

## Case Report

# Clinical and molecular effect on offspring of a marriage of consanguineous spinocerebellar ataxia type 7 mutation carriers: a family case report

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**Abstract:** Spinocerebellar ataxia type 7 (SCA7) is a genetic disorder characterized by degeneration of the cerebellum, brainstem, and retina that is caused by abnormal expansion of a CAG repeat located in the *ATXN7* gene encoding sequence on chromosome 3p21.1. Although SCA7 is an uncommon autosomal dominant ataxia, we previously found increased prevalence of the disease in a Southeastern Mexican population. In this study, we described to our knowledge for the first time a marriage of consanguineous SCA7 mutation carriers and their offspring effect. We characterized a severely affected infantile-onset female patient whose parents and two siblings exhibited no symptoms of the disease at time of diagnosis. A comprehensive clinical analysis of the proband showed a progressive cerebellar syndrome, including gait ataxia, movement disorders, and saccadic movements, as well as hyperreflexia, visual deterioration, urinary and cardiovascular dysfunction, and impaired nerve conduction. The SCA7 mutation was detected in the proband patient. Subsequently, genetic examination using four *ATXN7* gene-linked markers (three centromeric microsatellite markers [D3S1228, D3S1287, and D3S3635] and an intragenic Single Nucleotide Polymorphism [SNP-3145G/A]) revealed that the proband descends from a couple of consanguineous SCA7 mutation carriers. Genotyping analysis demonstrated that all offspring inherited only one mutant allele, and that the severe infantile-onset phenotype is caused by germinal expansion (from 37 to 72 CAG repeats) of the paternal mutant allele. Interestingly, the couple also referred a miscarriage. Finally, we found no CAA interruptions in the *ATXN7* gene CAG repeats tract in this family, which might explain, at least in part, the triplet instability in the proband.

**Keywords:** CAG repeats, CAA interruptions, consanguineous marriage, electrophysiological findings, infantile-onset phenotype, intergenerational transmission, spinocerebellar ataxia type 7

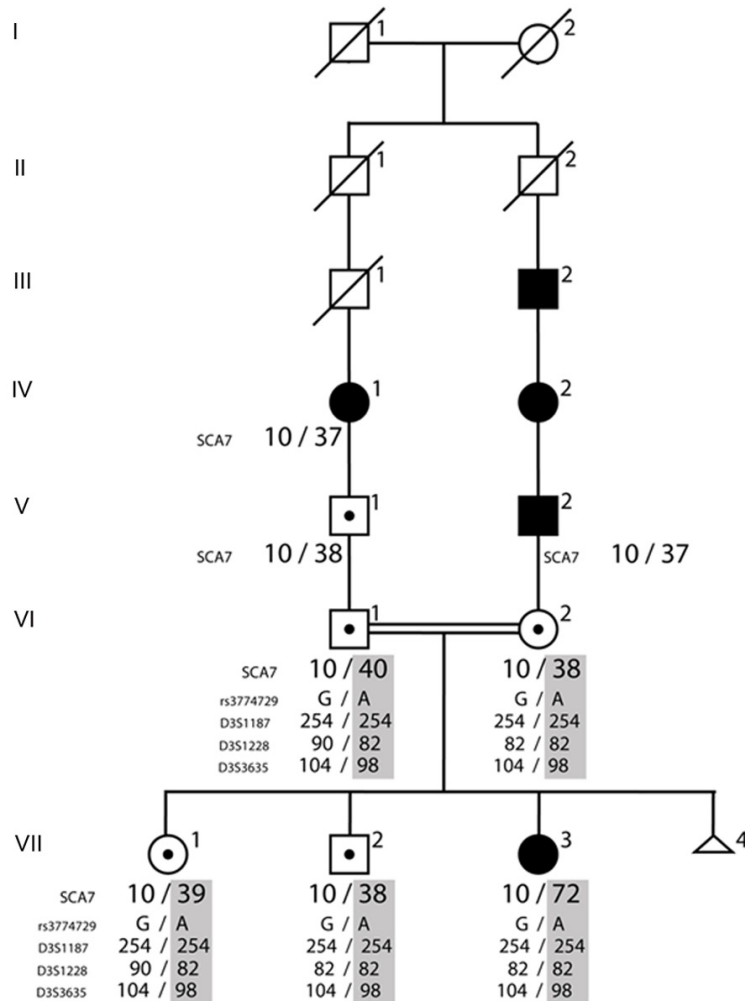
## Introduction

Autosomal dominant spinocerebellar ataxias (SCA) conform a group of rare and heterogeneous hereditary disorders with distinctive clinical characteristics, including gait ataxia, cerebellar dysarthria, dysmetria, adiadochokinesia, and postural tremor [1]. To date, at least 36 subtypes of SCA are known, and in eight of these (SCA1, 2, 3, 6, 7, 12, and 17, and Dentato-rubro-pallido-Luysian atrophy), the abnormal expansion of Cytosine-Adenine-Guanine (CAG) trinucleotide repeats were located in the coding

region of their respective genes underlying the disease [2, 3]. Expanded CAG repeats result in an elongated polyglutamine tract that interferes with normal protein function, as well as with other cellular processes.

In SCA7, one of the rarest forms of SCA [4, 5], the polymorphic tract of CAG repeats, ranges from 4-18 repeats in normal population, and from 36 up to 460 in affected individuals [6]. Larger alleles generally correlate with increased disease severity and earlier age-at-onset in successive generations of a given SCA genealogy.

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**Figure 1.** Pedigree of the SCA7 proband's family. Numbers below symbols indicate repeat number at the SCA locus. Filled symbols indicate clinically affected individuals and symbols with a central point mark indicate asymptomatic carriers. Genotyping analysis using four SCA7 gene-linked markers revealed the parental transmission on offspring of a marriage of consanguineous SCA7 mutation carriers. The analysis showed that in the proband (VII-3) and in her brother (VII-2), the D3S1228 marker was linked with their mutant allele from the father.

In our previous study, we reported a large series of patients with SCA7 in a Mexican population, which suggests a founder mutation effect in this particular group [7, 8]. In this study, we identify a severely affected infantile-onset female patient in this population whose parents and siblings exhibited no symptoms of the disease. We perform a detailed clinical and genetic analysis of this family and determined that the proband descends from consanguineous parents who are both carriers of the SCA7 mutation. Although all siblings inherited the mutation, the infantile-onset phenotype of the proband occurs on paternal transmission of the mutation

through large expansion of the CAG repeat, while the patient's two asymptomatic siblings inherited the mutant allele with no further expansion.

### Material and methods

#### Clinical evaluation

From a sample of 200 subjects recruited from the Tlaltetela community in the Mexican state of Veracruz, a population with high prevalence of SCA7 [7], we identified the proband's family. Neurological examination was carried out following Mayo Clinic procedures [9]. Ataxia-associated symptoms were assessed using the Scale and Rating of Ataxia (SARA) [10], while extracerebellar features were evaluated by the Inventory of Non-Ataxia Symptoms (INAS) [11]. Autonomic nervous system function was examined using the Scales for Outcomes in PD-Autonomic (SCOPA-AUT) [12]. Ophthalmological examination of patients included ocular fundoscopic and color vision tests; the latter was carried out employing the Ishihara pseudo-isochromatic plate. Signed informed consent was obtained from all studied subjects and the research protocol was approved by the National Institute of Rehabilitation (INR) Ethics and Investigation Committee.

#### Electrophysiological recording

Electrophysiological evaluation was performed as previously noted [13], including peripheral motor and sensitive nerve conduction studies, multimodal evoked potentials, and conventional electro-encephalographic recording.

#### Genotyping and sequencing of ATXN7 gene CAG repeats

Genomic DNA was extracted from peripheral blood leukocytes utilizing the Gentra Puregene blood kit (Qiagen, Hilden, NRW, Germany) and then amplified by fluorescent multiplex PCR [7, 14] on an Applied Biosystems thermal cycler

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(Applied Biosystems, Foster City, CA, USA), using specific oligonucleotides for amplification of the polymorphic CAG repeat tract on the *ATXN7* gene [7] and the Taq DNA polymerase enzyme (Roche Diagnostics GmbH, Mannheim, BW, Germany). Aliquots of PCR reactions were mixed with deionized formamide and the internal size standard (ABI GeneScan-500 TAMRA), heated, cooled on ice, and then electrophoresed at 15 kV through capillaries on the ABI PRISM 310 Genetic Analyzer (Applied Biosystems, CA, USA). Homozygous alleles were sequenced using the BigDye Terminator Cycle Sequencing Kit (Applied Biosystems), and analyzed on the ABI PRISM 310 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA). Heterozygous alleles were analyzed by PCR-based amplification, and then PCR products were purified with the QIAquick PCR purification Kit (Qiagen) and sequenced as mentioned previously.

### *Identification of parental transmission*

Four *ATXN7* gene-linked markers (three centromeric microsatellite markers [D3S1228, D3S1287, and D3S3635] and an intragenic Single Nucleotide Polymorphism [SNP-3145G/A]) [8, 15] were used for parental transmission analysis. Microsatellite markers were amplified by PCR and analyzed on the ABI PRISM 310 sequencer (Applied Biosystems). The SNP marker was analyzed using the 5' exonuclease-based real-time PCR assay on the StepOne™ thermal cycler (Applied Biosystems).

## Results

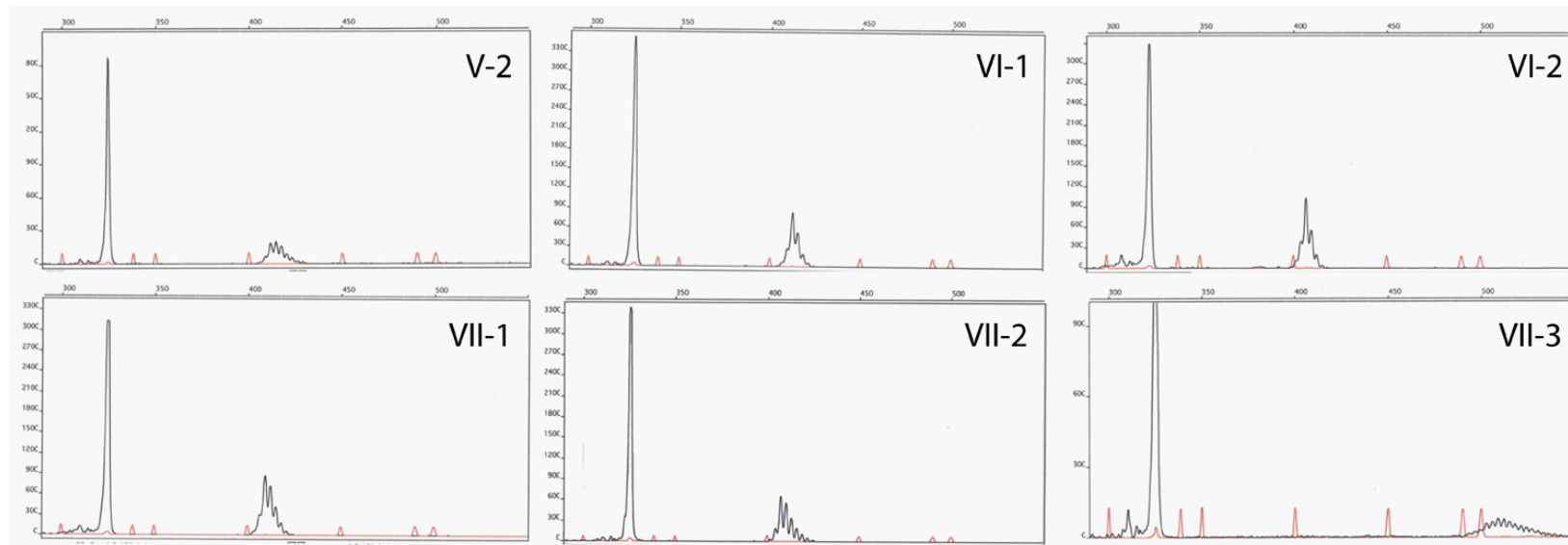
### *Family history and clinical features*

The proband was an 11 year-old-female (**Figure 1**, VII-3) who began with visual impairment at the age of 6 years, and with gait instability and incoordination in the upper limbs some months later. Progression of clinical manifestations confined the patient to a wheelchair at 9 years of age. Neurological examination demonstrated progressive cerebellar syndrome characterized by severe dysmetria and bilateral dysdiadochokinesis; the patient's poor coordination impeded application of the heel-to-shin test. In addition, the proband had truncal ataxia, postural and intention tremor, hypotonia, and her speech was unintelligible the majority of the time, while SARA score was 39, implying cerebellar

damage. Furthermore, the patient showed severe muscular atrophy, Babinsky signs, and prominent hyperreflexia in her upper and lower limbs, including crossed-supraclavicular, pectoral, and hip adductor. Other clinical features include significant cognitive disturbances, choreiform movements, as well as myokymia with an INAS score of 9. Sleep disturbances were frequent, with complaints that included insomnia, muscle cramps, and Restless legs syndrome (RLS). Ophthalmological examination revealed null visual acuity (no light perception), which impeded the patient from performing the Ishihara color blindness test. Fundoscopy revealed bilateral optic disk pallor and pigmentary changes in macular and peripheral retinal regions. The patient also exhibited oculomotor defects, including slowing saccadic movements and limited ocular movements (upward, horizontally, and downward). The Scales for Outcomes in Parkinson's disease-Autonomic (SCOPA-AUT) score demonstrated urinary and cardiovascular dysfunction as well as constipation and hypohidrosis. Neurophysiological assessment revealed significant reduction of distal and proximal motor amplitudes and prolonged latency for median and peroneal nerves. Sensory nerve conduction analysis showed no response for median and sural nerves, while Brainstem auditory evoked potential (BSAEP) examination revealed abnormal morphology and reproducibility of the potentials, which rendered difficult detection of the different components, while Visual evoked potentials (VEP) detected no bioelectric responses. Likewise, Somatosensory evoked potentials for the Tibial nerves (Tn-SSEP) were absent. Electroencephalogram analysis revealed a significant slow diffuse background with decreased occipital dominant rhythm. Finally, brain Magnetic resonance imaging (MRI) showed severe atrophy in cerebellum and brainstem, and mild cerebral atrophy.

With respect to the proband's siblings, her 12-year-old brother and 16-year old sister (**Figure 1**, VII-2 and VII-1, respectively) they did not exhibit any disease symptom, nor neurological signs of cerebellar dysfunction or ophthalmological defects, which was consistent with normal scores for SARA and INAS. Similarly, the proband's parents referred no disease-associated symptoms. Neurological examination of the proband's 36-year-old father (**Figure 1**, VI-1) revealed solely hyperreflexia, and a 20/25 visual acuity with no injury in macula and reti-

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**Figure 2.** Genotyping of CAG repeats at the ATXN7 gene by fluorescent PCR. Analysis of CAG repeats was carried by conventional PCR and the amplified products were separated by capillary electrophoresis. Electropherograms of SCA7 heterozygous patients carrying a normal allele of 10 CAG repeats and a mutant allele of diverse CAG repeats are shown (V-2: 37 CAG repeats; VI-1: 40 CAG repeats; VI-2: 38 CAG repeats; VII-1: 39 CAG repeats; VII-2: 38 CAG repeats; and VII-3: 72 CAG repeats).

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na. However, he exhibited early partial color blindness (> 3 errors) in response to the Ishihara test; analysis of paternal family history identified two members with confirmed diagnosis of SCA7 (**Figure 2**, V-1 and IV-1). The 35-year-old proband's mother (**Figure 1**, VI-2) exhibited no visual or motor symptoms; visual acuity and integrity of macula and retina were normal. Furthermore, the proband's mother's scores for SARA and INAS were normal. She referred a miscarriage in her last pregnancy (**Figure 1**, VII-4). The maternal family history revealed that her father (**Figure 1**, V-2) initiated with visual impairment and gait ataxia at age 54 years and developed the majority of SCA7-associated symptoms over time. Currently, the proband's maternal grandfather is 64 years of age and his symptomatology includes dysarthria, dysmetria of the extremities, dysdiadochokinesia, postural and intention tremor, hyperreflexia (crossed-supraclavicular, pectoral, and hip adductor and slowing saccadic movements), ophthalmoparesis, and ophthalmoplegia. Furthermore, ophthalmologic appraisal revealed decreased visual acuity (20/100) and granular maculopathy, while the Ishihara test demonstrated total color blindness. SARA (9) and SCOPA-AUT (20) scores implied mild cerebellar involvement and altered autonomic signs, respectively. While INAS score (2) did not show a greater involvement. In addition, this subject's numerous siblings presented characteristics compatible with SCA7.

### *Molecular diagnosis and analysis of parental transmission*

Molecular diagnosis performed on the SCA7 genealogy established that all members carry one mutant allele. **Figure 2** depicts electropherograms of each family member. The proband bears a normal allele with 10 CAG repeats and an expanded allele with 72 CAG repeats (**Figure 1**, VII-3). This mutant allele, which represents the largest expansion detected to date in this population, resulted in infantile-onset phenotype. The proband's father carries a 40-CAG repeat mutant allele, while her mother bears a 38-repeat allele (**Figure 1**, VI-1 and VI-2, respectively). The proband's siblings, VII-1 and VII-2, carry expanded alleles of 39 and 38 CAG repeats, respectively, while the proband's maternal grandfather (V-2) carries a 37-repeats mutant allele. Therefore, all asymptomatic members of the family are indeed carriers of the mutation.

Although the proband's parents denied that they were blood relatives, the fact that both carry the SCA7 mutation strongly suggests a consanguineous relationship between them. To approach this hypothesis, we performed a comprehensive familiar history evaluation and a linkage analysis using four *ATXN7* gene-linked markers: rs3774729; D3S1187; D3S1228, and D3S3635 [15]. **Figure 1** depicts haplotypes for each family member; alleles were named as previously noted [8]. We established that the proband's parents are indeed blood relatives and share a common ancestor five generation back (**Figure 1**). *ATXN7* gene-linked markers were also employed to analyze parent-offspring transmission of mutant alleles. We found that D3S1228 was the sole informative marker in this genealogy (**Figure 1**). In the proband and in her brother (VII-2), the D3S1228-82 allele, which is linked with the wild type *ATXN7* allele, were inherited from the mother; thus, their mutant allele must come from the father. Contrariwise, in the proband's sister (VII-1) the D3S1228-90 allele, which is linked with the wild-type *ATXN7* allele, comes from the father; therefore the mutant allele must be inherited from the mother. Overall, our result showed that paternal transmission caused expansion of the mutant allele and ultimately, early-onset and severe disease symptomatology exclusively in the proband, implying the existence of SCA7-modulating factors in this family.

Because the presence of CAA interruptions within the CAG repeats tract could modulate triplet stability in various polyglutamine diseases [16], we searched for such interruptions in the *ATXN7* gene of five family members with SCA7 and 30 normal subjects from Mexican general population. None of the normal or mutant *ATXN7* alleles contained CAA interruptions.

### **Discussion**

In this study, we described an infantile-onset female patient with severe SCA7 symptomatology, including, gait ataxia, cerebellar dysarthria, severe visual impairment, hyperreflexia, muscular atrophy, oculomotor defects, and movement disorders (chorea and myokymia). Electrophysiological and imaging findings were consistent with impaired nerve conduction and extensive and severe degeneration of the majority of the structures of the peripheral and central nervous system, including the autonomic nervous system.



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SCA7 is generally detected in adult patients with a family history consistent with autosomal dominant inheritance [17]. However, this patient with infantile disease onset had no familial history related with the disease at time of diagnosis, and detailed clinical evaluation uncovered no ataxic symptoms in the family's relatives, with the exception of hyperreflexia in her father. Genotyping in the SCA7 family solved the enigma; we identified an uncommonly large mutant allele (72 CAG repeats) in the infantile-onset patient and unexpectedly, the presence of one mutant allele in all family members. The low worldwide prevalence of SCA7 (< 1:100,000) renders the random occurrence of the *ATXN7* gene mutation in both parents practically impossible, raising the possibility of a consanguineous relationship between them. Genealogy and linkage analysis utilizing different *ATXN7* gene-linked polymorphic markers confirmed the notion that the offspring was the result of an unaware consanguineous marriage of a couple of SCA7 mutation carriers, with the identification of a common ancestor five generation back by family history.

It is noteworthy that the proband suffered a dramatic expansion of the mutant allele, from 40 to 72 repeats. Such germinal instability generally results in earlier age-at-onset and more severe symptomatology in the offspring of affected parents [18, 19], with alleles bearing 55-460 repeats giving rise to the infantile-onset phenotype and a rapid and fatal course of the disease [20, 21]. This background is consistent with the severe clinical presentation of the proband, as described above. It is thought that the extent of germinal expansions is influenced by the gender of transmitting parent, with larger expansions occurring through paternal transmission [22-25]. Intriguingly, although the proband and her brother inherited the paternal mutant allele from the father, the extent of the CAG repeats expansion and consequently, the SCA7 symptomatology, were markedly different in each one from each other. This implies that in addition to gender, there are other unknown factors that determine the germinal stability/expansion of CAG triplets in SCA7 (i.e., triplet stability-modifying genes, epigenetic factors, or somatic mosaicism). In various polyglutamine diseases, the presence of CAA interruptions could improve the stability of the CAG repeat tract [16]. However, we found no CAA

interruptions in the *ATXN7*-gene CAG repeat tract in any of the normal and mutant alleles analyzed, which is consistent with a previous report [24]. Finally, the couple of SCA7 carriers referred a miscarriage (**Figure 1**, VII-4) which might be caused by the presence of a homozygous mutation. Many authors have postulated that a homozygous mutation could be incompatible with life. Nevertheless, some homozygous cases have been reported in polyglutamine diseases [26, 27]. With only few reports of homozygotes worldwide, the controversy of gene dosage effect cannot be concluded. Other rational explanation for the miscarriage could be the presence of a largest abnormal CAG expansion; Monckton et al, revealed extraordinarily CAG repeat instability in male germline, and suggested that largest expansions might be associated with embryonic lethality or dysfunctional sperm [25].

This study reflects the importance of the molecular diagnostic. In many late onset neurodegenerative diseases it is frequent that the couples have already satisfied their reproductive needs by the time when the diagnosis of the condition is made. Therefore, reproductive options like prenatal testing or preimplantation genetic diagnosis cannot be offered to them. It highlights the importance of developing programs for pre-symptomatic diagnosis of these diseases, particularly in populations with a founder effect and an unusually elevated frequency of pre-symptomatic carriers. Genetic counseling is a valuable tool not only for couples wishing to have healthy descendents, but also for the rest of the family whom may be unaware of their own risk.

In summary, we presented a detailed clinical and genetic characterization of an infantile-onset patient with SCA7. We determined that the offspring is the result of a consanguineous marriage of a couple of SCA7 gene-mutation carriers and that the severe infantile-onset phenotype is caused by a germinal expansion of the paternal mutant allele.

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patients and members of SCA-affected families.

### Disclosure of conflict of interest

None.

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### References

- [1] Durr A. Autosomal dominant cerebellar ataxias: polyglutamine expansions and beyond. *Lancet Neurol* 2019; 9: 885-894.
- [2] Matilla-Dueñas A, Corral-Juan M, Volpini V, Sánchez I. The spinocerebellar ataxias: clinical aspects and molecular genetics. *AdvExp Med Biol* 2012; 724: 351-374.
- [3] Magaña JJ, Velázquez-Pérez L, Cisneros B. Spinocerebellar ataxia type 2: clinical presentation, molecular mechanisms, and therapeutic perspectives. *Mol Neurobiol* 2013; 47: 90-104.
- [4] David G, Abbas N, Stevanin G, Dürr A, Yvert G, Cancel G, Weber C, Imbert G, Saudou F, Antoniou E, Drabkin H, Gemmill R, Giunti P, Benomar A, Wood N, Ruberg M, Agid Y, Mandel JL, Brice A. Cloning of the SCA7 gene reveals a highly unstable CAG repeat expansion. *Nat Genet* 1997; 17: 65-70.
- [5] Moseley ML, Benzow KA, Schut LJ, Bird TD, Gomez CM, Barkhaus PE, Blindauer KA, Labuda M, Pandolfo M, Koob MD, Ranum LP. Incidence of dominant spinocerebellar and Friedreich triplet repeats among 361 families. *Neurology* 1998; 51: 1666-16671.
- [6] Garden GA, La Spada AR. Molecular pathogenesis and cellular pathology of spinocerebellar ataxia type 7 neurodegeneration. *Cerebellum* 2008; 7: 138-149.
- [7] Magaña JJ, Tapia-Guerrero YS, Velázquez-Pérez L, Cerecedo-Zapata C, Maldonado-Rodríguez M, Jano-Ito J, Leyva-García N, González-Piña R, Martínez-Cruz E, Hernández-Hernández O, Cisneros B. Analysis of CAG repeats in five SCA loci in Mexican population: epidemiological evidence of a SCA7 founder effect. *Clin Genet* 2014; 85: 159-165.
- [8] Magaña JJ, Gómez R, Maldonado-Rodríguez M, Velázquez-Pérez L, Tapia-Guerrero YS, Cortés H, Leyva-García N, Hernández-Hernández O, Cisneros B. Origin of spinocerebellar ataxia type 7 gene mutation in Mexican population. *Cerebellum* 2013; 12: 902-905.
- [9] Denny-Brown D, David MD, Tyler HR. *Handbook of Neurological Examination and Case Recording*. Cambridge, MA, USA: Harvard University Press; 1982.
- [10] Schmitz-Hubsch T, du Montcel ST, Baliko L, Berciano J, Boesch S, Depondt C, Giunti P, Globas C, Infante J, Kang JS, Kremer B, Mariotti C, Melegh B, Pandolfo M, Rakowicz M, Ribai P, Rola R, Schöls L, Szymanski S, van de Warrenburg BP, Dürr A, Klockgether T, Fancellu R. Scale for the assessment and rating of ataxia: development of a new clinical scale. *Neurology* 2006; 66: 1717-1720.
- [11] Schmitz-Hubsch T, Coudert M, Bauer P, Giunti P, Globas C, Baliko L, Filla A, Mariotti C, Rakowicz M, Charles P, Ribai P, Szymanski S, Infante J, van de Warrenburg BP, Dürr A, Timmann D, Boesch S, Fancellu R, Rola R, Depondt C, Schöls L, Zdienicka E, Kang JS, Döhlinger S, Kremer B, Stephenson DA, Melegh B, Pandolfo M, di Donato S, du Montcel ST, Klockgether T. Spinocerebellar ataxia types 1, 2, 3, and 6: disease severity and nonataxia symptoms. *Neurology* 2008; 71: 982-989.
- [12] Visser M, Marinus J, Stiggelbout AM, Van Hilten JJ. Assessment of autonomic dysfunction in Parkinson's disease: the SCOPA-AUT. *Movement Disord* 2004; 11: 1306-1312.
- [13] Velázquez-Pérez L, Sánchez-Cruz G, Canales-Ochoa N, Rodríguez-Labrada R, Rodríguez-Díaz J, Almaguer-Mederos L, Laffita Mesa J. Electrophysiological features in patients and presymptomatic relatives with spinocerebellar ataxia type 2. *J Neurol Sci* 2007; 263: 158-164.
- [14] Dorschner MO, Barden D, Stephens K. Diagnosis of five spinocerebellar ataxia disorders by multiplex amplification and capillary electrophoresis. *J Mol Diagn* 2002; 4: 108-113.
- [15] Stevanin G, David G, Dürr A, Giunti P, Benomar A, Abada-Bendib M, Lee MS, Agid Y, Brice A. Multiple origins of the spinocerebellar ataxia 7 (SCA7) mutation revealed by linkage disequilibrium studies with closely flanking markers, including an intragenic polymorphism (G314-5TG/A3145TG). *Eur J Hum Genet* 1999; 7: 889-896.
- [16] Laffita-Mesa JM, Velázquez-Pérez LC, Santos-Falcón N, Cruz-Mariño T, González-Zaldívar Y, Vázquez-Mojena Y, Almaguer-Gotay D, Almaguer Mederos LE, Rodríguez Labrada R. Unexpanded and intermediate CAG polymorphisms

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- at the SCA2 locus (ATXN2) in the Cuban population: evidence about the origin of expanded SCA2 alleles. *Eur J Hum Genet* 2012; 20: 41-49.
- [17] Horton LC, Frosch MP, Vangel MG, Weigel-Di-Franco C, Berson EL, Schmahmann JD. Spinocerebellar ataxia type 7: clinical course, phenotype-genotype correlations and neuropathology. *Cerebellum* 2013; 12: 176-93.
- [18] Mao R, Aylsworth AS, Potter N, Wilson WG, Breningstall G, Wick MJ, Babovic-Vuksanovic D, Nance M, Patterson MC, Gomez CM, Snow K. Childhood onset ataxia: testing for large CAG repeats in SCA2 and SCA7. *Am J Med Genet* 2002; 110: 338-345.
- [19] Cagnoli C, Stevanin G, Michielotto C, Gerbino-Promis G, Brussino A, Pappi P, Durr A, Dragone E, Viemont M, Gellera C, Brice A, Migone N, Brusco A. Large pathogenic expansions in the SCA2 and SCA7 genes can be detected by fluorescent repeat-primed polymerase chain reaction assay. *J MolDiagn* 2006; 8: 128-132.
- [20] Martin J, Van Regemorter N, Del-Favero J, Lofgren A, Van Broeckhoven C. Spinocerebellar ataxia type 7 (SCA7) - correlations between phenotype and genotype in one large Belgian family. *J Neurol Sci* 1999; 168: 37-46.
- [21] van de Warrenburg BP, Frenken CW, Ausems MG, Kleefstra T, Sinke RJ, Knoers NV, Kremer HP. Striking anticipation in spinocerebellar ataxia type 7: the infantile phenotype. *J Neurol* 2001; 248: 911-914.
- [22] David G, Dürr A, Stevanin G, Cancel G, Abbas N, Benomar A, Belal S, Lebre AS, Abada-Bendib M, Grid D, Holmberg M, Yahyaoui M, Hentati F, Chkili T, Agid Y, Brice A. Molecular and clinical correlations in autosomal dominant cerebellar ataxia with progressive macular dystrophy (SCA7). *Hum Mol Genet* 1998; 7: 165-170.
- [23] Gow LG, Castañeda MA, McKenna CK, Digre KB, Pulst SM, Perlman S, Lee MS, Gomez C, Fischbeck K, Gagnon D, Storey E, Bird T, Jeri FR, Ptáček LJ. Analysis of the dynamic mutation in the SCA7 gene shows marked parental effects on CAG repeat transmission. *Hum Mol Genet* 1998; 7: 525-32.
- [24] Michalik A, Martin JJ, Van Broeckhoven C. Spinocerebellar ataxia type 7 associated with pigmentary retinal dystrophy. *Eur J Hum Genet* 2004; 12: 2-15.
- [25] Monckton DG, Cayuela ML, Gould FK, Brock GJR, de Silva R, Ashisawa T. Very large (CAG)<sub>n</sub> DNA repeat expansions in the sperm of two spinocerebellar ataxia type 7 males. *Hum Mol Genet* 1999; 8: 2473-2478.
- [26] Rudnik-Schöneborn DS, Schneider-Gold C, Raabe U, Kress W, Zerres K, Schooser BG. Outcome and effect of pregnancy in myotonic dystrophy type 2. *Neurology* 2006; 66: 579-580.
- [27] Cruz-Mariño T, Gonzalez-Zaldivar Y, Laffita-Mesa JM, Almaguer-Mederos L, Aguilera-Rodriguez R, Almaguer-Gotay D, Rodriguez-Labrada R, Canales-Ochoa N, Macleod P, Velazquez-Pérez L. Uncommon features in Cuban families affected with Friedreich ataxia. *Neurosci Lett* 2010; 472: 85-89.