

RESEARCH ARTICLE

Open Access

Influence of environmental factors in the adherence of an atypical enteropathogenic *Escherichia coli* strain to epithelial cells

Fabiano T Romão¹, Rodrigo T Hernandes^{1,2}, Denise Yamamoto¹, Lika Osugui¹, Ana Flavia Popi¹ and Tânia A T Gomes^{1*}

Abstract

Background: Attachment is essential to maintain bacteria at their preferential intestinal colonization sites. There is little information on the influence of different environmental conditions in the interaction of atypical enteropathogenic *Escherichia coli* (aEPEC) strains with epithelial cells. In this study, we evaluated the effect of different glucose (5 and 25 mM) and CO₂ (0.03 and 5%) concentrations and presence of bile salts on the adhesiveness of the aEPEC strain 1551–2.

Results: We found that a CO₂-enriched atmosphere enhanced the adhesiveness of the aEPEC 1551–2 strain independently of glucose concentrations or presence of bile salts. Conversely, the presence of high glucose concentration altered the original localized adherence (LA) pattern observed at 5 mM glucose, which is characterized by the formation of compact bacterial clusters, to a hybrid adherence pattern (LA and an aggregative adherence-like pattern). In addition, at high glucose concentration, there was increased expression of the *fimA* gene, which encodes the major subunit of type 1 pilus (T1P), and an isogenic *fimA* mutant displayed only LA. The presence of bile salts did not interfere with the adhesion properties of the 1551–2 strain to HeLa cells.

Conclusions: Our data suggest that a CO₂-enriched atmosphere could favor aEPEC adhesion to the host cells, whereas enhanced T1P production under high glucose concentration could allow bacteria to access more extensive intestinal colonization sites in the host at the beginning of the infectious process.

Keywords: Atypical EPEC, Glucose, type I pilus, Adherence, Environmental factors, Diarrhea

Background

Enteropathogenic *Escherichia coli* (EPEC) is an important cause of gastroenteritis in humans [1]. The EPEC pathotype is subdivided into typical EPEC (tEPEC) and atypical EPEC (aEPEC), where the main difference between the two subgroups is the presence of the large virulence EAF (EPEC adherence factor) plasmid (pEAF) in tEPEC and its absence in aEPEC [2,3]. pEAF encodes bundle-forming pilus (BFP), a type IV fimbria, which mediates a localized adherence (LA) pattern *in vitro*, and is characterized by the formation of compact microcolonies on the surface of HeLa/HEp-2 cells [4,5].

The hallmark of EPEC pathogenesis is a histopathologic lesion termed attaching and effacing (AE) lesion, which is characterized by intimate bacterial adherence to enterocytes and effacement of microvilli [6]. The AE lesion is a consequence of the interaction between the outer membrane protein intimin and its receptor Tir (translocated intimin receptor), which is translocated through a type 3 secretion system (T3SS) into eukaryotic cells [7,8]. The genes involved in the AE lesion phenotype are localized in the pathogenicity island termed locus of enterocyte effacement or LEE region [9].

Attachment comprises a critical stage to avoid displacement of bacteria from its preferential site in the gut by the continuous flow of intestinal contents. The initial step in attachment of different diarrheagenic *E. coli* (DEC) pathotypes to the host intestinal epithelium is usually mediated

* Correspondence: tatg.amaral@unifesp.br

¹Departamento de Microbiologia, Imunologia e Parasitologia, Universidade Federal de São Paulo (UNIFESP), Escola Paulista de Medicina, São Paulo, SP, Brazil

Full list of author information is available at the end of the article

by fimbrial and afimbrial adhesins [10,11]. Previous studies have demonstrated that environmental conditions may regulate the expression of DEC virulence-encoding genes including adhesins and T3SS-dependent effectors [12-15]. In a tEPEC strain, serotype O111:H⁻, Puente and coworkers found that BFP expression was increased at higher glucose concentrations, suggesting that tEPEC preferentially colonizes the proximal small intestine, where the glucose concentration is greater than 25 mM [16]. This hypothesis was further confirmed in experiments employing an *in vitro* organ culture (IVOC) adhesion assay with intestinal tissues obtained from children [17].

Several studies in the literature have focused on identifying new putative adhesive structures, and the adherence pattern resulting from the interaction between these bacterial structures and host cells. However, particularly in aEPEC, very few studies have tried to understand how the intestinal environment may favor or prevent this interaction.

We previously identified an aEPEC strain, 1551-2 (serotype ONT:H⁻), which displays the LA pattern in HeLa cells in the absence of BFP [18]. In the present study we evaluated the adherence pattern and colonization efficiency of aEPEC 1551-2 in HeLa cells cultivated at different CO₂ and glucose concentrations and in the presence of bile salts.

Results

aEPEC 1551-2 strain adheres more efficiently to HeLa cells in an atmosphere supplemented with CO₂

Quantitative assays demonstrated that the different glucose concentrations tested (5 and 25 mM) did not promote substantial alterations in the adhesiveness of aEPEC 1551-2 ($P > 0.5$), as observed in Figure 1A. However, air atmosphere with 5% CO₂ favored bacterial-cell interaction, since the number of associated bacteria increased 2.3-fold at 5 mM glucose and 1.7-fold at 25 mM glucose, when compared with results obtained in assays performed in a normal air atmosphere (approximately 0.03% CO₂) ($P < 0.05$).

To exclude the possibility of differences in bacterial growth rate in the different environmental conditions tested, a growth curve (measuring the OD₆₀₀) was determined. Bacterial growth rates were not affected by the different conditions tested (data not shown), indicating that the difference observed in the quantitative adherence assays was due to the modification of the air atmosphere by the elevation of CO₂ concentration to 5%.

High glucose concentration modifies the adherence pattern of 1551-2 strain to HeLa cells

Regarding the aEPEC 1551-2 adherence pattern, the LA originally identified by Vieira and coworkers [18] was

only detected in the presence of low glucose, independently of the CO₂ concentration used (Figure 1B and D). On the other hand, at high glucose concentration, we identified a hybrid adherence pattern, in which LA was observed concurrently with an aggregative adherence-like (AA-like) pattern on the HeLa cell and on the glass slide surface (abiotic surface) (Figure 1C and E).

The AA-like pattern of 1551-2 strain is dependent on type 1 pilus (T1P) production at high glucose concentration

Since T1P is so far the only adhesin, besides intimin, identified in the 1551-2 strain [18-20], we hypothesized that the AA-like pattern would depend on T1P production. To investigate the contribution of T1P in the establishment of the AA-like pattern, we first evaluated the relative expression of the *fimA* gene (encodes for monomer that forms fimbriae) at two glucose concentrations (5 and 25 mM). By comparing the experiments performed in medium supplemented with different glucose concentrations, it was possible to observe an increase in *fimA* gene expression (1.75-fold and 2.03-fold, after 3 and 6 h of incubation, respectively) in experiments performed in medium with high glucose (Figure 2A).

Furthermore, we employed an isogenic *fimA* mutant (1551-2Δ*fimA*) to test the adherence pattern of this strain at high glucose. These experiments demonstrated that in contrast with the wild type aEPEC 1551-2 (Figure 2B), the 1551-2Δ*fimA* strain displayed only the LA pattern, while the AA-like pattern was completely abolished (Figure 2C). These findings clearly point to an association between glucose concentration and T1P production, since the isogenic mutant no longer produced the AA-like pattern, even in the presence of high glucose concentration (Figure 2C). Complementation of the *fimA* mutant with the pFimA recombinant plasmid restored its ability to produce the LA/AA-like pattern in the presence of high glucose (Figure 2E). The 1551-2Δ*fimA* strain carrying empty pBADMyc-His A vector (Table 1) produced LA on HeLa cells but was unable to produce the AA-like pattern (Figure 2D), confirming that L-arabinose did not affect the adherence phenotype of the complemented strain.

Presence of bile salts does not interfere with the adhesion properties of 1551-2 strain with HeLa cells

The presence of bile salts caused no significant alterations in the adhesive properties of aEPEC 1551-2 to epithelial cells (data not shown). Under all conditions tested, the adherence pattern and the amount of bacteria associated with HeLa cells were not significantly different, indicating that bile salts did not influence bacterial colonization.

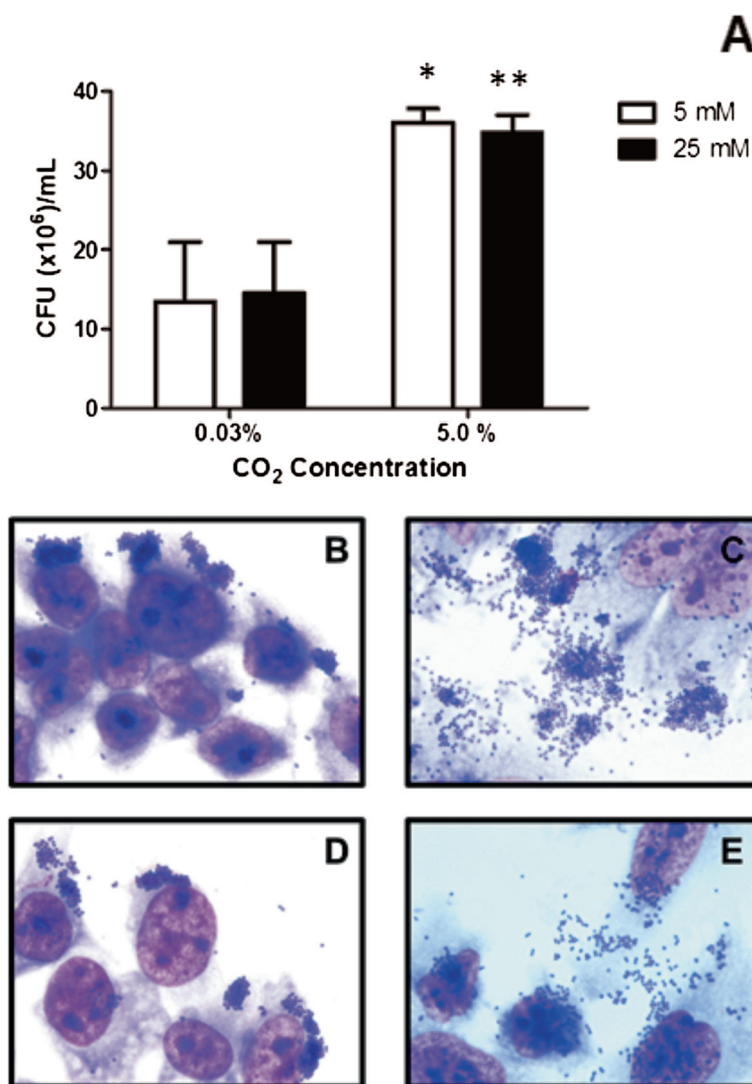


Figure 1 Comparison of quantitative and qualitative adherence of aEPEC 1551-2 strain to HeLa cells under different environmental conditions. **(A)** Number of adherent bacteria recovered after 6 h of incubation with HeLa cells, demonstrating the positive influence of CO₂ (5%) in the adhesiveness of aEPEC 1551-2. **(B, C, D and E)** Distinct adherence patterns observed in 1551-2 interaction with HeLa cells, after incubation in medium containing 5 mM **(B and D)** or 25 mM **(C and E)** glucose, in a normal air atmosphere **(B and C)** and in an atmosphere of 5% CO₂ **(D and E)**. The hybrid LA/AA-like pattern is observed over and around cells, only in assays performed in medium containing 25 mM glucose **(C and E)**. * and ** represent statistical differences observed between experiments performed under 0.03% and 5% CO₂ concentration, in medium containing low ($P = 0.03$) or high ($P = 0.002$) glucose concentration, respectively.

Discussion

In this study, we analyzed some environmental factors that could influence the colonization process of aEPEC 1551-2 strain in the intestinal environment, by mimicking characteristics (such as the gas atmosphere and glucose concentration) commonly found in different parts of the intestines.

We observed that the efficiency of bacterial association rose approximately two fold in a 5% CO₂ atmosphere, independently of the glucose concentration employed (Figure 1A). According to described by Babb (1977), hydrogen, nitrogen and CO₂ are the three prevailing

gases present in the intestine, with CO₂ concentrations varying between 5.1 and 29% [21]. Due to experimental limitations, only 0.03% and 5% CO₂ were tested.

Haigh and co-workers showed improved protein secretion by the tEPEC E2348/69 strain in the presence of 5% CO₂, independently of the presence of eukaryotic cells [22]. However, Kenny and coworkers questioned the actual influence of high CO₂ concentrations in improving bacterial protein secretion, since, in the adherence assays performed in a 5% CO₂ atmosphere, sodium bicarbonate is added to the culture medium to buffer the acidic pH induced by the higher CO₂ concentration [8]. Therefore,

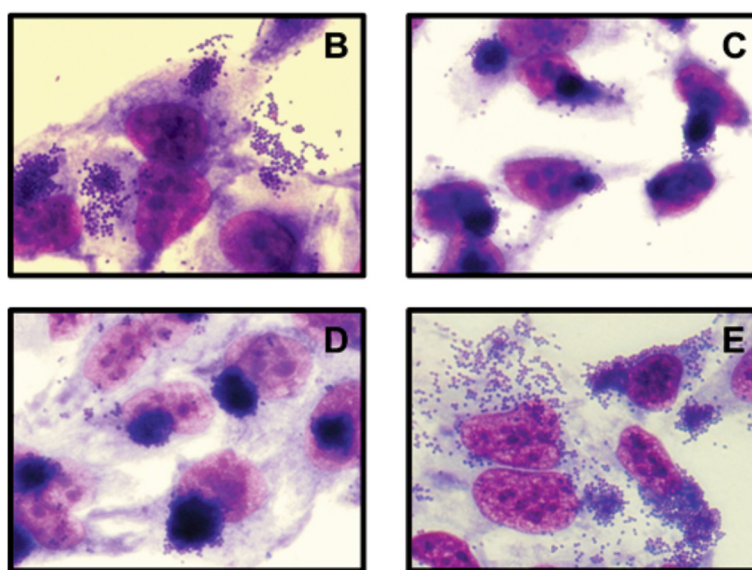
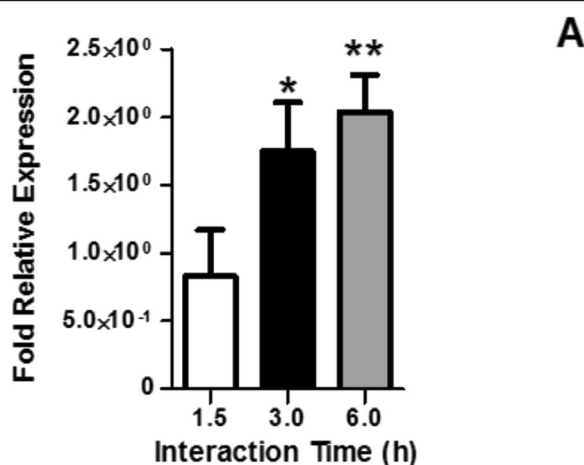


Figure 2 The aggregative adherence-like pattern shown by aEPEC 1551–2 at high glucose concentration is dependent on type 1 pili production. **(A)** Relative expression of *fimA* in presence of 5 mM (calibrator) and 25 mM glucose at 1.5 h, 3 h and 6 h. The data showed that at 3 and 6 h, there was an increase in *fimA* expression of $75 \pm 21\%$ and $103 \pm 15\%$, respectively. **(B, C, D and E)** Adherence patterns observed with the wild-type strain **(B)**, 1551-2Δ*fimA* **(C)**, 1551-2Δ*fimA* (pBAD) **(D)** and 1551-2Δ*fimA* (pFimA) **(E)**. The 1551-2Δ*fimA* mutant maintained the LA pattern on HeLa cells, but the AA-like pattern was no longer observed, pointing to the involvement of T1P in the establishment of this phenotype. * and ** indicate statistical differences observed between experiments performed at 5 and 25 mM glucose after 3 h ($P=0.02$) and 6 h ($P=0.002$) of incubation, respectively.

the higher protein concentrations observed in such situations could be due to the presence of sodium bicarbonate instead of CO_2 . Since CO_2 (or sodium bicarbonate) stimulate EPEC protein secretion, we believe that the increased adhesiveness of aEPEC 1551–2 in the atmosphere supplemented with 5% CO_2 could be related to increased secretion of the EPEC secreted proteins that form the T3SS-translocon (EspA, B and D), in view of the fact that this structure interacts with epithelial cells [23]. In addition, in our previous studies, the T3SS-translocon was shown to mediate aEPEC 1551–2 adherence to

HeLa cells in the absence of intimin [24]. Of note, intimin mediates the intimate bacterial attachment observed in the formation of AE lesions [1].

During the flow through the small intestine, glucose is rapidly absorbed, reaching a concentration higher than 25 mM in the duodenum, dropping to less than 5 mM in the ileum, until it is undetectable in the jejunum, depending on the host's diet [12]. Quantitative experiments to evaluate the efficiency of bacterial interaction with HeLa cells showed no statistical difference between the numbers of associated bacteria when assays were

Table 1 Bacterial strains and plasmids used in this study

<i>E. coli</i> strains and plasmids	Characteristics	Reference
Strains		
1551-2	aEPEC (ONT:H ⁻) isolated from a diarrheic child, which expresses LA on HeLa cells (Nal ^R)	[18]
1551-2Δ <i>fimA</i>	1551-2 <i>fimA</i> : <i>zeo</i> (Nal ^R Zeo ^R), T1P mutant	[24]
1551-2Δ <i>fimA</i> (pBAD)	1551-2 mutant harboring pBAD/Myc-His A vector (Nal ^R Zeo ^R Amp ^R)	[24]
1551-2Δ <i>fimA</i> (p <i>FimA</i>)	1551-2 mutant harboring p <i>FimA</i> (Nal ^R Zeo ^R Amp ^R)	[24]
Plasmid		
pBAD/Myc-His A	Cloning vector, harboring the araBAD promoter (pBAD) from <i>E. coli</i> , which is induced in the presence of L-arabinose.	Invitrogen
p <i>FimA</i>	pBAD/Myc-His A vector harboring the <i>fimA</i> gene from aEPEC 1551-2	[24]

Nal^R: nalidixic acid resistant; Zeo^R: zeocin resistant; Amp^R: ampicillin resistant; LA: Localized adherence.

performed in the presence of 5 or 25 mM glucose at the same CO₂ concentration (Figure 1A).

However, regardless of the CO₂ concentration used, two distinct adherence patterns were detected depending on the glucose concentration present in the culture medium. At 5 mM glucose, bacteria adhered in the LA pattern, originally reported by Vieira and coworkers [18], whereas at 25 mM glucose, a hybrid pattern consisting of LA and AA-like patterns was seen. Previous studies by our laboratory found that in contact with HeLa cells the 1551-2 strain produces T1P [20]. In addition, this strain lacks adhesin-encoding genes that are commonly found in other diarrhegenic *E. coli* pathotypes and extraintestinal pathogenic *E. coli* (ExPEC), as shown in previous studies [18-20,25].

In the present study, we showed that *fimA* expression increased 1.75-fold and 2.03-fold, in the period between 3 and 6 h, when 25 mM glucose was used as compared to 5 mM glucose. Various studies in the literature have shown that glucose influences the expression of fimbrial and non-fimbrial adhesins. Our results are in accordance with previous studies showing that adhesins such as the bundle-forming pilus (of tEPEC), colonization factor antigens (of enterotoxigenic *E. coli*) and toxin-coregulated pilus (of *Vibrio cholera*) are positively regulated by high glucose concentrations [16,26,27]. Müller and coworkers reported that two uropathogenic *E. coli* strains responded in opposite ways regarding fimbriation in the presence of cAMP receptor protein (CRP) and 3',5'-cyclic adenosine monophosphate (cAMP) complex by phase variation of *fimA* promoter [28]. CRP-cAMP is a signaling protein complex formed when there is a reduction in glucose concentration in the environment [28]. Therefore, the CRP-cAMP complex can induce the *fimA* promoter in the absence of glucose. However, in the presence of glucose and absence of CRP-cAMP, a complex of Lrp (leucine-responsive protein), DNA gyrase and FimB (an invertase of the T1P operon) raise the amount of fimbriated bacteria in the population by maintaining the

phase variation of *fimA* promoter in the ON position, reflecting a glucose-dependent event [28]. In our study, we found that the AA-like pattern of the 1551-2 strain depended on the presence of a high glucose concentration, leading us to hypothesize that the increased *fimA* expression in the later incubation periods corresponded to a glucose-dependent phenotype.

In contrast to the wild-type strain, the isogenic *fimA* mutant lost the ability to produce the AA-like pattern, even when the assay was performed in medium supplemented with high glucose concentration, confirming the influence of this sugar in the establishment of the AA-like pattern. The involvement of T1P in the AA pattern was first described in enteroaggregative *E. coli* (EAEC) prototype strain 042, by mutating the *fimD* gene (encoding the T1P usher). The *fimD* mutant strain was approximately 80% less adherent to HEp-2 cells than the wild-type strain [29]. Additionally, T1P has been implicated in biofilm formation in both EAEC (042) and aEPEC (1551-2) strains, reinforcing the involvement of this adhesin in the pathogenesis of diarrhegenic *E. coli* strains [24,29].

As suggested by Edwards and Puente, a combination of environmental signals may provide an intestinal map to identify an appropriate niche for bacterial colonization [12]. The differences in the adherence patterns on HeLa cells under some diverse environmental conditions could reflect the differential expression of adhesin-encoding genes in the distinct parts of the intestine. In view of the evidence that under high glucose concentration the modification in the adherence pattern from LA to LA/AA-like was not associated with an increase of the number of associated bacteria, we can hypothesize that T1P production can negatively interfere with microcolony formation in experiments performed in this circumstance. Therefore, the higher level of T1P production in 25 mM glucose could suggest that these fimbriae are involved in the first steps of the association of aEPEC1551-2 with the proximal small intestine, where the highest glucose concentration is detected [30].

Conclusions

Altogether, our data suggest that a CO₂-enriched atmosphere could favor aEPEC adhesion to the host cells, whereas enhanced T1P production under high glucose concentration could allow bacteria to access more extensive intestinal colonization sites in the host at the beginning of the infectious process.

Methods

Bacterial strain

The aEPEC 1551–2 strain (serotype ONT:H⁻) was isolated as part of standard patient care from a diarrheic child (23 months old), in the absence of other recognized pathogens, during an epidemiological study of diarrhea carried out at the Universidade Federal de São Paulo (UNIFESP), Brazil. This strain was initially reported to produce LA (in the 6-h assay) in HeLa cells [18]. The *fimA* mutant and complemented strains were obtained in a subsequent study from our laboratory [24]. The most relevant characteristics of the bacterial strains and plasmids employed in the present study are described in Table 1.

Bacterial strains were routinely grown aerobically in Luria-Bertani (LB) medium or Dulbecco's modified Eagle's medium (DMEM, Gibco, USA) at 37°C. When appropriate, strains were cultured in the presence of nalidixic acid (20 µg/ml) and/or zeocin (60 µg/ml). Assays employing the complemented strain were performed in the presence of 0.01% L-arabinose as previously described [24].

HeLa cells culture

HeLa cells were cultivated in DMEM supplemented with 10% fetal bovine serum (Gibco, USA) and in the presence of 1% antibiotic mixture (penicillin and streptomycin, Life Technologies, USA) in an atmosphere of 5% CO₂ at 37°C. To determine the adherence pattern and quantify the total number of associated bacteria (quantitative assay), HeLa cells were seeded in 24-well microplates, containing glass coverslips and the same media. Adherence assays were performed using semi-confluent cells (approximately 70% confluence) after two days of incubation.

Qualitative assays (adherence pattern)

Qualitative assays were performed as described earlier [20]. Briefly, HeLa cells were washed with phosphate buffered saline (PBS, pH 7.4). After washing, 1.0 ml of fresh medium (DMEM supplemented with 2% fetal bovine serum) was added to the cell monolayers, and the HeLa cells were then inoculated with suspensions (approximately 10⁸ CFU/mL) of bacteria grown in LB broth (overnight culture) diluted 1:50, and incubated at 37°C. During the interaction, we evaluated the effects of glucose concentration (5 or 25 mM), presence of bile salts (absence or presence of 0.015% w/v) and

CO₂ concentration (0.03 or 5%, with DMEM supplemented with 0.35 or 35 mM sodium bicarbonate, respectively). After 6 h of interaction (with a washing step after 3 h), the preparation was washed six times with PBS and then fixed with methanol (Merck, Germany) for 1 h, stained with May Grünwald-Giemsa stain, and examined by light microscopy for adherence pattern determination.

Quantitative assays

For determination of the total number of associated bacteria, HeLa cells were inoculated and the experiment was conducted as described above. During the interaction, we evaluated the same assay conditions as in the qualitative assays. After 6 h of interaction (with a washing step after 3 h), the preparations were washed six times with PBS. Next, the cells were lysed with 1% Triton X-100, and serial dilutions were plated on MacConkey agar plates. After approximately 18 h of incubation at 37°C, the resulting colonies were counted for determination of the total number of associated bacteria. All assays were performed in biological and technical triplicate, and the results represented the means ± standard errors.

Quantitative PCR assay

aEPEC 1551–2 was grown for 18 h in LB broth (or broth supplemented with antibiotics) and incubated statically at 37°C. The pre-inoculum was diluted 1:50 in DMEM supplemented with 5 or 25 mM glucose, and incubated statically for 1.5, 3.0 or 6.0 h, at 37°C, in an atmosphere of 0.03% CO₂. After incubation, the preparation was centrifuged at 5,900 g for 3 min, and the pellet was treated with Trizol (Invitrogen, USA) for total RNA extraction. cDNA was synthesized using SuperScript III First-Strand (Invitrogen, USA). The primers used for detection of *fimA* expression were: (forward) 5'-TC GATGCGGGATCTGTTGA-3' and (reverse) 5'-ACC GACGGCAGAGCTGGT-3'. All data were normalized to the expression levels of *rpoA* (RNA polymerase subunit A), by using the following primers: (forward) 5'-G CGCTCATCTTCTTCCGAAT-3' and (reverse) 5'-CG CGGTCGTGGTTATGTG-3', and analyzed with the comparative threshold (C_T) method. The expression levels of *fimA* at different glucose concentrations were compared by using the relative quantification method. Real-time data were expressed as fold change of the expression levels of *fimA* at different glucose concentrations. Data obtained with 25 mM glucose were compared by using the relative quantification method, using strains incubated with 5 mM glucose as the calibrator for each time point. Total RNAs of all treatments were obtained from three independent assays. Statistical differences were determined by the Student *t*-test, and *P* ≤ 0.05 was considered statistically significant.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

FTR performed all experiments in this study and drafted the manuscript. DY, LO, and AFP analyzed the results and formatted the data. RTH and TATG designed the experimental procedures, supervised all the experimental works, and corrected and produced the final version of the manuscript. All authors read and approved the final manuscript.

Acknowledgments

This study was supported by grants from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP 11/12664-5) and Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq, grants:304453/2011-0 and 480664/2012-9). FTR and DY received fellowships from Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES). Dr. A. Leyva helped with English editing of the manuscript.

Author details

¹Departamento de Microbiologia, Imunologia e Parasitologia, Universidade Federal de São Paulo (UNIFESP), Escola Paulista de Medicina, São Paulo, SP, Brazil. ²Departamento de Microbiologia e Imunologia, Instituto de Biociências, Universidade Estadual Paulista (UNESP), Botucatu, SP, Brazil.

Received: 15 July 2014 Accepted: 18 November 2014

Published online: 20 December 2014

References

- Kaper JB, Nataro JP, Mobley HL: **Pathogenic *Escherichia coli***. *Nat Rev Microbiol* 2004, **2**:123–140.
- Kaper JB: **Defining EPEC**. *Rev Microbiol* 1996, **27**:130–133.
- Trabulsi LR, Keller R, Gomes TA: **Typical and atypical enteropathogenic *Escherichia coli***. *Emerg Infect Dis* 2002, **8**:508–513.
- Girón JA, Ho AS, Schoolnik GK: **An inducible bundle-forming pilus of enteropathogenic *Escherichia coli***. *Science* 1991, **254**:710–713.
- Donnenberg MS, Girón JA, Nataro JP, Kaper JB: **A plasmid-encoded type IV fimbrial gene of enteropathogenic *Escherichia coli* associated with localized adherence**. *Mol Microbiol* 1992, **6**:3427–3437.
- Moon HW, Whipp SC, Argenzio RA, Levine MM, Giannella RA: **Attaching and effacing activities of rabbit and human enteropathogenic *Escherichia coli* in pig and rabbit intestines**. *Infect Immun* 1983, **41**:1340–1351.
- Jerse AE, Yu J, Tall BD, Kaper JB: **A genetic locus of enteropathogenic *Escherichia coli* necessary for the production of attaching and effacing lesions on tissue culture cells**. *Proc Natl Acad Sci U S A* 1990, **87**:7839–7843.
- Kenny B, DeVinney R, Stein M, Reinscheid DJ, Frey EA, Finlay BB: **Enteropathogenic *E. coli* (EPEC) transfers its receptor for intimate adherence into mammalian cells**. *Cell* 1997, **91**:511–520.
- McDaniel TK, Jarvis KG, Donnenberg MS, Kaper JB: **A genetic locus of enterocyte effacement conserved among diverse enterobacterial pathogens**. *Proc Natl Acad Sci U S A* 1995, **92**:1664–1668.
- Torres AG, Zhou X, Kaper JB: **Adherence of diarrheagenic *Escherichia coli* strains to epithelial cells**. *Infect Immun* 2005, **73**:18–29.
- Arenas-Hernandez MMP, Martínez-Laguna Y, Torres AG: **Clinical Implication of Enteroadherent *Escherichia coli***. *Curr Gastroenterol* 2012. [http://dx.doi.org/10.1007/s11894-012-0277-1]
- Edwards RA, Puente JL: **Fimbrial expression in enteric bacteria: a critical step in intestinal pathogenesis**. *Trends Microbiol* 1998, **6**:282–287.
- Beinke C, Laarmann S, Wachter C, Karch H, Greune L, Schmidt MA: **Diffusely adhering *Escherichia coli* strains induce attaching and effacing phenotypes and secrete homologs of Esp proteins**. *Infect Immun* 1998, **66**:528–539.
- Torres AG, Milflores-Flores L, Garcia-Gallegos JG, Patel SD, Best A, La Razione RM, Martínez-Laguna Y, Woodward MJ: **Environmental regulation and colonization attributes of the long polar fimbriae (LPF) of *Escherichia coli* O157:H7**. *Int J Med Microbiol* 2007, **297**:177–185.
- Torres AG, Tutt CB, Duval L, Popov V, Nasr AB, Michalski J, Scaletsky IC: **Bile salts induce expression of the afimbrial LDA adhesin of atypical enteropathogenic *Escherichia coli***. *Cell Microbiol* 2007, **9**:1039–1049.
- Puente JL, Bieber D, Ramer SW, Murray W, Schoolnik GK: **The bundle-forming pili of enteropathogenic *Escherichia coli*: transcriptional regulation by environmental signals**. *Mol Microbiol* 1996, **20**:87–100.
- Phillips AD, Frankel G: **Intimin-mediated tissue specificity in enteropathogenic *Escherichia coli* interaction with human intestinal organ cultures**. *J Infect Dis* 2000, **181**:1496–1500.
- Vieira MA, Andrade JR, Trabulsi LR, Rosa AC, Dias AM, Ramos SR, Frankel G, Gomes TA: **Phenotypic and genotypic characteristics of *Escherichia coli* strains of non-enteropathogenic *E. coli* (EPEC) serogroups that carry *eae* and lack the EPEC adherence factor and Shiga toxin DNA probe sequences**. *J Infect Dis* 2001, **183**:762–772.
- Gomes TA, Hernandez RT, Torres AG, Salvador FA, Guth BE, Vaz TM, Irino K, Silva RM, Vieira MA: **Adhesin-encoding genes from Shiga toxin-producing *Escherichia coli* are more prevalent in atypical than in typical enteropathogenic *E. coli***. *J Clin Microbiol* 2011, **49**:3334–3337.
- Hernandes RT, Velsko I, Sampaio SC, Elias WP, Robins-Browne RM, Gomes TA, Girón JA: **Fimbrial adhesins produced by atypical enteropathogenic *Escherichia coli* strains**. *Appl Environ Microbiol* 2011, **77**:8391–8399.
- Babb RR: **Intestinal gas (Medical Information)**. *West J Med* 1977, **127**:362–363.
- Haigh R, Baldwin T, Knutton S, Williams PH: **Carbon dioxide regulated secretion of the EaeB protein of enteropathogenic *Escherichia coli***. *FEMS Microbiol Lett* 1995, **129**:63–67.
- Cleary J, Lai LC, Shaw RK, Straatman-Iwanowska A, Donnenberg MS, Frankel G, Knutton S: **Enteropathogenic *Escherichia coli* (EPEC) adhesion to intestinal epithelial cells: role of bundle-forming pili (BFP), EspA filaments and intimin**. *Microbiology* 2004, **150**:527–538.
- Hernandes RT, De la Cruz MA, Yamamoto D, Girón JA, Gomes TA: **Dissection of the role of pili and type 2 and 3 secretion systems in adherence and biofilm formation of an atypical enteropathogenic *Escherichia coli* strain**. *Infect Immun* 2013, **81**:3793–3802.
- Gomes TA, Irino K, Girão DM, Girão VB, Guth BE, Vaz TM, Moreira FC, Chinarelli SH, Vieira MA: **Emerging enteropathogenic *Escherichia coli* strains?** *Emerg Infect Dis* 2004, **10**:1851–1855.
- Gaastra W, Syennerholm AM: **Colonization factors of enterotoxigenic *Escherichia coli* isolated from children in north India**. *J Infect Dis* 1996, **174**:768–776.
- Skorupski K, Taylor RK: **Cyclic AMP and its receptor protein negatively regulate the coordinate expression of cholera toxin and toxin-coregulated pilus in *Vibrio cholerae***. *Proc Natl Acad Sci U S A* 1997, **94**:265–270.
- Müller CM, Aberg A, Strasevičiene J, Emody L, Uhlin BE, Balsalobre C: **Type 1 fimbriae, a colonization factor of uropathogenic *Escherichia coli*, are controlled by the metabolic sensor CRP-cAMP**. *PLoS Pathog* 2009. [http://dx.doi.org/10.1371/journal.ppat.1000303]
- Moreira CG, Carneiro SM, Nataro JP, Trabulsi LR, Elias WP: **Role of type I fimbriae in the aggregative adhesion pattern of enteroaggregative *Escherichia coli***. *FEMS Microbiol Lett* 2003, **226**:79–85.
- Ferraris RP, Yasharpour S, Lloyd KC, Mirzayan R, Diamond JM: **Luminal glucose concentrations in the gut under normal conditions**. *Am J Physiol* 1990, **259**:G822–G837.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

