



Understanding the Role of Heat Shock Protein Isoforms in Male Fertility, Aging and Apoptosis

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Heat shock proteins (HSPs) play a role in the homeostasis, apoptosis regulation and the maintenance of the various other physiological processes. Aging is accompanied by a decrease in the resistance to environmental stress, while mitochondria are primary targets in the process of aging, their expression decreasing with age. Mitochondrion also plays a significant role in the process of spermatogenesis. HSPs have been shown to be involved in apoptosis with some of acting as apoptotic inhibitors and are involved in cytoprotection. In this review we discuss the roles of Hsp 27, 60, 70, and 90 in aging and male infertility and have concluded that these particular HSPs can be used as a molecular markers for mitochondrially- mediated apoptosis, aging and male infertility.

Key Words: Aging; Heat-shock proteins; Infertility, male; Mitochondria; Apoptosis

INTRODUCTION

Heat Shock Proteins (HSPs), originally identified as stress-responsive proteins, are the most prominent group of proteins involved in folding and unfolding of other proteins. HSPs are found virtually in all living organisms from bacteria to humans [1]. Hsp60, Hsp70, and Hsp90 (the most widely studied HSPs) refer to families of HSPs in the order of 60, 70, and 90 kilodaltons in size, respectively [2]. HSPs represent a highly conserved class of cytoprotective proteins specifically induced at the cellular level in response to one of several environmental stressors (heat shock, cellular energy depletion, oxidative stress or inflammation amongst other).

Some of the important factors involved in HSP induction are hyperthermia, oxidative stress, inflammation, cellular damage (via calcium overload), hypoxia and ischemia and cellular energy depletion [3]. In contrast, others HSPs are constitutively expressed at normal temperature and are only slightly induced by heat shock. They play a crucial role in regulating apoptosis. Hsp27, Hsp70 and Hsp90 are considered to be anti-apoptotic, since they were able to bind to some pro-apoptotic molecules, including cytochrome c and apoptotic protease activating factor 1(Apaf 1) [4]. In contrast, Hsp10 and Hsp60 are considered to be pro-apoptotic [5], although Chandra et al [6] have described a dual role and underlying mechanism for Hsp60 (i.e. Pro-apoptotic and Pro-survival).

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The functional diversity of the HSPs is indicative of their significance as biological molecules that play a significant role in the human aging and in male fertility. This review focuses on identifying the marker HSPs for aging and male fertility in humans.

Hsps 27, 60, 70 AND 90 ISOFORMS, AND SPERMATOGENESIS

Aging significantly alters male reproductive functioning and brings involutive changes in the testes and other organs. The effect of aging on the mammalian male reproductive organs has been principally analyzed in the testes. Spermatogenesis and steroidogenesis decrease with old age [7] and apoptosis increases with age, producing an accelerated germ cell loss [8]. HSPs are involved in the different developmental stages of spermatogenesis all of which involve dramatic transformations and cellular differentiation [9]. In testes showing maturational arrest, Hsp27 expression was strong Sertoli cells, weak in the spermatogonia and spermatocytes and absent in spermatids and Leydig cells. Adly et al [10] reported the differential expression patterns of Hsp27 in the human testes during normal spermatogenesis, indicating a possible role in this process. The seminiferous epithelium of human testes during normal spermatogenesis shows a cell type-specific expression of Hsp27, such that its expression is strong in the cytoplasm of the Sertoli cells, spermatogonia, and Leydig cells, moderate expression was observed in the spermatocytes, weak in the spermatids and absent in the spermatozoa. The altered expression of Hsp27 in testes showing abnormal spermatogenesis may be related to the pathogenesis of male infertility. Hsp60 has been detected in spermatogonia, primary spermatocytes, and Sertoli cells in the rat [11] and human testes [12]. HSPs have been detected on the surface of mouse, rat, bull, boar, and human sperm, and Hsp70 family members appear to be abundant components of the sperm surface [13,14]. Data from mouse, rat, bull, and indicate that Hsp70 protein is constitutively expressed in specific spermatogenic cell types during spermatogenesis [15,16]. Deficiency of Hsp70-2 in mice results in the arrest of spermatogenesis in meiotic prophase I and leads to infertility [17]. Two isoforms of Hsp90 have been identified in the

mouse and human [18] and are highly expressed in pre-meiotic spermatogenic cells [19]. In a recent study [20], it was reported that the aberrant expression of Hsp70 could disturb male fertility. There were significantly different expression levels of Hsp70-2 among normal, maturationally arrested and Sertoli cells-only testis tissues. Low expression of Hsp70-2 was detected in maturationally arrested testes compared to normal testes and Hsp70-2 could not be detected in Sertoli cell-only testes [21]. The Hsp70 proteins are among the most conserved proteins known and are highly homologous from bacteria to man [22].

Rong et al [23], have demonstrated differential Hsp90 expression in mouse testis and epididymis. Hsp90 is expressed in the mouse [24], pig [15], and rat [25] testes. Lee [18] has demonstrated high levels of Hsp90 transcripts in mouse meiotic prophase spermatogenic cells. Hsp90 is expressed at them spermatozoal surface and becomes tyrosine-phosphorylated during sperm capacitation [26]. Wu [27] showed that the expression pattern of Hsp90 in the rabbit testes was similar to that of Hsp70. Liu et al [28] examined Hsp90 alpha in the testicular biopsy specimens of 57 infertile men with maturational arrest. Hsp90 alpha is expressed in spermatogonia, spermatocytes, Sertoli cells, Leydig cells, and myoid cells in the normal testicular tissues, but highly expressed in the testicular tissues with maturational arrest.

ROLE OF HEAT SHOCK PROTEIN IN AGING

Assorted theories state that the length of life is related to an organism's resistance to intrinsic and extrinsic stress, which indicates that the life span is stress-dependent [29]. HSPs are not only induced in response to intrinsic and extrinsic stressors but also are the significant mediators of an organism's resistance to stress. This resistance is associated with promotion of an HSP gene, which in turn activates the HSP expression during aging, enhancing stress resistance and extending the life span. During aging, HSPs show tissue and disease specific expression patterns in unstressed aging animals [30,31]. The expression of HSP chaperones has a significant role in aging and longetivity. An unbalanced chaperone system in an aged organism can lead to the accumulation of aggregated proteins result-

ing mostly in folding diseases in the nervous system. Therefore, over-expression of chaperones often delays the onset or diminishes the symptoms of the disease [32], and increased chaperone induction can lead to increased longevity [33]. HSPs are increasingly being implicated in aging phenotypes and the control of life dyad across species [34].

Jonak et al [35] indicated a link between Hsp27 expression and age-dependent epidermal alterations. High levels of HspB1 (Hsp27) expression usually induces a protective pro-reducing state characterized by a decrease in the levels of reactive oxygen species (ROS) resulting in an HspB1-mediated up-regulation of reduced glutathione and mitochondrial membrane potential [36,37]. The anti-oxidative potential of HspB1 is therefore crucial to counteract the oxidative damages associated with many neurodegenerative [38] and inflammatory diseases, such as asthma induced airway inflammation [39]. During a stressed cell state, Hsp27 functions as a chaperone, while in an unstressed state; it is thought to provide cytoskeletal structural stability [40].

Hsp27 has been implicated in various neurodegenerative diseases. A highly induced expression of Hsp27 has been reported in the brains of aged persons and those with Alzheimer's disease (AD) patients. Hsp27 is present in proliferating astrocytes, neurofibrillary tangles, and Hirano bodies, some hippocampal neurons are also Hsp27 positive. However, in control brains Hsp27 immunoreactivity is restricted to blood vessels and to occasional astrocytes in the white matter. Similarly, patients suffering from other types of dementia (Parkinson/dementia complex, multiinfarct dementia, normal pressure hydrocephalus) showed a certain degree of expression Hsp27 in reactive astrocytes, but that was less than that in AD but more than that in controls. Occasional Hsp27 immunoreactive astrocytes were present in cases without dementia (parkinson's disease, lacunar state, or focal ischemic necrosis) [41].

Dysfunction of chaperones indicates deterioration in the quantity and quality and in addition to physical degeneration. Indentifying the role of Hsp during aging is not simple as aging is accompanied by a number of diseases [42]. Proteins of this family are also called chaperonins. Hsp70 gene polymorphism is observed individuals with shorter lifespan and induction of the Hsp70 gene has been observed in heat-treated isolated blood cells [43]. These results supports the hypothesis that increased longetivity is correlated with lower Hsp gene expression.

The HSP90 Protein family Hsp90 is important in the formation of steroid receptor complexes. To examine the role of chaperones in aging a stochastic model of the chaperone Hsp90 was developed [44]. Hsp90 constitutes upto 2% of the total number of cellular proteins and is highly conserved and abundant molecule [45]. Hsp90 is involved in aging. During the aging process, the normal increase in the concentration of Hsp90 in response to thermal stress is attenuated in lymphocytes [46], liver cells [47], and mesenchymal stem cells [48]. Diminished Hsp90 messenger RNA and protein synthesis following mitogen stimulation of lymphocytes with interleukin-2 has been observed in T lymphocytes from aged donors [49], and tissue from aged animals and blood from elderly humans show reduced production of stress proteins, including Hsp90, following thermal stress [50]. Hsp90 is further implicated in altered homeostasis of aging, which may be particularly relevant in cartilage due to the requirement of this tissue for accurate telomerase assembly and function [51] and given the loss of telomere length with age in chondrocytes [52].

There is increasing evidence for the involvement of HSPs and related heat shock factors in male-derived infertility. HSPs have been reported to be up-regulated in oligospermia and varicocele cases [53]. Several studies have strongly indicated that Ctehlamydia trachomatis may impair fertility by damaging the function of HSPs on the surface of spermatozoa [54].

HEAT SHOCK PROTEINS IN APOPTOSIS

Earlier reports have suggested that mitochondria can be affected by heat tension [55], and data from yeast [56], Antarctic bivalves [57], and rat cardiomyocytes [58] suggest that severe heat stress can structurally and functionally alterations in mitochondria. Left unchecked, dysfunctional mitochondria can cause cell death and eventually lead to deficits in organ function [59]. Haak et al [60] have stated that aging-related stultification of the mitochondrial stress response might have a broad negative influence on the power of aged organisms to tolerate physiological stress.

Aging-related impairments of the mitochondrial stress response may have a broad negative influence on the ability of aged organisms to tolerate physiological stress. Drosophila aging is characterized by a small but widespread downregulation of mitochondrial metabolism and electron transport chain genes [61,62], and this pattern is also observed in aging mammalian tissues [63]. Sustained oxidative damage to nucleic acids, proteins and lipids caused by ROS, is considered to be a major factor in the general functional decline of tissue associated with aging and age-associated degenerative diseases [64,65]. With age, the fluidity of cell membranes, those of mitochondria, decreases and this is associated with enhanced lipid peroxidation [66].

The main mitochondrial stress proteins are Hsp60 and mtHsp70 (mortalin) [67], all of which perform the vital functions of importing, transporting, refolding, and preventing aggregation of mitochondrial proteins [67-69]. Hsp60 is the main heat-inducible protein, although the expression of all three proteins can be upregulated during mitochondrial and cellular perturbation. It has been shown previously that mitochondrial protein degradation and import, two key functions of mitochondrial stress proteins, are impaired with aging, implying that the mi-

tochondrial stress response may be diminished in older organisms [69,70].

While hyperthermic challenge has been shown to induce apoptosis in young mice and rats [71,72], the high levels of cytochrome c release observed in a study [60] suggest that there is a strong activation of the apoptotic caspase cascade in older organisms. Additionally, the blunted Hsp60 levels in older mitochondria may contribute to an apoptotic response after a challenge, as this mitochondrial stress protein has been reported to play a role in suppressing apoptosis [69]. The release of cytochrome c, along with the decreased protein levels of Hsp60, may combine to promote apoptosis in aged animals after a stress induced disruption of normal function.

Mitochondrion have a key role in apoptosis since many of the endogenous cellular proteins that function as crucial determinants of cell death bring about their anti-apoptotic abilities by acting on mitochondria, thereby helping to prevent release of crucial pro-apoptotic proteins [73]. Experiments have demonstrated that Hsp72 and Hsp27 increase cell survival in response to apoptotic stimuli [74,75].

High temperatures can increase the rates of biochemical response which in turn can increase cell metabolism and might lead to increased oxidative processes. Levels of ROS have been shown to increase after exposure to both lethal (\geq 42°C) [76] and non-lethal (40°C) temperatures [77]. This might arise as a result of the mitochondrial

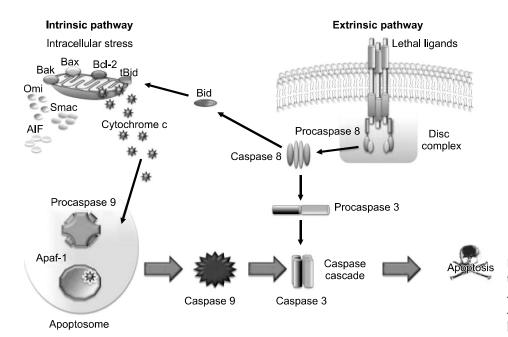


Fig. 1. Showing the intrinsic and the extrinsic apoptotic pathways. Adapted from Favoloro, et al. Aging (Albany NY) 2012;4:735-42 [78].

The World Journal of

respiratory chain dysfunction probably due to increased generation of ROS such as superoxide and hydrogen peroxide.

Cell death is an conserved evolutionary process characterized by a specific set of biochemical and morphological events, resulting in the ordered disassembly of the cell [78,79]. Caspase dependent apoptosis (Fig. 1) [80], occurs as molecular signaling cascade leading to the phenomenon of on blebbing. The resultant apoptotic cells are rapidly identified by phagocytic cells without induction of inflammation or tissue scarring [81].

Caspase-mediated cell death depends on activation of caspases that will then cleave a number of substrates [82] resulting in the biochemical and morphological changes typical of this kind of death. From a functional point of view we can distinguish two classes of caspases can be identified: upstream and downstream caspases. Activation of the up-stream caspases takes place when a sufficient number of enzyme molecules appear in finish adjacency and undergo conformational changes upon binding to the activation coordination compound, resulting in their cleavage and full activation [83]. Downstream caspases are acti-

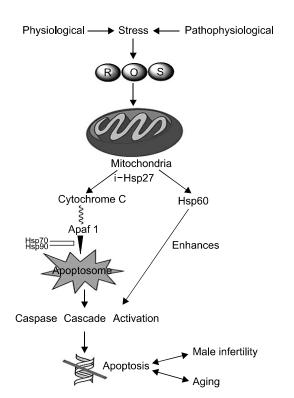


Fig. 2. Diagram summarizing the common mechanism for the role of HSPs in aging and male fertility.

vated by cleavage of the prodomain by upstream caspases. Two major molecular pathways lead to caspase activation and therefore to apoptosis the so-called extrinsic and intrinsic pathways. Multicellular organisms have evolved a series of molecular signaling events for responding to cellular damage and environmental strain. Included among these is the induction of a subfamily of HSPs designated as chaperonins (Hsp60, Hsp70, and Hsp90). Known functions of the chaperonins include folding, assembly and translocation of other proteins, Since apoptosis is considered a normal and necessary a part of many physiological processes (e.g. development, cell turnover, cell injury), it follows that one important facet of the heat shock stress response would be to modulate the balance between cell expiry and survival [84]. HSPs are robustly induced in response to various stressors (both intrinsic and extrinsic) and are key mediators of an organism's resistance to stress [85]. HSPs have been shown to block apoptosis by interfering with caspase activation. Over expression of Hsp27, Hsp70, Hsp60, or Hsp90, inhibits apoptosis and prevents caspase activation in many different cellular models following a variety of cellular stressors, including accumulation of misfolded proteins, or of ROS or DNA damage [86-88]. On the contrary, experimental depletion of Hsp27, Hsp60, Hsp70, or Hsp90, either by anti-sense constructions or siRNA strategies, increases the cells' sensitivity to apoptotic stimuli [89-91]. In some cellular contexts, Hsp70 exhaustion is sufficient to trigger apoptosis by activating caspase-3, in the absence of any additional stressful input [92]. Therefore, HSPs are either directly or indirectly involved in the regulation of caspase activity.

Hsps can block both the intrinsic and the extrinsic apoptotic pathways through the interaction with keystone protein s at three points: (i) upstream of mitochondria, thereby modulating signaling nerve pathways; (ii) at the mitochondrial level, controlling the release of apoptogenic particles; and (iii) and at the post-mitochondrial level, by blocking apoptosis at a later stage than any known survival enhancing drug or protein. HSPs therefore playing an important role in the maintenance and survival potential of cells by acting as anti-apoptotic proteins, a function that appears to be independent of their chaperoning activity. The death-inducing signaling complex is formed as a result of ligand binding to specific receptors in the intrinsic pathway, and caspases-8 is subsequently activated. In the intrinsic pathway, the release of cytochrome c from the mitochondria results in the formation of the apoptosome and activation of caspase-9. Downstream caspases such as caspases-3 are activated by caspase-8 and caspases-9, resulting in cell death. The two pathways are connected through the cleavage of the BH3 only protein; BH3 interacting domain (BID) [81].

Recent evidence has demonstrated that a significant pool of Hsp27 is located in the mitochondrial fraction of thermotolerant Jurkat cells [75]. Hsp27 is devoid of any endogenous ROS detoxifying activity, and can increase intracellular levels of glutathione [74]. It is a tripeptide with numerous functions within cells, including ROS detoxification and regulation of cell death.

Hsp70 also sequesters released apoptosis-inducing factor (AIF) from the mitochondria, thereby preventing caspase independent cell death [93]. Hsp27 and Hsp70 can modulate the death receptor pathway of apoptosis by hindering tBid translocation to mitochondria, which in turn inhibits cytochrome c release [94]. Hsp27, Hsp70, and Hsp90 can inhibit apoptosis upstream of mitochondria [95] and interfere with apoptosome formation, post-mitochondrial events, and caspase activation [96]. Furthermore, Hsp70 and phosphorylated Hsp27 can protect cells against oxidative stress. Hsp90 has been shown to be a negative regulator of caspase-2 activation [97]. Furthermore, Hsp70 and phosphorylated Hsp27 can protect cells against oxidative stress. Hsp90 has been shown to be a negative regulator of caspase-2 activation [97].

Hsp27 and Hsp70 can act at multiple control points of the apoptotic pathways to ensure that stress-induced damage does not inappropriately trigger cell death. Hsp70 and Hsp27 acts as endogenous inhibitors of apoptosis. Hsp70 is a decisive negative regulator of the mitochondrial pathway of caspase-mediated cell death that can block apoptosis at different stages, at a pre-mitochondrial stage by inhibiting stress-inducing signaling; at the mitochondrial stage by preventing mitochondrial membrane permeabilization through the occlusion of Bax translocation; and, at the post-mitochondrial level by interacting with AIF and Apaf-1 or by protecting essential nuclear protein from caspase-3 cleavage. Hsp27 can block cytochrome c induced caspase activation at different stages, namely at the pre-mitochondrial level by inhibiting cytochrome c waiver indirectly through its actions on F-actin, Bid or ROS and at the post-mitochondrial level through the sequestration of cytosolic cytochrome c. Hsp27 may also influence apoptosis by supporting the ubiquitination or degradation of proteins like IkBa or p27kip1 under stressful conditions. Hsp70 can also block caspase-independent pathways, because Hsp70 prevents cell death in conditions in which caspase activation does not occur [98]. In stressed cells, Hsp90 can bind to Apaf-1 thereby inhibiting the activation downstream [99]. Molecular chaperones of the Hsp90 gene family are considered indispensable regulators of protein folding.

Of the molecular chaperones regulated by heat shock factor 1, Hsp70 is the best understood regarding its regulation of signaling and cell death pathways. In addition to its role in promoting the refolding or clearance of misfolded or aggregated proteins, Hsp70 prevents apoptosis through its direct binding with Apaf1 [100].

CONCLUSION

The available literature indicates a clear expression and functional role of the HSPs during spermatogenesis or and process of aging without which the related mechanisms may alter. HSPs play an important role in apoptosis in mitochondrially-mediated aging and male infertility. Hsp27 and Hsp70 are found to be anti-apoptotic while Hsp60 and Hsp90 are pro-apoptotic. The differential expression of HSPs during the various stages of spermatogenesis in species from Drosophilla to human indicates significant role for Hsp27, 60, 70, and 90 in male infertility and also their prompt role in the apoptosis. Mitochondria are the main target site for apoptosis, which in turn leads to aging of the somatic cells and decreased fertility in a germ cells. Consequently, the differential expression of HSPs can be studied with the progression of age. Therefore, Hsp27, Hsp70, Hsp60, and Hsp90 can be used as the molecular markers of mitochondrially- mediated aging and male infertility.

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REFERENCES

- 1. De Maio A. Heat shock proteins: facts, thoughts, and dreams. Shock 1999;11:1-12.
- 2. Li Z, Srivastava P. Heat-shock proteins. Curr Protoc Immunol 2004 Feb; Appendix 1: Appendix 1T.
- 3. Simar D, Ruell P, Caillaud C. Hsp responses to exercising in a warm environment. [Internet]. Doha: ASPETAR; c2013 [cited 2014 Jul 20]. Available from: http://www. aspetar.com/ResearchEducationCentre/HSPResponses.aspx.
- 4. Li P, Nijhawan D, Budihardjo I, Srinivasula SM, Ahmad M, Alnemri ES, et al. Cytochrome c and dATP-dependent formation of Apaf-1/caspase-9 complex initiates an apoptotic protease cascade. Cell 1997;91:479-89.
- 5. Gupta S, Knowlton AA. HSP60, Bax, apoptosis and the heart. J Cell Mol Med 2005;9:51-8.
- 6. Chandra D, Choy G, Tang DG. Cytosolic accumulation of HSP60 during apoptosis with or without apparent mitochondrial release: evidence that its pro-apoptotic or pro-survival functions involve differential interactions with caspase-3. J Biol Chem 2007;282:31289-301.
- 7. Zirkin BR, Chen H. Regulation of Leydig cell steroidogenic function during aging. Biol Reprod 2000;63:977-81.
- 8. Kimura M, Itoh N, Takagi S, Sasao T, Takahashi A, Masumori N, et al. Balance of apoptosis and proliferation of germ cells related to spermatogenesis in aged men. J Androl 2003;24:185-91.
- 9. Meinhardt A, Wilhelm B, Seitz J. Expression of mitochondrial marker proteins during spermatogenesis. Hum Reprod Update 1999;5:108-19.
- 10. Adly MA, Assaf HA, Hussein MR. Heat shock protein 27 expression in the human testis showing normal and abnormal spermatogenesis. Cell Biol Int 2008;32:1247-55.
- 11. Werner A, Meinhardt A, Seitz J, Bergmann M. Distribution of heat-shock protein 60 immunoreactivity in testes of infertile men. Cell Tissue Res 1997;288:539-44.
- 12. Boulanger J, Faulds D, Eddy EM, Lingwood CA. Members of the 70 kDa heat shock protein family specifically recognize sulfoglycolipids: role in gamete recognition and mycoplasma-related infertility. J Cell Physiol 1995;165:7-17.
- 13. Miller D, Brough S, al-Harbi O. Characterization and cellular distribution of human spermatozoal heat shock proteins. Hum Reprod 1992;7:637-45.
- 14. Kamaruddin M, Kroetsch T, Basrur PK, Hansen PJ, King WA. Immunolocalization of heat shock protein 70 in bovine spermatozoa. Andrologia 2004;36:327-34.
- 15. Huang SY, Tam MF, Hsu YT, Lin JH, Chen HH, Chuang CK, et al. Developmental changes of heat-shock proteins in porcine testis by a proteomic analysis. Theriogenology 2005;64:1940-55.
- 16. Dix DJ, Allen JW, Collins BW, Mori C, Nakamura N,

- Poorman-Allen P, et al. Targeted gene disruption of Hsp70-2 results in failed meiosis, germ cell apoptosis, and male infertility. Proc Natl Acad Sci U S A 1996;93:3264-8.
- 17. Minami Y, Kawasaki H, Miyata Y, Suzuki K, Yahara I. Analysis of native forms and isoform compositions of the mouse 90-kDa heat shock protein, HSP90. J Biol Chem 1991;266:10099-103.
- 18. Lee SJ. Expression of HSP86 in male germ cells. Mol Cell Biol 1990;10:3239-42.
- 19. Erata GO, Koçak Toker N, Durlanik O, Kadioğlu A, Aktan G, Aykaç Toker G. The role of heat shock protein 70 (Hsp 70) in male infertility: is it a line of defense against sperm DNA fragmentation? Fertil Steril 2008;90:322-7.
- 20. Feng HL, Sandlow JI, Sparks AE. Decreased expression of the heat shock protein hsp70-2 is associated with the pathogenesis of male infertility. Fertil Steril 2001;76:1136-9.
- 21. Hunt C, Morimoto RI. Conserved features of eukaryotic hsp70 genes revealed by comparison with the nucleotide sequence of human hsp70. Proc Natl Acad Sci U S A 1985;82:6455-9.
- 22. Sarge KD, Cullen KE. Regulation of hsp expression during rodent spermatogenesis. Cell Mol Life Sci 1997;53:191-7.
- 23. Rong C, Han J, Du Z. Expression of heat shock protein 90 β and its regulation in the reproductive system of male mice. Nan Fang Yi Ke Da Xue Xue Bao 2013;33:491-5.
- 24. Moore SK, Kozak C, Robinson EA, Ullrich SJ, Appella E. Murine 86- and 84-kDa heat shock proteins, cDNA sequences, chromosome assignments, and evolutionary origins. J Biol Chem 1989;264:5343-51.
- 25. Ohsako S, Bunick D, Hayashi Y. Immunocytochemical observation of the 90 KD heat shock protein (HSP90): high expression in primordial and pre-meiotic germ cells of male and female rat gonads. J Histochem Cytochem
- 26. Ecroyd H, Jones RC, Aitken RJ. Tyrosine phosphorylation of HSP-90 during mammalian sperm capacitation. Biol Reprod 2003;69:1801-7.
- 27. Wu C. Heat shock transcription factors: structure and regulation. Annu Rev Cell Dev Biol 1995;11:441-69.
- 28. Liu Z, Wang G, Pan Y, Zhu C. Expression of androgen receptor and heat shock protein 90alpha in the testicular biopsy specimens of infertile patients with spermatogenic arrest. Zhonghua Nan Ke Xue 2004;10:662-6.
- 29. Muller FL, Lustgarten MS, Jang Y, Richardson A, Van Remmen H. Trends in oxidative aging theories. Free Radic Biol Med 2007;43:477-503.
- 30. Landis GN, Tower J. Superoxide dismutase evolution and life span regulation. Mech Ageing Dev 2005;126:365-79.
- 31. Macario AJ, Conway de Macario E. Sick chaperones, cellular stress, and disease. N Engl J Med 2005 6;353:1489-
- 32. Soti C, Csermely P. Chaperones come of age. Cell Stress Chaperones 2002;7:186-90.
- 33. Tatar M, Khazaeli AA, Curtsinger JW. Chaperoning extended life. Nature 1997;390:30.

- 34. Tower J. Hsps and aging. Trends Endocrinol Metab 2009;20:216-22.
- 35. Jonak C, Klosner G, Trautinger F. Heat shock proteins in the skin. Int J Cosmet Sci 2006;28:233-41.
- 36. Préville X, Salvemini F, Giraud S, Chaufour S, Paul C, Stepien G, et al. Mammalian small stress proteins protect against oxidative stress through their ability to increase glucose-6-phosphate dehydrogenase activity and by maintaining optimal cellular detoxifying machinery. Exp Cell Res 1999;247:61-78.
- 37. Yan LJ, Christians ES, Liu L, Xiao X, Sohal RS, Benjamin IJ. Mouse heat shock transcription factor 1 deficiency alters cardiac redox homeostasis and increases mitochondrial oxidative damage. EMBO J 2002;21:5164-72.
- 38. Facchinetti F, Dawson VL, Dawson TM. Free radicals as mediators of neuronal injury. Cell Mol Neurobiol 1998; 18:667-82.
- 39. Merendino AM, Paul C, Vignola AM, Costa MA, Melis M, Chiappara G, et al. Heat shock protein-27 protects human bronchial epithelial cells against oxidative stress-mediated apoptosis: possible implication in asthma. Cell Stress Chaperones 2002;7:269-80.
- 40. Bryantsev AL, Kurchashova SY, Golyshev SA, Polyakov VY, Wunderink HF, Kanon B, et al. Regulation of stress-induced intracellular sorting and chaperone function of Hsp27 (HspB1) in mammalian cells. Biochem J 2007;407: 407-17.
- 41. Renkawek K, Stege GJ, Bosman GJ. Dementia, gliosis and expression of the small heat shock proteins hsp27 and alpha B-crystallin in Parkinson's disease. Neuroreport 1999; 10:2273-6.
- 42. Cappello F, Conway de Macario E, Marino Gammazza A, Bonaventura G, Carini F, Czarnecka AM, et al. Hsp60 and human aging: Les liaisons dangereuses. Front Biosci (Landmark Ed) 2013;18:626-37.
- 43. Spector NL, Mehlen P, Ryan C, Hardy L, Samson W, Levine H, et al. Regulation of the 28 kDa heat shock protein by retinoic acid during differentiation of human leukemic HL-60 cells. FEBS Lett 1994;337:184-8.
- 44. Proctor CJ, Soti C, Boys RJ, Gillespie CS, Shanley DP, Wilkinson DJ, et al. Modelling the actions of chaperones and their role in ageing. Mech Ageing Dev 2005;126:119-
- 45. Buchner J. Hsp90 & Co. a holding for folding. Trends Biochem Sci 1999;24:136-41.
- 46. Rao DV, Watson K, Jones GL. Age-related attenuation in the expression of the major heat shock proteins in human peripheral lymphocytes. Mech Ageing Dev 1999;107:105-
- 47. Zhang HJ, Drake VJ, Morrison JP, Oberley LW, Kregel KC. Selected contribution: differential expression of stress-related genes with aging and hyperthermia. J Appl Physiol (1985) 2002;92:1762-9; discussion 1749.
- 48. Stolzing A, Sethe S, Scutt AM. Stressed stem cells: Temperature response in aged mesenchymal stem cells.

- Stem Cells Dev 2006;15:478-87.
- 49. Faassen AE, O'Leary JJ, Rodysill KJ, Bergh N, Hallgren HM. Diminished heat-shock protein synthesis following mitogen stimulation of lymphocytes from aged donors. Exp Cell Res 1989;183:326-34.
- 50. Hunter T, Poon RY. Cdc37: a protein kinase chaperone? Trends Cell Biol 1997;7:157-61.
- 51. Holt SE, Aisner DL, Baur J, Tesmer VM, Dy M, Ouellette M, et al. Functional requirement of p23 and Hsp90 in telomerase complexes. Genes Dev 1999;13:817-26.
- 52. Martin JA, Buckwalter JA. Aging, articular cartilage chondrocyte senescence and osteoarthritis. Biogerontology 2002;3:257-64.
- 53. Ferlin A, Speltra E, Patassini C, Pati MA, Garolla A, Caretta N, et al. Heat shock protein and heat shock factor expression in sperm: relation to oligozoospermia and varicocele. J Urol 2010;183:1248-52.
- 54. Mazzoli S, Cai T, Addonisio P, Bechi A, Mondaini N, Bartoletti R. Chlamydia trachomatis infection is related to poor semen quality in young prostatitis patients. Eur Urol 2010;57:708-14.
- 55. Venkataraman S, Wagner BA, Jiang X, Wang HP, Schafer FQ, Ritchie JM, et al. Overexpression of manganese superoxide dismutase promotes the survival of prostate cancer cells exposed to hyperthermia. Free Radic Res 2004;38: 1119-32.
- 56. Davidson JF, Schiestl RH. Mitochondrial respiratory electron carriers are involved in oxidative stress during heat stress in Saccharomyces cerevisiae. Mol Cell Biol 2001;
- 57. Heise K, Puntarulo S, Pörtner HO, Abele D. Production of reactive oxygen species by isolated mitochondria of the Antarctic bivalve Laternula elliptica (King and Broderip) under heat stress. Comp Biochem Physiol C Toxicol Pharmacol 2003;134:79-90.
- 58. Qian L, Song X, Ren H, Gong J, Cheng S. Mitochondrial mechanism of heat stress-induced injury in rat cardiomyocyte. Cell Stress Chaperones 2004;9:281-93.
- 59. Skulachev VP, Longo VD. Aging as a mitochondria-mediated atavistic program: can aging be switched off? Ann N Y Acad Sci 2005;1057:145-64.
- 60. Haak JL, Buettner GR, Spitz DR, Kregel KC. Aging augments mitochondrial susceptibility to heat stress. Am J Physiol Regul Integr Comp Physiol 2009;296:R812-20.
- 61. Landis GN, Abdueva D, Skvortsov D, Yang J, Rabin BE, Carrick J, et al. Similar gene expression patterns characterize aging and oxidative stress in Drosophila melanogaster. Proc Natl Acad Sci U S A 2004;101:7663-8.
- 62. Pletcher SD, Macdonald SJ, Marguerie R, Certa U, Stearns SC, Goldstein DB, et al. Genome-wide transcript profiles in aging and calorically restricted Drosophila melanogaster. Curr Biol 2002;12:712-23.
- 63. Zahn JM, Sonu R, Vogel H, Crane E, Mazan-Mamczarz K, Rabkin R, et al. Transcriptional profiling of aging in human muscle reveals a common aging signature. PLoS Genet

- 2006;2:e115.
- 64. Shigenaga MK, Hagen TM, Ames BN. Oxidative damage and mitochondrial decay in aging. Proc Natl Acad Sci U S A 1994;91:10771-8.
- 65. Ames BA, Shingenaga MK, Park EM. In: Davies KJA, editors. Oxidative damage and repair, chemical, biological and medical aspects. Elmstad, New York: Pergamon; 1991:181-7.
- 66. Laganiere S, Yu BP. Modulation of membrane phospholipid fatty acid composition by age and food restriction. Gerontology 1993;39:7-18.
- 67. Deocaris CC, Kaul SC, Wadhwa R. On the brotherhood of the mitochondrial chaperones mortalin and heat shock protein 60. Cell Stress Chaperones 2006;11:116-28.
- 68. Wadhwa R, Taira K, Kaul SC. An Hsp70 family chaperone, mortalin/mthsp70/PBP74/Grp75: what, when, and where? Cell Stress Chaperones 2002;7:309-16.
- 69. Bulteau AL, Szweda LI, Friguet B. Mitochondrial protein oxidation and degradation in response to oxidative stress and aging. Exp Gerontol 2006;41:653-7.
- 70. Rea IM, McNerlan S, Pockley AG. Serum heat shock protein and anti-heat shock protein antibody levels in aging. Exp Gerontol 2001;36:341-52.
- 71. Imao M, Nagaki M, Moriwaki H. Dual effects of heat stress on tumor necrosis factor-alpha-induced hepatocyte apoptosis in mice. Lab Invest 2006;86:959-67.
- 72. Sachidhanandam SB, Lu J, Low KS, Moochhala SM. Herbimycin A attenuates apoptosis during heat stress in rats. Eur J Pharmacol 2003;474:121-8.
- 73. Concannon CG, Gorman AM, Samali A. On the role of Hsp27 in regulating apoptosis. Apoptosis 2003;8:61-70.
- 74. Mehlen P, Schulze-Osthoff K, Arrigo AP. Small stress proteins as novel regulators of apoptosis. Heat shock protein 27 blocks Fas/APO-1- and staurosporine-induced cell death. J Biol Chem 1996;271:16510-4.
- 75. Samali A, Robertson JD, Peterson E, Manero F, van Zeijl L, Paul C, et al. Hsp27 protects mitochondria of thermotolerant cells against apoptotic stimuli. Cell Stress Chaperones 2001;6:49-58.
- 76. Moriyama-Gonda N, Igawa M, Shiina H, Urakami S, Wada Y, Terashima M. Heat-induced cellular damage and tolerance in combination with adriamycin for the PC-3 prostate cancer cell line: relationships with cytotoxicity, reactive oxygen species and heat shock protein 70 expression. Eur Urol 2000;38:235-40.
- 77. Pallepati P, Averill-Bates DA. Mild thermotolerance induced at 40°C protects HeLa cells against activation of death receptor-mediated apoptosis by hydrogen peroxide. Free Radic Biol Med 2011;50:667-79.
- 78. Kerr JF, Wyllie AH, Currie AR. Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. Br J Cancer 1972;26:239-57.
- 79. Steller H. Mechanisms and genes of cellular suicide. Science 1995;267:1445-9.
- 80. Favaloro B, Allocati N, Graziano V, Di Ilio C, De Laurenzi

- V. Role of apoptosis in disease. Aging (Albany NY) 2012;
- 81. Wickman G, Julian L, Olson MF. How apoptotic cells aid in the removal of their own cold dead bodies. Cell Death Differ 2012;19:735-42.
- 82. Ai X, Butts B, Vora K, Li W, Tache-Talmadge C, Fridman A, et al. Generation and characterization of antibodies specific for caspase-cleaved neo-epitopes: a novel approach. Cell Death Dis 2011;2:e205.
- 83. Lüthi AU, Martin SJ. The CASBAH: a searchable database of caspase substrates. Cell Death Differ 2007;14:641-50.
- 84. Xanthoudakis S, Roy S, Rasper D, Hennessey T, Aubin Y, Cassady R, et al. Hsp60 accelerates the maturation of pro-caspase-3 by upstream activator proteases during apoptosis. EMBO J 1999;18:2049-56.
- 85. Morimoto RI. Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. Genes Dev 2008;22:1427-38.
- 86. Garrido C, Brunet M, Didelot C, Zermati Y, Schmitt E, Kroemer G. Heat shock proteins 27 and 70: anti-apoptotic proteins with tumorigenic properties. Cell Cycle 2006;5: 2592-601.
- 87. Mosser DD, Caron AW, Bourget L, Meriin AB, Sherman MY, Morimoto RI, et al. The chaperone function of hsp70 is required for protection against stress-induced apoptosis. Mol Cell Biol 2000;20:7146-59.
- 88. Mosser DD, Morimoto RI. Molecular chaperones and the stress of oncogenesis. Oncogene 2004;23:2907-18.
- 89. Kamada M, So A, Muramaki M, Rocchi P, Beraldi E, Gleave M. Hsp27 knockdown using nucleotide-based therapies inhibit tumor growth and enhance chemotherapy in human bladder cancer cells. Mol Cancer Ther 2007;6:299-308.
- 90. Aghdassi A, Phillips P, Dudeja V, Dhaulakhandi D, Sharif R, Dawra R, et al. Heat shock protein 70 increases tumorigenicity and inhibits apoptosis in pancreatic adenocarcinoma. Cancer Res 2007;67:616-25.
- 91. Compton SA, Elmore LW, Haydu K, Jackson-Cook CK, Holt SE. Induction of nitric oxide synthase-dependent telomere shortening after functional inhibition of Hsp90 in human tumor cells. Mol Cell Biol 2006;26:1452-62.
- 92. Gurbuxani S, Bruey JM, Fromentin A, Larmonier N, Parcellier A, Jäättelä M, et al. Selective depletion of inducible HSP70 enhances immunogenicity of rat colon cancer cells. Oncogene 2001;20:7478-85.
- 93. Matsumori Y, Hong SM, Aoyama K, Fan Y, Kayama T, Sheldon RA, et al. Hsp70 overexpression sequesters AIF and reduces neonatal hypoxic/ischemic brain injury. J Cereb Blood Flow Metab 2005;25:899-910.
- 94. Marin-Vinader L, Shin C, Onnekink C, Manley JL, Lubsen NH. Hsp27 enhances recovery of splicing as well as rephosphorylation of SRp38 after heat shock. Mol Biol Cell 2006;17:886-94.
- 95. Liao W, Li X, Mancini M, Chan L. Proteasome inhibition induces differential heat shock protein response but not

- unfolded protein response in HepG2 cells. J Cell Biochem 2006;99:1085-95.
- 96. Gusarova V, Caplan AJ, Brodsky JL, Fisher EA. Apoprotein B degradation is promoted by the molecular chaperones hsp90 and hsp70. J Biol Chem 2001;276:24891-900.
- 97. Garrido C, Solary E. A role of HSPs in apoptosis through "protein triage"? Cell Death Differ 2003;10:619-20.
- 98. Steel R, Doherty JP, Buzzard K, Clemons N, Hawkins CJ, Anderson RL. Hsp72 inhibits apoptosis upstream of the mitochondria and not through interactions with Apaf-1. J

- Biol Chem 2004;279:51490-9.
- 99. Pandey P, Saleh A, Nakazawa A, Kumar S, Srinivasula SM, Kumar V, et al. Negative regulation of cytochrome c-mediated oligomerization of Apaf-1 and activation of procaspase-9 by heat shock protein 90. EMBO J 2000;19:4310-
- 100. Kim HE, Jiang X, Du F, Wang X. PHAPI, CAS, and Hsp70 promote apoptosome formation by preventing Apaf-1 aggregation and enhancing nucleotide exchange on Apaf-1. Mol Cell 2008;30:239-47.