransmission (Beaulieu *et al.*, 2007b). In contrast, D₃ receptor-deficient mice exhibit a reduction of Akt phosphorylation in response to dopaminergic drugs. This suggests that D₂ receptors are critical for the inhibition of Akt by dopamine, whereas the D₃ receptors appear to potentiate the D₂ receptor-mediated dopamine response (Beaulieu *et al.*, 2007b).

The role of β Arr2 in mediating the regulation of Akt and GSK3 by D₂ receptors is supported by direct *in vivo* biochemical observations in pharmacological and genetic models of enhanced dopaminergic neurotransmission (Beaulieu *et al.*, 2004; 2005). Amphetamine and apomorphine have been shown to inhibit the phosphorylation and activation of Akt in the striatum of wild-type mice, whereas these two drugs failed to inhibit Akt in β Arr2-KO mice. Furthermore, regulation of Akt and GSK3 signalling observed in mice with genetically increased dopaminergic tone caused by a lack of DAT, was absent in double mutant mice deficient for both DAT and β Arr2, suggesting an important role of this scaffolding protein in Akt regulation by dopamine (Beaulieu *et al.*, 2005). Further characterization of the molecular mechanisms underlying the regulation of Akt by D₂ receptors, following receptor stimulation has shown that β Arr2 is involved in the formation of a protein complex composed of Akt, β Arr2 and protein phosphatase 2A (PP2A) (Beaulieu *et al.*, 2005). Formation of this complex allows PP2A to dephosphorylate and inactivate Akt, resulting in the activation of GSK3 (Beaulieu *et al.*, 2004; 2005).

It is worth mentioning that the formation of the Akt : β Arr2 : PP2A signalling complex in response to D₂ receptor activation represents a mechanism through which dopamine can trigger the inactivation of PI3K/Akt signalling in a regulated fashion. Importantly, the Akt : β Arr2 : PP2A signalling complex dissociates in response to lithium, thus provid-

ing a probable explanation for the early behavioural observations of the antagonistic effect of lithium on dopaminergic behaviours as well as a reasonable mechanism for the activation of Akt by lithium (Beaulieu and Caron, 2008a; O'Brien *et al.*, 2011; Pan *et al.*, 2011). The details of the mechanism(s) by which lithium triggers this dissociation are not yet fully understood. Current evidence suggests that lithium may affect the stability of this complex by acting on several of its components, possibly in a synergistic fashion. First, lithium has been shown to interfere with the interaction of Akt1 and β Arr2 (Beaulieu *et al.*, 2008b). Direct investigation of the Akt- β Arr2 interaction using recombinant proteins have demonstrated that this interaction is dependent upon the presence of magnesium ions and that excess magnesium can prevent the dissociation of Akt and β Arr2 upon treatment with a therapeutic dose of lithium (1 mM). Second, GSK3 β has also been shown to interact with β Arr2. Recent evidence obtained from transgenic mice overexpressing *Xenopus* GSK3 β in neurons indicate that activated GSK3 can act as a feed-forward mechanism for its own activation (Figure 2) by stabilizing the Akt : β Arr2 : PP2A signalling complex (O'Brien *et al.*, 2011). According to this model, direct inhibition of GSK3 by lithium would thus constitute a mechanism that can promote the disassembly of the Akt : β Arr2 : PP2A.

The effect of β Arr2-mediated Akt/GSK3 signalling on dopaminergic behaviours is supported by several experimental observations *in vivo*. β Arr2-KO mice have been shown to display spontaneous locomotor hypoactivity, reduced apomorphine-induced climbing and amphetamine-induced hyperlocomotion (Gainetdinov *et al.*, 2004; Beaulieu *et al.*, 2005). These mice also have a reduced responsiveness to the dopamine-dependent locomotor effects of morphine (Bohn *et al.*, 2003). In addition, novelty-driven locomotor hyperactivity, a phenotype that is typical of hyperdopaminergic DAT-KO mice, is less pronounced in double mutant mice lacking both β Arr2 and DAT (Beaulieu *et al.*, 2005). Administration of lithium exerts multiple actions on behaviours in DAT-KO and normal mice, including suppression of spontaneous locomotor activity, but this was not observed in β Arr2-KO mice (Beaulieu *et al.*, 2004; 2005). In line with these data, mice lacking Akt1 demonstrate an enhanced sensitivity to amphetamine with regard to the disruption of sensorimotor gating in the pre-pulse inhibition (PPI) test, which is used to model psychosis in rodents (Emamian *et al.*, 2004). As described above, Akt1 is inhibited following the stimulation of D₂ receptors, thus the increased behavioural effect of amphetamine in Akt1-KO mice is likely to result from the involvement of Akt in dopaminergic behavioural responses.

Genetic suppression of GSK3 activity also inhibits locomotor hyperactivity related to excessive dopaminergic tone in amphetamine-treated mice (Beaulieu *et al.*, 2004). Similarly, several GSK3 inhibitors as well as GSK3 β haploinsufficiency can block amphetamine-induced hyperactivity (Beaulieu *et al.*, 2004; Gould *et al.*, 2004; Kalinichev and Dawson, 2011). In contrast, mice overexpressing GSK3 β show pronounced locomotor hyperactivity (Prickaerts *et al.*, 2006), and transgenic mice expressing a GSK3 β mutant that lacks an inhibitory phosphorylation site (thus is constitutively active) demonstrate increased novelty-driven and amphetamine-induced hyperactivity (Polter *et al.*, 2010).

More recent evidence obtained using strains of cell type-specific conditional GSK3 β -KO mice have generated a more nuanced portrait of the contribution of GSK3 β in the regulation of dopaminergic behaviour. Ablation of GSK3 β expression specifically in D₁ or D₂ receptor-expressing striatal neurons (Urs *et al.*, 2012) confirmed the selective contribution of GSK3 β to the acute action of amphetamine on locomotion in D₂ but not D₁ receptor-expressing neurons. The antagonistic action of the D₂ receptor partial agonist aripiprazole and lithium on amphetamine-induced locomotion is also curbed in mice lacking GSK3 β in D₂ receptor-expressing neurons. In contrast, haloperidol-induced catalepsy is not affected by diminished GSK3 β expression in either D₁ or D₂ receptor-expressing striatal neurons, whereas the disruptive effects of amphetamine on sensory motor gating is abolished by in either D₁ or D₂ receptor-expressing neuron-selective GSK3 β gene inactivation. Taken together, these observations confirm the role of β Arr2-mediated regulation of GSK3 β in D₂ receptor-expressing neurons in the effects of amphetamine, lithium and aripiprazole on locomotion. The fact that haloperidol-induced catalepsy remains intact in mice lacking GSK3 β suggests the involvement of at least two separate signalling pathways mediating the effects of antipsychotics and strengthens the rationale for the development of biased D₂ receptor antagonists to selectively target these pathways in schizophrenia (Beaulieu *et al.*, 2007a; Beaulieu, 2012). Further confirmation of these hypotheses should come from repeating these experiments in mice lacking β Arr2 in specific neuronal populations.

Selective ablation of GSK3 β post-natally in forebrain pyramidal neurons revealed other functions of GSK3 β in the regulation of dopamine-associated behaviours (Latapy *et al.*, 2012). The locomotor effects of amphetamine are marginally increased in these mice, which suggests a minor role of cortical neurons in the modulation of amphetamine action and further indicates that the opposing effect of GSK3 β inhibition on amphetamine-induced locomotion is mediated by GSK3 β in subcortical structures. Additionally, these mice display reduced anxiety and enhanced social interactions. Investigation of the possible contribution of GSK3 β in behavioural responses to social defeat stress (Wilkinson *et al.*, 2011; Latapy *et al.*, 2012) using either conditional forebrain KO mice, GSK3 β haplo-insufficient mice or mice expressing a dominant negative GSK3 in the nucleus accumbens also revealed a role for subcortical GSK3 β inhibition in mediating resilience to this form of stress. This emphasizes the need to further examine the contribution of GSK3-mediated dopamine receptor signalling in coping behaviours.

Beyond its potential involvement in the action of lithium, β Arr2-mediated D₂ receptor signalling can also contribute to effects of antipsychotics. Characterization of the effects of different antipsychotics using BRET in transfected HEK293 cells revealed that first-generation antipsychotics (chlorpromazine, haloperidol), as well as second- (clozapine, quetiapine, olanzapine, risperidone, ziprasidone) and third- (aripiprazole) generation antipsychotics potently antagonize quinpirole-induced β Arr2 recruitment to D₂ receptors (Masri *et al.*, 2008). In contrast, strong differences existed in the potency of these drugs in preventing inhibition of cAMP synthesis by D₂ receptors. Of interest, D₂ receptor partial agonist aripiprazole displayed partial D₂ receptor agonist

activity for cAMP-mediated signalling in the absence of quinpirole while functioning as an antagonist for cAMP when quinpirole was applied concomitantly. Because second- and third-generation antipsychotics are characterized by fewer extrapyramidal side effects, this study led to the hypothesis that identification of functionally selective D₂ receptor antagonists that specifically prevent β Arr2 recruitment to D₂ receptors may pave the way for the development of new antipsychotics that would have fewer side effects while retaining their therapeutic activity.

This hypothesis led to the development of new aripiprazole derivative compounds: UNC9975, UNC0006 and UNC9994, which display antipsychotic-like activity in rodents (Allen *et al.*, 2011). In the absence of a full agonist, these three compounds have the distinction of acting as partial D₂ agonist for β Arr2 recruitment without affecting cAMP. It should be noted, however, that these compounds may not be fully functionally selective as Allen *et al.* also reported that they can act as neutral antagonists for cAMP-mediated D₂ signalling. It is also noteworthy that aripiprazole behaves as a partial agonist for β Arr2 recruitment when applied alone on cells (Allen *et al.*, 2011) while acting as an antagonist of β Arr2 recruitment when simultaneously applied with quinpirole (Masri *et al.*, 2008). It is thus possible that the UNC compounds may display different pharmacological properties when applied alone *in vitro* and in the context of an active dopamine tone *in vivo* where they might antagonize both cAMP and β Arr2 mediated D₂ receptor signalling through a combination of neutral antagonism and partial agonism.

Overall, β Arr2-mediated D₂ receptor signalling provides interesting avenues for the development of new drugs targeting dopamine neurotransmission. However, it is unclear at the moment whether this type of intervention will be more suited for clinical interventions in schizophrenia or bipolar disorder. Indeed, this form of signalling is directly targeted by lithium (Beaulieu *et al.*, 2004; 2008b), a drug that has very limited efficacy for the treatment of schizophrenia.

Protein phosphatase metallo-dependent (PPM/PP2C) and G $\alpha_{i/o}$ mediated regulation of huntingtin (HTT) protein phosphorylation by D₂ receptors

Recent investigation has revealed a role of D₂ receptors in the regulation of the phosphorylation of the HTT protein on Ser⁴²¹ (Marion *et al.*, 2014). It is known that phosphorylation of HTT on this residue by Akt in response to IGF-1 leads to reduction of the formation of nuclear inclusions and HTT toxicity (Humbert *et al.*, 2002; Rangone *et al.*, 2004). Intriguingly, D₂ receptor stimulation reduces the phosphorylation of HTT on this residue in heterologous cells and in the mouse striatum (Marion *et al.*, 2014). The molecular mechanism of this regulation appears not to involve the regulation of Akt by D₂ receptors. Instead, the regulation of HTT phosphorylation by D₂ receptors involves the activation of G $\alpha_{i/o}$ and the formation of a protein complex between HTT and D₂ receptors. Indeed, treatment of transfected cells with the G $\alpha_{i/o}$ inhibitor, Pertussis toxin, prevented the dephosphorylation of HTT in response to D₂ receptor stimulation. Furthermore, the study revealed the formation of a protein complex comprising D₂

receptors, HTT and two members of the PPM/PP2C family. The first of these phosphatases, PPM1A, was shown to interact directly with HTT *in vivo* whereas the second phosphatase, PPM1B as well as HTT interact directly with the D₂ receptors. While it is not clear at the moment if PPM1A and B both participate in HTT dephosphorylation and the contribution of cAMP-mediated mechanisms has remained unexplored, the potential involvement of this mechanism in the regulation of HTT toxicity certainly warrants further investigations.

Transactivation of RTK by dopamine receptors

RTKs are a major family of cell surface receptors involved in many functions in neuronal and non-neuronal cell types (Lemmon and Schlessinger, 2010). Members of this family include, among others, the BDNF receptor neurotrophic tyrosine kinase, receptor, type 2 (TrkB), EGF/neuregulin family receptors (ErbB family) and receptors for insulin and insulin-like growth factor 1 (IGFR1). Activation of RTKs by their cognate ligands enhances receptor dimer formation, internalization, and recruitment of monomeric receptors to the cell surface. RTK activation generally results in a concomitant rapid activation of several signalling pathways, including PI3K/Akt, Ras/MAPK and PLC-mediated signalling (Figure 2).

In addition to direct activation by their ligands, RTKs can also be transactivated by GPCRs (Eguchi *et al.*, 1998; Maudsley *et al.*, 2000; Rajagopal *et al.*, 2004). However, the molecular mechanisms of this transactivation are not clearly understood. Independent investigations conducted in different systems have underscored the possible contribution of both G protein- and arrestin-mediated mechanisms involving either direct activation of RTK by intracellular processes or autocrine/paracrine RTK activation following ligand shedding in response to GPCR activation.

Dopamine receptors have been shown to transactivate RTKs in different experimental systems (Figure 3). The D₄ receptor was shown to transactivate the platelet-derived growth factor β (PDGF β) receptor, and D₂ receptors were able to transactivate IGF receptors in heterologous cell systems (Chi *et al.*, 2010; Mannoury la Cour *et al.*, 2011). Furthermore, both D₁ and D₂ receptors can transactivate ErbB-1 in transfected CHO-K1 cells (Swift *et al.*, 2011) and primary neuron cultures (Iwakura *et al.*, 2011; Yoon and Baik, 2013). Finally, D₁, D₂ and potentially D₁-D₂ receptor heteromers have been shown to transactivate the BDNF receptor in cultured striatal neurons (Iwakura *et al.*, 2008; Swift *et al.*, 2011; Barbeau *et al.*, 2013). Systemic administration of the D₁-class receptor agonist SKF 38393 also increased TrkB activation at 3 and 6 h following drug injection in 4-day-old rats (Iwakura *et al.*, 2008). The treatment of rats with the D₁ receptor antagonist SCH23390 reduced striatal TrkB activation, suggesting that transactivation of TrkB by D₁ receptors occurs in response to normal endogenous dopamine tone.

The mechanisms by which dopamine receptors transactivate RTKs are not fully understood. Quantitative pharmacological characterization of ErbB-1 receptors by various GPCRs has shown that these phenomena are not restricted by the coupling of the GPCR to a different G protein (Swift *et al.*, 2011). Increased release of the RTK ligand BDNF does not appear to be essential for the transactivation of TrkB by D₁

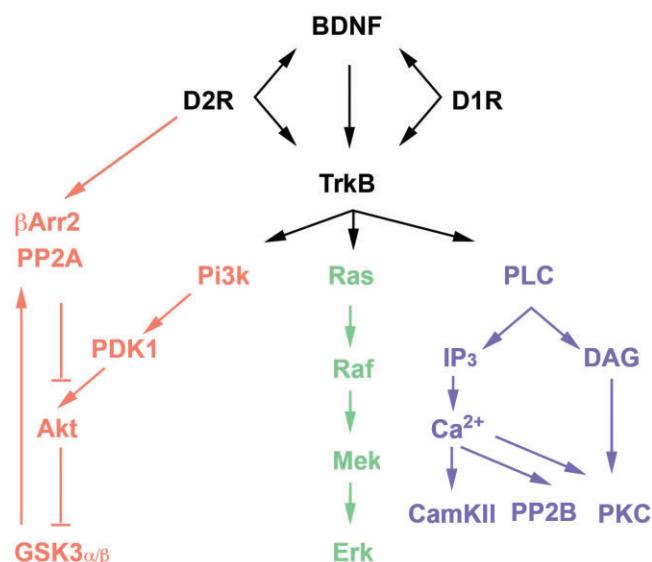


Figure 3

Mechanisms and signalling events involved in the transactivation of RTK by dopamine receptors. Raf, proto-oncogene serine/threonine-PK; Ras, rat sarcoma family of small GTPases.

receptors (Iwakura *et al.*, 2008). However, release of EGF appears to play a role in the transactivation of ErbB-1 by D₂ receptors in cultured neurons (Iwakura *et al.*, 2011; Yoon and Baik, 2013). Interestingly, stimulation of either frontal cortex D₅ receptors (Perreault *et al.*, 2013) or striatal D₁-D₂ receptors (Hasbi *et al.*, 2009) has also been reported to increase BDNF levels, therefore potentially leading to increased TrkB activation in response to dopamine receptor stimulation.

Transactivation of RTKs by dopamine receptors can have a major effect on our understanding of dopamine receptor signalling *in vivo*. RTKs are coupled to several signalling mechanisms that can elicit cellular responses, which are beyond the direct effect of G protein or arrestin-mediated cellular responses. For instance, although D₂ receptor stimulation leads to a β Arr2-dependent inactivation of Akt and concomitant activation of GSK3 *in vivo*, the opposite has been reported to occur in heterologous cell systems and, in some cases, cultured neurons (Brami-Cherrier *et al.*, 2005; Beaulieu, 2012). It has recently been shown that activation of Akt by recombinant D₂ receptors in transfected cells can be attributed to the transactivation of IGFR and concomitant activation of PI3K-mediated signalling by this RTK (Mannoury la Cour *et al.*, 2011). It is also possible that some instances of MAPK and PLC-mediated signalling in response to dopamine receptor activation may also result from RTK transactivation (Figure 3). The apparent involvement of RTK ligand release in the transactivation of some RTKs by dopamine receptors also raises the possibility that activation of dopamine receptors in dopaminergic neurons may elicit paracrine signalling responses to dopamine in either non-dopaminergic neurons or non-neuronal cells, possibly leading to indirect regional effects of dopamine receptor stimulation. This type of regional responses may be important considering the potential role of TrkB, and probably other RTKs, in regulating drug-induced reward (Lobo *et al.*, 2013).

Dopamine receptor oligomerization

Historically, GPCRs are believed to function as monomeric units, but now there is mounting evidence indicating that several GPCRs can exist in oligomeric forms (Perreault *et al.*, 2014). Regarding dopamine receptors, they can form both homomers and heteromers with several receptors, including other GPCRs and ionotropic glutamate receptors (Guo *et al.*, 2008; Van Craenenbroeck *et al.*, 2011; Perreault *et al.*, 2014). Some of these interactions may be regulated via mechanisms likely orchestrated by AC and cAMP (Woods and Jackson, 2013). A study by the Javitch group suggested that D₂ receptor homodimers and a G protein exist as the minimal single functional unit, which is maximally activated by the binding of an agonist to only one protomer and is either negatively or positively modulated by the ligand to the other protomer of an agonist or an inverse agonist respectively (Han *et al.*, 2009). This allosteric modulation between the two protomers of the complex is mediated through intermolecular interactions by the direct association among receptors and not by downstream effects. The development of RET-based techniques has been of fundamental importance in the discovery and characterization of many homomers and heteromers and is now considered to be the preferred biophysical method in describing complex formations (Milligan, 2004; Pflieger and Eidne, 2006; Marullo and Bouvier, 2007; Salahpour *et al.*, 2012). Both BRET and FRET rely on the principle of a non-radiative energy transfer between a donor protein and a fluorescent acceptor. In case of FRET, the donor is also a fluorescent protein (e.g., CFP), whereas in BRET, the donor is an enzyme (*Renilla* luciferase) that produces bioluminescence upon the degradation of a substrate (coelenterazine *h* or derivatives) (Pflieger and Eidne, 2006). Because the energy transfer is only possible when the donor and the acceptor are closer than 10 nm, when two proteins fused to a donor and an acceptor produce a BRET or FRET signal, it is an indication of a physical contact (Pflieger and Eidne, 2006; Marullo and Bouvier, 2007; Lohse *et al.*, 2012). However, a simple BRET or FRET signal is insufficient to distinguish true heterodimerization from a random collision; thus many experimental approaches have been adopted to characterize a putative heterodimer, such as saturation curves, competition assays and others (Marullo and Bouvier, 2007; Salahpour and Masri, 2007). Although these approaches have been extremely useful to study various complexes in cellular systems, these methods cannot simply be applied directly in native tissue, although there are some examples of successful applications *in vivo* using FRET with selective fluorescent ligands (Albizu *et al.*, 2010) or antibodies (Perreault *et al.*, 2010). After the *in vitro* description of the heterodimer and the characterization of its functional features, it is more common to prove the existence of the complex in native tissue using indirect evidence, such as identifying the 'biochemical fingerprint' and reproducing the specific characteristics of the complex and/or analysing physiological or behavioural responses to co-activation of the receptors (Ferre and Franco, 2010). In this section, we limit our discussion only to the heterodimers formed by dopamine receptors that have at least partial validation in studies in native tissue and/or *in vivo*.

D₁-D₂ receptor heterodimer

As mentioned earlier, D₁ and D₂ receptors can form a heterodimer complex that has been shown to exist in a heterologous system and in primary striatal neurons as well as *in vivo* in the rodent brain (Perreault *et al.*, 2013; 2014). Several techniques have been used to characterize this complex, ranging from classic biochemical approaches, including co-immunoprecipitation of the two proteins (Lee *et al.*, 2004), to more accurate techniques such as quantitative FRET (Dziedzicka-Wasylewska *et al.*, 2006; Hasbi *et al.*, 2009; Perreault *et al.*, 2010). The expression and cellular localization of D₁-D₂ receptor heterodimers has been characterized not only in cells, but also *in vivo*, primarily in rat striatum. Studies from BAC transgenic mice demonstrate that the majority of the D₁- and D₂ receptor-expressing neurons are segregated in two different populations, although a small percentage of neurons express both receptors, ranging from a 6% in caudate putamen to a 15–30% in nucleus accumbens (Bertran-Gonzalez *et al.*, 2008; Perreault *et al.*, 2010). These MSNs expressing D₁ and D₂ receptors are interesting in that they express both dynorphin and enkephalin; thus, one might consider them to be a third, distinct subset on MSNs. Among these, not all of the neurons show constitutive D₁-D₂ receptor heterodimer formation. Although a small proportion of caudate putamen MSNs revealed a D₁-D₂ receptor complex, in most of the neurons (90%) expressing both D₁ and D₂ receptors in the nucleus accumbens, these receptors are present as heterodimers (Perreault *et al.*, 2010). As mentioned earlier, this occurs under basal conditions, and it has been shown that several stimuli and pathological conditions could alter the state and the proportion of D₁-D₂ heterodimers (Dziedzicka-Wasylewska *et al.*, 2006; Perreault *et al.*, 2010). A peculiar aspect of this heterodimer is that it has a unique pharmacology that is distinct from that of its single protomer (Figure 1). Activation of the D₁-D₂ receptor complex induces the recruitment of the G $\alpha_{q/11}$ protein, leading to the release of calcium from the internal stores (Rashid *et al.*, 2007a,b; Hasbi *et al.*, 2009). It appears that D₁ and D₂ receptors are both necessary for this pathway, thus the application of dopamine or a combination of two selective D₁ and D₂ receptor agonists are able to increase intracellular calcium, whereas treatment with either a D₁ or D₂ receptor antagonist can abolish this effect (Hasbi *et al.*, 2009).

A selective D₁-D₂ receptor heteromer agonist (SKF 83959) has also been described: (Rashid *et al.*, 2007b). It should be noted, however, that a recent report argues the selectivity of this compound by showing that this effect is dependent on the level of G $\alpha_{q/11}$ expression in cells and there could be a contribution from the G $\beta\gamma$ subunits and GRK2 on the calcium increase induced by the D₁-D₂ receptor heterodimer (Chun *et al.*, 2013), while another report questioned in general the ability of SKF 83959 to influence PLC (Lee *et al.*, 2014). It has been suggested that this heteromer may play a role in brain disorders such as addiction (Perreault *et al.*, 2010), schizophrenia (Dziedzicka-Wasylewska *et al.*, 2008) and major depression (Pei *et al.*, 2010), although additional evidence is needed to support these hypotheses.

D₁-D₃ receptor heterodimers

D₁ and D₃ receptors are co-expressed in the majority of the substance P – expressing GABAergic medium spiny neurons,

suggesting that there could be functional crosstalk between these two receptors. Two independent studies have demonstrated that D₁ and D₃ receptors can form a constitutive heterodimer (Fiorentini *et al.*, 2008; Marcellino *et al.*, 2008). Using BRET and FRET techniques in transfected cells, it has been shown that D₁ and D₃ receptors can physically interact with no change in complex formation upon agonist treatment. Moreover, using co-immunoprecipitation, it was possible to isolate the D₁–D₃ complex from striatal membranes, confirming the existence of this heteromer in the brain. This cooperativity between D₁ and D₃ receptors is also evident in behavioural experiments. It is known that activation of D₁ receptors stimulates locomotor activity, whereas the role of D₃ receptors is less clear. In reserpinized mice (a model to isolate postsynaptic effects), D₃ receptor agonists can potentiate the stimulatory effects of D₁, but not D₂ receptor agonists. Furthermore, this potentiation can be counteracted with a D₃ receptor antagonist and is not present in D₃ receptor KO mice (Marcellino *et al.*, 2008). It has been suggested that the functional synergy of the D₁–D₃ dimer could be important for processes related to drug addiction and L-DOPA-induced dyskinesia in Parkinson's disease. It will be of interest to further characterize the physiological relevance of the D₁–D₃ heterodimer in other dopamine-related functions and pathologies.

D₂–D₄ receptor heterodimers

Both the long and the short D₂ receptor isoforms can associate with D₄ receptors, (Borroto-Escuela *et al.*, 2011b; Gonzalez *et al.*, 2012b). Using BRET, co-immunoprecipitation and proximity ligation assay, it has been shown in cell culture that D_{2L} receptors can exist in a heterodimeric form with the major variants of the D₄ receptors: D_{4.2}, D_{4.4} and D_{4.7}, with D_{4.7} being the least effective in forming the complex (Borroto-Escuela *et al.*, 2011b). Ferrè and colleagues showed with BRET that D_{2S} receptors can also associate in a heterodimer complex with the two variants D_{4.2} and D_{4.4}, but not with the variant D_{4.7} (Gonzalez *et al.*, 2012b). This study revealed a biochemical fingerprint for this heteromer that could potentiate D₄ receptor activation of MAPK. In the mouse striatum, although the single activation of either D₂ or D₄ receptors had no effect on MAPK, co-administration of D₂ and D₄ receptor agonists induced a strong ERK phosphorylation response (Gonzalez *et al.*, 2012b). This synergistic activity was lost in knock-in mice carrying the D_{4.7} variant, demonstrating the lack of mutual functional interaction between these two receptors. Moreover, the same synergistic effect was observed as regard to the ability of D₄ receptors to modulate glutamate release in the striatum (Gonzalez *et al.*, 2012b).

Dopamine receptor/NMDA receptor heteromer

Many studies have reported D₁ receptor-mediated modulation of NMDA activity, primarily through a G protein-dependent mechanism involving the cAMP/PKA pathway and proteins such as DARPP-32 (Blank *et al.*, 1997). However, there is evidence indicating that NMDA receptors and D₁ receptors could also interact physically. A first study demonstrated the existence of this complex in hippocampal extracts by co-immunoprecipitation (Lee *et al.*, 2002). It has been described that there are two sites of interaction in the carboxy-terminal domain of D₁ receptor: one interacts with

the GluN1 (NR1) subunit and the other with the GluN2A (NR2A) subunit. Each of these sites is responsible for a specific functional characteristic of the complex. These data were confirmed by another group in BRET studies that showed D₁ receptors and GluN1 forming a constitutive heterodimer in cells (Fiorentini *et al.*, 2003). Interestingly, the D₁–GluN1 complex is formed early in the ER and is translocated to the membrane only upon association with the GluN2B (NR2B) subunit (Fiorentini *et al.*, 2003). When at the cell membrane, the D₁ receptors involved in this complex lose the ability to desensitize and internalize upon stimulation, suggesting a dual role of this receptor in native tissue depending on micro-domain localization and aggregation with NMDA receptors, as previously reported (Dumartin *et al.*, 1998). The activation of D₁ receptors leads to a decrease of NMDA receptor functionality, as measured by NMDA-mediated currents in HEK cells and hippocampal neurons. This loss of activity is likely due to a decrease of NMDA receptor expression to the plasma membrane mediated by GluN2A subunit (Lee *et al.*, 2002). D₁ receptor agonists could reduce the cytotoxicity induced by an overactivation of the NMDA receptors. This effect appears to be mediated through a PI3K mechanism dependent on the GluN1 subunit instead of the reduction of the calcium influx (Lee *et al.*, 2002). In another study (Nai *et al.*, 2010), D₁ receptor activation in hippocampal slices increased the NMDA-dependent long-term potentiation, as previously reported (Huang and Kandel, 1995). This effect could be abolished by disrupting the heteromer with specific interfering peptides. This interaction can have important functional consequence on cognition, as disruption of the D₁–NMDA complex led to working memory impairment (Nai *et al.*, 2010).

Another reported interaction with NMDA receptors is between D₂ receptors and GluN2B (Liu *et al.*, 2006). It has been demonstrated that in both the dorsal striatum and nucleus accumbens, D₂ receptors and GluN2B are clustered in the PSD and form a complex that is more prevalent upon systemic cocaine treatment. Cocaine is also able to produce a selective decrease in the phosphorylation of Ser¹³⁰³ on the GluN2B subunit. This decrease is D₂ receptor-dependent, blocked by antagonist treatment, enhanced by agonist treatment and mediated by CaMKII. Cocaine treatment induces a D₂ receptor-dependent decrease in CaMKII activity, and its association with GluN2B results in the decrease of phosphorylation of Ser¹³⁰³. Functionally, cocaine application to striatal neurons causes a decrease in NMDA-mediated currents, an effect that can be abolished by heteromer disruption using a selective peptide that prevents the binding between the third intracellular loop of D₂ receptors and the carboxy-terminal tail of GluN2B. Furthermore, disruption of the D₂–GluN2B heteromer prevents phosphorylation of GluN2B and reduces cocaine-stimulated locomotor activity.

Adenosine receptor/dopamine receptor complexes

Many studies have shown that adenosine and dopamine exert opposing effects in the basal ganglia, with adenosine receptor agonists generally suppressing motor response and antagonists inducing motor activation (Ferre *et al.*, 1997). Because the adenosine A₁ receptor is expressed mainly in striato-nigral neurons, which also express dopamine D₁ receptors, and the adenosine A_{2A} receptor is expressed

predominantly in striato-pallidal neurons, which express D₂ receptors, it is suggested that A₁-D₁ and A_{2A}-D₂ heteromers might exist and have discrete localization in the basal ganglia (Fuxe *et al.*, 2010). Thus, heterodimer formation has been reported and extensively characterized for the A₁-D₁ complex (Gines *et al.*, 2000; Toda *et al.*, 2003) and the A_{2A}-D₂ complex (Hillion *et al.*, 2002; Canals *et al.*, 2003) using different biophysical approaches. In both complexes, both A₁ and A_{2A} receptor activation can antagonize the cAMP responses upon either D₁ or D₂ receptor stimulation in transfected cells (Ferre *et al.*, 1991; Gines *et al.*, 2000). For these reasons, it has been proposed that these complexes could be of importance as novel approach to treat some dopamine-related diseases such as Parkinson's disease and schizophrenia. Particularly, A_{2A} receptor antagonists, which can act as potentiators of the dopamine response, could be applied as adjunct therapy with D₂ receptor agonists and L-DOPA for the locomotor impairments present in Parkinson's disease patients (Fuxe *et al.*, 2007). In fact, A_{2A} receptor antagonists have been shown to improve motor deficits in several animal models of Parkinson's disease as well as demonstrate anti-parkinsonian properties in clinical trials (Fuxe *et al.*, 2008) and reduce L-DOPA-induced dyskinesia (Chase *et al.*, 2003). At the same time, A_{2A} receptor agonists showed an antipsychotic-like profile in different schizophrenia models, both in rodents (Rimondini *et al.*, 1997) and non-human primates (Andersen *et al.*, 2002). The mechanism proposed on the basis of these effects is the ability of A_{2A} receptor agonists to reduce D₂ receptor agonist binding and G protein coupling, particularly in the nucleus accumbens. Because the A_{2A}-D₂ heteromer can internalize as one unit, these agonists can also reduce D₂ receptor availability at the plasma membrane (Hillion *et al.*, 2002).

Other dopamine receptor heterodimers

Another example how dopamine receptors can alter their signalling pathways comes from a study on the D₂-ghrelin receptor heterodimer (Kern *et al.*, 2012). The ghrelin receptor is a GPCR expressed in different regions of the brain that can be activated by the stomach peptide ghrelin (Kojima *et al.*, 1999). In the hypothalamus, the ghrelin receptor and D₂ receptors are co-expressed, and using time-resolved FRET experiments, it has been shown that these two receptors form a heterodimer in native tissue (Kern *et al.*, 2012). Moreover, when co-expressed with the ghrelin receptor, D₂ receptors are able to elicit a calcium response dependent on PLC activation, IP₃ receptors and intracellular calcium stores. Interestingly, G α_q is not responsible for this increase, but rather the G $\beta\gamma$ subunit, derived from G α_i activation, mediates this effect likely through the direct association with GRK2 and direct stimulation of PLC (Inglese *et al.*, 1994; Koch *et al.*, 1994). This heterodimer formation appears to have an important role in appetite, as the suppression of food intake induced by a dopamine D₂ receptor agonist was prevented by a ghrelin antagonist and was absent in ghrelin-KO mice, demonstrating the importance of this heterodimer in physiology.

One interesting example how dimerization can alter the physiological functions of each protomer is the case of the D₄- β_1 adrenoceptor and D₄- α_{1B} adrenoceptor heteromers (Gonzalez *et al.*, 2012a). D₄ receptors are highly expressed in the pineal gland, and their expression follows a circadian rhythm, being high at the beginning of the light cycle and

low at the end (Bai *et al.*, 2008). The role of the pineal gland is to translate light inputs from the retina by producing and secreting melatonin, a product of serotonin. In this brain region, β_1 and α_{1B} adrenoceptors are the primary receptors that control this mechanism. In transfected cells, Gonzalez and colleagues proved the existence of these two heterodimers using BRET, and they also used a proximity ligation assay to demonstrate their presence *in vivo* in the pineal gland. Similar to the circadian nature of D₄ receptors, these complexes were found only at the beginning of the light cycle (Gonzalez *et al.*, 2012a). A distinctive biochemical property of these heterodimers was the modulation of ERK and Akt activity. When co-expressed in the same cells or when naturally present in native tissue, treatment with a D₄ receptor agonist was able to reduce ERK and Akt phosphorylation induced by β_1 and α_{1B} adrenergic receptors. Similarly, D₄ receptor antagonists prevent MAPK and Akt activation induced by β_1 or α_{1B} adrenoceptors, and adrenoceptor antagonists could block D₄ receptor stimulation of these pathways, showing a cross-antagonist property of this heterodimer. This functional interaction between D₄, β_1 and α_{1B} receptors was also reflected by the production and release of serotonin and melatonin, with a circadian regulation mediated by D₄ receptors and its modulation of β_1 and α_{1B} adrenoceptor activity (Gonzalez *et al.*, 2012a).

One GPCR that has emerged as a novel modulator of the dopamine system is the trace amine-associated receptor 1 (TA₁, known also as TAAR1) (Borowsky *et al.*, 2001; Sotnikova *et al.*, 2009; Espinoza *et al.*, 2011). The TA₁ receptor is expressed in several areas innervated by dopaminergic terminals such as the basal ganglia and frontal cortex, as well as in regions containing monoaminergic nuclei, including dopaminergic neurons in the ventral tegmental area (Lindemann *et al.*, 2008). Several studies have shown that the TA₁ receptor can modulate dopaminergic activities such as D₂ receptor function and the firing rate of dopaminergic neurons (Lindemann *et al.*, 2008; Bradaia *et al.*, 2009; Espinoza *et al.*, 2011). Moreover, TA₁ receptor agonists have been shown to influence effects of a wide range of dopaminergic agents and related behaviours such as amphetamine-induced hyperactivity (Revel *et al.*, 2011) and cocaine self-administration (Revel *et al.*, 2012), and are effective as antipsychotic compounds in several models of schizophrenia against positive, negative and cognitive symptoms (Revel *et al.*, 2013). One mechanism through which TA₁ receptors can influence the dopamine system is by forming a heterodimer with D₂ receptors. Using BRET, it has been demonstrated that TA₁ receptors can form a constitutive heterodimer with D₂, but not with D₁ receptors (Espinoza *et al.*, 2011; 2013), and that this heteromer can be disrupted by the D₂ receptor antagonist haloperidol. This functional interaction exerts its effect in modulating the cAMP pathway and haloperidol could potentiate TA₁ receptor-mediated cAMP signalling in cells. Furthermore, haloperidol's effects, such as *c-Fos* induction in the striatum and catalepsy, were reduced in TA₁ receptor-KO mice indicating prominent interaction between D₂ and TA₁ receptors at the levels of postsynaptic structures (Espinoza *et al.*, 2011). The D₂-TA₁ receptor interaction can also play an important role in presynaptic regulation of dopaminergic transmission by modulating D₂ receptor autoreceptor functions via TA₁ receptors (Leo *et al.*, 2014).

Other developments and emerging trends

Newly identified functions mediated by dopamine receptors

Recent studies that often employed newly developed approaches and tools have revealed novel functions mediated by dopamine receptors and clarified the contribution of specific subtypes to previously established functions. Several observations regarding the functional role of dopamine receptors have been made using transgenic techniques. In addition to gene KO studies, which have been described in previous reviews (Sibley, 1999; Holmes *et al.*, 2004; Beaulieu and Gainetdinov, 2011), mice overexpressing D₂ and D₃ receptors have been recently characterized. Particularly, mice selectively overexpressing D₂ receptors in the striatum have persistent abnormalities in prefrontal cortex function and deficits in working memory and behavioural flexibility (Kellendonk *et al.*, 2006), as well as motivation (Simpson *et al.*, 2012) and timing precision (Ward *et al.*, 2009) deficits that are often found of schizophrenia models. These mutants also demonstrated altered dendritic morphology of medium spiny neurons via involvement of K_v2 channels (Cazorla *et al.*, 2012), and a deficit in inhibitory GABA-mediated transmission and dopamine sensitivity in the prefrontal cortex (Li *et al.*, 2011b). Taken together, these studies indicated an important role for the striatal processes in the pathogenesis of the cognitive symptoms of schizophrenia. Based on this model, overstimulation or excessive D₂ receptor activity in the striatum leads to altered functioning of prefrontal cortex neurons via several mechanisms, culminating in deficiencies in executive function and working memory – key components of schizophrenia-related deficits (Simpson *et al.*, 2010). In contrast, mice overexpressing D₃ receptors in the striatum have less pronounced, but still significant phenotype. They do not demonstrate cognitive deficits, but show disrupted motivation, suggesting that targeting D₃ receptors may have effect on motivational symptoms, which are not improved by the currently available antipsychotics in schizophrenic patients (Simpson *et al.*, 2014).

In another study, a role of subpopulation of D₁ receptor-expressing cholinergic neurons in putative antipsychotic action of the M₄ muscarinic receptor agonist xanomeline was demonstrated (Dencker *et al.*, 2011). In mutant mice lacking the M₄ muscarinic receptors only in D₁ dopamine receptor-expressing cells, the antipsychotic-like effects of xanomeline were completely abolished suggesting that M₄ muscarinic receptors co-localized with D₁ receptors are involved.

Several recent observations have indicated that the direct modulation of the DAT function by D₂ receptors might occur (Chen *et al.*, 2013) and this interaction has physiological or pathological relevance (Bowton *et al.*, 2010; Owens *et al.*, 2012); however, no alterations in DAT function were found in D₂ autoreceptor KO mice (Bello *et al.*, 2011). Based on studies involving mice that constitutively express only the short isoform D_{2S} or lacking both isoforms, it has been proposed that D_{2L} is the major postsynaptic isoform expressed in medium spiny GABA neurons while D_{2S} is predominantly expressed on presynaptic terminals and involved in autoreceptor function (Lindgren *et al.*, 2003). However, recent study

with virus-mediated receptor restoration in D₂ receptor KO mice has indicated that both these alternatively spliced forms of the D₂ receptors are equally capable of acting as postsynaptic receptors and autoreceptors (Neve *et al.*, 2013). Interestingly, while the role of presynaptic D₂ receptors in autoreceptor regulation of dopamine neuron firing rate, as well as dopamine synthesis and release is well established (Bello *et al.*, 2011), a recent study involving cell-specific KO mice has shown a significant contribution of postsynaptic D₂ receptors in the local feedback regulation of dopaminergic transmission in the dorsal striatum as well (Anzalone *et al.*, 2012). As D₃ receptors are also shown to contribute to the regulation of dopamine release and are expressed both at the presynaptic terminals and postsynaptic structures (Gainetdinov *et al.*, 1996; Joseph *et al.*, 2002; Gross *et al.*, 2013), it would be of interest to explore if postsynaptic D₃ receptors could be involved in the regulation of presynaptic transmission via similar local feedback mechanisms.

Application of optogenetic techniques has allowed for more precise anatomical and cellular dissection of the role of specific dopamine receptors in dopamine-related functions. An important advance in understanding the complex interplay of basal ganglia output by direct and indirect pathway projection neurons in regulating movement has been achieved (Kravitz *et al.*, 2010; Freeze *et al.*, 2013). Application of optogenetic and pharmacological approaches in transgenic mice has highlighted an important role of striatal D₂ receptors in the concerted balance of the striatal output system and structural plasticity (Cazorla *et al.*, 2014). Distinct roles for striatal neurons in the direct and indirect pathways in reinforcement have been shown by demonstrating that optogenetic stimulation of D₁ receptor-expressing neurons induce persistent reinforcement whereas stimulation of D₂ receptor-expressing neurons induce transient punishment (Kravitz *et al.*, 2012). The role of medial prefrontal D₁ receptor-expressing neurons in the control of food intake (Land *et al.*, 2014) and temporal control (Narayanan *et al.*, 2012) has also been demonstrated. Recent evidence indicates that optogenetic inhibition of D₁ but not D₂ receptor-expressing medium spiny neurons in the nucleus accumbens alters cocaine-mediated regulation of the T-lymphoma invasion and metastasis 1 (Tiam1) protein, which has been implicated in structural and synaptic plasticity (Chandra *et al.*, 2013). Furthermore, using a combination of optogenetic and chemogenetic approaches based on designer receptors exclusively activated by designer drugs (DREADD) technology (Lee *et al.*, 2013), it has been shown that strengthening the accumbal indirect pathway via D₂ receptors promotes resilience to compulsive cocaine use (Bock *et al.*, 2013).

A role of D₂ receptor-related signalling in suppressing human osteoclastogenesis has been recently identified (Hanami *et al.*, 2013), which may provide a plausible mechanism for the skeletal effects of antipsychotics observed in children and adolescents (Calarge *et al.*, 2013). A complex role of D₁ and D₂ receptors has been demonstrated for epileptogenesis (Bozzi and Borrelli, 2013), and cochlear functions (Maison *et al.*, 2012). Evidence gained from humans and mice also indicates that D₄ receptors may contribute to longevity (Grady *et al.*, 2013). Studies involving D₄ receptor KO mice and selective D₄ receptor agonists and antagonists in animal models of drug-seeking and drug-taking behaviours have sug-

gested that treatments based on antagonism of the D₄ receptor may be effective approaches for the management of psychostimulant and nicotine abuse (Di Ciano *et al.*, 2014). That presynaptic D₂ receptors might be involved in cocaine- and nicotine-induced structural plasticity in mesencephalic dopaminergic neurons via mechanism involving ERK and Akt signalling is also intriguing (Collo *et al.*, 2012; 2013). A novel mechanism for D₁ receptor-mediated regulation of ERK signalling involving the tyrosine phosphatase Shp-2, that is required for ERK activation by tyrosine kinase receptors, has been recently shown (Fiorentini *et al.*, 2011), and this mechanism appears to be involved in L-DOPA-induced dyskinesia in the experimental model of Parkinson's disease (Fiorentini *et al.*, 2013).

Recent evidence also indicated that dopamine receptors are involved in immune system regulation and processes related to inflammation and autoimmune reactions. It has been shown that astrocytic D₂ receptors modulate innate immunity through α B-crystallin, which results in suppression of neuroinflammation (Shao *et al.*, 2013). A role of D₂ receptors in renal inflammation has been demonstrated as well (Zhang *et al.*, 2012). It has been also observed that stimulation of D₅ receptors expressed on dendritic cells potentiates Th17-mediated immunity, thus indicating that D₅ receptors are able to modulate the development of an autoimmune response *in vivo* (Prado *et al.*, 2012).

Although the role of retinal dopamine and dopamine receptors in the regulation of vision and related processes is well established, recent studies have uncovered previously unappreciated mechanisms. Studies involving KO mice lacking specific dopamine receptors have shown that in addition to the role of dopamine in suppressing rod-driven signals in bright light, it also enhances the same signals under dim illumination via D₁ receptor-dependent sensitization of rod bipolar cells by GABA (Herrmann *et al.*, 2011). A role of both D₁ and D₄ receptors in various dimensions of light-adapted vision has been demonstrated as well (Jackson *et al.*, 2012). In a mouse model of type 1 diabetes, which develops early visual dysfunction because of dopamine deficiencies, acute treatment with either D₁ or D₄ receptor agonists improved overall retinal and visual function (Aung *et al.*, 2014). Retinal dopamine plays a critical role in the development of myopia predominantly via the D₂ receptor, but there is recent evidence suggesting that the balance of D₂ and D₁ receptor activation is important (Feldkaemper and Schaeffel, 2013). Evidence from KO mice suggests that not only retinal, but also central dopamine could play a role in the regulation of retinal function via differential involvement of D₁ and D₂ receptors (Lavoie *et al.*, 2014a). Furthermore, D₂ receptor-regulated GSK3 signalling also appears to contribute to electroretinogram anomalies observed in subjects at high genetic risk for schizophrenia and bipolar disorder, suggesting that electroretinograms can serve as a biomarker for central dopamine abnormalities related to psychiatric disorders (Lavoie *et al.*, 2014b).

It is well known that dopamine acting through D₂ receptors controls lactotroph proliferation and prolactin levels; however, the role of the D_{2S} and D_{2L} receptor isoforms was unknown. Recent investigations have shown that the presence of either the D_{2S} or D_{2L} isoforms *in vivo* prevents hyperprolactinaemia, the development of lactotroph hyperplasia,

and tumourigenesis, all of which are observed when both isoforms are deleted in mice (Radl *et al.*, 2013). However, the protective function of a single D₂ receptor isoform is overridden when single isoform-KO mice are challenged by chronic estrogen treatments, suggesting that signalling from both isoforms is necessary in conditions that simulate pathological states (Radl *et al.*, 2013).

Selective disruption of D₂ receptors in pituitary lactotrophs in conditional mutant mice results in an increase in body weight and adiposity (Perez Millan *et al.*, 2014), indicating that D₂ receptors might be involved in the metabolic side effects of antipsychotic drug treatment such as obesity, cardiovascular complications and increased incidence of type 2 diabetes (Reynolds and Kirk, 2010). Interestingly, the D₂ receptor agonist bromocriptine has been recently approved for clinical use for the management of type 2 diabetes mellitus, particularly in forms associated with cardiovascular deficits (Grunberger, 2013).

The general role of D₂ receptors and dopamine in endocrine tumours is supported by reports that D₂ receptor agonists are effective in controlling hormone secretion and cell proliferation in experimental studies (Gatto and Hofland, 2011). Furthermore, dopamine agonists have been found to be efficacious in a subgroup of patients with pituitary adenomas and a few reported cases of carcinoids (Gatto and Hofland, 2011). It has also been shown that dopamine, acting via D₂ receptors, blocks stress-mediated ovarian carcinoma growth (Moreno-Smith *et al.*, 2011) in part through the antiangiogenic effect of dopamine and in part through D₁ receptors that stimulates vessel stabilization by increasing pericyte recruitment to tumour endothelial cells (Moreno-Smith *et al.*, 2013). Interestingly, recent unbiased screening attempts to identify novel anti-cancer therapies yielded dopaminergic compounds such as the D₂ receptor antagonist thioridazine, which selectively targets cancer stem cells by inducing differentiation to overcome neoplastic self-renewal (Sachlos *et al.*, 2012), and perphenazine, which was found to be effective in T-cell acute leukaemia by inducing PP2A-mediated apoptosis (Gutierrez *et al.*, 2014).

Influence on glutamate and GABA transmission

One recurrent topic that has received significant attention recently is related to signalling mechanisms involved in the convergence of dopamine receptor-mediated signalling with glutamatergic and GABA neurotransmission. It is well known that, as a slow neurotransmitter, dopamine exerts its actions through the modulation of the effects of fast neurotransmitters such as glutamate and GABA. This multi-transmitter interaction is critical for many vital functions mediated by dopamine as well as related disorders such as schizophrenia. As presented in Figures 1 and 2, both D₁ and D₂ receptors have multiple potential signalling mechanisms that may be involved in this modulation. Dopamine receptors can alter the phosphorylation of critical subunits of glutamate AMPA receptors (GluA1) and NMDA receptors (GluN2B), via the G α subunit-mediated cAMP-PKA-DARPP32 signalling cascade (Dell'anno *et al.*, 2013; Hobson *et al.*, 2013; Song *et al.*, 2013; Flores-Barrera *et al.*, 2014; Murphy *et al.*, 2014), as well as G α_q and G $\beta\gamma$ subunit-mediated PLC signalling and subsequent Ca²⁺-dependent events (Jenkins and Traynelis, 2012; Jenkins

et al., 2014). Activation of the G protein-independent Akt/GSK3 signalling cascade by D₂ receptors could lead to significant alterations in glutamatergic signalling mediated by NMDA and AMPA receptors (Li *et al.*, 2009; Li and Gao, 2011a) as well as changes in kinesin-mediated AMPA receptor trafficking (Du *et al.*, 2010). Similar mechanisms can also affect GABA transmission (Li *et al.*, 2012). As discussed earlier, the direct interaction of dopamine receptors with GluN1 and GluN2B subunits of NMDA receptors has been demonstrated (Fiorentini *et al.*, 2003; Liu *et al.*, 2006). Several mechanisms of direct and indirect interaction of D₃ receptors with glutamate neurotransmission have also been documented (Sokoloff *et al.*, 2013).

Further understanding of the mechanisms of the intricate interaction of dopamine receptor-mediated signalling events with glutamate and GABA signalling could manifest into exciting new molecular targets for novel pharmacological approaches to dopamine-related disorders.

Search for biased ligands of dopamine receptors

The growing realization of the complexity of signalling mediated by GPCRs in general, and dopamine receptors in particular, has provided a theoretical framework for the development of pathway-specific biased ligand pharmacology of D₂ receptors (Beaulieu *et al.*, 2007a; Beaulieu and Gainetdinov, 2011). In addition to the studies described earlier (Masri *et al.*, 2008; Allen *et al.*, 2011), a structure-activity analysis of pathway-specific biased agonism at D₂ receptors based on aripiprazole derivatives (Chen *et al.*, 2012) and search for novel potential antipsychotic cariprazine derivatives (Shonberg *et al.*, 2013) and other D₂ receptor targeting compounds (Hiller *et al.*, 2013) have been performed. These studies have recently resulted in identification of several functionally selective D₂ receptor agonists that are biased towards G protein- versus β Arr2-dependent signalling (Free *et al.*, 2014; Möller *et al.*, 2014). Future characterization of the biochemical and behavioural effects of these biased compounds could eventually result in the development of improved antipsychotic drugs with reduced propensity of undesirable side effects. It should be noted that although the majority of current studies are focused on the development of biased ligands targeting either G protein-dependent or β Arr2-dependent signalling mechanisms, it might be expected that biased ligands that, for example, will specifically target one, but not another modality of G protein-independent signalling could be identified in future (Figures 1 and 2).

Allosteric modulators of dopamine receptors

Another exciting direction in the pharmacology of GPCRs that is gaining attention recently is related to the development of allosteric modulators (Nickols and Conn, 2014). Although classic drug discovery approaches targeting GPCRs have traditionally focused on developing ligands for orthosteric sites, which bind endogenous ligands, more recent efforts have been aimed at modulating receptor function via allosteric modulators, which target a site distinct from the orthosteric site. It is expected that the development of such positive or negative allosteric modulators or 'bitopic' ligands that interact with both the allosteric and the orthosteric sites could eventually result in increased drug selectivity for thera-

peutic action and potentially decreased adverse side effects. Thus, a specific allosteric modulator of D₂ receptors, the peptidomimetic 3(R)-[(2(S)-pyrrolidinylcarbonyl) amino]-2-oxo-1-pyrrolidineacetamide (PAOPA), has been recently characterized as binding to a site on the D₂ receptor that is distinct from the endogenous ligand binding site (Tan *et al.*, 2013). PAOPA can influence several signalling and cellular events characteristic of D₂ receptor activation (Basu *et al.*, 2013), and in preclinical models, can attenuate schizophrenia-like behavioural manifestations without causing significant motor abnormalities common with many current antipsychotics (Tan *et al.*, 2013).

Based on the crystal structure of D₃ receptors (Chien *et al.*, 2010) a virtual screen targeting allosteric sites of this receptor has been recently performed (Lane *et al.*, 2013). This screening resulted in the identification of a number of allosteric ligands for D₃ receptors, including chemically diverse compounds with a variety of functional activity profiles and high affinities and ligand efficiencies. It is believed that identification of the allosteric structural features of D₃ receptors that are essential to selectivity and efficacy could be critical for the identification of highly selective D₃ receptor ligands, which are notorious for their difficulty in development (Newman *et al.*, 2012). Allosteric modulation of certain ligands can also occur at the level of receptor heterodimers as demonstrated by the ability of a newly identified dopamine agonist to negatively modulate adenosine A_{2A} receptor binding properties by interacting with the A_{2A}-D₂ receptor heteromer (Trincavelli *et al.*, 2012). Interestingly, A_{2A} receptor agonist-induced modulation of D₂ receptor agonist-induced β Arr2 recruitment has been described, suggesting the involvement of a possible A_{2A}-D₂- β Arr2 complex in this allosteric modulation (Borroto-Escuela *et al.*, 2011a).

New dopamine receptor-based drugs

In addition to the clinically approved compounds targeting dopamine receptors that were discussed in our recent review (Beaulieu and Gainetdinov, 2011), several new compounds have either been approved for clinical use or are in the last stages of clinical trials. Essentially all of these compounds are antipsychotics targeting D₂ receptors. It should be noted that no clinically approved antipsychotic drugs exist to date that are not D₂ receptor blockers. Among the newer antipsychotics recently approved for the treatment of schizophrenia, the most conspicuous are iloperidone, asenapine, lurasidone and blonanserin, considered to be atypical antipsychotics (George *et al.*, 2013). These compounds at least partially target D₂ receptors with additional influence on other receptors, particularly 5-HT_{2A} receptors. Although they exert clear antipsychotic activity, essentially all of them produce metabolic side effects and hyperprolactinaemia, which require appropriate monitoring (Wang *et al.*, 2014). In addition to the olanzapine-fluoxetine combination, quetiapine, lurasidone is now FDA-approved for the acute treatment of bipolar depression (McIntyre *et al.*, 2013). Among several emerging antipsychotic compounds, cariprazine has attracted the most attention because this drug has partial agonist activity at D₂ and D₃ receptors, with a six- to eightfold higher affinity for the human D₃ receptor over the D₂ receptor. Cariprazine is at latest stages of development, and an application has been submitted to the FDA for approval as a treatment for schizo-

phrenia, bipolar mania and depression (Veselinovic *et al.*, 2013). Another interesting compound is brexpiprazole, a novel drug candidate in clinical development for psychiatric disorders with high affinity for D₂ dopamine, 5-HT_{2A} and adrenoceptors (Maeda *et al.*, 2014). Brexpiprazole was showing significant antipsychotic-like properties in several preclinical models suggesting that brexpiprazole is a 5-HT-dopamine activity modulator with a unique pharmacology that might provide novel treatment option for psychiatric disorders.

D₂ receptor antibodies in autoimmune encephalitis

One exciting direction in the current neuropsychiatry is the emergence of novel class of disorders related to autoantibodies against critical synaptic proteins such as NMDA receptors and D₂ receptors (Lancaster and Dalmau, 2012). Remarkably, some patients with these autoantibodies have symptoms virtually indistinguishable from schizophrenia or other neuropsychiatric or movement disorders. Antibodies to surface D₂ receptors were found in patients with autoimmune movement and psychiatric disorders (Dale *et al.*, 2012). D₂ receptor autoantibodies were found also in patients with Sydenham chorea (Cox *et al.*, 2013), and recent study reported that herpes simplex encephalitis relapse with chorea was associated with autoantibodies against D₂ receptors and the NMDA receptors (Mohammad *et al.*, 2014). This recently emerging field is growing monthly, with more patients identified, and it is expected that the autoimmune theory of neuropsychiatric and movement disorders will have significant support in the future.

Conclusions

The studies described here show the significant progress made in understanding dopamine receptor functions, the complexity of their signalling mechanisms and potential new applications of dopamine receptor-based pharmacological strategies. Using a variety of the most up-to-date approaches, multidimensional analysis of dopamine receptor biology will eventually provide an opportunity for the precise targeting of desired components of post-receptor intracellular processes either via receptor-related mechanisms or post-receptor signalling cascades, thereby providing an exciting opportunity to target pathological processes with minimal propensity of developing side effects. Such approaches involving 'biased agonism', allosteric-based targeting of receptors and heteromers and downstream intracellular signalling events could eventually result in emergence of a new generation of dopamine receptor-based therapies for a variety of dopamine-related disorders.

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Author contributions

All authors contributed equally to this work.

Conflict of interest

The authors declare no conflict of interest in writing this paper.

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