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Tumor matrix protein collagen XIa1 in cancer

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Abstract

The extracellular matrix is increasingly recognized as an essential player in cancer development and progression. Collagens are one of the most important components of the extracellular matrix, and have themselves been implicated in many aspects of neoplastic transformation. Collagen XI is a minor collagen whose main physiologic function is to regulate the diameter of major collagen fibrils. The α 1 chain of collagen XI (colXI α 1), has known pathogenic roles in several musculoskeletal disorders. Recent research has highlighted the importance of colXI α 1 in many types of cancer, including its roles in metastasis, angiogenesis, and drug resistance, as well as its potential utility in screening tests and as a therapeutic target. High levels of colXI α 1 overexpression have been reported in multiple expression profile studies examining differences between cancerous and normal tissue, and between beginning and advanced stage cancer. Its expression has been linked to poor progression-free and overall survival. The consistency of this data across cancer types is particularly striking, including colorectal, ovarian, breast, head and neck, lung, and brain cancers. This review discusses the role of collagen XI α 1 in cancer and its potential as a target for cancer therapy.

Keywords

Collagen; extracellular matrix; colXIa1; COL11A1; cancer

Introduction

The extracellular matrix (ECM) is an essential component of the cancer cell niche. The ECM is a complex macromolecular network composed of biochemically distinct elements, including polysaccharides, proteoglycans, proteins, and glycoproteins. It provides structural

Conflict of Interest statement

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None

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support for cells in the form of the basement membrane, a specialized type of matrix essential for many cellular processes. In addition, the ECM forms the interstitial matrix, which is important in structural tissue support as well as in regulating and integrating cell behavior^{1,2}.

The role of the ECM in tumor progression is becoming increasingly clear; a flood of recent research has illustrated that dysregulation of its various components plays an essential role in generating and maintaining the tumor microenvironment^{1,2}. For example, tumor ECM is often stiffer and more highly crosslinked than normal stroma, promoting abnormal cell behavior^{3–5}. Tumor cells are especially sensitive to changes in the stiffness of their environment, specifically increased collagen crosslinking, which then helps to drive the malignant phenotype³. A normal, functional ECM is essential for maintaining cellular architecture and polarity, which is universally lost in neoplastic growth. Abnormal ECM also promotes abnormal behavior in stromal cells, including the fibroblasts, immune cells, and endothelial cells that help make up the tumor microenvironment, and thus contributes to the formation and perpetuation of the neoplastic niche^{6,7}.

One of the most important components of the ECM, and the most abundant protein in the body, is collagen. Currently there are 28 known collagens, which are trimeric molecules consisting of three polypeptide alpha chains (which may or may not be identical) forming a triple helix structure common to all collagens. The collagen family is diverse, and therefore is divided into three subgroups based on molecular structure and supramolecular assemblies: fibrillar collagens, non-fibril forming collagens, and fibril-associated collagens^{8–10}. Fibrillar collagens are the most abundant, and are capable of forming highly ordered fibrils in the ECM. Fibril-associated collagens, containing interrupted triple helices, associate with and help regulate fibrillar collagens. Non-fibril forming collagens, which include type IV collagen found in the basement membrane, do not form or associate with fibrils⁸.

Many previous reports have revealed collagen XI as a player in human disease. Collagen XI is a minor fibrillar collagen most abundantly found in cartilage, but which has also been found in odontoblasts, trabecular bone, skeletal muscle, placenta, lung, and neoepithelium of the brain¹¹. Collagen XI copolymerizes with both collagen II and collagen IX, and is essential in maintaining proper fibril diameter and function in connective tissue; absence or mutation in the alpha chain of collagen XI results in abnormally thickened cartilage fibrils¹². Collagen XI, like all collagens, is a heterotrimer consisting of $\alpha 1$, $\alpha 2$, and $\alpha 3$ chains, located on different chromosomes, which are synthesized as procollagens and proteolytically cleaved to yield mature trimers¹¹. The $\alpha 1$ and $\alpha 2$ chains are genetically distinct, while the $\alpha 3$ chain is a hyperglycosylated form of the $\alpha 1$ chain of collagen II. Mutations in the gene encoding the $\alpha 1$ chain of collagen XI (colXI $\alpha 1$) have been implicated in many musculoskeletal disorders, including Stickler syndrome, characterized by opthalamic, articular, orofacial, and auditory abnormalities^{13,14}; fibrochondrogenesis, a lethal form of dwarfism¹⁵; as well as osteoarthritis¹⁶, lumbar disc herniation¹⁷, limbus vertebra¹⁸, and Achilles tendinopathy¹⁹.

Recent progress has highlighted an important role for collagen XI in many aspects of neoplastic transformation. This review will focus on the role of collagen XIa1 in cancer.

Dysregulation of collagen XIa1 in cancer

Normal, physiologic expression of collagen XI is very low or nonexistent in most tissues^{20–22}. Therefore, changes in collagen XI α 1 (colXI α 1) expression associated with cancer are excellent putative markers both of neoplastic change and disease progression.

ColXIa1 was found to be the most highly overexpressed gene in high-stage (versus lowstage) cancer in a meta-analysis of microarray data from multiple cancers²³. This analysis generated a metastasis-associated gene expression signature of which colXIa1 was the top hit, and which was common to multiple cancers, including ovarian, colon, breast, and lung²³. This metastasis-associated gene expression signature corresponds to a stromal desmoplastic reaction commonly produced by tumor-associated fibroblasts (TAFs). This reaction is frequently present in high stage cancers either in the process of or which have already invaded into the surrounding tissues and/or metastasized to distant sites. Specifically, colXIa1 mRNA expression correlated with cancer staging in this group of cancers, suggesting that it alone can be used as a proxy for the metastasis-associated gene expression signature. Expression of this metastasis-associated gene signature by primary tumors may be an early marker for invasive potential^{23,24}. To validate this data, human neuroblastoma cells were xenografted into immunocompromised mice. The resulting tumors also expressed high levels of colXIa1 as assessed by quantitative PCR, as well as the other epithelial-to-mesenchymal transition (EMT)-associated genes in the signature²⁴. This signature was only expressed by the xenografted human cells, and not the surrounding mouse stroma, suggesting that signals for EMT derive specifically from the tumor²⁴. A subset of murine adjpocyte markers was downregulated in association with the upregulation of EMT signature genes, implicating adipocytes in the development of cancer cell invasion. Similarly, highly migratory human glioma cells were used to assess markers for cell invasion and their epigenetic controls, as compared to non-migratory breast cancer cells. ColXIa1 mRNA was highly upregulated in the glioma cells as compared to the breast cancer cells. This was associated with both hyperacetylation and methylation of specific lysine residues on histone 3, both of which are markers for transcriptional activation 25 . ColXIa1 is also highly expressed at the protein level in human gliomas versus paired normal tissue²⁶.

Colorectal carcinoma

The role of colXIa1 in cancer was first identified in sporadic colorectal cancer, which expresses high levels of colXIa1 mRNA, while there is no expression in normal colonic tissue. In this setting, colXIa1 seems to be co-expressed with collagen Va2, the other minor fibrillar collagen with which collagen XI shares a high degree of sequence homology, and which is also not expressed in normal adult colon²⁰. ColXIa1 is also overexpressed in colon polyps from patients with familial adenomatous polyposis, an inherited mutation in the tumor suppressor APC which causes colorectal carcinoma. The authors suggest that colXIa1 overexpression could be related to the APC/ β -catenin pathway, which is dysregulated not only in familial adenomatous polyposis but also in the majority of sporadic colorectal cancers²¹. This aberrant expression has been linked to a mutation in exon 54 in the colXIa1 gene, discovered by analyzing mutations in exfoliated tumor cells from the stool of

colorectal cancer patients. The mutation may act as a potential non-invasive screening test for colorectal cancer²⁷.

Ovarian carcinoma

In a study looking at gene expression in human epithelial ovarian cancer tissue samples, colXIa1 was the most highly expressed of the studied genes, and expression correlated with stage of disease²⁸. High colXIa1 at both the mRNA and protein level was also associated with cancer recurrence or persistence, as well as lower overall and disease-free survival^{28,29}. ColXIa1 was similarly identified as part of a 10-gene panel from 403 high-grade serous ovarian cancer samples predictive of poor clinical outcome³⁰. Furthermore, colXIa1 expression (both mRNA and protein) differs by tumor site and stage: the lowest levels were expressed in primary ovarian cancer samples, moderate levels in concurrent metastases, and highest levels in recurrent/persistent metastases from the same patient³⁰. ColXIa1 was also the most highly overexpressed gene in a microarray study of metastatic versus primary ovarian serous papillary carcinoma, where it was upregulated by an average of 8.23-fold in omental metastases³¹. ColXI α 1 was identified as one of 12 genes highly expressed in ovarian tumor vasculature, and may serve as a tumor biomarker³². Markers of tumor blood supply may act as important drug targets, following in the footsteps of the VEGF inhibitor bevacizumab, which has been successful as part of a multi-therapy regimen in treating many types of solid tumors.

Breast carcinoma

Multiple studies have found that colXI α 1 is much more highly expressed in invasive ductal carcinoma than its precursor, the premalignant lesion ductal carcinoma in situ, indicating that it may play a role in local invasion of cancer cells^{33–35}. Additionally, increased colXI α 1 expression has been reported in primary breast tumors compared to paired lymph node metastases^{36,37}. Overexpression of colXI α 1, along with other similar extracellular matrix genes, at primary tumor sites suggests that it is important in modulating the ECM to facilitate tumor cell dissemination and therefore metastasis; this is in agreement with the findings from the metastasis-associated gene signature discussed above. ColXI α 1 was also overexpressed in a "high risk" group of breast cancer patients, who were axillary lymph node-positive and had distant metastases within follow up time (43 months), versus those in the "low risk" group, who were also node-positive but had no metastases within this time³⁶. However, another study found that colXI α 1 protein was actually downregulated in primary breast tumors that had metastasized versus those that had not³⁸. The apparent discrepancy may be due to the use of immunohistochemistry to assess protein level, versus the study describing higher expression in primary tumors, which was based on mRNA data.

Aerodigestive tract tumors

Similar to the above findings, in a microarray study of head and neck squamous cell carcinoma (HNSCC), colXIa1 was one of two genes of 12,000 studied found to be highly upregulated (by an average of 476-fold) in all HNSCC samples studied, and had no expression in normal tissue, making it an ideal marker for HNSCC³⁹. This finding was confirmed by another study in which colXIa1 was upregulated by an average of 6.63 fold in HNSCC tumors versus normal adjacent tissue⁴⁰. An additional study demonstrated a 7.5-

fold increase in colXI α 1 mRNA levels in metastatic HNSCC tumors compared to nonmetastatic tumors⁴¹. A genome-wide analysis examining chromosomal alterations occurring in environmental insult-initiated esophageal squamous cell carcinoma found significant amplification of colXI α 1 gene compared to matched normal tissue, which may be used as an early biomarker for esophageal cancer⁴². Single nucleotide polymorphisms in the colXI α 1 gene have been linked to papillary thyroid cancer, with certain alleles conferring a protective effect⁴³.

ColXIa1 is also overexpressed in non-small cell lung cancer, as well as correlating with pathological stage, presence of lymph node metastasis, and poor prognosis⁴⁴. In transitional cell carcinoma of the bladder, colXIa1 is one of seven genes with differential expression between superficial and muscle-invasive tumors⁴⁵. ColXIa1 expression is also capable of differentiating premalignant from malignant stomach cancers⁴⁶. See Table 1 for a summary of colXIa1 dysregulation across cancer types.

ColXIa1 and tumor-associated stroma

TAFs are the most abundant cell type within the stroma of many solid malignancies. ColXI α 1 was found to be more highly expressed by TAFs isolated from HNSCC explants than in normal fibroblasts derived from cancer-free patients⁴⁰. Immunohistochemical studies in pancreatic cancer have also suggested that colXI α 1 may be a marker capable of differentiating TAFs from normal activated fibroblasts, a distinction which has been elusive so far⁴⁷. In ovarian cancer studies, colXI α 1 expression was mainly confined to stromal cells, specifically to the intra/peritumoral stromal cells, and not expressed >1 mm from epithelial tumor cells, indicating again that colXI α 1 is a specific marker for TAFs, and potentially for malignant cells undergoing EMT³⁰. Taken together, this strongly implicates stroma-secreted colXI α 1 in the ECM modulation that is critical in facilitating tumor spread.

ColXIa1 is highly expressed in the stroma of breast cancer, with anti-procolXIa1 antibodies capable of recognizing invasive ductal carcinoma-associated TAFs and therefore differentiating breast malignancies from benign lesions, which have very little or no procolXIa1 expression^{48,49}. Similarly, resection margins from gastric carcinoma patients exhibit high colXIa1 levels⁵⁰. Expression is also limited to stroma in colorectal cancer²⁰. However, in several of the studies discussed above, the source of increased colXIa1 was the epithelial compartment, not the surrounding stroma, suggesting a complex interplay between these two factors in facilitating tumor invasion.

ColXIa1 as a therapeutic target

Although colXIa1 is clearly active in promoting cancer progression and metastasis, descriptive studies, even with human tissue, are limited in terms of functional or mechanistic significance. Thus it is noteworthy that functional studies in various cancer types have confirmed that dysregulation of colXIa1 expression is indeed playing a relevant role in neoplastic progression (Figure 1). ColXIa1 siRNA-mediated knockdown in both ovarian cancer cell lines and HNSCC cell lines significantly suppressed invasion and slowed proliferation. This effect was not observed when normal, non-cancerous cells were transfected with colXIa1 siRNA, further confirming its potential as a therapeutic target^{28,40}.

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In the ovarian cancer cell lines, colXIa1 knockdown was associated with decreased matrix metalloproteinase (MMP) 3 mRNA expression and activity. This effect was mediated through Ets-1, a transcription factor that regulates expression of many matrix-modulating genes, including several MMPs, and which is known to be expressed in many types of cancer. ColXIa1 knockdown decreased Ets-1 binding to the MMP3 promoter, indicating that the effect of colXIa1 on cell invasion may be mediated through MMP3 upregulation²⁸. When $colXIa_1$ -knockdown cells were inoculated into mice, they showed impaired tumor spread and invasion compared to mice injected with cells expressing wild-type colXIa1^{28,30}. One study in a mouse model of pancreatic cancer suggests that colXIa1 may be a downstream target of the Hedgehog signaling pathway, which is activated in most pancreatic cancers. In pancreatic cancer cells, overexpression of Gli1, the major transcription factor activated by the Hedgehog pathway, resulted in increased expression of colXIa151. Studies in pancreatic and ovarian cancer have also suggested that colXIa1 is a downstream target of the TGF- β signaling pathway, which is itself regulated by the ECM and is commonly dysregulated in cancer. TGF-β upregulates colXIa1 expression through transcription factor NF-Y^{28,30,52}. Treatment with TGF- β inhibitors drastically reduced colXI α 1 expression in ovarian cancer cell lines, as well as cell invasion 28 .

ColXIα1 has also been linked with therapeutic resistance, which is a significant problem across cancer types. The stroma is often hypothesized as the source of resistance to many targeted therapies that seem promising in vitro but fail in clinical trials. The colXIα1-driven metastasis-associated gene expression signature discussed above was also associated with resistance to neoadjuvant chemotherapy in breast cancer²³. High levels of colXIα1 protein secretion have been linked with resistance to platinum-based therapies in ovarian cancer²⁹. Therefore, therapeutic strategies targeting collagen XI may help sensitize cells to conventional treatments.

Precisely how colXI α 1 is mediating these effects is unknown. Limited research has identified integrin $\alpha 2\beta$ 1 and the discoidin domain receptor family as putative collagen XI receptors; however, very little is known about downstream signaling^{53,54}. One potential colXI α 1 binding partner is nucleolin, a nuclear transport protein which also has autolytic activity. This activity results in cleavage products capable of post-transcriptionally regulating MMP9, which, like MMP3, may be involved in colXI α 1 would be therapeutically beneficial. Approaches including antisense targeting or modulation of transcription or translation for colXI α 1 may hold the most promise.

Conclusion

The expression pattern of colXIa1 in cancer is obviously complex and incompletely understood. While most studies point to a direct relationship between colXIa1 expression and cancer progression, specifically as it pertains to metastasis, there are some seemingly discordant findings. The expression of colXIa1 changes as the tumor evolves, and differences in study timepoints and the source of the colXIa1, whether it be the tumor itself or the surrounding stroma, may account for large expression differences. A limited number of functional studies confirm that colXIa1 is playing a role in cancer proliferation, invasion,

and metastasis, as well as in resistance to therapeutics. More in-depth studies are needed to determine mechanisms through which $colXI\alpha1$ influences cancer cell behavior. What data is available suggests that collagen XI\alpha1, with its low expression in normal tissue and high expression in cancer, and its mechanistic significance, may be an ideal target for future therapies across multiple cancer types.

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References

- 1. Barkan D, Green JE, Chambers AF. Extracellular matrix: a gatekeeper in the transition from dormancy to metastatic growth. Eur J Cancer. 2010; 46:1181–8. [PubMed: 20304630]
- 2. Hynes RO. The extracellular matrix: not just pretty fibrils. Science. 2009; 326:1216–9. [PubMed: 19965464]
- Levental KR, Yu H, Kass L, et al. Matrix crosslinking forces tumor progression by enhancing integrin signaling. Cell. 2009; 139:891–906. [PubMed: 19931152]
- 4. Lo CM, Wang HB, Dembo M, et al. Cell movement is guided by the rigidity of the substrate. Biophys J. 2000; 79:144–52. [PubMed: 10866943]
- Paszek MJ, Zahir N, Johnson KR, et al. Tensional homeostasis and the malignant phenotype. Cancer Cell. 2005; 8:241–54. [PubMed: 16169468]
- Quante M, Tu SP, Tomita H, et al. Bone marrow-derived myofibroblasts contribute to the mesenchymal stem cell niche and promote tumor growth. Cancer Cell. 2011; 19:257–72. [PubMed: 21316604]
- Orimo A, Gupta PB, Sgroi DC, et al. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. Cell. 2005; 121:335–48. [PubMed: 15882617]
- Carter EM, Raggio CL. Genetic and orthopedic aspects of collagen disorders. Curr Opin Pediatr. 2009; 21:46–54. [PubMed: 19253462]
- 9. Grassel S, Bauer RJ. Collagen XVI in health and disease. Matrix Biol. 2013; 32:64–73. [PubMed: 23149016]
- Ricard-Blum S. The collagen family. Cold Spring Harb Perspect Biol. 2011; 3:a004978. [PubMed: 21421911]
- Yoshioka H, Iyama K, Inoguchi K, et al. Developmental pattern of expression of the mouse alpha 1 (XI) collagen gene (Col11a1). Dev Dyn. 1995; 204:41–7. [PubMed: 8563024]
- Hida M, Hamanaka R, Okamoto O, et al. Nuclear factor Y (NF-Y) regulates the proximal promoter activity of the mouse collagen alpha1(XI) gene (Col11a1) in chondrocytes. In Vitro Cell Dev Biol Anim. 2014; 50:358–66. [PubMed: 24092017]
- Domet MJ. The general pediatrician and the craniofacial defects team. Ear Nose Throat J. 1986; 65:296–304. [PubMed: 2427307]
- Vijzelaar R, Waller S, Errami A, et al. Deletions within COL11A1 in Type 2 stickler syndrome detected by multiplex ligation-dependent probe amplification (MLPA). BMC Med Genet. 2013; 14:48. [PubMed: 23621912]
- Bekdache GN, Begam MA, Chedid F, et al. Fibrochondrogenesis: prenatal diagnosis and outcome. J Obstet Gynaecol. 2013; 33:663–8. [PubMed: 24127948]
- Rodriguez-Fontenla C, Calaza M, Evangelou E, et al. Assessment of osteoarthritis candidate genes in a meta-analysis of nine genome-wide association studies. Arthritis Rheumatol. 2014; 66:940–9. [PubMed: 24757145]

- Mio F, Chiba K, Hirose Y, et al. A functional polymorphism in COL11A1, which encodes the alpha 1 chain of type XI collagen, is associated with susceptibility to lumbar disc herniation. Am J Hum Genet. 2007; 81:1271–7. [PubMed: 17999364]
- Koyama K, Nakazato K, Min S, et al. COL11A1 gene is associated with limbus vertebra in gymnasts. Int J Sports Med. 2012; 33:586–90. [PubMed: 22510797]
- Hay M, Patricios J, Collins R, et al. Association of type XI collagen genes with chronic Achilles tendinopathy in independent populations from South Africa and Australia. Br J Sports Med. 2013; 47:569–74. [PubMed: 23624467]
- 20. Fischer H, Stenling R, Rubio C, et al. Colorectal carcinogenesis is associated with stromal expression of COL11A1 and COL5A2. Carcinogenesis. 2001; 22:875–8. [PubMed: 11375892]
- 21. Fischer H, Salahshor S, Stenling R, et al. COL11A1 in FAP polyps and in sporadic colorectal tumors. BMC Cancer. 2001; 1:17. [PubMed: 11707154]
- Imamura Y, Scott IC, Greenspan DS. The pro-alpha3(V) collagen chain. Complete primary structure, expression domains in adult and developing tissues, and comparison to the structures and expression domains of the other types V and XI procollagen chains. J Biol Chem. 2000; 275:8749–59. [PubMed: 10722718]
- Kim H, Watkinson J, Varadan V, et al. Multi-cancer computational analysis reveals invasionassociated variant of desmoplastic reaction involving INHBA, THBS2 and COL11A1. BMC Med Genomics. 2010; 3:51. [PubMed: 21047417]
- 24. Anastassiou D, Rumjantseva V, Cheng W, et al. Human cancer cells express Slug-based epithelialmesenchymal transition gene expression signature obtained in vivo. BMC Cancer. 2011; 11:529. [PubMed: 22208948]
- Chernov AV, Baranovskaya S, Golubkov VS, et al. Microarray-based transcriptional and epigenetic profiling of matrix metalloproteinases, collagens, and related genes in cancer. J Biol Chem. 2010; 285:19647–59. [PubMed: 20404328]
- An JH, Lee SY, Jeon JY, et al. Identification of gliotropic factors that induce human stem cell migration to malignant tumor. J Proteome Res. 2009; 8:2873–81. [PubMed: 19351187]
- Suceveanu AI, Suceveanu A, Voinea F, et al. Introduction of cytogenetic tests in colorectal cancer screening. J Gastrointestin Liver Dis. 2009; 18:33–8. [PubMed: 19337631]
- 28. Wu YH, Chang TH, Huang YF, et al. COL11A1 promotes tumor progression and predicts poor clinical outcome in ovarian cancer. Oncogene. 2013
- Teng PN, Wang G, Hood BL, et al. Identification of candidate circulating cisplatin-resistant biomarkers from epithelial ovarian carcinoma cell secretomes. Br J Cancer. 2014; 110:123–32. [PubMed: 24178762]
- Cheon DJ, Tong Y, Sim MS, et al. A collagen-remodeling gene signature regulated by TGF-beta signaling is associated with metastasis and poor survival in serous ovarian cancer. Clin Cancer Res. 2014; 20:711–23. [PubMed: 24218511]
- Tothill RW, Tinker AV, George J, et al. Novel molecular subtypes of serous and endometrioid ovarian cancer linked to clinical outcome. Clin Cancer Res. 2008; 14:5198–208. [PubMed: 18698038]
- 32. Buckanovich RJ, Sasaroli D, O'Brien-Jenkins A, et al. Tumor vascular proteins as biomarkers in ovarian cancer. J Clin Oncol. 2007; 25:852–61. [PubMed: 17327606]
- Vargas AC, McCart Reed AE, Waddell N, et al. Gene expression profiling of tumour epithelial and stromal compartments during breast cancer progression. Breast Cancer Res Treat. 2012; 135:153– 65. [PubMed: 22718308]
- Castellana B, Escuin D, Peiro G, et al. ASPN and GJB2 Are Implicated in the Mechanisms of Invasion of Ductal Breast Carcinomas. J Cancer. 2012; 3:175–83. [PubMed: 22514560]
- 35. Knudsen ES, Ertel A, Davicioni E, et al. Progression of ductal carcinoma in situ to invasive breast cancer is associated with gene expression programs of EMT and myoepithelia. Breast Cancer Res Treat. 2012; 133:1009–24. [PubMed: 22134623]
- 36. Feng Y, Sun B, Li X, et al. Differentially expressed genes between primary cancer and paired lymph node metastases predict clinical outcome of node-positive breast cancer patients. Breast Cancer Res Treat. 2007; 103:319–29. [PubMed: 17123152]

- Ellsworth RE, Seebach J, Field LA, et al. A gene expression signature that defines breast cancer metastases. Clin Exp Metastasis. 2009; 26:205–13. [PubMed: 19112599]
- Halsted KC, Bowen KB, Bond L, et al. Collagen alpha1(XI) in normal and malignant breast tissue. Mod Pathol. 2008; 21:1246–54. [PubMed: 18660795]
- Sok JC, Kuriakose MA, Mahajan VB, et al. Tissue-specific gene expression of head and neck squamous cell carcinoma in vivo by complementary DNA microarray analysis. Arch Otolaryngol Head Neck Surg. 2003; 129:760–70. [PubMed: 12874079]
- 40. Sok JC, Lee JA, Dasari S, et al. Collagen type XI alpha1 facilitates head and neck squamous cell cancer growth and invasion. Br J Cancer. 2013; 109:3049–56. [PubMed: 24231953]
- Schmalbach CE, Chepeha DB, Giordano TJ, et al. Molecular profiling and the identification of genes associated with metastatic oral cavity/pharynx squamous cell carcinoma. Arch Otolaryngol Head Neck Surg. 2004; 130:295–302. [PubMed: 15023835]
- 42. Chattopadhyay I, Singh A, Phukan R, et al. Genome-wide analysis of chromosomal alterations in patients with esophageal squamous cell carcinoma exposed to tobacco and betel quid from high-risk area in India. Mutat Res. 2010; 696:130–8. [PubMed: 20083228]
- Park HJ, Choe BK, Kim SK, et al. Association between collagen type XI alpha1 gene polymorphisms and papillary thyroid cancer in a Korean population. Exp Ther Med. 2011; 2:1111–1116. [PubMed: 22977629]
- 44. Chong IW, Chang MY, Chang HC, et al. Great potential of a panel of multiple hMTH1, SPD, ITGA11 and COL11A1 markers for diagnosis of patients with non-small cell lung cancer. Oncol Rep. 2006; 16:981–8. [PubMed: 17016581]
- 45. Ewald JA, Downs TM, Cetnar JP, et al. Expression microarray meta-analysis identifies genes associated with Ras/MAPK and related pathways in progression of muscle-invasive bladder transition cell carcinoma. PLoS One. 2013; 8:e55414. [PubMed: 23383328]
- 46. Zhao Y, Zhou T, Li A, et al. A potential role of collagens expression in distinguishing between premalignant and malignant lesions in stomach. Anat Rec (Hoboken). 2009; 292:692–700.
 [PubMed: 19306436]
- Garcia-Pravia C, Galvan JA, Gutierrez-Corral N, et al. Overexpression of COL11A1 by cancerassociated fibroblasts: clinical relevance of a stromal marker in pancreatic cancer. PLoS One. 2013; 8:e78327. [PubMed: 24194920]
- 48. Fuentes-Martinez N, Garcia-Pravia C, Garcia-Ocana M, et al. Overexpression of proCOL11A1 as a stromal marker of breast cancer. Histol Histopathol. 2014
- 49. García Pravia CFMN, García Ocaña M, Del Amo J, De los Toyos JR, et al. Anti-proCOL11A1, a new marker of infiltrating breast cancer. Br J Surg. 2009; S5:11.
- Aquino PF, Fischer JS, Neves-Ferreira AG, et al. Are gastric cancer resection margin proteomic profiles more similar to those from controls or tumors? J Proteome Res. 2012; 11:5836–42. [PubMed: 23145836]
- Feldmann G, Habbe N, Dhara S, et al. Hedgehog inhibition prolongs survival in a genetically engineered mouse model of pancreatic cancer. Gut. 2008; 57:1420–30. [PubMed: 18515410]
- 52. Gaspar NJ, Li L, Kapoun AM, et al. Inhibition of transforming growth factor beta signaling reduces pancreatic adenocarcinoma growth and invasiveness. Mol Pharmacol. 2007; 72:152–61. [PubMed: 17400764]
- Tuckwell DS, Reid KB, Barnes MJ, et al. The A-domain of integrin alpha 2 binds specifically to a range of collagens but is not a general receptor for the collagenous motif. Eur J Biochem. 1996; 241:732–9. [PubMed: 8944759]
- Shrivastava A, Radziejewski C, Campbell E, et al. An orphan receptor tyrosine kinase family whose members serve as nonintegrin collagen receptors. Mol Cell. 1997; 1:25–34. [PubMed: 9659900]
- Brown RJ, Mallory C, McDougal OM, et al. Proteomic analysis of Col11a1-associated protein complexes. Proteomics. 2011; 11:4660–76. [PubMed: 22038862]

Highlights

- Collagen XI is a minor collagen with pathogenic roles in musculoskeletal diseases
- ColXIa1 is overexpressed at mRNA and protein levels in many cancer types
- ColXIa1 is highly expressed in tumor stroma, especially by tumor-associated fibroblasts
- ColXIa1 has functional roles in cancer development and may be a therapeutic target

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Figure 1.

Collagen XIa1 in cancer promotion. ColXIa1 is a downstream target of the TGF- β pathway, acting through transcription factor NF-Y. Once translated, procolXIa1 is transported into the ECM, where its N and C termini are cleaved by proteinases. Mature colXIa1 can then regulate MMP3 expression through transcription factor Ets-1, perhaps through putative cell surface receptors integrin a2 β 1 and/or DDR1/2. ColXIa1 also binds nucleolin, a cell surface receptor, which autolytically degrades itself into fragments which post-transcriptionally regulate MMP9. Ultimately, colXIa1 promotes cell proliferation, invasion and metastasis, and therapeutic resistance.

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Dysregulation of colXIa1 in cancer.

yel Reference	45	38	38	34	36	33	49	27	21	20	50	46	n 26	25	39	40	41	42	44	28	31 28
Measurement le	mRNA	protein	protein	mRNA	mRNA	mRNA	Protein	DNA	mRNA	mRNA	protein	mRNA	mRNA, protei	mRNA	mRNA	mRNA	mRNA	DNA	mRNA	mRNA	mRNA mRNA
Clinical association	muscle invasion	metastasis	tumor stroma	local invasion	primary tumor; high risk of metastasis	local invasion	tumor-associated fibroblasts	malignancy	malignancy	malignancy	resection margin	malignancy	malignancy	motility	malignancy	malignancy, metastasis	metastasis	malignancy	pathological stage; lymph node metastasis, poor prognosis	pathological stage; lower overall survival, lower progression-free survival	pathological stage; lower overall survival, lower progression-free survival poor prognosis
Control	papillary transitional cell carcinoma tissue	nonmetastasized primary tumor	normal stroma	DCIS, normal breast tissue, mouse breast carcinomas	paired lymph node metastases	DCIS and normal adjacent stroma	normal breast tissue	none	paired normal colon tissue	Normal colon tissue	gastric tumor and normal gastric tissue	paired premalignant and normal gastric tissue	paired normal neural tissue	non-migratory tumor cells	paired normal mucosal tissue	paired normal mucosal tissue	nonmetastatic HNSCC, normal mucosa	paired normal tissue	normal lung tissue	internal	internal internal
Dysregulation	Overexpressed	Underexpressed	Underexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Mutation	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	Overexpressed	DNA amplification	Overexpressed	Overexpressed	Overexpressed Overexpressed
Sample type	muscle-invasive transitional cell carcinoma	IDC	tumor stroma	IDC	IDC	IDC	Multiple breast cancer types	sloughed tumor cells	FAP polyps	multiple CRC types	tumor resection margins	multiple gastric cancer types	gliobastoma	tumor cells	HNSCC, multiple sites	HNSCC, multiple sites	oral cavity/oropharynx SCC	esophageal SCC	NSCLC	epithelial ovarian cancer	epithelial ovarian cancer serous and endometrioid ovarian cancer
Cancer type	Bladder	Breast	Breast	Breast	Breast	Breast	Breast	Colorectal	Colorectal (inherited)	Colorectal (sporadic)	Gastric	Gastric	Glioma	Glioma	HNSCC	HNSCC	HNSCC	HNSCC	Lung	Ovarian	Ovarian Ovarian

Cancer type	Sample type	Dysregulation	Control	Clinical association	Measurement level	Reference
Ovarian	serous ovarian cancer	Overexpressed	internal	metastasis; lower overall survival	mRNA, protein	30
Ovarian	epithelial ovarian cancer vasculature	Overexpressed	healthy ovarian vascular tissue	angiogenesis	mRNA, protein	32
Pancreatic	PDAC	Overexpressed	chronic pancreatitis and normal pancreas tissue	tumor-associated fibroblasts	mRNA, protein	47
Thyroid	peripheral blood	ANS	peripheral blood from healthy subjects	increased/reduced cancer risk	DNA	43

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