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## Chemoprevention of Cigarette Smoke–Induced Alterations of MicroRNA Expression in Rat Lungs

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### Abstract

We previously showed that exposure to environmental cigarette smoke (ECS) for 28 days causes extensive downregulation of microRNA expression in the lungs of rats, resulting in the overexpression of multiple genes and proteins. In the present study, we evaluated by microarray the expression of 484 microRNAs in the lungs of either ECS-free or ECS-exposed rats treated with the orally administered chemopreventive agents *N*-acetylcysteine, oltipraz, indole-3-carbinol, 5,6-benzoflavone, and phenethyl isothiocyanate (as single agents or in combinations). This is the first study of microRNA modulation by chemopreventive agents in nonmalignant tissues. Scatterplot, hierarchical cluster, and principal component analyses of microarray and quantitative PCR data showed that none of the above chemopreventive regimens appreciably affected the baseline microRNA expression, indicating potential safety. On the other hand, all of them attenuated ECS-induced alterations but to a variable extent and with different patterns, indicating potential preventive efficacy. The main ECS-altered functions that were modulated by chemopreventive agents included cell proliferation, apoptosis, differentiation, *Ras* activation, *P53* functions, NF- $\kappa$ B pathway, transforming growth factor–related stress response, and angiogenesis. Some micro-RNAs known to be polymorphic in humans were downregulated by ECS and were protected by chemopreventive agents. This study provides proof-of-concept and validation of technology that we are further refining to screen and prioritize potential agents for continued development and to help elucidate their biological effects and mechanisms. Therefore, microRNA analysis may provide a new tool for predicting at early carcinogenesis stages both the potential safety and efficacy of cancer chemopreventive agents.

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#### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

## Introduction

MicroRNAs (miRNA) provide a major epigenetic mechanism that regulates translation of expressed genes into proteins. These small noncoding RNAs play a role in crucial biological processes, such as cell growth (1), apoptosis (2), development (3), differentiation (4), and endocrine homeostasis (5). MiRNAs have been investigated in a variety of diseases, including diabetes, heart diseases, Alzheimer's disease, and viral infections (6). The most active area and the starting point for the pathogenetic role of miRNAs is cancer research (7, 8), to such an extent that alterations in miRNA genes have been proposed to be involved in the pathophysiology of many, perhaps, all human cancers (9).

Less attention has been paid to the possible occurrence of miRNA alterations in healthy tissues as a consequence of exposures to environmental and life-style factors, including carcinogens and noxious agents, drugs, and food components. Recently, we provided evidence that exposure of rats to environmental cigarette smoke (ECS) results in extensive downregulation, in the lung, of the expression of several miRNAs involved in a variety of cell functions, such as stress response, apoptosis, proliferation, angiogenesis, and gene transcription (10). These findings were confirmed in a study evaluating miRNA expression in the human airway epithelium of smokers (11) and by our further studies in mice (12). The results of these studies support the view that cigarette smoke mainly works as a tumor promoter by triggering a variety of epigenetic mechanisms (13, 14).

In the present study, we evaluated miRNA expression as a new tool for assessing the ability of chemopreventive agents to modulate physiologic miRNA profiles as well as alterations induced in the lung following exposure of rats to ECS. The investigated chemopreventive agents, all of them administered orally, included dietary agents, such as phenethyl isothiocyanate (PEITC) and indole-3-carbinol (I3C), the synthetic flavone 5,6-benzoflavone (BF), and pharmacologic agents, such as *N*-acetyl-L-cysteine (NAC) and oltipraz (OPZ). Due to the interest of pursuing a combined chemoprevention strategy, combinations of PEITC with I3C and of NAC with OPZ were also investigated.

## Materials and Methods

### Animals and treatments

Adult male Sprague-Dawley rats (Harlan Italy), weighing 315 to 320 g, were divided into 16 groups, each composed of eight animals. Eight groups had their whole bodies exposed to ECS for 28 consecutive d, as previously described (15), whereas the remaining rats (Sham exposed) were kept for the same period of time in filtered air. The rats belonging to 14 groups were treated with chemopreventive agents, starting 3 d before exposure to ECS. NAC was given with the drinking water, whereas all other agents were supplemented to the diet. In particular, as detailed in a previous article (15), the experimental groups were as follows: *group 1*, rats kept in filtered air (Sham); mean diet consumption, 25.0 g/d; body weight gain during the 28 d of the experiment, 17.8%; *group 2*, rats treated with OPZ (400 mg/kg diet); mean dietary consumption and mean intake of chemopreventive agent, 19.4 g/d and 7.8 mg/rat/d, respectively; body weight gain, 13.2%; *group 3*, rats treated with PEITC (500 mg/kg diet; 19.2 g/d and 9.6 mg/rat/d); body weight gain, 14.2%; *group 4*, rats treated

with BF (500 mg/kg diet; 20.1 g/d and 10.1 mg/rat/d); body weight gain, 13%; *group 5*, rats treated with I3C (2,500 mg/kg diet; 25 g/d and 62.5 mg/rat/d); body weight gain, 15.7%; *group 6*, rats treated with NAC (1,000 mg/kg body weight; 25 g/d and 300 mg/rat/d); body weight gain, 15.1%; *group 7*, rats treated with a combination of PEITC and I3C (22.5 g/d and 11.3 and 56.3 mg/rat/d, respectively); body weight gain, 15.4%; *group 8*, rats treated with a combination of OPZ and NAC (22.8 g/d and 9.1 and 300 mg/rat/d, respectively); body weight gain, 16.4%; *group 9*, ECS-exposed rats. body weight gain, 11.7%; *group 10*, ECS-exposed rats treated with OPZ (400 mg/kg diet; 18.9 g/d and 7.6 mg/rat/d); body weight gain, 6.3%; *group 11*, ECS-exposed rats treated with PEITC (500 mg/kg diet; 19.6 g/d and 9.8 mg/rat/d); body weight gain, 6.7%; *group 12*, ECS-exposed rats treated with BF (500 mg/kg diet; 19.4 g/d and 9.7 mg/rat/d); body weight gain, 6.4%; *group 13*, ECS-exposed rats treated with I3C (2,500 mg/kg diet; 25 g/d and 62.5 mg/rat/d); body weight gain, 11.8%; *group 14*, ECS-exposed rats treated with NAC (1,000 mg/kg body weight; 25.0 g/d and 300 mg/rat/d); body weight gain, 13.6%; *group 15*, ECS-exposed rats treated with a combination of PEITC and I3C (20.9 g/d and 10.5 and 52.3 mg/rat/d, respectively); body weight gain, 7.7%; *group 16*, ECS-exposed rats treated with a combination of OPZ and NAC (21.8 g/d and 8.7 and 300 mg/rat/d, respectively); body weight gain, 8.6%.

At the end of the experiment, the rats were starved for 24 h, anesthetized deeply with diethyl ether, and killed by cervical dislocation. The lungs were collected, immersed in an RNA-stabilizing buffer, and frozen at  $-80^{\circ}\text{C}$ .

The housing and all treatments of animals were in accordance with U.S. NIH and Italian and institutional guidelines.

### RNA extraction and miRNA analysis

RNA extraction, quantification, and evaluation of integrity, microarray profiling for 484 rodent miRNAs, and real-time quantitative PCR (qPCR) for *miR-let7c* were done as previously described (10). The whole list of miRNAs included in the microarray used is available at the Gene Expression Omnibus database (registration number requested).

### Analysis of data

Local background was subtracted from raw data, which were then log transformed, normalized, and analyzed by GeneSpring software version 7.2 (Agilent Technologies). Expression data were median centered by using the Gene-Spring normalization option, which normalizes both per gene and per array. Quadruplicate data generated for each miRNA were compared among the various experimental groups by volcano plot analysis, which evaluates both fold variations and statistical significance of differences by ANOVA following Bonferroni multiple testing correction. Global miRNA expression profiles of the various experimental groups were compared by hierarchical cluster analysis and by bidimensional perchloric acid.

## Results

### Overall microarray evaluation of miRNA expression in the lung of rats treated with chemopreventive agents

As inferred from the scatterplots shown in Fig. 1 (*top*), the oral administration of either individual chemopreventive agents (BF, NAC, OPZ, or PEITC) or of combined agents (OPZ + NAC or PEITC + I3C) did not appreciably affect miRNA expression profiles in rat lung. In fact, only a negligible number of miRNAs was dysregulated >2-fold following treatment with the chemopreventive regimens tested, compared with Sham-exposed rats. The large majority of the miRNAs falling outside the 2-fold variation interval were poorly expressed in the lung. None of these variations, which are likely to be ascribed to the experimental fluctuations characteristic of low-intensity signals, reached the statistical significance threshold. Note that the sample from rats treated with I3C alone could not be tested due to insufficient amounts of RNA.

### Overall microarray evaluation of miRNA expression in the lung of rats exposed to ECS and treated with chemopreventive agents

The scatterplots shown in Fig. 1 (*bottom*) highlight a general trend to modulation of ECS-altered miRNA profiles by all chemopreventive agents tested. In fact, compared with rats exposed to ECS in the absence of chemopreventive treatments, all the agents tended to upregulate miRNA expression, thus counteracting the downregulation induced by ECS alone.

As shown at a glance in Fig. 1, this protective effect varied among the chemopreventive regimens tested. Treatment-related differences in modulating miRNA profiles were well detectable by performing unsupervised hierarchical cluster analysis (Fig. 2). It is evident that Sham and ECS were allocated far away in the dendrogram and that, with the exception of I3C, treatment of ECS-exposed rats with all other chemopreventive agents were able to reallocate their miRNA profiles in the same dendrogram cluster that included untreated rats (Sham).

### Modulation of the expression of individual miRNAs in the lung of rats exposed to ECS and treated with chemopreventive agents

Table 1 lists the 25 miRNAs whose expression was significantly modulated by chemopreventive agents in rat lung, compared either with unexposed rats (Sham) or ECS-exposed rats. All miRNAs, excepting *miR-294*, were significantly downregulated by ECS, in the absence of chemopreventive agents (Table 1, *first column*). Most chemopreventive agents attenuated this effect to a variable extent.

In particular, three general situations can be envisaged when comparing the values recorded in ECS-exposed rats receiving a chemopreventive agent with those recorded in ECS-exposed rats in the absence of chemopreventive agents. The first situation, indicating a poor protective effect, was that the chemopreventive agent did not significantly affect ECS-induced miRNA alterations (absence of superscripts \*, ‡, or b in Table 1). This situation was observed for 6 of 25 miRNAs in BF-treated rats (24.0%), 4 in NAC-treated rats (16.0%), 6

in OPZ-treated rats (24.0%), 4 in PEITC-treated rats (16.0%), 14 in I3C-treated rats (56.0%), 3 in rats treated with both OPZ and NAC (12.0%), and 2 in rats treated with both PEITC and I3C (8.0%). The second situation, reflecting a moderate protective effect, was that the values recorded in ECS-exposed rats treated with a chemopreventive agent were significantly different from both Sham and ECS (‡ superscripts in Table 1). This situation was observed for 9 of 25 miRNAs in BF-treated rats (36.0%), 7 in NAC-treated rats (28.0%), 6 in OPZ-treated rats (24.0%), 3 in PEITC-treated rats (12.0%), 4 in I3C-treated rats (16.0%), 5 in rats treated with both OPZ and NAC (20.0%), and 7 in rats treated with both PEITC and I3C (28.0%). The third situation, reflecting a strong protective effect, was that the values recorded in ECS-exposed rats treated with a chemopreventive agent were significantly different from ECS only (superscripts §, b, and a in Table 1). This situation was observed for 10 of 25 miRNAs in BF-treated rats (40.0%), 14 in NAC-treated rats (56.0%), 13 in OPZ-treated rats (52.0%), 18 in PEITC-treated rats (72.0%), 7 in I3C-treated rats (28.0%), 17 in rats treated with both OPZ and NAC (68.0%), and 16 in rats treated with both PEITC and I3C (64.0%). Based on these data, it seems that PEITC alone, PEITC + I3C, and OPZ + NAC were the most effective treatments in counteracting miRNA downregulation by ECS. However, it should be noted that, in the case of the combined treatment with PEITC and I3C, miRNA expression often exceeded the baseline situation.

The striking upregulation of *miR-294* in ECS-exposed rats was attenuated by treatment with either I3C, NAC + OPZ, or PEITC + I3C. In all three cases, the expression level was no longer significantly different from Sham but did not reach the significance threshold also compared with ECS.

In addition to statistical significance criteria, we assumed an arbitrary threshold value of 0.90 for (ECS + chemopreventive agent)/Sham ratios as a biological indicator of a strong protective effect of a chemopreventive agent toward ECS-induced miRNA alterations (bold values in Table 1).

The biological functions of those ECS-dysregulated miRNAs whose expression was modified by the investigated chemopreventive agents were inferred by selecting genes having a context score of >0.31, as reported in Targetscan,<sup>5</sup> Miranda,<sup>6</sup> and Sanger Institute<sup>7</sup> databases. Table 2 shows an outline of the main functions regulated by miRNAs whose ECS-altered expression was normalized by the investigated chemopreventive agents.

### Principal component analysis of miRNA expression profiles in rat lung

Principal component analysis (PCA) took into account the data obtained in the 15 experimental groups by analyzing the expression of all 484 miRNAs tested. PCA evaluates the dispersion of data according to the two main components of variance, thus resulting in a bidimensional scattering of the experimental groups, which fall in four quadrants that cluster similar miRNA profiles.

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<sup>5</sup><http://www.targetscan.org>

<sup>6</sup><http://www.microrna.org>

<sup>7</sup><http://microrna.sanger.ac.uk>

The results of PCA are reported in Fig. 3. It is evident that ECS and Sham fall far away, in two opposite quadrants. All chemopreventive agents, in the absence of exposure to ECS, fall in the same quadrant where Sham is allocated, thus reflecting the poor alterations of miRNA expression exerted by all chemopreventive regimens tested. On the other hand, when given to ECS-exposed rats, all chemopreventive agents tended to depart from ECS and to approach Sham. In the case of the combined treatment with PEITC and I3C, however, PCA allocation was the farthest away from ECS, but at the same time it was far away from Sham, thus reflecting a nonphysiologic modulation of ECS-related miRNA expression.

### Confirmation by qPCR of *let-7c* expression in rat lung

As shown in Table 1, microarray analyses indicated that the expression of *let-7c* was markedly downregulated in ECS-exposed rats and was modulated, to a variable extent, by the chemopreventive agents tested. In addition, due to the major relevance of *let-7c* in pulmonary carcinogenesis, we further analyzed this miRNA by qPCR in the lung of Sham-exposed rats and of ECS-exposed rats, either untreated or treated with the chemopreventive agents. Exposure to ECS caused a downregulation of *let-7c* expression, as inferred from the increased PCR cycle needed to attain the positivity threshold, compared with Sham (Fig. 4). This effect was attenuated by all but one (BF) chemopreventive agents tested. PEITC and OPZ, when tested individually, had a moderate effect, whereas PEITC + I3C, OPZ + NAC, NAC, and I3C were all very close to the Sham expression level (Fig. 4).

### Discussion

Due to the significant difficulties encountered in clinical chemoprevention trials, animal models are very useful to evaluate potential cancer protective effects of dietary and pharmacologic agents, and to prioritize agents for clinical trials. Modulation of early molecular biomarkers, also including those investigated by using “omics” technologies, is of particular interest both for understanding the mechanisms of action of chemopreventive agents and for assessing their safety and efficacy. The following general criteria should be met: (a) as a molecular indicator of safety, an optimal chemopreventive agent should not substantially alter the baseline expression of genes, miR-NAs, and proteins; and (b) as an indicator of efficacy, an optimal chemopreventive agent should be able to counteract the molecular alterations induced by carcinogens and to restore the physiologic situation.

We previously investigated the gene expression profiles of the same chemopreventive agents in the same rat model as those evaluated in the present study (15), producing data that support and complement our present miRNA results. NAC, OPZ, and their combination clustered close to negative controls, thus denoting little alteration of the physiologic situation. At the same time, however, in ECS-exposed lungs, treatments with either OPZ or NAC clustered close to positive controls, thus denoting poor inhibition of ECS-related transcriptome alterations. In contrast, PEITC was the most effective agent in this analysis. The picture from these analyses is complicated by the circumstance that agents appearing to better counteract ECS-related alterations in gene expression also change the physiologic situation (15). In a further study, the lung postmitochondrial fractions of mice treated with Sham, NAC, ECS, or ECS + NAC were analyzed by antibody microarray for the levels of

518 proteins. The oral administration of NAC did not change the basal level of any protein >2-fold and reduced the number of ECS-induced proteins from 50 (9.7%) to 42 (8.1%; ref. 16).

In principle, as supported by the present study, miRNA expression compares favorably to gene and protein expression because the number of existing miRNAs is relatively small, and accordingly, mirnome coverage by microarray analysis is quite broad. In addition, each miRNA targets hundreds of genes (17), thus resulting in an effective global regulation of cell functions. In contrast, only a very small fraction of expressed genes is translated into proteins (18), and the number of detectable proteins is orders of magnitude lower than the number of existing proteins (19). In the interpretation of our data, it should be taken into account that they reflect the net effects of ECS and chemopreventive agents on the lung cell population, which is composed of over 40 cell types involved in a multitude of functions (20). Therefore, we cannot rule out that individual cell types may react in a different way to exogenous stimuli.

The results obtained in the present *in vivo* study provide evidence, for the first time, that mirnome analysis is a promising tool to investigate cancer chemopreventive agents. None of the chemopreventive regimens tested was found to alter the baseline expression of the 484 analyzed miRNAs to an appreciable extent. On the other hand, the mirnome profiles in the lung of ECS-exposed rats were considerably dysregulated, mainly in the sense of downregulation. These data agree with the results obtained in a previous study (10), where we discussed the correspondence between downregulation of specific miRNAs and upregulation of targeted proteins. In particular, several functions out of those that we found to be dysregulated by ECS at the mirnome level had previously been reported to be altered at both transcriptome and proteome levels. These functions include stress response (16), protein repair (16, 21), *RAS* activation (22), *P53* suppression (21, 23, 24), cell proliferation (16, 23), apoptosis (16, 24, 25), and angiogenesis (16).

The two functions that were most frequently modulated by chemopreventive agents were angiogenesis and transforming growth factor- $\beta$  (TGF- $\beta$ )-related response. In fact, all chemopreventive agents counteracted the proangiogenic effect of ECS, which is exerted by downregulating miRNA silencing proangiogenic genes (10). In particular, BF tended to normalize *miR-10a*; OPZ tended to normalize *let-7b*, *let-7c*, *miR-10a*, and *miR-123-prec*; PEITC tended to normalize *let-7a*, *let-7c*, *miR-123-prec*, and *miR-222*; and NAC tended to normalize *miR-123-prec*. Of the two combinations tested, PEITC + I3C affected several angiogenesis-related miRNAs, including *let-7a*, *let-7b*, *let-7c*, *let-7f*, *miR-10a*, *miR-123-prec*, *miR-145-prec*, and *miR-222-prec*, whereas OPZ + NAC tended to attenuate *let-7a*, *let-7b*, *let-7c*, *let-7f*, *miR-123-prec*, and *miR-222-prec*. The antiangiogenic effect is likely to be related to the antioxidant properties of the chemopreventive agents tested. Reactive species, such as hydrogen peroxide and nitric oxide, are involved in the production of factors inducing vessel growth, and their proangiogenic effect is attenuated by factors protecting them from oxidative stress (26, 27). PEITC + I3C, PEITC, I3C, OPZ, and OPZ + NAC, in decreasing rank of activity, counteracted the ECS-downregulated *miR-26a* silencing of TGF- $\beta$ . The effect of I3C on *miR-26a* is consistent with the known ability of this agent to induce *p21/p27*, which results in an inhibition of cyclin-dependent kinases

upregulated during *TGF-β*-mediated antiproliferative response (28). Modulation of ECS-induced cell proliferation by the combination of OPZ and PEITC occurred through regulation of several miRNAs, including those of the *let-7* family, *miR-191-prec*, and *miR-222-prec*, which explains the ability of PEITC to slow down the cell cycle through multiple mechanisms, among which the covalent binding to tubulin (29).

In addition, each one of the chemopreventive agents tested specifically modulated other biological functions. The most effectively modulated miRNAs in ECS-exposed rats treated with NAC, an analogue and precursor of reduced glutathione, are involved in NF-κB-mediated stress response (*miR-146-prec*) and *P53* functions (*miR-34b*, *miR-34c*, and *miR-140s*). NAC, besides being a potent scavenger of free radicals, works through multiple mechanisms, also including its ability to inhibit activation and nuclear translocation of NF-κB, to modulate the post-translational increase of *P53* expression, and to decrease *TGF-β* biological activity (30, 31). In particular, attenuation by NAC of ECS-induced miRNA downregulation is consistent with the observed attenuation of ECS-induced proteins or functions in rodent lung. In fact, NAC modulated ECS-altered proteins involved in cell replication (cyclin F), stress response (FOS, IκKα, IκKβ, and IκKγ), and apoptosis (caspase-14; ref. 16), and related functions (30).

In addition to modulation of *TGF-β* expression (*miR-26a*), cell proliferation (*let-7b* and *let-7c*), and angiogenesis (*let-7b*, *let-7c*, *miR-10a*, and *miR-123-prec*), OPZ normalized a miRNA involved in *P53* function (*miR-34c*). OPZ is known to interfere with signal transduction pathways that play a central role in cell proliferation (32), to induce wild-type *P53* protein and the *P53* pathway (33), and to suppress *TGF-β* (34).

The ECS-downregulated miRNAs affected by PEITC are involved in a variety of functions, including stress response (*miR-125b*), *TGF-β* expression (*miR-26a*), NF-κB activation (*miR-146-prec*), *Ras* activation (*let-7a*, *let-7c*, and *miR-192*), cell proliferation (*let-7a*, *let-7c*, and *miR-222-prec*), cell apoptosis (*miR-99b*), and angiogenesis (*let-7a*, *let-7c*, *miR-123-prec*, and *miR-222-prec*). These findings are consistent with literature data, showing that PEITC activates the *MAPK* stress response pathway (35, 36), and is a potent inducer of apoptosis (25). In fact, either alone or in combination with I3C, PEITC was the only agent able to protect *miR-99b* from ECS-induced downregulation. *MiR-192* is involved in *H-Ras*-mediated oncogenic transformation, which causes elevated generation of reactive oxygen species and renders the cells highly sensitive to PEITC (37).

I3C affected miRNAs involved in *P53* functions (*miR-34b*), *TGF-β* expression (*miR-26a*), *ErbB2* activation (*miR-125a-prec*), and angiogenesis (*miR-10a*). The effect on *miR-34b* explains at the mirnome level the mechanisms of I3C, which is known to arrest the cell cycle by increasing activated *P53* protein in both prostate cancer cells (38) and mammary epithelial cells (39). I3C was the only individual agent that protected *miR-125a-prec* from ECS downregulation. *MiR-125a-prec* recognizes as a main target the *ErbB2* oncogene, which plays a fundamental role in breast cancer (40) and is also expressed in lung carcinomas (41). I3C exerts antiproliferative and apoptotic effects in breast cancer cells expressing *ErbB2* (42). It is noteworthy that I3C was the only chemopreventive agent that had some effect on the ECS-related overexpression of *miR-294*, which is involved in gene



transcription and recognizes transcriptional repressor genes (e.g., zinc finger protein 697; AT-rich interactive domain 4A) as main targets. Accordingly, the increase of *miR-294* expression results in a global increase of gene transcription as a response to ECS (10), a mechanism that is attenuated by I3C.

Combined chemoprevention is a very promising strategy in the pharmacologic and dietary prevention of cancer. The combination of NAC and OPZ affected miRNAs that were either individually modulated by both agents (*miR-34c* and *miR-123-prec*) or by one of them only (*let-7b*, *let-7c*, *miR-26a*, and *miR-146-prec*). In addition, this combined chemopreventive regimen affected a series of miRNAs involved in cell proliferation (*let-7a*, *let-7f*, and *miR-222-prec*), *Ras* activation (*miR-192*), and angiogenesis (*miR-222-prec*). Finally, the combination of PEITC and I3C had profound effects on almost all miRNAs downregulated by ECS, which were not affected by the individual agents, thus suggesting that these agents display more than additive effects. Note however that, as previously discussed, the resulting modulation departed from the physiologic situation.

MiRNA genes are frequently located near mouse cancer susceptibility loci (43). We found that chemopreventive agents are effective in protecting from ECS some miRNAs known to be polymorphic in humans (44, 45). This was the case for *let-7a* (PEITC), *miR-125b* (PEITC), *miR-140s* (NAC), and *miR-146-prec* (NAC, PEITC). Accordingly, it is likely that the efficacy of these chemopreventive agents, when administered to humans, may be influenced by the genetic polymorphisms of these miRNAs. This issue could help to explain the wide interindividual variability observed in the efficacy of these chemopreventive agents when tested in clinical trials (46, 47). It has been predicted that single nucleotide polymorphisms that reside in the miRNA target site may become a gold mine for molecular epidemiology (48). Likewise, the study of polymorphic miRNAs is likely to become an essential tool to develop and qualify chemopreventive agents for clinical trials.

As documented in this study, all of these agents have complex patterns of effects on miRNAs, which become even more complex for their combinations. Objectively comparing and prioritizing between agents for clinical development will require, in part, future studies that can interpret the many complex miRNA patterns associated with agent effects on carcinogenesis. For example, patterns of angiogenesis-related miRNA effects ranged from that of NAC, which modulated only one angiogenesis miRNA (*miR-123-prec*), to that of PEITC, which modulated several (*let-7a*, *let-7c*, *miR-222*, and *miR-123-prec*). It will be important to sort out the critical patterns of angiogenesis-related effects (and all other carcinogenesis-related effects) in animal studies of lung cancer development and, ultimately, in early-phase human trials before proceeding to large-scale, definitive clinical testing.

In conclusion, our present study is the first to look at chemopreventive effects on miRNAs and shows the promise of miRNA profiles for predicting the future clinical efficacy and safety of novel chemopreventive agents. This study provides proof-of-concept and technology validation, and we are refining the technology to both screen and prioritize potential agents for continued drug development and to provide biological and mechanistic insights into specific promising agents. Objective prioritization of agents will require sophisticated statistical modeling. A valuable contribution of our future work will be to look

at correlations between miRNA results and tumor results in the same model. Our future studies also will examine miRNA polymorphisms—for example, in smokers who develop or do not develop lung cancer—for profiles that may identify individuals who are sensitive (or resistant) to specific agents and thus advance the evolution of personalized prevention.

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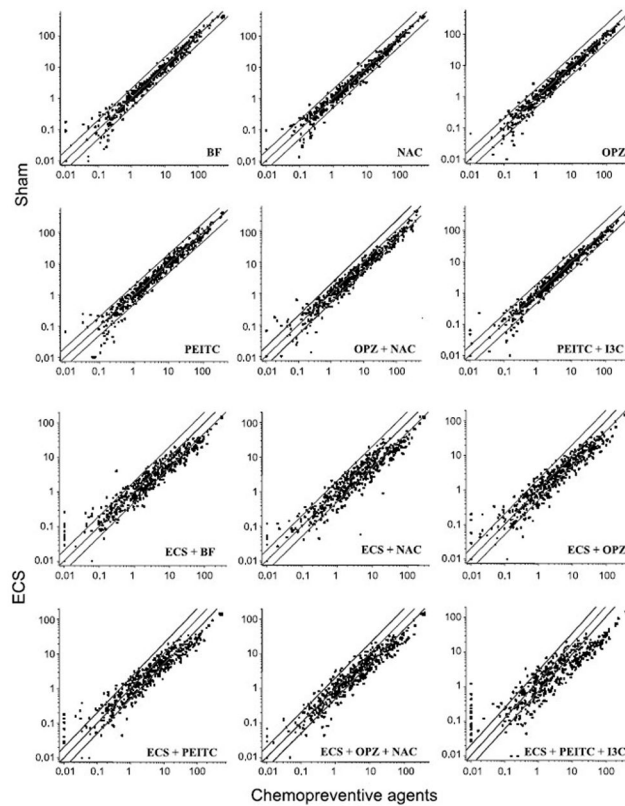
NIH National Cancer Institute (contract N01-CN-53301).

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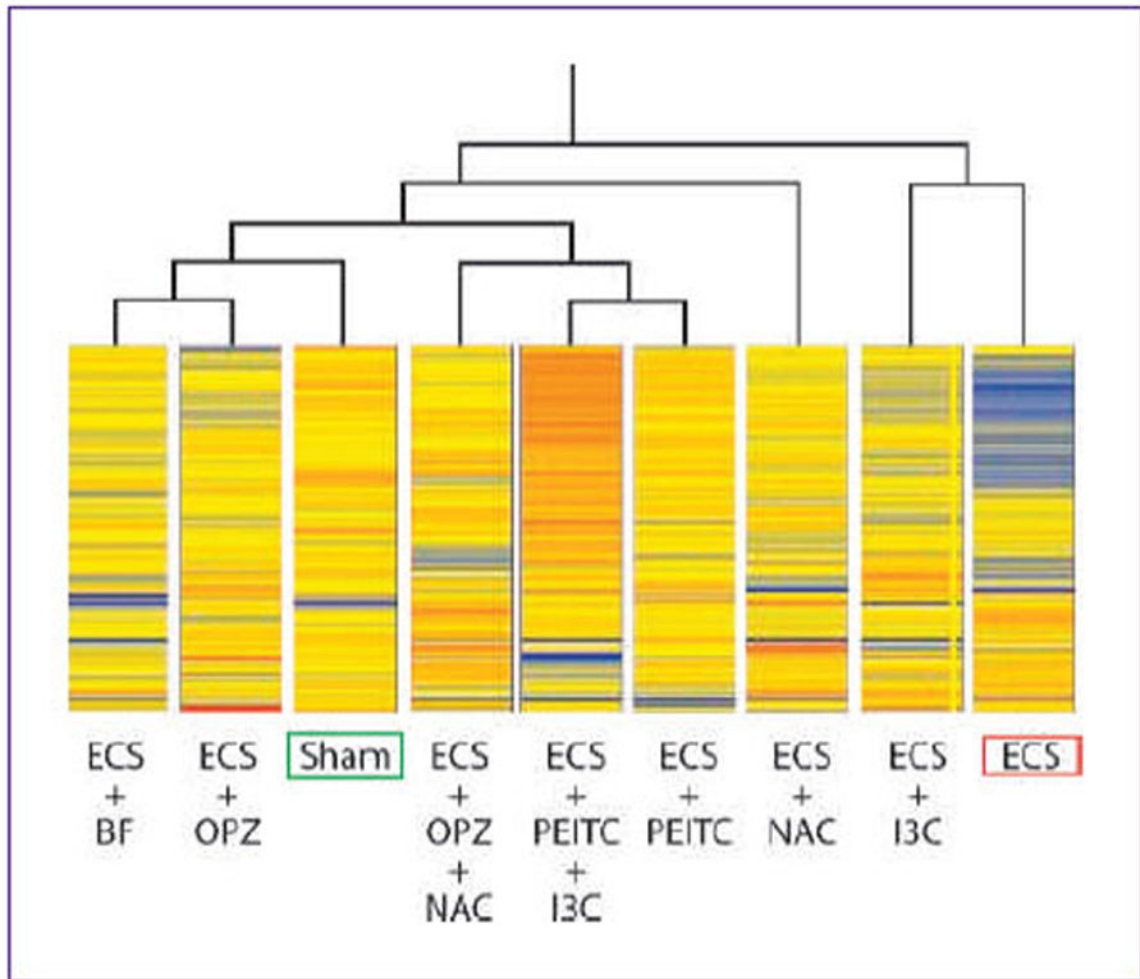
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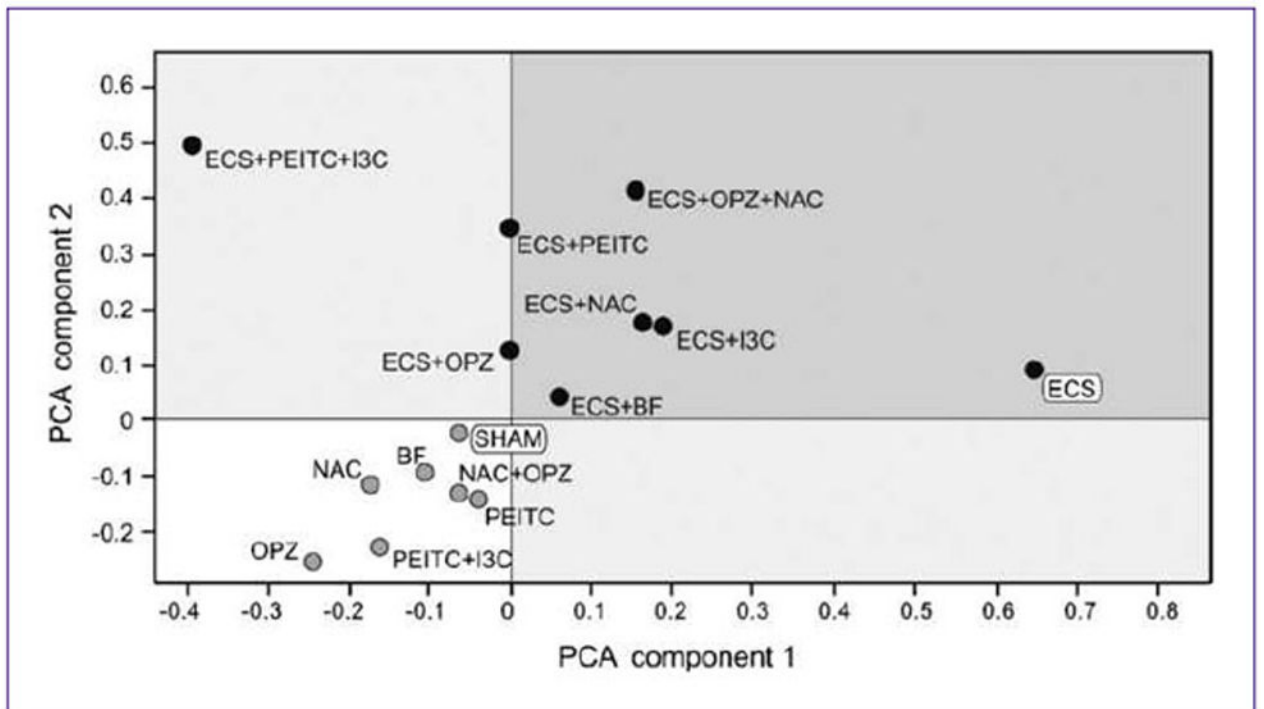
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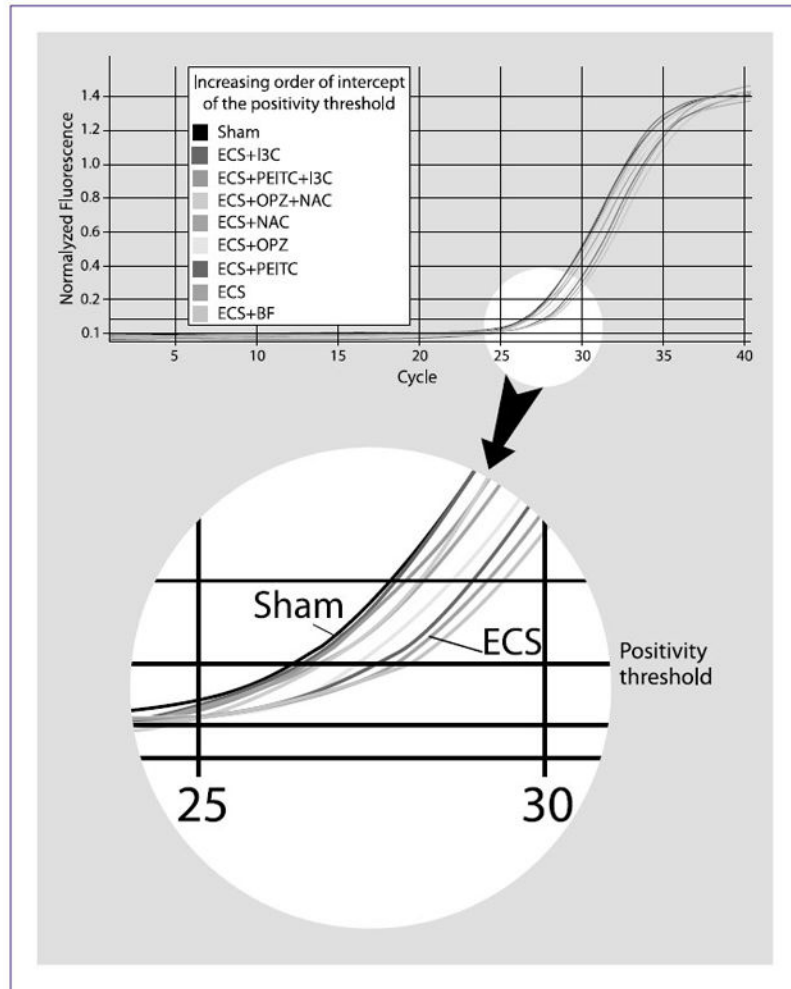
**Fig. 1.** Scatterplots relating the expression of 484 miRNAs in the lung of rats treated with various chemopreventive regimens to the expression of the same miRNAs in the lung of rats, either Sham-exposed (*top*) or ECS-exposed (*bottom*). *Central diagonal line in each panel*, equivalence of expression between the groups compared on the X and Y scales; *two outer diagonal lines in each panel*, 2-fold variation intervals.



**Fig. 2.** Hierarchical cluster analysis linking the expression profiles of 484 miRNAs among variously treated experimental groups. *Each line within each column*, the intensity of expression of each miRNA on a color scale, from blue (lowest) to red (highest).



**Fig. 3.** Bidimensional PCA comparing the expression profile of 484 miRNAs among variously treated experimental groups.



**Fig. 4.** Real-time qPCR amplification curves (*top*) and positivity threshold (*bottom*) of *let-7c* expression in the lung of variously treated rats.



Table 1

MiRNAs whose expression was significantly modulated by chemopreventive agents in rat lung, compared either with unexposed rats (Sham) or ECS-exposed rats

miRNA	ECS/Sham	ECS+BF/Sham	ECS+NAC/Sham	ECS+OPZ/Sham	ECS+OPZ+NAC/Sham	ECS+PEITC/Sham	ECS+BC/Sham	ECS+PEITC+BC/Sham
<i>let-7a</i>	0.24 (0.00-0.77)*	0.52 (0.27-1.64) <sup>†</sup>	0.73 (0.15-1.10) <sup>†</sup>	0.85 (0.40-1.19) <sup>†</sup>	<b>1.14</b> (0.82-1.13) <sup>‡</sup>	<b>0.91</b> (0.69-1.18) <sup>†</sup>	0.77 (0.23-1.14)	<b>1.55</b> (0.64-2.64) <sup>‡</sup>
<i>let-7b</i>	0.24 (0.00-0.85) <sup>†</sup>	0.53 (0.00-1.22) <sup>‡</sup>	0.56 (0.00-1.25) <sup>‡,§</sup>	<b>1.00</b> (0.94-1.06) <sup>†</sup>	<b>0.90</b> (0.50-1.31) <sup>‡</sup>	0.84 (0.32-1.35) <sup>‡</sup>	0.60 (0.00-1.28) <sup>‡,§</sup>	<b>1.35</b> (0.40-2.31) <sup>†</sup>
<i>let-7c</i>	0.22 (0.00-0.80) <sup>§</sup>	0.69 (0.15-1.10) <sup>†</sup>	0.72 (0.05-1.02) <sup>†</sup>	<b>0.96</b> (0.71-1.17) <sup>‡</sup>	<b>1.33</b> (0.67-2.07) <sup>‡</sup>	<b>1.05</b> (0.85-1.22) <sup>‡</sup>	0.73 (0.27-1.16)	<b>1.23</b> (0.70-1.81) <sup>‡</sup>
<i>let-7f</i>	0.19 (0.00-0.74)*	0.38 (0.00-0.85) <sup>‡,§</sup>	0.31 (0.00-0.82) <sup>‡,§</sup>	0.57 (0.11-1.07) <sup>‡</sup>	<b>1.02</b> (0.42-1.19) <sup>‡</sup>	0.89 (0.25-1.15) <sup>‡</sup>	0.70 (0.00-0.90)	<b>1.10</b> (0.70-1.84) <sup>‡</sup>
<i>miR-10a</i>	0.27 (0.00-0.89)*	<b>1.40</b> (0.67-2.06) <sup>‡</sup>	0.51 (0.13-1.08) <sup>†</sup>	<b>1.02</b> (0.49-1.20) <sup>‡</sup>	0.85 (0.26-1.15) <sup>†</sup>	0.68 (0.41-1.19) <sup>†</sup>	<b>0.96</b> (0.72-1.17) <sup>†</sup>	<b>1.80</b> (0.62-3.26) <sup>§,  </sup>
<i>miR-20a</i>	0.32 (0.00-0.99)*	0.81 (0.30-1.17) <sup>†</sup>	<b>1.02</b> (0.58-1.20) <sup>‡</sup>	<b>1.12</b> (0.76-1.39) <sup>‡</sup>	<b>1.08</b> (0.71-1.17) <sup>‡</sup>	<b>1.34</b> (0.70-1.84) <sup>‡</sup>	<b>1.24</b> (0.78-1.39) <sup>‡</sup>	<b>1.71</b> (0.64-2.66) <sup>  </sup>
<i>miR-30a</i>	0.31 (0.00-0.96) <sup>§</sup>	0.69 (0.05-1.33)	0.76 (0.17-1.35)	0.64 (0.00-1.31)	0.67 (0.02-1.32)	0.84 (0.33-1.35)	0.65 (0.0-1.31)	<b>1.69</b> (0.19-3.18) <sup>†</sup>
<i>miR-30c</i>	0.12 (0.00-0.57) <sup>  </sup>	0.63 (0.08-1.05) <sup>‡</sup>	0.58 (0.00-1.27) <sup>‡,§</sup>	0.61 (0.00-1.29) <sup>‡,§</sup>	0.76 (0.34-1.18) <sup>‡</sup>	0.60 (0.00-1.28) <sup>‡,§</sup>	0.87 (0.41-1.19) <sup>†</sup>	<b>0.95</b> (0.55-1.20) <sup>‡</sup>
<i>miR-34b</i>	0.33 (0.00-0.99)*	0.76 (0.17-1.35) <sup>†</sup>	<b>1.13</b> (0.59-1.67) <sup>‡</sup>	0.69 (0.05-1.33)	0.57 (0.00-1.26) <sup>‡,§</sup>	0.57 (0.00-1.25) <sup>‡,§</sup>	<b>0.99</b> (0.92-1.07) <sup>‡</sup>	0.81 (0.27-1.35) <sup>†</sup>
<i>miR-34c</i>	0.32 (0.00-0.97)*	0.85 (0.36-1.34) <sup>†</sup>	<b>1.12</b> (0.62-1.61) <sup>‡</sup>	<b>0.93</b> (0.59-1.28) <sup>‡</sup>	<b>0.97</b> (0.75-1.20) <sup>‡</sup>	0.62 (0.00-1.29)	0.59 (0.00-1.27)	<b>2.05</b> (0.01-4.10) <sup>†</sup>
<i>miR-99b</i>	0.29 (0.00-0.92)*	0.68 (0.04-1.33) <sup>‡,§</sup>	0.65 (0.00-1.31) <sup>‡,§</sup>	0.78 (0.20-1.35) <sup>†</sup>	0.61 (0.00-1.28) <sup>‡,§</sup>	<b>0.97</b> (0.73-1.21) <sup>‡</sup>	0.79 (0.21-1.35) <sup>†</sup>	<b>1.56</b> (0.26-2.86) <sup>‡,§</sup>
<i>miR-122a</i>	0.25 (0.00-0.85)*	0.52 (0.00-1.21) <sup>*,†</sup>	0.61 (0.00-1.29) <sup>†</sup>	0.50 (0.00-1.19) <sup>†,§</sup>	0.53 (0.00-1.22) <sup>†,§</sup>	0.59 (0.00-1.27) <sup>†,§</sup>	0.50 (0.00-1.19) <sup>†,§</sup>	<b>0.98</b> (0.80-1.16) <sup>‡</sup>
<i>miR-123-prec</i>	0.32 (0.00-0.97)*	0.59 (0.00-1.27) <sup>*,†</sup>	<b>0.91</b> (0.52-1.30) <sup>†</sup>	<b>1.01</b> (0.85-1.18) <sup>‡</sup>	<b>1.27</b> (0.38-2.06) <sup>‡</sup>	<b>1.19</b> (0.53-1.84) <sup>‡</sup>	0.63 (0.00-1.30) <sup>‡,§</sup>	<b>2.07</b> (0.01-4.13) <sup>*,‡</sup>
<i>miR-124a-prec</i>	0.14 (0.00-0.62)*	0.61 (0.10-1.18)	0.70 (0.06-1.33) <sup>†</sup>	0.63 (0.09-1.18)	0.80 (0.48-1.25) <sup>†</sup>	0.73 (0.12-1.35) <sup>†</sup>	0.88 (0.76-1.16) <sup>†</sup>	<b>1.19</b> (0.64-1.78) <sup>‡</sup>
<i>miR-125a-prec</i>	0.12 (0.00-0.59) <sup>§</sup>	0.57 (0.00-1.26) <sup>‡,§</sup>	0.61 (0.00-1.28) <sup>‡,§</sup>	0.69 (0.04-1.33) <sup>†</sup>	0.73 (0.02-1.14) <sup>†</sup>	0.71 (0.08-1.34) <sup>†</sup>	<b>1.01</b> (0.12-1.19) <sup>†</sup>	<b>1.12</b> (0.65-1.73) <sup>‡</sup>
<i>miR-125b</i>	0.30 (0.00-0.69) <sup>§</sup>	0.61 (0.00-1.11) <sup>†,§</sup>	0.76 (0.32-1.25) <sup>†,§</sup>	0.88 (0.35-1.25) <sup>‡,§</sup>	0.76 (0.22-1.23) <sup>†,§</sup>	<b>0.90</b> (0.52-1.25) <sup>‡</sup>	0.81 (0.35-1.18)	<b>1.84</b> (0.43-3.25) <sup>‡,§</sup>
<i>miR-140s</i>	0.23 (0.00-0.81) <sup>§</sup>	0.66 (0.07-1.17) <sup>§</sup>	<b>0.90</b> (0.48-1.32) <sup>†</sup>	0.76 (0.17-1.35) <sup>†</sup>	0.81 (0.27-1.35) <sup>†</sup>	0.89 (0.45-1.33) <sup>†</sup>	0.78 (0.36-1.25)	<b>1.55</b> (0.27-2.84) <sup>‡,§</sup>
<i>miR-145-prec</i>	0.22 (0.00-0.80) <sup>  </sup>	0.54 (0.00-1.23) <sup>†</sup>	0.82 (0.29-1.35) <sup>†</sup>	0.64 (0.00-1.30) <sup>†</sup>	0.70 (0.06-1.33) <sup>†</sup>	0.84 (0.32-1.35) <sup>†</sup>	0.48 (0.00-1.17) <sup>§</sup>	<b>1.72</b> (0.18-3.25) <sup>‡</sup>
<i>miR-146-prec</i>	0.28 (0.00-0.90)*	0.89 (0.44-1.33) <sup>†</sup>	<b>1.03</b> (0.80-1.25) <sup>‡</sup>	0.61 (0.00-1.29) <sup>†,§</sup>	<b>1.00</b> (0.90-1.09) <sup>‡</sup>	<b>1.23</b> (0.50-1.96) <sup>‡</sup>	0.63 (0.00-1.30) <sup>§</sup>	<b>2.24</b> (0.50-3.54) <sup>*,‡</sup>
<i>miR-191-prec</i>	0.11 (0.00-0.54)*	0.52 (0.00-1.21) <sup>*,†</sup>	0.70 (0.07-1.34) <sup>†</sup>	0.62 (0.00-1.29) <sup>†,§</sup>	0.56 (0.00-1.27) <sup>†,§</sup>	0.85 (0.35-1.35) <sup>‡</sup>	0.40 (0.00-1.08) <sup>*</sup>	<b>1.43</b> (0.34-2.51) <sup>§,  </sup>
<i>miR-192</i>	0.24 (0.00-0.84)*	0.68 (0.03-1.33) <sup>§</sup>	0.70 (0.06-1.33)	0.59 (0.00-1.27)	<b>0.96</b> (0.68-1.23) <sup>†</sup>	<b>1.05</b> (0.74-1.36) <sup>†</sup>	0.83 (0.30-1.35)	<b>1.35</b> (0.40-2.31) <sup>†</sup>
<i>miR-219-prec</i>	0.19 (0.00-0.74)*	0.64 (0.00-1.31)	0.73 (0.12-1.35)	0.57 (0.00-1.26)	0.86 (0.39-1.34)	0.63 (0.00-1.30)	0.43 (0.00-1.12) <sup>§</sup>	0.75 (0.15-1.35)
<i>miR-222-prec</i>	0.26 (0.00-0.75) <sup>§</sup>	0.59 (0.00-1.12) <sup>†</sup>	0.76 (0.04-1.15) <sup>†</sup>	0.84 (0.22-1.23) <sup>‡</sup>	<b>1.09</b> (0.79-1.31) <sup>‡</sup>	<b>1.23</b> (0.75-1.39) <sup>‡</sup>	0.53 (0.00-1.07) <sup>†,§</sup>	<b>1.69</b> (0.52-2.49) <sup>‡</sup>

miRNA	ECS/Sham	ECS+BF/Sham	ECS+NAC/Sham	ECS+OPZ/Sham	ECS+OPZ+NAC/Sham	ECS+PEITC/Sham	ECS+I3C/Sham	ECS+PEITC+I3C/Sham
<i>miR-223-prec</i>	0.21 (0.00–0.77) <sup>§</sup>	0.44 (0.00–1.13) <sup>‡,§</sup>	0.50 (0.00–1.19) <sup>‡,§</sup>	0.49 (0.00–1.18) <sup>‡,§</sup>	0.73 (0.12–1.35) <sup>‡</sup>	0.84 (0.34–1.35) <sup>‡</sup>	0.32 (0.00–0.97) <sup>*</sup>	<b>1.45</b> (0.33–2.57) <sup>‡</sup>
<i>miR-294</i>	10.7 (0.00–24.76) <sup>§</sup>	8.76 (0.00–17.78) <sup>§</sup>	9.31 (0.00–21.50) <sup>§</sup>	10.51 (0.00–24.36) <sup>§</sup>	7.67 (0.00–19.97)	9.34 (0.00–21.66) <sup>§</sup>	5.59 (0.00–10.20)	7.17 (0.00–18.78)

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NOTE: The reported values are the ratio of miRNA expression in the lung of ECS-exposed rats, either untreated or treated with chemopreventive agents, to that in the lung of Sham-exposed rats. The numbers between brackets are the 95% confidence intervals. Bold values identify the (ECS + chemopreventive agents)/Sham ratios higher than 0.90, which was arbitrarily assumed as demonstrative of a strong protective effect of chemopreventive agents toward ECS-induced miRNA alterations.

\* Statistical analysis:  $P < 0.01$  compared with Sham.

<sup>‡</sup> Statistical analysis:  $P < 0.05$  compared with ECS.

<sup>‡</sup> Statistical analysis:  $P < 0.001$  compared with ECS.

<sup>§</sup> Statistical analysis:  $P < 0.05$  compared with Sham.

// Statistical analysis:  $P < 0.01$  compared with ECS.

<sup>¶</sup> Statistical analysis:  $P < 0.001$  compared with Sham.

**Table 2**

Main functions regulated by those miRNAs whose ECS-altered expression was normalized by chemopreventive agents (bold values in Table 1)

ECS-altered miRNA	MiRNA-regulated function	Protective chemopreventive agent(s)
<i>let-7a</i>	Cell proliferation, <i>Ras</i> activation, angiogenesis	PEITC + I3C > OPZ + NAC > PEITC
<i>let-7b</i>	Cell proliferation, <i>Ras</i> activation, angiogenesis	PEITC + I3C > OPZ > OPZ + NAC
<i>let-7c</i>	Cell proliferation, <i>Ras</i> activation, angiogenesis	OPZ + NAC > PEITC + I3C > PEITC > OPZ
<i>let-7f</i>	Cell proliferation, <i>Ras</i> activation, angiogenesis	PEITC + I3C > OPZ + NAC
<i>miR-10a</i>	Angiogenesis	PEITC + I3C > BF > OPZ > I3C
<i>miR-26a</i>	<i>TGF</i> expression	PEITC + I3C > PEITC > I3C > OPZ > OPZ + NAC
<i>miR-30a</i>	Cell adhesion, protein repair, NF-κB activation, <i>EGF</i> activation, cell proliferation	PEITC + I3C
<i>miR-30c</i>	Cell adhesion, protein repair, NF-κB activation, <i>EGF</i> activation, cell proliferation	PEITC + I3C
<i>miR-34b</i>	<i>P53</i> effector	NAC > I3C
<i>miR-34c</i>	<i>P53</i> effector	PEITC + I3C > NAC > OPZ + NAC > OPZ
<i>miR-99b</i>	Apoptosis	PEITC + I3C > PEITC
<i>miR-122a</i>	Stress response	PEITC + I3C
<i>miR-123-prec</i>	Angiogenesis	PEITC + I3C > OPZ + NAC > PEITC > OPZ > NAC
<i>miR-124a-prec</i>	Stress response	PEITC + I3C
<i>miR-125a-prec</i>	<i>ErbB2</i> activation	PEITC + I3C > I3C
<i>miR-125b</i>	Stress response	PEITC + I3C > PEITC
<i>miR-140s</i>	<i>P53</i> effector	PEITC + I3C > NAC
<i>miR-145-prec</i>	Protein repair, angiogenesis	PEITC + I3C
<i>miR-146-prec</i>	NF-κB activation	PEITC + I3C > PEITC > NAC > OPZ + NAC
<i>miR-191-prec</i>	Cell proliferation	PEITC + I3C
<i>miR-192</i>	<i>Ras</i> activation	PEITC + I3C > PEITC > OPZ + NAC
<i>miR-219-prec</i>	<i>Elk-1</i> and <i>Fos</i> activation	None
<i>miR-222-prec</i>	Cell proliferation, angiogenesis	PEITC + I3C > PEITC > OPZ + NAC
<i>miR-223-prec</i>	Protein repair, <i>Ras</i> activation	PEITC + I3C
<i>miR-294</i>	Gene transcription	None