



# Simple chained guide trees give poorer multiple sequence alignments than inferred trees in simulation and phylogenetic benchmarks

Multiple sequence aligners typically work by progressively aligning the most closely related sequences or group of sequences according to guide trees. In PNAS, Boyce et al. (1) report that alignments reconstructed using simple chained trees (i.e., comb-like topologies) with random leaf assignment performed better in protein structure-based benchmarks than those reconstructed using phylogenies estimated from the data as guide trees. The authors state that this result could turn decades of research in the field on its head. In light of this statement, it is important to check immediately whether their result holds under evolutionary criteria: recovery of homologous sequence residues and inference of phylogenetic trees from the alignments (2). We have done this and the results are entirely opposed to Boyce et al.'s findings (1).

Simulation entails simplifying assumptions, but provides a baseline for which the truth is known with certainty. Using ALF (3), we simulated over 100 different evolutionary scenarios, each containing 1,024 homologous sequences evolved along trees generated from birth–death processes. We then applied the same aligners as Boyce et al. (1) (ClustalOmega, Mafft, Muscle) and additionally Prank (4), using as guide trees: (i) chained tree with random leaf assignment of Boyce et al. (1); (ii) balanced tree with leaf assignment optimized using the traveling salesman problem heuristic as tested by Boyce et al. (1); (iii) default tree estimated by each aligner; (iv) least-squares distance tree estimated using specialized phylogenetic software; and (v) the true tree, known from simulation.

With all aligners, using better trees consistently yielded alignments with more homologous columns (Fig. 1A). In particular, chained trees with random leaf assignments yielded the worst alignments under that measure,

with only about half as many correct alignment columns.

To confirm these results on empirical data, we performed a similar analysis on gene families of 1,024 homologous sequences each, sampled from the OMA (Orthologous Matrix) database. Based on the alignments obtained with the various guide trees, we inferred trees and compared their congruence with the National Center for Biotechnology Information taxonomy, assuming that more accurate alignments should yield more accurate trees, which in turn should have a higher congruence with the known biology (5). Here too, there is a clear correlation between the accuracy of the input guide trees and that of the resulting trees (Fig. 1B).

So why can the structure-based benchmark used by Boyce et al. (1) yield results that are so diametrically at odds with simulation-based and phylogeny-based ones? One clue may be that structural benchmarks exclusively consider highly compact, highly conserved core regions, which are atypical outside of structural contexts. In Balibase, used by Boyce et al. (1), the core regions constitute only 18.8% of all alignment columns; the benchmark is thus uninformative about the alignment of the vast majority of the protein sequences. In these conserved regions—50,787 columns in total—only four columns contain gaps; the benchmark provides virtually no information about the placement of insertions and deletions either.

For evolutionary analyses, the conclusion is clear: guide trees closer to the correct evolutionary history of the sequences result in better alignments.

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