


Research Paper

Database of Gene Co-Regulation (dGCR): A Web Tool for Analysing Patterns of Gene Co-regulation across Publicly Available Expression Data

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Abstract

The database of **Gene Co-Regulation (dGCR)** is a web tool for the analysis of gene relationships based on correlated patterns of gene expression over publicly available transcriptional data. The motivation behind dGCR is that genes whose expression patterns correlate across many experiments tend to be co-regulated and hence share biological function. In addition to revealing functional connections between individual gene pairs, extended sets of co-regulated genes can also be assessed for enrichment of gene ontology classes and interaction pathways. This functionality provides an insight into the biological function of the query gene itself. The dGCR web tool extends the range of expression data curated by existing co-regulation databases and provides additional insights into gene function through the analysis of pathways, gene ontology classes and co-regulation modules.

Key words: Global gene expression; connectivity map; microarray.

Introduction

Global gene expression serves as a quantitative high content descriptor of biological states. This together with the vast amount of publicly available data [1, 2] provides an ideal basis for comparing biological perturbations through their associated transcriptional profiles across a wide range of experiments. The approach has been successfully applied in the area of drug repositioning with the connectivity map (CMAP) initiative [3, 4] and has led to the development of transcription based search engines such as GEM-TREND [5], ProfileChaser [6], Gene Expression Atlas [7] and SPIED [8, 9], that have thrown up intriguing connections between diverse biological systems. These methodologies are not driven by gene discovery and the pinpointing of a single gene underlying a given pathology, for example. Rather, the transcriptomes are compared globally, with the regu-

lation of individual genes not making a profound contribution to the correlation. However, expression data can be a useful tool in assigning shared biological function to a pair of genes that show correlated expression changes over a relatively large dataset. In other words, robust patterns of co-expression can inform co-regulation and therefore function. Currently, there are many gene analysis portals based around correlated gene expression patterns. The Human Gene Co-expression [10] web tool taps into the gene expression variation within a set of immortalised human cell lines with varied genotypes to define patterns of co-expression for over 4,000 genes. Whereas, Gene friends [11] provides co-expression patterns based on over 1,000 murine arrays. CoXpresDB [12] has extended the analysis to include data from a variety of species in an effort to increase the robustness of

gene pair co-regulation through conservation across species. The data underlying this analysis is from a relatively large set of arrays, ~10,000, and from 11 species. BIOGPS [13] is a gene annotation and analysis portal with links to expression data over human, mouse and rat arrays corresponding to various tissue types. These expression patterns are compared to define ranked lists of correlating genes. The neuroblast application of Allen Brain Atlas [14] uses *in situ* data from multiple well defined mouse brain and spinal cord regions to define gene expression patterns and compare these to inform shared function.

The essentially automated compilation of transcriptional data into the unified searchable platform SPIED allowed for the quantitative comparison of query expression profiles against a theoretically unlimited amount of deposited transcriptional data. The data underlying SPIED is effectively a large matrix over experimental conditions and genes. Furthermore, the matrix can be thought of as a series of expression patterns for the genes, extending to over 200,000 conditions, and these can be correlated resulting in a gene co-regulation matrix. The dGCR web tool presented here was designed to interrogate this matrix. In particular, dGCR is based on the analysis of the most regulated genes across the database facilitating a simple binary statistic fisher test measure of the correlation in gene expression patterns. Thus, gene co-regulation can be defined over a vast collection of diverse experimental conditions and the robustness assessed over blocks of data corresponding to species type and platform technology.

The details underlying the construction of dGCR are given in the Methods section. In the Results sections the use of dGCR is illustrated with examples and an analysis of the global picture of gene co-regulation presented. Specifically, a detailed analysis of the co-regulation matrix shows a significant overlap between the different platforms and hence a high degree of internal consistency. It is further shown that protein/gene interaction networks, gene ontology classes and pathway gene sets are significantly enriched for co-regulated genes. Finally, hierarchical clustering derived gene modules, demarcating distinct functional networks, are derived from the co-regulation matrix.

Results

The user inputs a gene name and then selects the required gene symbol from a candidate list of possible alternatives, see Figure 1. The number of top scoring co-regulating genes to be displayed can be extended from the default of 50. Also, co-regulation can be restricted to be measured over specific species or plat-

form technologies. Once the chosen gene is selected the output lists the top co-regulated genes together with details of the platforms where co-regulation is significant, see Figure 2. Web links enable the user to query literature on the given genes. The gene list as a whole is usually enriched for informative gene ontology (GO) classes and pathways. With this in mind the output page hosts buttons that link to enriched GO and pathway classes that can reveal functional aspects of the query gene, see Figure 3. The example gene queries correspond to the two isoforms of DAG lipase, DAGLA and DAGLB. These enzymes have diverse functions, being implicated in neuronal development, adult neurogenesis, synaptic retrograde signalling and immune responses [15]. It is of interest to see to what extent gene co-regulation can throw light on the different functions of the two isoforms. From GO and pathway analysis it is clear that the α isoform is involved in synaptic function whereas the β isoform is involved in the immune response, see Figure 3. Here, dGCR was queried across the whole database, without restrictions on species or platform type. The synaptic role of DAGLA has long been established [16] and the role of DAGLB in the immune response has recently emerged [17]. A further analysis consists in placing the query gene in a module defined through hierarchical clustering, see Methods. Here novel connections can emerge where two genes are weakly correlated ($\sum \log(p) > -100$, see Methods), but belong to the same module by virtue of shared co-regulated genes. In the present query case, DAGLA belongs to a module of 107 genes and the receptor for its hydrolysis product, 2-AG, the cannabinoid receptor (CB1, CNR1), is in the same module [16].

A global analysis

The gene expression samples making up dGCR come from 26 array platforms, corresponding to Affymetrix (www.affymetrix.com), Illumina (www.illumina.com) and Agilent (www.agilent.co.uk) technologies and with probes specific to human, mouse and rodent material. One of the reasons for the effectiveness of dGCR is that patterns of gene co-regulation are highly conserved across these platforms. This is quantified in the Supplementary Material: Table S1, detailing the number of genes present for each platform, the number of co-regulated pairs (defined by a stringent significance threshold of the Fisher exact test, $\log(p) < -100$) and the number of these shared between the platforms. For example, comparing the human array GPL96 with the mouse array GPL1261, there are 9,963 common genes with 165,240 and 130,005 co-regulated pairs respectively and 26,424 of these are shared.

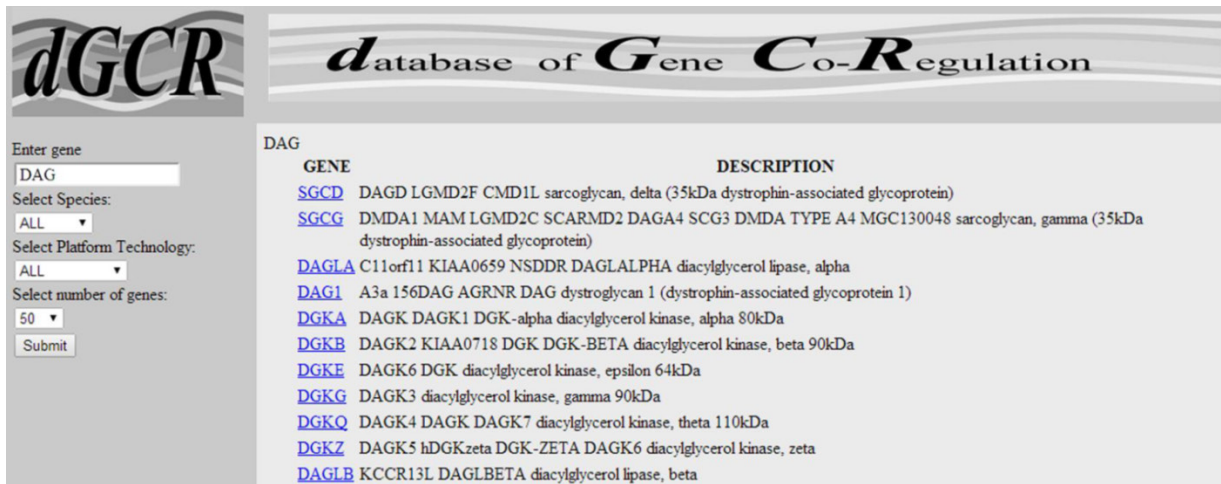


Figure 1. The dGCR web page screen shot with query 'DAG'. The possible genes are listed with links for further analysis. At left the user selects the number of co-regulating genes (50-500) and may restrict the analysis to a given species. The two genes that will be analysed here are the two isoforms of DAG lipase, DAGL α (DAGLA) and DAGL β (DAGLB).

Figure 2. The genes most co-regulated with the two isoforms of DAG lipase. The scores are the sums of the fisher log-odds scores for each platform. Each gene entry is hyperlinked to explanatory web material. The correlation values and significance scores (in brackets) are given for the individual platforms in the right column. Moving the mouse over the individual scores reveals the platform where the correlation obtains.

Genes co-regulating with DAGLA

Enriched [Gene Ontology Classes](#) [Pathways](#)
Genes in same [Module](#)

GENE	score	platforms	CORRELATION DETAILS
CCK	-206.88	5	0.43(-19.24) 0.47(-13.51) 0.96(-114.11) 0.98(-49.40) 0.52(-10.62)
CAMK2A	-193.37	8	0.37(-10.82) 0.73(-8.31) 0.33(-8.09) 0.91(-101.54) 0.74(-25.67) 0.53(-9.50) 0.92(-9.86) 0.83(-19.58)
SLC30A3	-173.90	4	0.74(-12.53) 0.94(-109.64) 0.87(-41.77) 0.71(-9.96)
ITPKA	-171.07	5	0.59(-15.66) 0.91(-101.44) 0.64(-20.80) 0.62(-12.05) 0.90(-21.12)
DLGAP2	-169.66	3	0.56(-10.92) 0.98(-116.69) 1.00(-42.05)
CAMKV	-168.82	4	0.94(-107.56) 0.88(-40.09) 0.73(-12.03) 0.54(-9.14)
CACNG3	-163.69	3	0.74(-8.16) 0.97(-119.37) 0.89(-36.16)
SATB2	-158.17	4	0.37(-10.54) 0.95(-104.26) 0.78(-32.94) 0.53(-10.43)
KCNH3	-155.35	4	0.47(-8.67) 0.98(-104.75) 0.91(-32.89) 0.57(-9.04)
FBXL16	-154.49	6	1.00(-8.90) 0.65(-11.02) 0.93(-88.65) 0.51(-10.48) 0.89(-12.02) 0.91(-23.42)
CTXN1	-152.63	3	0.94(-115.29) 0.67(-29.21) 0.51(-8.13)

Genes co-regulating with DAGLB

Enriched [Gene Ontology Classes](#) [Pathways](#)
Genes in same [Module](#)

GENE	score	platforms	CORRELATION DETAILS
ZFP36L2	-260.24	3	0.70(-152.80) 0.66(-85.67) 0.74(-21.77)
STK32B	-226.23	3	0.76(-177.58) 0.68(-37.82) 0.73(-10.83)
INPP5D	-188.73	6	0.75(-112.12) 0.45(-18.87) 0.58(-12.76) 0.90(-10.03) 0.89(-8.29) 0.77(-26.66)
SCARB1	-184.44	5	0.75(-93.43) 0.51(-22.90) 0.82(-27.89) 0.91(-20.83) 0.78(-19.39)
SHANK3	-171.72	2	0.71(-156.10) 0.39(-15.62)
NAV1	-168.74	4	0.57(-119.24) 0.22(-9.23) 0.45(-12.53) 0.76(-27.74)
RANGAP1	-168.23	7	0.76(-58.16) 0.66(-29.11) 0.85(-22.04) 1.00(-8.38) 0.96(-30.89) 1.00(-10.69) 0.90(-8.96)
MLXIP	-167.12	2	0.75(-106.09) 0.65(-61.03)
WFS1	-164.48	5	0.72(-92.61) 0.67(-29.17) 0.45(-8.43) 0.74(-15.50) 0.95(-18.77)
STK38	-156.23	3	0.63(-119.46) 0.42(-24.09) 1.00(-12.68)
SMARCA4	-149.42	2	0.65(-115.35) 0.45(-34.07)

A**Gene Ontology Classes**

GO term	<i>p</i>	GENES
GO:0045202: synapse (276)	$e^{-16.60}$	RDMBP2; CHRMI
GO:0030425: dendrite (166)	$e^{-14.81}$	DDN; ADCY2; C
GO:0007399: nervous system development (384)	$e^{-13.84}$	GFRA2; ENCI
GO:0030054: cell junction (395)	$e^{-13.61}$	GRIN2A; CF
GO:0005216: ion channel activity (311)	$e^{-13.08}$	GABRA5;
GO:0006811: ion transport (556)	$e^{-12.93}$	KCNH2
GO:0005244: voltage-gated ion channel activity (154)	$e^{-12.38}$	SCN2
GO:0005516: calmodulin binding (143)	$e^{-9.97}$	NRX
GO:0007268: synaptic transmission (201)	$e^{-8.40}$	NP
GO:0006810: transport (1548)	$e^{-6.08}$	CAI

B**Pathways**

Pathway	<i>p</i>	GENES
REACTOME NEURONAL SYSTEM (275)	$e^{-19.40}$	CACNG3; ADC GRIN2A; KCN STX1A; CAM
REACTOME TRANSMISSION ACROSS CHEMICAL SYNAPSES (183)	$e^{-17.08}$	CACNG3; AF KCNJ4; SLC
REACTOME NEUROTRANSMITTER RECEPTOR BINDING AND DOWNSTREAM TRANSMISSION IN THE POSTSYNAPTIC CELL (134)	$e^{-13.18}$	CACNG3; KCNJ4; C
KEGG CALCIUM SIGNALING PATHWAY (171)	$e^{-9.14}$	CHRM ITPKA
REACTOME GABA RECEPTOR ACTIVATION (52)	$e^{-7.81}$	ADC
REACTOME GPCR DOWNSTREAM SIGNALING (763)	$e^{-6.95}$	AD DA
REACTOME G ALPHA Q SIGNALLING EVENTS (171)	$e^{-6.70}$	CF
REACTOME P75 NTR RECEPTOR MEDIATED SIGNALLING (77)	$e^{-6.68}$	NR
REACTOME GASTRIN CREB SIGNALLING PATHWAY VIA PKC AND MAPK (191)	$e^{-6.31}$	CH

C**Gene Ontology Classes**

GO term	<i>p</i>	GENES
GO:0005096: GTPase activator activity (168)	$e^{-9.22}$	RANGAP1; SIP1; LRRK
GO:0007155: cell adhesion (561)	$e^{-7.18}$	CD300A; SCARB1; ZYX
GO:0006955: immune response (467)	$e^{-6.45}$	GBP1; PPBP; LTB4R2
GO:0000166: nucleotide binding (1800)	$e^{-6.03}$	STK38; OBP1; DHR
GO:0005624: membrane fraction (610)	$e^{-5.20}$	HPS4; LAMP1; F
GO:0005856: cytoskeleton (541)	$e^{-4.30}$	NAV1; ARHG
GO:0016020: membrane (1782)	$e^{-4.18}$	JUP; DHR53
GO:0005886: plasma membrane (1873)	$e^{-3.91}$	CD300A; T
GO:0016301: kinase activity (670)	$e^{-3.56}$	CRK; LRI
GO:0005488: binding (769)	$e^{-3.13}$	FAM65A;

D**Pathways**

Pathway	<i>p</i>	GENES
ST INTEGRIN SIGNALING PATHWAY (80)	$e^{-9.54}$	ITGA6;
PID CXCR4 PATHWAY (98)	$e^{-8.77}$	INPP5
KEGG PATHWAYS IN CANCER (320)	$e^{-6.36}$	JUP
KEGG NEUROTROPHIN SIGNALING PATHWAY (123)	$e^{-5.38}$	Y
PID PDGFRBPATHWAY (124)	$e^{-5.36}$	
KEGG CHEMOKINE SIGNALING PATHWAY (184)	$e^{-4.3}$	
KEGG FOCAL ADHESION (190)	e^{-4}	
REACTOME PLATELET ACTIVATION SIGNALING AND AGGREGATION (195)	e^{-4}	
REACTOME SIGNALLING BY NGF (204)	e^{-4}	

Figure 3. The user can analyse the co-regulated gene set for the enrichment of gene ontology classes, shown in A and C, and pathways, shown in B and D. Both analyses point to a synaptic function for DAGL α and immunological function for DAGL β .

Pathways and Gene Ontology classes

A complementary global analysis consists of directly scoring pathway sets for the mutual co-regulation of the genes. This analysis is relevant to transcriptional data analysis in general as pathway [18] and GO set enrichment [19] are ubiquitously applied in the field. To this end we scored all pairs of genes within a given pathway for their co-regulation across the full platform set. On average the intra-pathway co-regulation scores are highly significant. Explicitly, all pairs of genes within the given pathway pair are assigned correlation scores corre-

sponding to the summed log Fisher scores across all the array platforms and the average over all pairs is highly significant, with $(\sum \log(p)) = -182$. The distribution of co-regulation scores is compared to that of a random collection of gene sets in Figure 5A. GO classes are defined through shared molecular function, biological process and cellular location. There is thus an obvious basis for genes in the same class to be co-regulated. To quantify this we scored the available GO sets for co-regulation against the dGCR database in a similar way to that of the pathway analysis already described. Again the average intra-pathway

co-regulation score is highly significant ($\langle \sum \log(p) \rangle = -187$). The distribution of co-regulation scores is compared to that of a random collection of gene sets in Figure 5A.

Protein and gene interaction networks

It is interesting to compare co-regulation with biological interaction at the protein level as there is an obvious basis for interacting proteins to be correlated

in their expression patterns. BioGRID [20] hosts a comprehensive dataset of protein interaction data. Interactions fall into many categories and varying degrees of certainty. Taking each category in turn we collected the co-regulation scores for the genes corresponding to the given protein pairs and found that these are shifted towards high significance relative to random pairs. See Figure 5B.

Module #23

ABHD12B	ADAM23	ADCY2	ADFP	AK5	ANKRD24	ANKS1B	ASCL1	ASTN1	BAIAP2
BAI3	BCAN	BCL9	B3GAT1	CABP1	CACNA2D3	CACNG3	CACNG7	CAMKV	CAMK2A
CNR1	CNTNAP2	COL19A1	COL22A1	CRMP1	CSPG5	CTNND2	CX3CL1	C1QL2	DAGLA
DCLK2	DCX	DDN	DGKB	DLX5	DNM3	EPHB6	FAM124A	FAM131B	FBN3
FGF13	FIGN	FOXP2	FRAS1	FSTL4	GABRB1	GABRB3	GAP43	GFAP	GPM6A
GRM3	GSG1L	HPCA	H2AFY2	IL17RD	ITPKA	KCNB1	KCNJ4	KIF1A	KIF26A
KIF5C	KLHL13	LASS1	LGR5	LHX8	LMO3	LRP4	LRRN1	MARK1	MKRN3
MPPED2	MT3	MUMIL1	NCALD	NCAN	NCDN	NGEF	NPAS2	NRCAM	NTSR2
OLIG1	OMG	PCDH9	PDE2A	PDE9A	PPP2R2B	PTPRZ1	RAVER2	RGS12	RTN1
SALL2	SCG3	SCN3B	SEMA6A	SOX11	SPTBN2	SST	STOX2	SYT10	TAC1
TERT	TTYH1	TUBB2B	VANGL2	WASF1	ZNF462	ZNF618			

Figure 4. Screen shot of module #23. The module consists of 107 genes and DAGL α is shown highlighted in blue. The module links the post-synaptic 2-AG synthesising enzyme, DAGL α , to the pre-synaptic 2-AG receptor, the cannabinoid receptor (CBI, CNR1), circled in red.

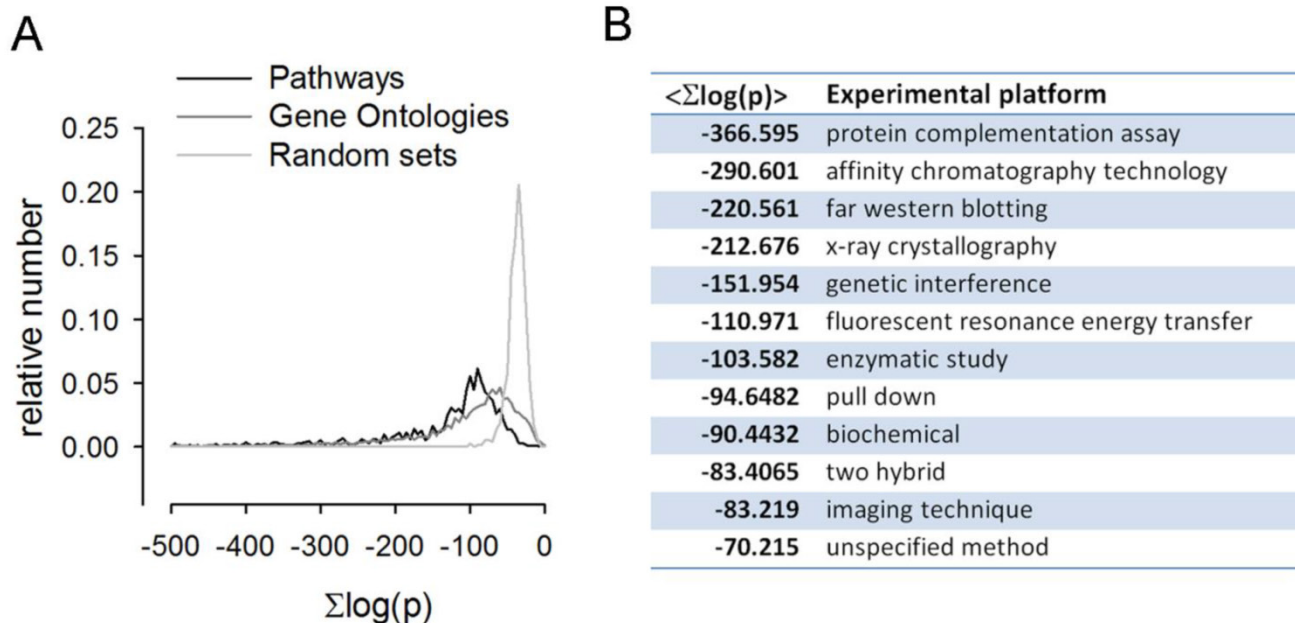


Figure 5. Pathways, GO sets and interacting proteins were scored for mutual co-regulation. The summed log-odds co-regulation scores across the array platforms for each gene pair in a given pathway and GO set were compiled and the distributions plotted in A. It is clear that for both pathway sets and gene ontology sets the distributions are significantly shifted towards high scores relative to the random gene sets distribution. The co-regulation score averages across the protein interacting pairs from the 12 distinct experimental platforms in the BioGrid database are shown in the table B.

Discussion and Conclusions

Patterns of gene co-regulation can offer an insight into shared biological function. In contrast to pathway and gene ontology sets, co-regulation is established purely quantitatively, based on multiple gene expression studies across different species and cell/tissue types. The dGCR web tool presented here is a simple to use interface for accessing patterns of co-regulation and should enable users to uncover novel relationships between query genes and offer novel insights into the underlying biology. The database constitutes a complimentary analysis to that of SPIED, which is queried with expression profile data and offers a quantitative comparison of biological perturbations across multiple experiments. Whereas, dGCR is based on a re-working of the same expression data so that comparisons of genes are scored across samples as opposed to comparisons of samples being scored across genes. The idea that co-regulation informs function underlies the many gene analysis portals detailed in the introduction. The novelty with dGCR is to extend the methodology to include at present a quarter of publicly available transcriptional data for human and rodent species. The data processing underlying dGCR is automatic, based on a simple normalisation procedure, and it is hoped to extend the database beyond the current 200,000 samples and include other species.

Methods

dGCR is based on the expression data collected in SPIED [8, 9], compiled from the data hosted by NCBI GEO (www.ncbi.nlm.nih.gov/geo), consisting of over 200,000 array samples covering Affymetrix, Agilent and Illumina platform technologies and human, mouse and rat material. Here, expression data was converted to fold profiles defined relative to the probe average across the sample series. The probe profiles were then mapped to profiles defined in terms of non-redundant gene sets. For genes with multiple probes the gene level was assigned the level of the probe with greatest fold magnitude. SPIED was populated with gene lists ranked according to fold magnitude. In addition to the full database an abridged database consisting of the top 1000 regulated genes (500 up and 500 down regulated) was also created for rapid querying and for gene set enrichment analysis. This latter database facilitated a simple expression pattern comparison between genes, where the up/down status of a given pair could be followed over an extensive set of samples and the extent of co-regulation determined. This database served as the basis for dGCR. Specifically, for a given pair of genes

that are both regulated over a set of samples corresponding to independent transcription profile experiments, the correlation was given by $\frac{n_{++}+n_{--}-(n_{+-}+n_{-+})}{n_{++}+n_{--}+n_{+-}+n_{-+}}$, where n are the sample numbers and the +/- indices refer to the gene's up/down status. The significance was measured by an exact Fisher test.

Querying dGCR returns a list of genes ranked based on the summed log-odds correlation scores, $\sum \log(p)$, across either the full set of platforms, in the default mode, or the platforms corresponding to the given choice of species or technology. The output also lists the measure of correlation and the corresponding significance for the individual platforms.

The dGCR database consists of gene pair co-regulation data for separate platforms. The internal consistency of the co-regulation matrices derived from multiple platforms was addressed with a simple enrichment analysis scoring the likelihood of shared gene pairs in the top slice of co-regulation significance. There is a high degree of overlap between the platforms forming the database. Details of the overlap between the top scoring gene pair sets corresponding to the different platforms are shown in Supplementary Material: Table S1.

Gene cluster modules were generated by first selecting co-regulated gene pairs with summed significance scores across the platforms of below -100, with the co-regulation being significant in at least 5 platforms. This resulted in 14,535 genes with 3.1×10^6 co-regulation associations i.e. a network with 14,535 nodes and 3.1×10^6 edges. Gene pairs were then ranked based on the similarity of the respective edges, measured by a Tanimoto score. Specifically, the similarity score is

$T_{nm} = \frac{\sum_l e_{nl}e_{ml}}{\sum_l e_{nl} + \sum_l e_{ml} - \sum_l e_{nl}e_{ml}}$, where n, m, l are the nodes and the edges are $e_{nm} = 0, 1$. If edges have the same score then the one with nodes having the most edges is ranked higher. The clustering proceeds by seeding the first cluster with the highest ranked edge. The next edge is added to this cluster if there is a shared node otherwise a new cluster is seeded with this edge. If an edge connects two clusters then these clusters are merged. That is, clusters are defined hierarchically. This results in all genes eventually ending up in the same cluster. However, by restricting the merger of clusters each gene can be assigned to a distinct module and genes with otherwise weak connections may be assigned function by virtue of their association with a module. One way of doing this is to ignore edges linking clusters above a given threshold size. When clusters with more than 10 nodes are not merged 470 distinct modules emerge, ranging in size

from 202 to 2 genes. These modules are given in the Supplementary Material: Table S2. The dGCR output page has a link to the module to which the query belongs.

Gene set enrichment is measured by the likelihood relative to random mixing with a fisher test. The GO terms used were populated with genes from all evidence categories and taken from the three headings: biological process, cellular component or molecular function. The pathway gene sets correspond to the MsigDB canonical pathway sets and number 1,320 (www.broadinstitute.org/gsea/msigdb/collections.jsp) [21]. The gene ontology sets are from the gene ontology project hosted at www.geneontology.org [22]. These consist of 9335 GO term sets.

Supplementary Material

Table S1: The overlap between co-regulation matrices from the various platforms used for the dGCR database.

Table S2: Gene lists for the 470 modules based on restricted hierarchical clustering.

<http://www.jgenomics.com/v03p0029s1.xls>

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Availability of dGCR

dGCR is available at www.dGCR.org.

Authors' contributions

GW is the sole author of the present study.

Competing Interests

The authors have declared that no competing interest exists.

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