



Stress and the commensal microbiota: importance in parturition and infant neurodevelopment

Tamar L. Gur^{1,2,3*}, Brett L. Worly^{1,2} and Michael T. Bailey^{4,5,6,7}

¹ Psychiatry and Behavioral Health, The Ohio State University Wexner Medical Center, Columbus, OH, USA

² Department of Obstetrics and Gynecology, The Ohio State University Wexner Medical Center, Columbus, OH, USA

³ Department of Neuroscience, The Ohio State University Wexner Medical Center, Columbus, OH, USA

⁴ Division of Biosciences, College of Dentistry, The Ohio State University, Columbus, OH, USA

⁵ Institute for Behavioral Medicine Research, The Ohio State University Wexner Medical Center, Columbus, OH, USA

⁶ Department of Pediatrics, The Ohio State University Wexner Medical Center, Columbus, OH, USA

⁷ Comprehensive Cancer Center, The Ohio State University Wexner Medical Center, Columbus, OH, USA

Edited by:

Stefan Oskar Reber, University of Ulm, Germany

Reviewed by:

John F. Cryan, University College Cork, Ireland

Casimiro Cabrera Abreu, Queen's University and Providence Care, Canada

*Correspondence:

Tamar L. Gur, The Ohio State University Wexner Medical Center, 5038 Graves Hall, 333W 10th Street, Columbus, OH 43210, USA
e-mail: tamar.gur@osumc.edu

The body is colonized by an enormous array of microbes that are collectively called the microbiota. During quiescent periods, microbial communities within the gut are relatively resistant to change. However, several factors that disrupt homeostasis can also significantly change gut microbial community structure. One factor that has been shown to change the composition of the gut microbiota is exposure to psychological stressors. Studies demonstrate that the commensal microbiota are involved in stressor-induced immunomodulation, but other biological effects are not yet known. This review discusses emerging evidence that the microbiota can impact the brain and behavior and indicates that stressor-induced alterations in the composition of gut microbial communities contribute to stressor-induced behavioral changes. This review will also discuss the evidence that such effects are most evident early in life, where both stress and the microbiota have been linked to birth outcomes, such as prematurity, and neurodevelopment. When considered together, a paradigm emerges in which stressor-induced alterations in commensal microbial populations significantly impact parturition and infant neurodevelopment.

Keywords: microbiota, psychological stress, neurodevelopment, prematurity, neuroimmune, anxiety, depression

INTRODUCTION

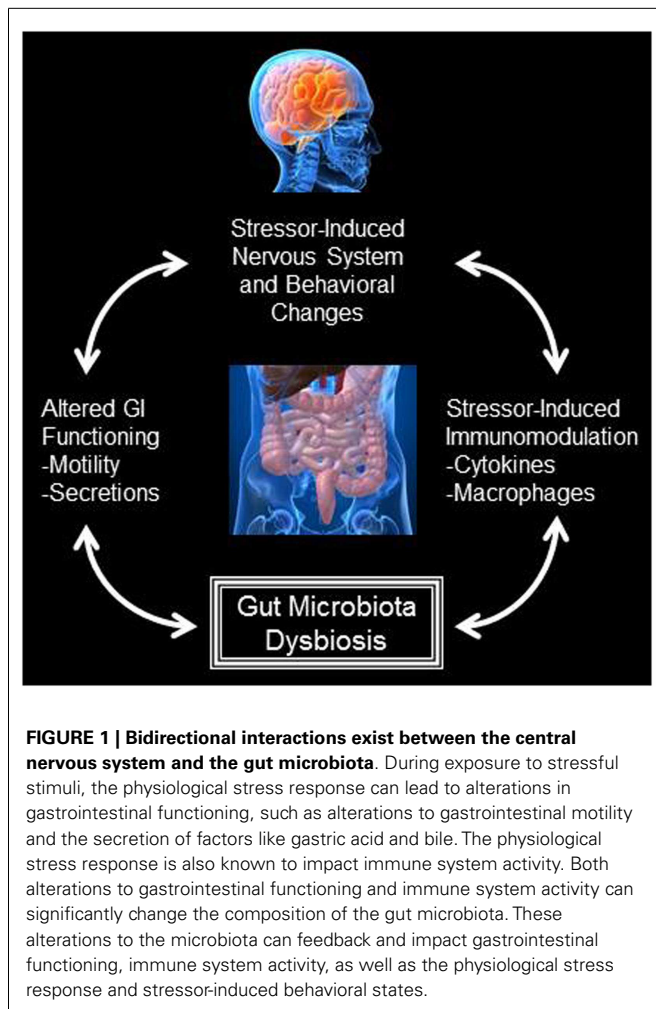
The hypothesis that the origins of adult disease are developmental, beginning *in utero* is called the “Barker hypothesis” after one of its leading proponents and the author of a study demonstrating increased risk of cardiovascular disease in infants born underweight (1). It states that adverse influences early in development, such as poor nutrition or infection, result in permanent physiological changes and in increased disease risk in adulthood. This is an area of increasing research in many fields, including Psychiatry, Immunology, and Endocrinology, which are now exploring whether development of diseases such as asthma, diabetes, and anxiety contains a developmental, intrauterine element. Neurodevelopment is exquisitely sensitive to perturbations in the maternal milieu, including diet, infection, and stress, with potentially long-lasting behavioral consequences. Disorders such as schizophrenia, anxiety, depression, and autism have been found to be associated with *in utero* and early neonatal exposure to these stimuli (2). Infants exposed to antenatal stress demonstrate increased risk of developing a host of childhood and adult diseases. While alterations in the hypothalamic–pituitary–adrenal (HPA) axis and immune function have been the target of investigation as underlying mechanisms conferring increased risk, the microbiome is an emerging candidate as a potential mediator of stress-induced pathogenesis (Figure 1).

Every surface of the body naturally harbors unique microbial communities comprised of archaea, protists, viruses, and bacteria.

To date, bacteria residing on mucosal surfaces, including the oral cavity, reproductive tract, and gastrointestinal tract, are the best characterized. These bacteria form highly ordered microbial communities as a result of ecological successions that select microbes that are best adapted for their given niche. Although these microbial communities are relatively resistant to change, it is recognized that factors such as alterations in diet and the administration of antibiotics can result in modifications in microbial community structure. Studies from this laboratory, as well as others, have demonstrated that psychosocial stressors can also impact microbial community structure in the gut. This review will briefly describe studies that have linked stressor-induced alterations in gut microbial community structure to alterations in immune system activity and behavioral responses. The potential impact of these interactions on pregnancy outcome and on infant development will also be discussed.

NEUROENDOCRINE-BACTERIAL INTERACTIONS

The field of psychoneuroimmunology has amply demonstrated that the physiological response to different types of stressor significantly impacts immune system reactivity to antigenic challenge [reviewed in Ref. (3)]. Primary mediators of the stress response, including endogenous glucocorticoids such as cortisol in humans and corticosterone in rodents, can affect immune system reactivity by suppressing the expression and activity of key transcription factors, such as NF- κ B (4). Likewise, stressor-induced



activation of the sympathetic nervous system, resulting in the release of endogenous catecholamine hormones (namely epinephrine from the adrenal medulla, and norepinephrine from adrenergic nerve terminals), can significantly increase or decrease immune cell activity depending on the leukocyte subset and the adrenergic receptor that is bound. As details of the importance of neuroendocrine mediators for leukocyte reactivity emerged (5), findings demonstrating that bacterial pathogens themselves also respond to neuroendocrine hormones began to emerge (6–12).

The growth of many types of bacteria, including both infectious and commensal organisms can be significantly impacted by neuroendocrine hormones. For example, the growth of commensal and of pathogenic *E. coli* can be increased over 10,000-fold by simply adding norepinephrine to a serum-based microbial medium (8, 10, 12–14). It is now recognized that a wide variety of neuroendocrine hormones can impact a vast array of bacteria in culture [reviewed in Ref. (10, 13)], however, demonstrating that direct neuroendocrine–bacterial interactions occur *in vivo* has been more challenging.

One of the first studies assessing the effects of neuroendocrine hormones on bacterial growth involved increasing norepinephrine

levels *in vivo* using 6-hydroxydopamine, which lyses sympathetic nerve terminals. This resulted in an approximately 10,000-fold increase in commensal *E. coli* levels in the cecum of mice (11). The effects of norepinephrine on bacterial growth was also evident in an ileal loop model, where growing *Salmonella enterica* with norepinephrine prior to inoculation into the ileal loop significantly increased pathogen growth and associated disease (15). Findings that neuroendocrine mediators associated with the stress response could significantly impact bacterial growth led us to test whether stressor exposure could significantly change the levels of bacteria cultured from the intestines.

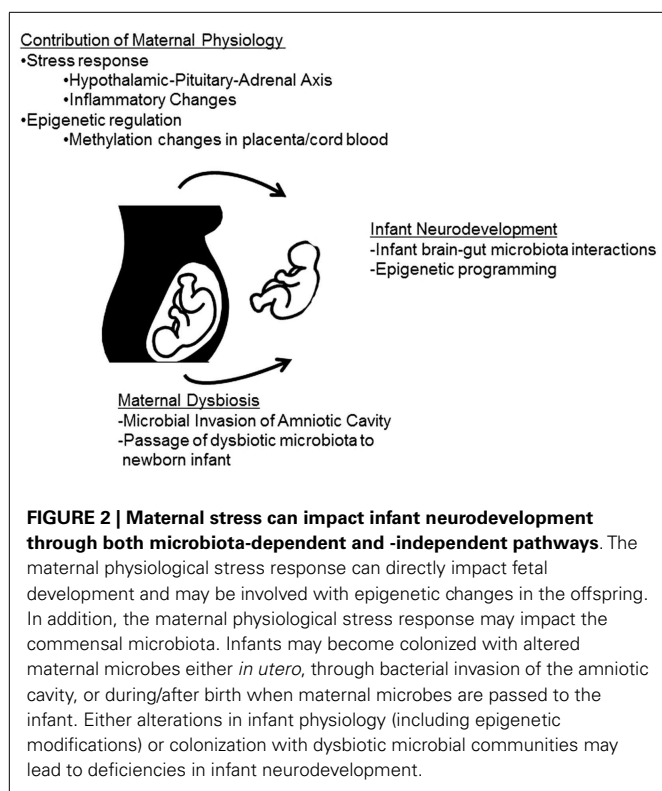
STRESSOR EXPOSURE AND THE COMMENSAL MICROBIOTA

Tannock and Savage demonstrated almost 40 years ago that moving mice into a cage lacking bedding, food, and water reduced the number of lactobacilli that could be cultured from the gastrointestinal tract (16). While this suggested that the stress response associated with new housing led to the differences, it was difficult to interpret these data due to the food and water deprivation. Thus, infant rhesus monkeys, with *ad libitum* access to food and water were exposed to a maternal separation stressor and the number of lactobacilli that could be cultured from the intestines was assessed. Exposure to the stressor significantly reduced lactobacilli levels, and the magnitude of the reduction was associated with stress-indicative behaviors. In general, monkeys which showed the greatest behavioral signs of distress also had the lowest levels of lactobacilli (17).

Stressor-induced reductions in lactobacilli have also been identified in humans during the stress of school examinations (18) and in murine studies utilizing prolonged restraint (19) or a short-lasting social stressor (19). The biological importance of stressor-induced alterations of the microbiota is not well understood. However, studies demonstrate that some aspects of stressor-induced increases in immune system reactivity are dependent upon the microbiota. For example, exposure to a social stressor, known as social disruption (SDR), significantly increases IL-6 levels in the blood and increases the reactivity of splenic macrophages to microbial stimulation. These effects, however, did not appear in germ-free (GF) mice or mice treated with antibiotics to reduce the microbiota (20). Similarly, exposing rats to repeat tail shock significantly increases cytokine levels in the blood; treatment with antibiotics to reduce the microbiota attenuated stressor-induced increases, but only for cytokines whose activation is dependent upon the inflammasome (21, 22). When considered together, these studies demonstrate the importance of the microbiota for stressor-induced immunopotential, and suggest that microbial products act as a danger signal to prime the immune system for enhanced reactivity (23).

STRESS, THE MICROBIOME, AND PRETERM BIRTH

While it is becoming increasingly evident that stressor-induced alterations in the microbiota of adult animals can significantly impact host physiology, these effects are transient and return to baseline after termination of the stressor. However, the microbiota have exaggerated and prolonged effects when perturbed during gestation or early in infancy. Interest in the connection between stress and the human microbiome, and its impact specifically on female



reproduction and the developing fetus is evolving, but still in a nascent stage (Figure 2).

Preterm birth (PTB) is the first critical juncture to look at in the reproductive cycle when trying to discern the effects of stress and the microbiome on women, as stress is a well-known risk factor that affects the pillars of PTB: immune response/inflammation, and the HPA axis (24). There is considerable debate about stress, inflammation, cortisol, and PTB, and exact mechanisms are ripe for evaluation. Microbial invasion of the amniotic cavity and the associated host inflammatory response is a leading etiology of PTB (25–28). The mechanisms by which micro-organisms gain access to the decidua and amniotic cavity are not completely known, but are thought to involve invasion from microbes locally (e.g., lower gut or genital tract) or hematogenously (25). The ability of genital pathogens to invade the amnion and induce PTB has been well studied. However, the source of hematogenous microbes and their ability to invade the amniotic cavity remains unclear. Oral microbes have been proposed as a key source of microbes passed to the amniotic cavity via hematogenous transmission (29). This is based on recent evidence that the placenta harbors a commensal microbiome with a community structure that is more closely reflective of oral microbial communities than microbial communities found elsewhere in the body (30). In addition, periodontal diseases are often, but not always (31), related to increased risk of PTB (32). It is thought that during some cases of periodontal infection, oral pathogens, such as *Fusobacterium nucleatum* translocate from the oral cavity into circulation where they eventually reach and invade the pregnant uterus (32). Given the strong association between stress and PTB, as well as oral microbiota and PTB, it is

important to consider the possibility that oral microbes play a critical role in linking the stress response to PTB. While it is known that stressor exposure can change the composition of the gut microbiota, and the ability of microbes to translocate from the lumen of the intestines to the interior of the body (19, 33–35), to date, it is not known whether stressor exposure also impacts the composition of the oral microbiota, or their ability to translocate to other organs. Moreover, the relationship between the various commensal microbial communities and the intrauterine environment is not well understood in part due to the lack of studies involving laboratory animal models, and this is certainly an important area of future studies.

STRESS, THE MICROBIOME, AND PARTURITION

With information suggesting that early-life exposures can be formative for neurodevelopmental, allergic, and immunological response later in life, attention can be turned to parturition for potential influencing factors specifically with the microbiome. Two studies suggest that the neonatal fecal microbiome is altered due to cesarean section at day 1 of life, and continues up to 6 weeks of age (36, 37). Meta-analysis suggests that a 20% increase in asthma rates exists among babies born with cesarean section, although substantial heterogeneity exists in the data (38). Breastfed babies have a different, more “beneficial” gut microbiota (39, 40), obtain different prebiotic human milk oligosaccharides (41), and acquire maternal intestinal bacteria from breast milk (42). In addition to having different routes of delivery, it is interesting to note that babies born via cesarean section are also born to mothers with higher levels of stress. Cesarean section, especially unplanned cesarean section, is perceived as severely stressful (43, 44) and can lead to significant increases in glucocorticoid and catecholamine neuroendocrine hormones (45, 46). Thus, while the mode of delivery is undoubtedly important, delivery via cesarean section may have additional effects on the baby due to the high physiological stress response in the mother. For example, cesarean section is associated with a different human milk microbiome, potentially as a result of the stress of cesarean section. While never described before for stress, it is recognized that overall health of the mother can impact the milk microbiome; obese women’s breast milk microbiome has been shown to be less diverse than healthy weight controls (47). Thus, it is likely that stress and hormonal alterations impact breast milk microbiome composition. Interestingly, early changes in the human gut microbiome with perinatal antibiotics were found to have an association with increased disease risks early in life (48).

NEONATAL MICROBIOME AND NEURODEVELOPMENT

There is accumulating evidence that the microbiome can influence behavior [Reviewed in Ref. (49)], supporting the concept of a microbiome–gut–brain axis. While specific mechanisms underlying this influence remain unknown, the presence of the microbiome in the placenta and amnion suggest proximity and capacity to influence the developing fetus. Studies examining the impact of microbiome on the developing central nervous system (CNS) have utilized GF mice that have decreased anxiety, as well as increased motor activity compared to conventional mice. These behavioral changes were accompanied by increased turnover of dopamine, norepinephrine, and serotonin in the striatum, though not, it

should be noted, in the hippocampus or frontal cortex. Levels of brain-derived neurotrophic factor (BDNF), a protein with a significant role in neurodevelopment, and the development of depression and anxiety, were found to be reduced in the hippocampus and amygdala. These are brain regions implicated in the pathogenesis of anxiety, suggesting a mechanistic substrate for the behavioral changes (50). Because BDNF has a significant role in neurodevelopment, these findings have tantalizing implications regarding the influence of stress, alterations in the microbiome, neonatal colonization, and neurodevelopmental disorders.

Not all aspects of neurodevelopment are influenced by the microbiota, and some components of microbial influences on neurodevelopment are gender-specific. For example, GF mice have also demonstrated significant deficits in social behavior (51), some of which, specifically social avoidance, were reversed with re-colonization post-weaning. However, social cognition did not improve with re-colonization, suggesting that social cognition is as amenable to microbial-based interventions.

The effects that the microbiota have on the developing CNS appear to be gender-dependent. GF male mice have increased levels of serotonin and its metabolites in their hippocampus, and these did not normalize with introduction of a regular microbiome following weaning. Immunological and neuroendocrine effects were also found, with GF mice demonstrating a blunted immune response, based on TNF- α production, with a larger effect in female mice. GF animals also had a stronger corticosterone response in relation to stress, with female mice showing a smaller response. Male GF mice showed decreased expression of BDNF; however, female mice did not (52).

When considered together, data from laboratory animals indicate that maternal microbes that are passed to the infant are necessary for normal neurodevelopment. Disrupting these microbial communities, or their passage to the infant, can in turn impact neurodevelopment. Moreover, there are significant sex differences in the impact of microbial colonization on neurodevelopment. While it is not yet clear how this occurs, sex difference mechanisms in mice may be due to estrous cycle hormones and the CNS serotonergic system, as the estrogen receptor (ER beta) has a role in the hippocampal serotonin concentration. It is unclear whether the estrogen and estrogen receptor have a larger impact, or whether the microbiome has a larger impact on serotonin, and further studies are needed.

NEONATAL MICROBIOME AND HEALTH OUTCOMES

Animal studies support the idea that alterations in neonatal life and gut microbiome can have substantial impact on long-lasting health. Rat offspring deprived of their mother showed increased permeability of the colonic mucosa and a 10- to 100-fold increase in bacterial adherence to colonic tissue and spleen translocation. Of note, these changes were prevented by injecting rat pups with a corticotropin-releasing hormone receptor antagonist daily during maternal separation, suggesting that the HPA axis mediated this effect (53). Further evidence of maternal influence on the offspring is provided by the finding that, in rats, maternal diet influenced offspring gut architecture (54) and high-fat diet in rat mothers led to decreases in maltase and sucrase in offspring, while *E. coli* introduced to rat mothers in pregnancy and lactation led

to increased offspring intestinal permeability, systemic inflammation, and obesity for this next generation (55). Furthermore, *E. coli* introduced in rat mothers led to changes in offspring weight and gut microbiota (56). Maternal administration of antibiotics in the prenatal period was associated with increased gut permeability and systemic inflammation (54) as well as increased visceral sensitivity (57). In rodents, maternal separation, a highly validated model of early-life stress, also lead to an increase in visceral sensitivity, with concomitant alterations in stress response and microbial community structure (58). A swine model for perinatal disturbance was adopted with oral antibiotics to sows. Short- and long-term changes were seen in paracellular permeability (59). Together, these studies suggest that altering either the maternal or perinatal flora has long-lasting implications on the gut of the offspring, as well as immunological repercussions. Mechanisms for this change are still being evaluated, but one potential route is that perinatal changes in bacterial colonization alter gastrointestinal heat shock protein expression with permanent implications on health (60).

HOW THE MICROBIOME MAY IMPACT NEURODEVELOPMENT AND FUTURE DIRECTIONS

While clear lines of evidence exist supporting an effect of stress on the microbiome, as well as the ability of stress to modulate neurodevelopment and behavioral changes, a delineated mechanism of action by the microbiome upon the CNS is an area of intense scrutiny and investigation (61–63). To date, the HPA axis and alterations in the immune response have accumulated the strongest evidence of associations with alterations in the microbiome.

The immune system, in addition to being directly influenced by stress, is a direct target of both microbiota and probiotic agents, and also has bidirectional communication with the CNS, making it an appealing candidate. The microbiome, through the innate immune system, alters levels of both pro- and anti-inflammatory cytokines, which are capable of directly impacting CNS function (64, 65). Stress also has a clear impact on the HPA axis, which is capable of regulating the inflammatory cascade, and growth of microbiota, as discussed above. However, directly tethering this phenomenon to the developing CNS is a more arduous task. While studies reviewed above demonstrate alterations in important players in CNS development, such as BDNF, with alterations (or lack of) microbiota, many steps in this process remain elusive.

Another potential mediator of the effect of stress on neurodevelopment, via the microbiome, is epigenetic regulation (66). This refers to heritable changes in gene expression not due to changes in DNA sequence, and epigenetic modification is a process known to be especially sensitive to early-life experiences. Thus, *in utero* and early-life alterations in microbiome could potentially utilize epigenetic processes to exert long-term behavioral changes, such as those described following psychosocial stressors. Indeed, a growing body of literature supports a major role of epigenetic modification in the neurobiology of psychiatric disorders (67). How might the microbiome influence epigenetics? Microbiotas are involved in the breakdown of nutrients, and in that process create metabolites with neuroactive properties, including amino acids and monoamines (68). Moreover, they are a key source of butyrate, which is a histone deacetylase (HDAC) inhibitor, a key

step in transcriptional regulation. However, whether these substances are able to make a substantial impact on CNS function itself, and how they would access the CNS from the periphery, is open to debate and requires further examination.

As a whole, the ability of the microbiome to impact the developing CNS, and participate in the effect of psychosocial stress is an exciting concept, because it is susceptible to targeting with pre- and probiotics, which has vast implications for both neurodevelopment and other health outcomes. While much work remains to be done to elucidate mechanisms, this daunting task is imperative to complete.

REFERENCES

- Barker DJ, Osmond C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. *Lancet* (1986) 1:1077–81. doi:10.1016/S0140-6736(86)91340-1
- Bale TL, Baram TZ, Brown AS, Goldstein JM, Insel TR, McCarthy MM, et al. Early life programming and neurodevelopmental disorders. *Biol Psychiatry* (2010) 68:314–9. doi:10.1016/j.biopsych.2010.05.028
- Padgett DA, Glaser R. How stress influences the immune response. *Trends Immunol* (2003) 24:444–8. doi:10.1016/S1471-4906(03)00173-X
- Bellavance MA, Rivest S. The HPA – immune axis and the immunomodulatory actions of glucocorticoids in the brain. *Front Immunol* (2014) 5:136. doi:10.3389/fimmu.2014.00136
- Padro CJ, Sanders VM. Neuroendocrine regulation of inflammation. *Semin Immunol* (2014) 26(5):357–68. doi:10.1016/j.smim.2014.01.003
- Lyte M, Arulanandam BP, Frank CD. Production of shiga-like toxins by *Escherichia coli* O157:H7 can be influenced by the neuroendocrine hormone norepinephrine. *J Lab Clin Med* (1996) 128:392–8. doi:10.1016/S0022-2143(96)80011-4
- Lyte M. Induction of gram-negative bacterial growth by neurochemical containing banana (*Musa x paradisiaca*) extracts. *FEMS Microbiol Lett* (1997) 154:245–50. doi:10.1111/j.1574-6968.1997.tb12651.x
- Lyte M, Arulanandam B, Nguyen K, Frank C, Erickson A, Francis D. Norepinephrine induced growth and expression of virulence associated factors in enterotoxigenic and enterohemorrhagic strains of *Escherichia coli*. *Adv Exp Med Biol* (1997) 412:331–9. doi:10.1007/978-1-4899-1828-4_54
- Lyte M, Freestone PP, Neal CP, Olson BA, Haigh RD, Bayston R, et al. Stimulation of *Staphylococcus epidermidis* growth and biofilm formation by catecholamine inotropes. *Lancet* (2003) 361:130–5. doi:10.1016/S0140-6736(03)12231-3
- Lyte M. Microbial endocrinology and infectious disease in the 21st century. *Trends Microbiol* (2004) 12:14–20. doi:10.1016/j.tim.2003.11.004
- Lyte M, Bailey MT. Neuroendocrine-bacterial interactions in a neurotoxin-induced model of trauma. *J Surg Res* (1997) 70:195–201. doi:10.1006/jrsr.1997.5130
- Lyte M, Nguyen KT. Alteration of *Escherichia coli* O157:H7 growth and molecular fingerprint by the neuroendocrine hormone noradrenaline. *Microbios* (1997) 89:197–213.
- Freestone PP, Sandrini SM, Haigh RD, Lyte M. Microbial endocrinology: how stress influences susceptibility to infection. *Trends Microbiol* (2008) 16:55–64. doi:10.1016/j.tim.2007.11.005
- Lyte M, Erickson AK, Arulanandam BP, Frank CD, Crawford MA, Francis DH. Norepinephrine-induced expression of the K99 pilus adhesin of enterotoxigenic *Escherichia coli*. *Biochem Biophys Res Commun* (1997) 232:682–6. doi:10.1006/bbrc.1997.6356
- Pullinger GD, Carnell SC, Sharaff FF, van Diemen PM, Dziva F, Morgan E, et al. Norepinephrine augments *Salmonella enterica*-induced enteritis in a manner associated with increased net replication but independent of the putative adrenergic sensor kinases QseC and QseE. *Infect Immun* (2010) 78:372–80. doi:10.1128/IAI.01203-09
- Tannock GW, Savage DC. Influences of dietary and environmental stress on microbial populations in the murine gastrointestinal tract. *Infect Immun* (1974) 9:591–8.
- Bailey MT, Coe CL. Maternal separation disrupts the integrity of the intestinal microflora in infant rhesus monkeys. *Dev Psychobiol* (1999) 35:146–55. doi:10.1002/(SICI)1098-2302(199909)35:2<146::AID-DEV7>3.0.CO;2-G
- Knowles SR, Nelson EA, Palombo EA. Investigating the role of perceived stress on bacterial flora activity and salivary cortisol secretion: a possible mechanism underlying susceptibility to illness. *Biol Psychol* (2008) 77:132–7. doi:10.1016/j.biopsycho.2007.09.010
- Galley JD, Nelson MC, Yu Z, Dowd SE, Walter J, Kumar PS, et al. Exposure to a social stressor disrupts the community structure of the colonic mucosa-associated microbiota. *BMC Microbiol* (2014) 14:189. doi:10.1186/1471-2180-14-189
- Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG, Lyte M. Exposure to a social stressor alters the structure of the intestinal microbiota: implications for stressor-induced immunomodulation. *Brain Behav Immun* (2011) 25:397–407. doi:10.1016/j.bbi.2010.10.023
- Maslanik T, Tannura K, Mahaffey L, Loughridge AB, Benninson L, Ursell L, et al. Commensal bacteria and MAMPs are necessary for stress-induced increases in IL-1beta and IL-18 but not IL-6, IL-10 or MCP-1. *PLoS One* (2012) 7:e50636. doi:10.1371/journal.pone.0050636
- Maslanik T, Mahaffey L, Tannura K, Beninson L, Greenwood BN, Fleshner M. The inflammasome and danger associated molecular patterns (DAMPs) are implicated in cytokine and chemokine responses following stressor exposure. *Brain Behav Immun* (2013) 28:54–62. doi:10.1016/j.bbi.2012.10.014
- Fleshner M. Stress-evoked sterile inflammation, danger associated molecular patterns (DAMPs), microbial associated molecular patterns (MAMPs) and the inflammasome. *Brain Behav Immun* (2013) 27:1–7. doi:10.1016/j.bbi.2012.08.012
- Shapiro GD, Fraser WD, Frasn MG, Seguin JR. Psychosocial stress in pregnancy and preterm birth: associations and mechanisms. *J Perinat Med* (2013) 41:631–45. doi:10.1515/jpm-2012-0295
- DiGiulio DB. Diversity of microbes in amniotic fluid. *Semin Fetal Neonatal Med* (2012) 17:2–11. doi:10.1016/j.siny.2011.10.001
- Goldenberg RL, Hauth JC, Andrews WW. Intrauterine infection and preterm delivery. *N Engl J Med* (2000) 342:1500–7. doi:10.1056/NEJM200005183422007
- Goncalves LF, Chaiworapongsa T, Romero R. Intrauterine infection and prematurity. *Ment Retard Dev Disabil Res Rev* (2002) 8:3–13. doi:10.1002/mrdd.10008
- Romero R, Gomez R, Chaiworapongsa T, Conoscenti G, Kim JC, Kim YM. The role of infection in preterm labour and delivery. *Paediatr Perinat Epidemiol* (2001) 15(Suppl 2):41–56. doi:10.1046/j.1365-3016.2001.00007.x
- Cetin I, Pileri P, Villa A, Calabrese S, Ottolenghi L, Abati S. Pathogenic mechanisms linking periodontal diseases with adverse pregnancy outcomes. *Reprod Sci* (2012) 19:633–41. doi:10.1177/1933719111432871
- Aagaard K, Ma J, Antony KM, Ganu R, Petrosino J, Versalovic J. The placenta harbors a unique microbiome. *Sci Transl Med* (2014) 6:237ra65. doi:10.1126/scitranslmed.3008599
- Abati S, Villa A, Cetin I, Dessole S, Luglie PF, Strohmenger L, et al. Lack of association between maternal periodontal status and adverse pregnancy outcomes: a multicentric epidemiologic study. *J Matern Fetal Neonatal Med* (2013) 26:369–72. doi:10.3109/14767058.2012.733776
- Madianos PN, Lief S, Murtha AP, Boggess KA, Auten RL Jr, Beck JD, et al. Maternal periodontitis and prematurity. Part II: maternal infection and fetal exposure. *Ann Periodontol* (2001) 6:175–82. doi:10.1902/annals.2001.6.1.175
- Bailey MT, Engler H, Sheridan JF. Stress induces the translocation of cutaneous and gastrointestinal microflora to secondary lymphoid organs of C57BL/6 mice. *J Neuroimmunol* (2006) 171:29–37. doi:10.1016/j.jneuroim.2005.09.008
- Bailey MT, Dowd SE, Parry NM, Galley JD, Schauer DB, Lyte M. Stressor exposure disrupts commensal microbial populations in the intestines and leads to increased colonization by *Citrobacter rodentium*. *Infect Immun* (2010) 78:1509–19. doi:10.1128/IAI.00862-09
- Reber SO, Peters S, Slattery DA, Hofmann C, Scholmerich J, Neumann ID, et al. Mucosal immunosuppression and epithelial barrier defects are key events in murine psychosocial stress-induced colitis. *Brain Behav Immun* (2011) 25:1153–61. doi:10.1016/j.bbi.2011.03.004
- Biasucci G, Rubini M, Riboni S, Morelli L, Bessi E, Retetangos C. Mode of delivery affects the bacterial community in the newborn gut. *Early Hum Dev* (2010) 86(Suppl 1):13–5. doi:10.1016/j.earlhumdev.2010.01.004
- Dominguez-Bello MG, Costello EK, Contreras M, Magris M, Hidalgo G, Fierer N, et al. Delivery mode shapes the acquisition and structure of the initial microbiota across multiple body habitats in newborns. *Proc Natl Acad Sci USA* (2010) 107:11971–5. doi:10.1073/pnas.1002601107

38. Thavagnanam S, Fleming J, Bromley A, Shields MD, Cardwell CR. A meta-analysis of the association between caesarean section and childhood asthma. *Clin Exp Allergy* (2008) **38**:629–33. doi:10.1111/j.1365-2222.2007.02780.x
39. Fallani M, Young D, Scott J, Norin E, Amarri S, Adam R, et al. Intestinal microbiota of 6-week-old infants across Europe: geographic influence beyond delivery mode, breast-feeding, and antibiotics. *J Pediatr Gastroenterol Nutr* (2010) **51**:77–84. doi:10.1097/MPG.0b013e3181d1b11e
40. Roger LC, Costabile A, Holland DT, Hoyles L, McCartney AL. Examination of faecal *Bifidobacterium* populations in breast- and formula-fed infants during the first 18 months of life. *Microbiology* (2010) **156**:3329–41. doi:10.1099/mic.0.043224-0
41. Shen Q, Tuohy KM, Gibson GR, Ward RE. In vitro measurement of the impact of human milk oligosaccharides on the faecal microbiota of weaned formula-fed infants compared to a mixture of prebiotic fructooligosaccharides and galactooligosaccharides. *Lett Appl Microbiol* (2011) **52**:337–43. doi:10.1111/j.1472-765X.2011.03005.x
42. Albesharar R, Ehrmann MA, Korakli M, Yazaji S, Vogel RF. Phenotypic and genotypic analyses of lactic acid bacteria in local fermented food, breast milk and faeces of mothers and their babies. *Syst Appl Microbiol* (2011) **34**:148–55. doi:10.1016/j.syapm.2010.12.001
43. Ryding EL, Wijma K, Wijma B. Psychological impact of emergency cesarean section in comparison with elective cesarean section, instrumental and normal vaginal delivery. *J Psychosom Obstet Gynaecol* (1998) **19**:135–44. doi:10.3109/01674829809025691
44. Schindl M, Birner P, Reingrabner M, Joura E, Husslein P, Langer M. Elective cesarean section vs. spontaneous delivery: a comparative study of birth experience. *Acta Obstet Gynecol Scand* (2003) **82**:834–40. doi:10.1034/j.1600-0412.2003.00194.x
45. Loughran PG, Moore J, Dundee JW. Maternal stress response associated with caesarean delivery under general and epidural anaesthesia. *Br J Obstet Gynaecol* (1986) **93**:943–9. doi:10.1111/j.1471-0528.1986.tb08013.x
46. Scheinin M, Scheinin H, Ekblad U, Kanto J. Biological correlates of mental stress related to anticipated caesarean section. *Acta Anaesthesiol Scand* (1990) **34**:640–4. doi:10.1111/j.1399-6576.1990.tb03163.x
47. Cabrera-Rubio R, Collado MC, Laitinen K, Salminen S, Isolauri E, Mira A. The human milk microbiome changes over lactation and is shaped by maternal weight and mode of delivery. *Am J Clin Nutr* (2012) **96**:544–51. doi:10.3945/ajcn.112.037382
48. Bedford Russell AR, Murch SH. Could peripartum antibiotics have delayed health consequences for the infant? *BJOG* (2006) **113**:758–65. doi:10.1111/j.1471-0528.2006.00952.x
49. Dinan TG, Cryan JF. Melancholic microbes: a link between gut microbiota and depression? *Neurogastroenterol Motil* (2013) **25**:713–9. doi:10.1111/nmo.12198
50. Diaz HR, Wang S, Anuar F, Qian Y, Bjorkholm B, Samuelsson A, et al. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci U S A* (2011) **108**:3047–52. doi:10.1073/pnas.1010529108
51. Desbonnet L, Clarke G, Shanahan F, Dinan TG, Cryan JF. Microbiota is essential for social development in the mouse. *Mol Psychiatry* (2014) **19**:146–8. doi:10.1038/mp.2013.65
52. Clarke G, Grenham S, Scully P, Fitzgerald P, Moloney RD, Shanahan F, et al. The microbiome-gut-brain axis during early life regulates the hippocampal serotonergic system in a sex-dependent manner. *Mol Psychiatry* (2013) **18**:666–73. doi:10.1038/mp.2012.77
53. Gareau MG, Jury J, Yang PC, MacQueen G, Perdue MH. Neonatal maternal separation causes colonic dysfunction in rat pups including impaired host resistance. *Pediatr Res* (2006) **59**:83–8. doi:10.1203/01.pdr.0000190577.62426.45
54. Fak F, Ahrne S, Molin G, Jeppsson B, Westrom B. Microbial manipulation of the rat dam changes bacterial colonization and alters properties of the gut in her offspring. *Am J Physiol Gastrointest Liver Physiol* (2008) **294**:G148–54. doi:10.1152/ajpgi.00023.2007
55. Fak F, Karlsson CL, Ahrne S, Molin G, Westrom B. Effects of a high-fat diet during pregnancy and lactation are modulated by *E. coli* in rat offspring. *Int J Obes (Lond)* (2012) **36**:744–51. doi:10.1038/ijo.2011.118
56. Karlsson CL, Molin G, Fak F, Johansson Hagslatt ML, Jakesevich M, Hakansson A, et al. Effects on weight gain and gut microbiota in rats given bacterial supplements and a high-energy-dense diet from fetal life through to 6 months of age. *Br J Nutr* (2011) **106**:887–95. doi:10.1017/S0007114511001036
57. O'Mahony SM, Felice VD, Nally K, Savignac HM, Claesson MJ, Scully P, et al. Disturbance of the gut microbiota in early-life selectively affects visceral pain in adulthood without impacting cognitive or anxiety-related behaviors in male rats. *Neuroscience* (2014) **277**:885–901. doi:10.1016/j.neuroscience.2014.07.054
58. O'Mahony SM, Marchesi JR, Scully P, Codling C, Ceolho AM, Quigley EM, et al. Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol Psychiatry* (2009) **65**(3):263–7. doi:10.1016/j.biopsych.2008.06.026
59. Boudry G, Morise A, Seve B, Huerou-Luron LE. Effect of milk formula protein content on intestinal barrier function in a porcine model of LBW neonates. *Pediatr Res* (2011) **69**:4–9. doi:10.1203/PDR.0b013e3181fc9d13
60. Lalles JP. Long term effects of pre- and early postnatal nutrition and environment on the gut. *J Anim Sci* (2012) **90**(Suppl 4):421–9. doi:10.2527/jas.53904
61. Funkhouser LJ, Bordenstein SR. Mom knows best: the universality of maternal microbial transmission. *PLoS Biol* (2013) **11**(8):e1001631. doi:10.1371/journal.pbio.1001631
62. Jašarević E, Rodgers AB, Tracy L. A novel role for maternal stress and microbial transmission in early life programming and neurodevelopment neurobiology of stress. *Neurobiol Stress* (2015) **1**:81–8.
63. Borre YE, O'Keefe GW, Clarke G, Stanton C, Dinan TG, Cryan JF. Microbiota and neurodevelopmental windows: implications for brain disorders. *Trends Mol Med* (2014) **20**(9):509–18. doi:10.1016/j.molmed.2014.05.002
64. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW. From inflammation to sickness and depression: when the immune system subjugates the brain. *Nat Rev Neurosci* (2008) **9**:46–56. doi:10.1038/nrn2297
65. Dantzer R, O'Connor JC, Lawson MA, Kelley KW. Inflammation-associated depression: from serotonin to kynurenine. *Psychoneuroendocrinology* (2011) **36**:426–36. doi:10.1016/j.psyneuen.2010.09.012
66. Stilling RM, Dinan TG, Cryan JF. Microbial genes, brain & behaviour – epigenetic regulation of the gut-brain axis. *Genes Brain Behav* (2014) **13**:69–86. doi:10.1111/gbb.12109
67. Tsankova N, Renthal W, Kumar A, Nestler EJ. Epigenetic regulation in psychiatric disorders. *Nat Rev Neurosci* (2007) **8**:355–67. doi:10.1038/nrn2132
68. Lyte M. Probiotics function mechanistically as delivery vehicles for neuroactive compounds: microbial endocrinology in the design and use of probiotics. *Bioessays* (2011) **33**:574–81. doi:10.1002/bies.201100024

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 02 October 2014; accepted: 12 January 2015; published online: 02 February 2015.

Citation: Gur TL, Worly BL and Bailey MT (2015) Stress and the commensal microbiota: importance in parturition and infant neurodevelopment. *Front. Psychiatry* **6**:5. doi: 10.3389/fpsy.2015.00005

This article was submitted to *Affective Disorders and Psychosomatic Research*, a section of the journal *Frontiers in Psychiatry*.

Copyright © 2015 Gur, Worly and Bailey. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.