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The Isothiocyanato Moiety. An Ideal Protecting Group for Stereoselective Sialic Acid Glycoside Synthesis and Subsequent Diversification^{**}

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Abstract

The preparation of a crystalline, peracetyl adamantanyl thiosialoside donor protected by an isothiocyanate group is described. On activation at -78 C in the presence of typical carbohydrate acceptors this donor gives high yields of the corresponding sialosides with exquisite α -selectivity. The high selectivity extends to the 4-O-benzyl-protected 3-OH acceptors that are typically less reactive and selective than galactose 3,4-diols. Treatment of the α -sialosides with tris(trimethylsilyl)silane or allyltris(trimethylsilyl)silane sialosides replaces the C5-N5 bond by a C-H or a C-C bond. Reaction of the isothiocyanate-protected sialosides with thioacids achieves conversion into amides. Reaction of the isothiocyanate with an amine gives a thiourea, which can be converted to a guanidine. The very high α -selectivities observed with the new donor and the rich chemistry of the isothiocyante function considerably extend the scope for optimization at the sialoside 5-position.

Keywords

glycosylation; isothiocyanate; guanidine; thiourea; deamination

In recent years major steps have been taken toward the establishment of high-yielding and highly α -selective chemical sialidation reactions.^[1] For the most part advances have centered around modification of the N-5 protecting group,^[2] culminating in the discovery of the 4-*O*-,5-*N*-oxazolidine systems^[3] and their *N*-acetyl variants,^[3c, 4] which afford excellent yields and selectivities. Nevertheless, the potential applications of sialic acid glycosides and their oligomers in medicine,^[5] and the consequent need for larger scale synthesis, drive the continued search for improved methods.

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The N-5 position also plays a prominent role in the development of sialic acid glycosides with improved properties for application as therapeutic agents and/or vaccines.^[6] Such N-5 modified systems are either produced chemoenzymatically,^[6b, 7] or chemically by removal of the N-5 protecting group post-glycosylation followed by derivatization.^[6a, 6c, 8] We now reveal that protection of N-5 in the form of an isothiocyanate provides a sialyl donor that is not only exquisitely α -selective in its coupling reactions but which also, by taking advantage of the versatile chemistry^[9] of the isothiocyanate group, affords facile access to an unprecedented range of functionality in the so-formed glycosides. The isothiocyanate **2**, previously obtained in low yield as a by-product in the synthesis of an *N*-acetyl-4-*O*,5-*N*-oxazolidinthione-proected sialyl donor,^[10] was procured in 59% yield by treatment of the β -S-adamantanyl thiosialoside **1**^[4b, 10] with HCl in ether, followed by phenyl chlorothionoformate and aqueous sodium bicarbonate, and finally acetic anhydride in pyridine (Scheme 1). Donor **2** is a stable, readily handled white crystalline solid.

Activation of donor **2** at -78 °C in 2/1 dichloromethane and acetonitrile in the presence of 1.2 equivs of various acceptors (Scheme 2) afforded the corresponding glycosides **3-7** (Table 1). The anomeric configuration of the products was assigned on the basis of the heteronuclear ${}^{3}J_{C1,H3ax}$ coupling constant, which followed the typical pattern.^[11] An authentic sample of the β -anomer of **5** was obtained in 20% yield by coupling **2** with methyl 2,4,6-tri-*O*-benzyl- α - $_{D}$ -galactopyranoside **10** in pure dichloromethane at -30 °C (Figure 1), and enabled confirmation of its absence in the couplings conducted at -78 °C in the dichloromethane/acetonitriloe mixture. A by-product of this latter reaction conducted at -30 °C, obtained in 27% yield, was methyl 3-*O*-(1-adamantanyl)-2,4,6-tri-*O*-benzyl- β - $_{D}$ -galactopyranoside **13** (Figure 1) arising from formation of the 1-adamantanyl cation in the reaction mixture at the higher temperature.

The formation of a single anomer of **5** at -78 °C (Table 1, entry 3) is especially noteworthy. Typically, 4-*O*-protected galactopyranosyl 3-OH acceptors give only poor selectivity in sialidation reactions, even with the oxazolidinone-proected donors,^[4] hence the common use of the more reactive and selective 3,4-galactosyl diols. The excellent selectivities obtained with the isothiocyanate-protected donor are all the more remarkable when contrasted with the coupling reactions of related azide-protected sialyl donors **14** (Figure 1),^[12] which are reported to be competent donors for coupling to primary alcohols,^[13] but to be much less selective with secondary alcohols.^[2, 13c]

Two possibilities were envisaged for the greater selectivity of the isothiocyanate **2** over the structurally-related azides **14**. First, as the isothiocyanate group is considerably more polar than the azido and isocyanate groups (dipole moments of C₆H₅N₃, C₆H₅N=C=O, and PhN=C=S in Debye units, respectively: 1.82, 2.43, 2.69^[14]), it is possible that the isothiocyanate simply serves as a strongly electron-withdrawing group and promotes S_N2 glycosylation as has been proposed^[15] for the oxazolidinone system. Alternatively, consistent with current models for the through-space stabilization of glycosyl oxocarbenium ions,^[16] it is possible that a transient intermediate sialyl oxocarbenium ion preferentially adopts the ⁵H₄ conformation **15** benefitting from stabilization by the pseudoaxial 4-*O*-acetate and the isothiocyanate groups, with the latter providing significant steric shielding to the β-face (Figure 1). In a competition experiment designed to begin to probe this question a

1:1:1 mixture of the isothiocyanate **2**, the *N*-acetyloxazolidinone **16** (Figure 2), and acceptor **10** was activated at -78 °C by the addition of 0.2 equiv of triflic acid. After standard work up the disaccharides **5** and **17** (Figure 2) were isolated in 3 and 51% yield, respectively, the latter as a 4:1 α : β -mixture. Consistent with this result, donors **2** and **16** were recovered from this experiment in 73 and 17% yield, respectively. While this experiment does not exclude the involvement of oxocarbenium ions such as **15**, it establishes the isothiocyanate-protected donor **2** is less reactive than the *N*-acetyloxazolidinone-protected donor **16** under the usual conditions consistent with the highly electron-withdrawing nature of the isothiocyanate moiety.

Turning to the post-glycosylation derivatization of the isothiocyanate group, in a modification of the Saegusa-Barton^[17] radical deamination protocol heating of disaccharide **5** with tris(trimethylsilyl)silane^[18] and AIBN in toluene at reflux afforded the 5-deamino- α -sialoside **18** (Scheme 3). Acetylation of the residual alcohol in **4** followed by AIBN-initiated reaction with allyltris(trimethylsilyl)silane^[19] in toluene at reflux gave the 5-deamino-5-allyl- α -sialoside **19** as a single equatorial diastereoisomer consistent with earlier reports^[20] on radical C-C-bond formation at the 4-position of glucopyranosides (Scheme 3). Reaction of *N*-Boc-L-methionine thioacid and benzyloxy thioacetic acid, both derived by deprotection of the corresponding 9-fluorenylmethyl thioesters,^[21] with disaccharide **5** gave the modified sialosides **20** and **21** in good yield (Scheme 3). The greater ease of reaction of thioacids with isothiocyanate to amide transformation, the residual alcohol in disacccharide **7** was acetylated and the product 7-Ac allowed to react with benzyloxy thioacetic acid in the presence of piperidine in DMF at 40 °C to give the disialoside **22** containing a protected glycolyl amide and an acetamide (Scheme 3).

In a further demonstration of the possibilities afforded by the isothiocyanate group, disaccharide **5** was treated first with 2-phenylethylamine to give the thiourea **23** in 90% yield. Subsequent reaction with methyl iodide gave an isothiourea **24**, which on heating with ammonia in DMF gave the guanidine **25** (Scheme 4).

Finally, selected disaccharides were subjected to a two-step deprotection protocol involving the saponification of all esters followed by hydrogenolysis over palladium-charcoal in aqueous buffer (Scheme 5 and Table 2). In this manner novel sialosides either completely lacking subsitution at the 5-position (Table 2, entry 1) or in which the amido function has been replaced by an alkyl chain (Table 2, entry 2) become available for the first time. A variety of N5 amides can readily produced, as exemplified by the important N-glycoyl chain (Table 2, entry 3), and even guanidines (Table 2, entry 4) may be easily accessed in this manner.

Overall, the crystalline sialyl donor **2** affords very highly stereoselective access to a range of sialyl saccharides. Because of the richness of isothiocyanate chemistry, such isothiocyanate-protected saccharides offer direct introduction of a range of standard and novel functionality at the 5-position post-glycosylation, frequently in a single reaction step. In combination with the stereospecific oxidative deamination methods recently developed in our laboratory,^[24] this chemistry opens up the 5-position of the sialic acid glycosides, beyond the modified

amides accessible by current methods, as a promising locus for the optimization of their diverse biological properties.

Experimental Section

General coupling protocol

A solution of donor **2** (0.15 mmol), acceptor (0.18 mmol), and activated 4 Å acid-washed powdered molecular sieves (300 mg, 2.0 g/mmol) in anhydrous $CH_2Cl_2:MeCN$ (2:1, 2 mL) was stirred for 5 h under Ar, and then cooled to -78 °C, followed by addition of NIS (42 mg, 0.18 mmol) and TfOH (2 μ L, 0.02 mmol). The reaction mixture was stirred at -78 °C for 5 h and then quenched with DIPEA (7 μ L). The mixture was diluted with CH_2Cl_2 , filtered through Celite, washed with 20% aqueous $Na_2S_2O_3$ solution, dried over Na_2SO_4 , and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel eluting with EtOAc:hexane mixtures to afford the desired coupled product.

General Protocol for Amide Formation from Isothiocyanates

To the required 9-fluorenylmethyl (thioester (0.03 mmol) at room temperature was added piperidine (0.21 mmol) in DMF (500 μ L). The reaction mixture was stirred for 15 min, then diluted with CHCl₃ (3 mL). The resulting solution was washed with 1*N* HCl aq. (3 mL) and brine (3 mL), dried over Na₂SO₄, and concentrated *in vacuo*. The residue was dried under high vacuum, and dissolved in dry CH₂Cl₂ (0.5 mL) before addition of the isothiocyanate (0.02 mmol). The reaction mixture was stirred for 36 h at room temperature before the volatiles were removed *in vacuo*. The residue was purified by column chromatography on silica gel eluting with EtOAc:hexane mixtures to afford the corresponding amide.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

References

- 1. a) Hanashima S. Trends in Glycosci Glycobiol. 2011; 23:111–121.b) Adak AK, Yu CC, Liang CF, Lin CC. Curr Op Chem Biol. 2013; 17:1030–1038.
- 2. De Meo C, Priyadarshani U. Carbohydr Res. 2008; 343:1540-1552. [PubMed: 18452900]
- a) Tanaka H, Nishiura Y, Takahashi T. J Am Chem Soc. 2006; 128:7124–7125. [PubMed: 16734441] b) Farris MD, De Meo C. Tetrahedron Lett. 2007; 48:1225–1227.c) Hsu CH, Chu KC, Lin YS, Han JL, Peng YS, Ren CT, Wu CY, Wong CH. Chem Eur J. 2010; 16:1754–1760. [PubMed: 20066711]
- 4. a) Crich D, Li W. J Org Chem. 2007; 72:2387–2391. [PubMed: 17338570] b) Crich D, Li W. J Org Chem. 2007; 72:7794–7797. [PubMed: 17824651]
- a) Angata T, Varki A. Chem Rev. 2002; 102:439–469. [PubMed: 11841250] b) Chen X, Varki AP. ACS Chem Biol. 2010; 5:163–176. [PubMed: 20020717] c) Varki A, Gagneux P. Ann N Y Acad Sci. 2012; 1253:16–36. [PubMed: 22524423] d) Boons, GJ.; Demchenko, AV. Carbohydrate-Based Drug Discovery. Wong, CH., editor. Vol. 1. Wiley-VCH; Weinheim: 2003. p. 55-102.
- 6. a) Wang Q, Guo Z. ACS Med Chem Lett. 2011; 2:373–378. [PubMed: 21691430] b) Rillahan CD, Schwartz E, McBride R, Fokin VV, Paulson JC. Angew Chem Int Ed. 2012; 51:11014–11018.c) Rillahan CD, Macauley MS, Schwartz E, He Y, McBride R, Arlian BM, Rangarajan J, Fokin VV, Paulson JC. Chem Sci. 2014; 5:2398–2406. [PubMed: 24921038]

- a) Sugiarto G, Lau K, Qu J, Li Y, Lim S, Mu S, Ames JB, Fisher AJ, Chen X. ACS Chem Biol. 2012; 7:1232–1240. [PubMed: 22583967] b) Song X, Yu H, Chen X, Lasanajak Y, Tappert MM, Air GM, Tiwari VK, Cao H, Chokhawala HA, Zheng H, Cummings RD, Smith DF. J Biol Chem. 2011; 286:31610–31622. [PubMed: 21757734]
- 8. a) Pazynina G, Tyrtysh T, Nasonov V, Belyanchikov I, Paramonov A, Malysheva N, Zinin AI, Kononov LO, Bovin N. Synlett. 2013:226–230.b) Hsu Y, Ma HH, Lico LS, Jan JT, Fukase K, Uchinashi Y, Zulueta MML, Hung SC. Angew Chem Int Ed. 2014; 53:2413–2416.c) Boltje TJ, Heise T, Rutjes FPT, van Delft FL. Eur J Org Chem. 2013d) Rich JR, Withers SG. Angew Chem Int Ed. 2012; 51:8640–8643.
- 9. a) Witczak ZJ. Adv Carbohydr Chem Biochem. 1986; 44:91–145. [PubMed: 3544702] b) Fernandez JMG, Mellet CO. Adv Carbohydr Chem Biochem. 2000; 55:35–135.c) Braverman S, Cherkinsky M, Birsa ML. Science of Synthesis. 2005; 18:65–320.
- 10. Rajender S, Crich D. J Carbohydr Chem. 2013; 32:324-335.
- a) Czarniecki MF, Thornton ER. J Am Chem Soc. 1977; 99:8273–8279.b) Hori H, Nakajima T, Nishida Y, Ohrui H, Meguro H. Tetrahedron Lett. 1988; 29:6317–6320.c) Haverkamp J, Spoormaker T, Dorland L, Vliegenthart JFG, Schauer R. J Am Chem Soc. 1979; 101d) Prytulla S, Lauterwein J, Klessinger M, Thiem J. Carbohydr Res. 1991; 215:345–349.e) Kancharla PK, Crich D. J Am Chem Soc. 2013; 135:18999–19007. [PubMed: 24261615]
- 12. Schneider R, Freyhardt CC, Schmidt RR. Eur J Org Chem. 2001:1655-1661.
- a) Yu CS, Niikura K, Lin CC, Wong CH. Angew Chem Int Ed. 2001; 40:2900–2903.b) Mukaiyama T, Mandai H, Jona H. Chem Lett. 2002:1182–1183.c) Lu KC, Tseng SY, Lin CC. Carbohydr Res. 2002; 337:755–760. [PubMed: 11950471]
- 14. a) Cheng CL, Le Fevre RJW, Ritchie GLD. J Chem Soc B. 1971:435–437.b) Förner W, Badawi HM. J Theo Comp Chem. 2010; 9:511–529.
- a) Kancharla PK, Navuluri C, Crich D. Angew Chem Int Ed. 2012; 51:11105–11109.b) Kancharla PK, Kato T, Crich D. J Am Chem Soc. 2014; 136:5472–5480. [PubMed: 24606062]
- a) Smith DM, Woerpel KA. Org Biomol Chem. 2006; 4:1195–1201. [PubMed: 16557303] b) Jensen HH, Bols M. Acc Chem Res. 2006; 39:259–265. [PubMed: 16618093]
- a) Saegusa T, Kobayashi S, Ito Y, Yasuda N. J Am Chem Soc. 1968; 90:4182–4182. [PubMed: 5655885] b) Barton DHR, Bringmann G, Lamotte G, Motherwell WB, Hay Motherwell RS, Porter AEA. J Chem Soc Perkin Trans 1. 1980:2657–2664.
- Ballestri M, Chatgilialoglu C, Clark KB, Griller D, Giese B, Kopping B. J Org Chem. 1991:678– 683.
- a) Kosugi M, Kurata H, Kawata Ki, Migita T. Chem Lett. 1991:1327–1328.b) Chatgilialoglu C, Ferreri C, Ballestri M, Curran DP. Tetrahedron Lett. 1996; 37:6387–6390.
- 20. a) Giese B, Witzel T. Angew Chem Int Ed. 1986; 25:450–451.b) Gupta V, Kahne D. Tetrahedron Lett. 1993; 34:591–594.
- 21. Crich D, Sana K, Guo S. Org Lett. 2007; 9:4423-4426. [PubMed: 17900128]
- 22. a) Kricheldorf HR, Leppert E. Makromol Chem. 1973; 167:47–68.b) Gonda J, Bednárikova M. Tetrahedron Lett. 1997; 38:5569–5572.c) Gonda J, Zavacká E, Buděšínský M, Císa ova I, Podlaha J. Tetrahedron Lett. 2000; 41:525–529.d) Gonda J, Martinková M, Walko M, Zavacká E, Buděšínský M, Císa ova I. Tetrahedron Lett. 2001; 42:4401–4404.e) Schoepfer J, Marquis C, Pasquier C, Neier R. J Chem Soc Chem Commun. 1994:1001–1002.f) Crich D, Sasaki K. Org Lett. 2009; 11:3514–3517. [PubMed: 19719195]
- 23. a) Hakimelahi GH, Just G. Tetrahedron Lett. 1980; 21:2119–2122.b) Rakotomanomana N, Lacombe JM, Pavia A. Carbohydr Res. 1990; 197:318–323.c) Shangguan N, Katukojvala S, Greenberg R, Williams LJ. J Am Chem Soc. 2003; 125:7754–7755. [PubMed: 12822965]
- 24. Navuluri C, Crich D. Angew Chem Int Ed. 2013; 52:11549–11552.

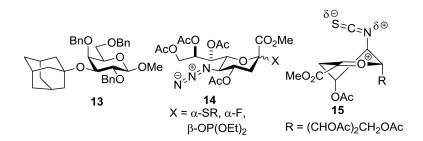


Figure 1. Structures of the Adamantanyl Ether 13, of Azido-Protected Sialyl Donors 14, and of the ${}^{5}H_{4}$ Conformer the Hypothetical Oxocarbenium Ion 15

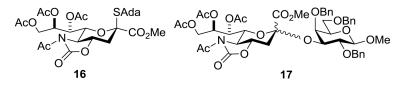
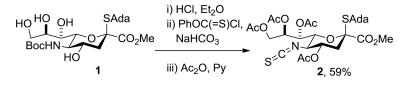
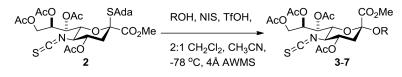


Figure 2. Structures of the *N*-acetyloxazolidionone-protected donor 16 and glycoside 17 from the competition experiment



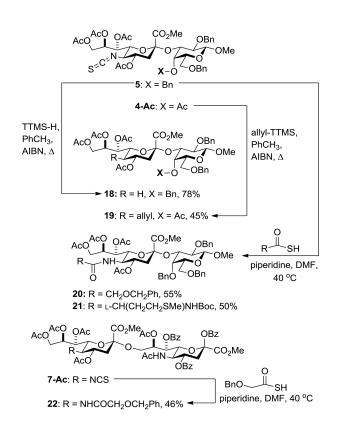
Scheme 1.

Synthesis of the Isothiocyanate **2**. Abbreviations: Ada = 1-adamantanyl, Boc = *tert*-butyloxycarbonyl, Py = pyridine



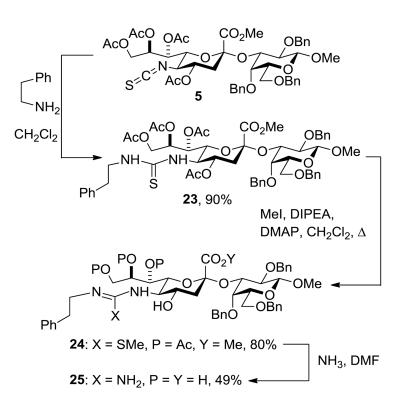
Scheme 2.

Glycosylation with Isothiocyanate 2. Abbreviations: NIS = N-iodosuccinimide, TfOH = trifluoromethanesulfonic acid, AWMS = acid-washed molecular sieves.



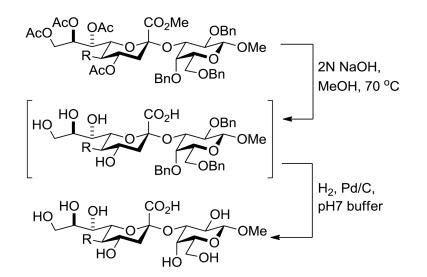
Scheme 3.

Formation of Desamino and Amido Derivatives from Isothiocyanate **5**. Abbreviation: TTMS = tris(trimethylsilyl)silyl



Scheme 4.

Synthesis of Thiourea and Guanidine Derivatives. Abbreviations: DIPEA = diisopropylethylamine, DMAP = 4-dimethylaminopyridine, DMF = dimethylformamide



Scheme 5. Deprotection of Selected Disaccharides

Table 1

Glycosylation reactions with isothiocyanate $2^{[a][b]}$

Entry	Acceptor	Product, Yield, selectivity
1	$\begin{array}{c} & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & &$	
2		- Offin - 00 - 0
3		COBn COSn COSn t → 0, COSn t → 0, COSn S. X + laothocyanyl saityl, 87%, n− only
4[c]		$\begin{array}{c} COIn \\ & COIn \\ COIn \\ COIn \\ & $
5		HD QBz OBz Admit Colle 0 Obz H

[a]Bn = benzyl, Bz = benzyl, isothiocyanyl sialyl = [methyl (4,7,8,9-tetra-O-acetyl-3-deoxy-5-isothiocyanyl D-glycero-D-galacto- α , β -nonulopyranosid)onate].

[b] unless otherwise stated all reactions were conducted at -78 °C with 1.2 equiv of acceptor in 2:1 dichloromethane:acetonitrile.

[c] After glycosylation the crude reaction mixture was acetylated to facilitate purification.

Table 2

Deprotection Reactions

Entry	Substrate	Product, Yield, selectivity
1	18	HO HO OH CO2H OH HO HO CO2H OH HO HO COH 26, 91%
2[a]	19	HO HO OH CO2H OH HO HO CO2H OH HO HO COH
3	20	HO HO CON HO HO HO CON HO HO HO CON HO CON 28, 91%
4	25	но но он со он оме нух ни ос он ос он ph N HO HO COH 29,52%

[a]Concomitant hydrogenation of the allyl group took place in the course of the hydrogenolytic debenzylation.