

Effect of indomethacin on cell cycle proteins in colon cancer cell lines

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Abstract

AIM: To study the effect of indomethacin (IN) on human colon cancer cell line SW480 with p53 mutant and SW480 transfected wild-type p53 (wtp53/SW480) *in vitro* and investigate molecular mechanism of anti-tumor effect of IN on colon cancer.

METHODS: SW480 cells and wtp53/SW480 cells were treated with different concentrations of IN respectively, the expressions of CDK₂, CDK₄ and p21^{WAF1/CIP1} protein were detected by Western blotting.

RESULTS: IN gradually down-regulated the expression of CDK₂, CDK₄ protein of wtp53/SW480 cells in a dose-dependent manner, and inhibitory effect reached the maximum level at 600 μmol/L; IN up-regulated the expression of p21^{WAF1/CIP1} protein in a dose-dependent manner at a certain concentration range, and the expression reached the maximum level at 400 μmol/L, and returned to the base level at 600 μmol/L. The expression of CDK₂, CDK₄ and p21^{WAF1/CIP1} protein of SW480 cells did not change.

CONCLUSION: IN exerts antitumor effect partly through down regulation of the expression of CDK₂, CDK₄ protein and up regulation of the expression of p21^{WAF1/CIP1}.

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Key words: CDK₂; CDK₄; p21^{WAF1/CIP1}; SW480; Colon cancer

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INTRODUCTION

Nonsteroidal anti-inflammatory drugs (NSAIDs) have been shown to reduce the incidence and mortality from colorectal cancer^[1,2]. Indomethacin (IN), a NSAID, is related with the decrease of incidence rate and mortality rate of colon cancer when administered on a long-term basis. Previous studies demonstrated that IN inhibit growth and metastasis of tumor by non-selectively inhibiting cyclooxygenase (COX)^[3-5]. It is found in recent years that inhibiting cell proliferation and inducing cell apoptosis is another mechanism by which IN inhibit tumor^[6-8]. In this study, Western blot analysis was employed to examine the effect of IN on the expression of cyclin-dependent kinase (CDK) CDK₂, CDK₄ and cyclin-dependent kinase inhibitor (CKI) p21^{WAF1/CIP1} in wild-type p53 transfected colon cancer cell line SW480 (wtp53/SW480), so as to provide experimental evidence to elucidate the anti-tumor mechanism of IN.

MATERIALS AND METHODS

Reagents

p53-Mutated human colon cancer SW480 cell line was purchased from ATCC. wtp53/SW480 was SW480 transfected with wild-type p53 allele according to Bampoe and Shetty ascribed previously^[9,10], wtp53/PLXSN recombinant plasmid was kindly provided by Dr. Zhou Xiao, wtp53/PLXSN plasmid was transfected into SW480 cells. Eight-hundred milligrams per liter of G418 was added 48 h after transfection and kept at 200 mg/L concentration for G418 resistance screening. G418-resistant clones of wtp53/SW480 cells were selected randomly. The cells were amplified in media with 200 mg/L G418. The blank plasmid PLXSN was transfected into SW480 cells for the control. Total RNA was extracted from wtp53/SW480 cells, SW480 cells and blank plasmid transfected SW480 cells using Trizol isolation kit according to the manufacturer's instructions. cDNA was synthesized from total RNA in 20 μL reaction mixture using a RT-PCR kit (GIBICO-BRL), including 25 mmol/L MgCl₂ 24 μL, 10× RT buffer 2 μL, 10 mmol/L dNTP 2 μL, Rnasin 40 U, AMV 200 U, Oligo(dT) 151 μL. The mixture was incubated at 42 °C for 60 min, 95 °C for 5 min. Fifty microliters of PCR reaction mixture contained 5 μL product of the reverse transcription as a template, 10×PCR buffer 5 μL, sense and anti-sense primers 50 pmol, 10 mmol dNTP 2 μL, Taq DNA polymerase 2 U. The PCR condition was 94 °C for 3 min pre-denaturing, then 30 cycles at 94 °C for 50 s, 53 °C for 30 s, and 72 °C for 40 s, and further extension at 72 °C for 7 min, PCR products were resolved on 1% agarose gels and visualized by ethidium

bromide staining. RPMI-1640 was purchased from GIBICO-BRL, IN was dissolved in DMSO (Sigma) to make 200 mmol/L stock solution and kept at -20°C . Rabbit anti-CDK₂, CDK₄, and p21^{WAF1/CIP1} polyclonal antibodies were purchased from Santa Cruz; anti- β -actin antibody was kindly provided by Dr. Zhi-Ming He from the Cancer Research Institute, Xiangya Medical College, horseradish peroxidase-conjugated goat anti-rabbit and goat anti-mouse second antibodies, NC membranes were from Huamei Biotech Company (Luo Yan, China); BCATM Protein Assay Reagent Kit was from Pierce Chemical Co. (Rockford, IL), ECL chemiluminescence kit was from NENTM life science.

Cell culture

SW480 and wtp53/SW480 were maintained in RPMI-1640 medium containing 10% inactive fetal bovine serum, 50 IU/L penicillin and 50 $\mu\text{g/L}$ streptomycin at 37°C under an atmosphere of 5 mL/L CO₂.

Drug treatment

In experimental group, the cells were treated with IN at final concentration of 100, 200, 400 and 600 $\mu\text{mol/L}$; in the control group, the cells were treated only with medium or DMSO. The cells were harvested in 24 h treatment.

Protein extraction

The cells were washed twice with PBS, followed by 5 000 r/min centrifugation at 4°C for 10 min, lysed with lysis buffer (50 mmol/L Tris-Cl pH 6.8, 1 mmol/L EDTA, 2% SDS, 5 mmol/L DTT, 10 mmol/L PMSF) on ice, denaturation at boiling water, renaturation on ice, disruption with ultrasound for 30 s, 12 000 g centrifugation again for 30 min and discard supernatant. The protein concentration was analyzed with Protein Assay Reagent Kit, and aliquots of cell lysates were stocked at -70°C .

Western blotting

Aliquots of cell lysates containing 80 μg of total proteins were separated by SDS-polyacrylamide gel for 2-3 h, and transferred to nitrocellulose filters. The filters were blocked with PBS-T buffer containing 5% skimmed milk for 4-6 h, incubated with rabbit anti-CDK₂ (1:200), anti-CDK₄ (1:200), anti-p21^{WAF1/CIP1} (1:500) polyclonal antibodies respectively for 1 h, then the filters washed with PBS-T for 4 min \times 15 min and followed by the addition of horseradish peroxidase-linked goat anti-rabbit IgG and ECL visualization of the bands.

For internal control, the same filters were washed with PBS-T for 30 min \times 3 min, followed by incubation with mouse anti- β -actin in 5% skimmed milk (1:10 000 dilution) for 1 h, incubation with horseradish peroxidase-linked goat anti-mouse IgG for 1 h, and ECL visualization of the bands.

RESULTS

Effect of IN on CDK₂, CDK₄ and p21^{WAF1/CIP1} protein expression in SW480 cells

After 24 h of action of IN at different concentration, it was found by Western blot analysis that the protein expression of not only CDK₂ and CDK₄ but also p21^{WAF1/CIP1} were not

obviously changed in a certain range of IN concentration (Figure 1).

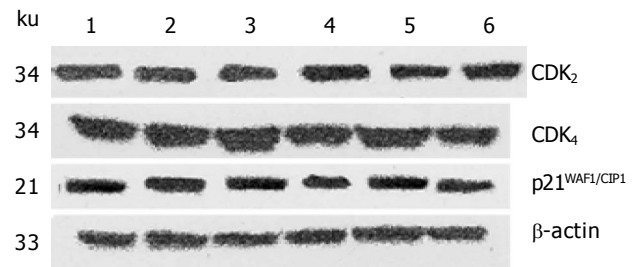


Figure 1 The effect of IN on CDK₂, CDK₄ and p21^{WAF1/CIP1} protein expression in SW480 cells. Lane 1: Control group; lane 2: DMSO group; lane 3: IN 100 $\mu\text{mol/L}$; lane 4: IN 200 $\mu\text{mol/L}$; lane 5: IN 400 $\mu\text{mol/L}$; lane 6: IN 600 $\mu\text{mol/L}$.

Effect of IN on CDK₂, CDK₄ and p21^{WAF1/CIP1} protein expression in wtp53/SW480 cells

After 24 h of action of IN at different concentration, it was found by Western blot analysis that the expression of CDK₂ and CDK₄ was decreased with the increase in IN concentration in certain range. Figure 2 shows that IN can significantly down-regulate the protein expression of CDK₂ and CDK₄ in a dose-dependent manner in a certain range, and the inhibitory effect was most evident at a IN concentration of 400 and 600 $\mu\text{mol/L}$; CDK₄ seems to be more effective. Meanwhile, the protein expression of p21^{WAF1/CIP1} was increased with the augment of IN concentration in a dose-dependent manner in certain range, most evidently at a IN concentration of 400 $\mu\text{mol/L}$, and reduced to basic level at a IN concentration of 600 $\mu\text{mol/L}$.

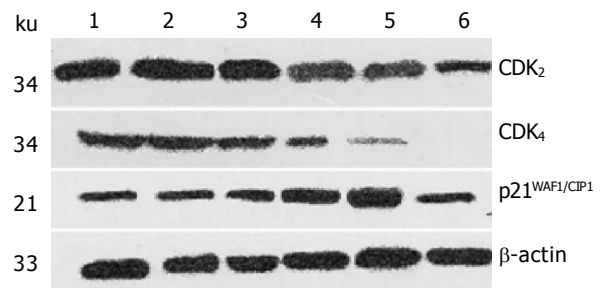


Figure 2 The effect of IN on CDK₂, CDK₄ and p21^{WAF1/CIP1} protein expression in wtp53/SW480 cells. Lane 1: Control group; lane 2: DMSO group; lane 3: IN 100 $\mu\text{mol/L}$; lane 4: IN 200 $\mu\text{mol/L}$; lane 5: IN 400 $\mu\text{mol/L}$; lane 6: IN 600 $\mu\text{mol/L}$.

DISCUSSION

Inhibition of tumor cells proliferation and induction of tumor cells apoptosis are associated with the blocking of the programmed progression of cell cycle^[11-13]. NSAIDs inhibit proliferation and induce apoptosis in human colorectal cancer cells *in vitro*^[14]. Our previous studies have

shown that IN inhibit proliferation of colon cancer cells by increasing the ratio of cells in G₀/G₁ phase in tumor cells, reducing the ratio of cells in S phase and G₂/M phase and thus lead to cell apoptosis, western blot analysis was employed to investigate the effect of IN on colon cancer cell line HCT116 contained with wild-type p53 and found it could down-regulate the expression of CDK₂ and CDK₄, and up-regulate p21^{WAF1/CIP1} expression.

For its instability in solution and a short half-life of only few minutes, p53 protein is difficult to be detected by western blot. In this study, Western blot analysis was employed to detect the expression of CDK₂, CDK₄ and p21^{WAF1/CIP1} protein. To eliminate the error caused by the volume of protein, β-actin protein was used as control, for its stable expression and abundant content in the same class of cells which could ensure the reliability of objective protein and β-actin protein.

The results of the study showed that: IN down-regulated the expression of CDK₂ and CDK₄ protein, especially caused dramatic decreases in Cdk₄ activities, which leads to a G₁ phase cell cycle arrest, in the meantime, up-regulated p21^{WAF1/CIP1} protein in wtp53/SW480 cells, in a dose-dependent manner within a certain range of concentration; while it had no effect on the expression of CDK₂, CDK₄ and p21^{WAF1/CIP1} protein in SW480 cells, suggesting that the existence of wild-type p53 was required for the down-regulation of CDK₂ and CDK₄ expression by IN^[15]. Therefore, we can speculate that up-regulation of p21^{WAF1/CIP1} protein expression inhibits CDK₂ and CDK₄ expression as well as their activity, p21^{WAF1/CIP1} being an important physiological regulator of Cdk₄ complexes^[16], it increased with the augment of IN concentration, most evidently at 400 μmol/L, and reduced to a basic level at 600 μmol/L, the most effective being 400 μmol/L; the drop of CDK₂ and CDK₄ activity in turn decreases the activity of CDK₂ or CDK₄ complex and induces cell cycle arrest in G₂ phase, indicating the effect of IN on colon cancer cells transfected with wild-type p53, namely wtp53/SW480 cells, being to induce cell apoptosis depending on p53 in a dose-dependent manner. Based on the results of previous studies and current research, we conclude IN induces cell cycle arrest in G₂ phase in a p53-dependent way and inhibits cell proliferation in a p53-p21^{WAF1/CIP1} dependent way.

As it is well known, cell cycle is dominated by a regulatory network. A cell proliferation-regulating system composed of cyclins, CDKs and CKIs, can accelerate the progression of cell cycle and thus promote cell proliferation. The main regulatory point to start cell cycle is in G₁ phase, the core in cell cycle regulation are cyclin and CDK^[17], and the formation of CyclinD/CDK₄ and CyclinE/CDK₂ is the rate-limiting step for progression of G₁ phase. Activation of CDK₂ and CDK₄ can yield many cell cycle related proteins, thus prompting progression from G₁ phase to S phase^[18,19]; p21^{WAF1/CIP1}, a major member of CKIs family, which is closely related with growth and differentiation of tumor cells, is an extensive inhibitor for CDK, it can bind and block Cyclin/CDK complex and thus inhibit the activity of CDK₂ and CDK₄, stop the progression of cell cycle and induce G₂ phase arrest^[11]. Wild-type p53 and p21^{WAF1/CIP1}, which are functionally related, are the inhibitors for CDKs-

cyclin complex^[20,21], so abnormality in the p53 or p21^{WAF1/CIP1} gene will lead to abnormal cell proliferation^[22-24]. There is a high frequency of p53 mutation in colon cancer, and mutant p53 can neither stop cell growth nor induce cell apoptosis, therefore, colon cancer is resistant to conventional chemotherapy drugs and not sensitive to assistant treatment of DNA damage. Transfection of wild-type p53 together with chemotherapy and radiotherapy can evidently induce apoptosis, reduce the dosage of chemotherapy and radiotherapy and decrease side effects^[25,26]. Thus, it is held by some that for many tumors with p53 mutation or deletion which has a very poor sensitivity to radiotherapy and chemotherapy, gene transfection should be considered to enhance the sensitivity^[27-29]. So the tumor-suppressing protein p53 mediate chemotherapy drugs induced apoptosis through the regulatory effect of p21 protein on cell cycle progression^[30,31]. *In vitro* experiments have proved that IN inhibits cell proliferation, alters cell cycle and arrests G₁ phase possibly through down-regulating CDK₂ and CDK₄ protein expression and up-regulating p21^{WAF1/CIP1} protein, giving elementary experimental evidence to elucidate the mechanism by which IN inhibits colon cancer, but much needs to be done to elucidate the anti-tumor mechanism of IN in detail^[32] and on a network basis.

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