

Draft Genome Sequences of the Three *Pectobacterium*-Antagonistic Bacteria *Pseudomonas brassicacearum* PP1-210F and PA1G7 and *Bacillus simplex* BA2H3

Slimane Khayi,^{a,b} Yannick Raoul des Essarts,^{a,c} Samuel Mondy,^a Mohieddine Moumni,^b Valérie Hélias,^{c,e} Amélie Beury-Cirou,^d Denis Faure^a

CNRS, Institut des Sciences du Végétal, UPR2355, Saclay Plant Sciences, Gif-sur-Yvette, France^a; Faculté des Sciences, Département de Biologie, Université Moulay Ismail, Meknès, Morocco^b; Fédération Nationale des Producteurs de Plants de Pomme de Terre-Recherche Développement Promotion du Plant de Pomme de Terre (FN3PT-RD3PT), Paris, France^c; Comité Nord Plant de Pomme de Terre (CNPPT), Semences, Innovation, Protection Recherche et Environnement (SIPRE), Achicourt, France^d; UMT Innoplant (FN3PT-INRAIGEPP1349), Le Rheu, France^e

***Pectobacterium* spp. are bacterial pathogens causing soft rot diseases on a wide range of plants and crops. We present in this paper the draft genome sequences of three bacterial strains, *Pseudomonas brassicacearum* PP1-210F and PA1G7 and *Bacillus simplex* BA2H3, which exhibit antagonistic activities against the *Pectobacterium* plant pathogens.**

Received 11 December 2014 Accepted 18 December 2014 Published 29 January 2015

Citation Khayi S, Raoul des Essarts Y, Mondy S, Moumni M, Hélias V, Beury-Cirou A, Faure D. 2015. Draft genome sequences of the three *Pectobacterium*-antagonistic bacteria *Pseudomonas brassicacearum* PP1-210F and PA1G7 and *Bacillus simplex* BA2H3. *Genome Announc.* 3(1):e01497-14. doi:10.1128/genomeA.01497-14.

Copyright © 2015 Khayi et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Denis Faure, faure@isv.cnrs-gif.fr.

Pectobacterium atrosepticum, *Pectobacterium wasabiae*, and *Pectobacterium carotovorum*, including *P. carotovorum* subsp. *brasilienis* and *carotovorum*, are worldwide pathogens responsible for blackleg and soft rot diseases on potato plants and tubers (1–3). Two biocontrol strategies against *Pectobacterium* phytopathogens have been developed, those of antibiosis and antivirulence (4). The biocontrol strain *Rhodococcus erythropolis* R138 targets the expression of the virulence functions in *Pectobacterium* spp. because of its capacity to disrupt the quorum-sensing signals *N*-acylhomoserine lactones (5). The genome sequence of the *R. erythropolis* antivirulence agent R138 was published recently (6). This antivirulence agent does not inhibit the growth of *Pectobacterium* spp. In contrast, we isolated three *Pectobacterium*-antagonistic bacteria, *Pseudomonas brassicacearum* PP1-210F and PA1G7 and *Bacillus simplex* BA2H3, which exhibit an ability to inhibit the growth of *Pectobacterium* strains *in vitro*. An assessment of their antagonistic abilities in the greenhouse and field settings is under way. Some other strains belonging to the *Pseudomonas fluorescens*, *P. brassicacearum*, and *B. simplex* species were previously described for their biocontrol activities against different microbial pathogens (7–10).

The genomic DNA of each bacterium was subjected to the next-generation Illumina HiSeq 2000 version 3 technology. A shotgun long jumping distance mate-pair library was constructed,

with an insert size of 8,000 bp. The sequencing of the library was carried out using a 2 by 100-bp paired-end read module by Eurofins Genomics (France). Assembly was performed by CLC Genomics 5.5 (CLC bio). The sequence reads were trimmed on quality (threshold 0.05), and minimal size (>60 nucleotides). Contigs were generated by *de novo* assembly (CLC parameters, automatic determination of the word and bubble sizes with no scaffolding). Scaffolding of the contigs was performed using SSPACE basic version 2.0 (11). For the finishing, automatic gap closure was processed using GapFiller version 1.11 (12). The remaining gaps were resolved by the mapping of mate pairs, using as a reference the 8 kb from each of the contig ends (read length, 0.9; identity, 0.95). Next, using homemade script and fastq select.tcl from the MIRA3 package, the mapped reads for both orientations (R1 and R2) were retrieved and *de novo* assembled (using the CLC parameters). The sequences were annotated using the Rapid Annotations using Subsystems Technology (RAST) pipeline (13). The detailed statistics for the three draft genome sequences are summarized in Table 1.

Nucleotide sequence accession numbers. The whole-genome shotgun projects for these bacteria have been deposited at DDBJ/EMBL/GenBank under the accession numbers [AYJR000000000](https://www.ncbi.nlm.nih.gov/nuccore/AYJR000000000) (*P. brassicacearum* PP1-210F), [AXBR000000000](https://www.ncbi.nlm.nih.gov/nuccore/AXBR000000000) (*B. simplex* BA2H3), and [JBON000000000](https://www.ncbi.nlm.nih.gov/nuccore/JBON000000000) (*P. brassicacearum* PA1G7). The versions de-

TABLE 1 Statistics for the 3 draft genome sequences

| Organism | Accession no. | Genome size (bp) | N_{50} (bp) | No. of contigs | No. of scaffolds | G+C content (%) | No. of CDSs ^a | No. of tRNAs | No. of rRNAs |
|-----------------------------------|--|------------------|---------------|----------------|------------------|-----------------|--------------------------|--------------|--------------|
| <i>B. simplex</i> BA2H3 | AXBR000000000 | 5,542,531 | 339,104 | 34 | 11 | 40.2 | 5,856 | 75 | 31 |
| <i>P. brassicacearum</i> PP1-210F | AYJR000000000 | 6,772,045 | 210,148 | 51 | 5 | 60.4 | 6,045 | 67 | 15 |
| <i>P. brassicacearum</i> PA1G7 | JBON000000000 | 6,789,417 | 301,959 | 53 | 8 | 60.5 | 6,052 | 57 | 13 |

^a CDSs, coding DNA sequences.

scribed in this paper are versions AYJR01000000 (*P. brassicacearum* PP1-210F), AXBR01000000 (*B. simplex* BA2H3), and JBON01000000 (*P. brassicacearum* PA1G7).

ACKNOWLEDGMENTS

S.K. received a Ph.D. grant from Paris-Sud University (Paris-Saclay University) and the Ministry of Higher Education of Morocco (no. H011/007); Y.R.D.E. received a Ph.D. grant from FN3PT-RD3PT and the Association Nationale de la Recherche et de la Technologie (ANRT-CIFRE no. 1282/2011).

This work was supported by cooperative projects between France and Morocco (PRAD 14-02, Campus France no. 30229 ZK), and between CNRS, FN3PT-RD3PT, and CNPPT-SIPRE. This project received a French State grant from LABEX Saclay Plant Sciences (reference ANR-10-LABX-0040-SPS) managed by the French National Research Agency under the Investments for the Future program (reference no. ANR-11-IDEX-0003-02).

REFERENCES

1. Pérombelon MCM. 2002. Potato diseases caused by soft rot erwinias: an overview of pathogenesis. *Plant Pathol* 51:1–12.
2. Duarte V, De Boer SH, Ward LJ, de Oliveira AMR. 2004. Characterization of atypical *Erwinia carotovora* strains causing blackleg of potato in Brazil. *J Appl Microbiol* 96:535–545. <http://dx.doi.org/10.1111/j.1365-2672.2004.02173.x>.
3. Pitman AR, Harrow SA, Visnovsky SB. 2010. Genetic characterisation of *Pectobacterium wasabiae* causing soft rot disease of potato in New Zealand. *Eur J Plant Pathol* 126:423–435. <http://dx.doi.org/10.1007/s10658-009-9551-y>.
4. Faure D, Dessaux Y. 2007. Quorum sensing as a target for developing control strategies for the plant pathogen *Pectobacterium*. *Eur J Plant Pathol* 119:353–365. <http://dx.doi.org/10.1007/s10658-007-9149-1>.
5. Cirou A, Mondy S, An S, Charrier A, Sarrazin A, Thoison O, DuBow M, Faure D. 2012. Efficient biostimulation of native and introduced quorum-quenching *Rhodococcus erythropolis* populations is revealed by a combination of analytical chemistry, microbiology, and pyrosequencing. *Appl Environ Microbiol* 78:481–492. <http://dx.doi.org/10.1128/AEM.06159-11>.
6. Kwasiborski A, Mondy S, Beury-Cirou A, Faure D. 2014. Genome sequence of the quorum-quenching *Rhodococcus erythropolis* strain R138. *Genome Announc* 2(2):e00224-14. <http://dx.doi.org/10.1128/genomeA.00224-14>.
7. Levenfors JP, Eberhard TH, Levenfors JJ, Gerhardson B, Hökeberg M. 2008. Biological control of snow mould (*Microdochium nivale*) in winter cereals by *Pseudomonas brassicacearum*, MA250. *BioControl* 53:651–665. <http://dx.doi.org/10.1007/s10526-007-9102-4>.
8. Ortet P, Barakat M, Lalaoua D, Fochesato S, Barbe V, Vacherie B, Santaella C, Heulin T, Achouak W. 2011. Complete genome sequence of a beneficial plant root-associated bacterium, *Pseudomonas brassicacearum*. *J Bacteriol* 193:3146. <http://dx.doi.org/10.1128/JB.00411-11>.
9. Zhou T, Chen D, Li C, Sun Q, Li L, Liu F, Shen Q, Shen B. 2012. Isolation and characterization of *Pseudomonas brassicacearum* J12 as an antagonist against *Ralstonia solanacearum* and identification of its antimicrobial components. *Microbiol Res* 167:388–394. <http://dx.doi.org/10.1016/j.micres.2012.01.003>.
10. Schwartz A, Ortiz I, Maymon M, Herbold C, Fujishige N, Vijanderaan J, Vilella W, Hanamoto K, Diener A, Sanders E, DeMason D, Hirsch A. 2013. *Bacillus simplex*—a little known PGPB with anti-fungal activity—alters pea legume root architecture and nodule morphology when coinoculated with *Rhizobium leguminosarum* bv. *viciae*. *Agronomy* 3:595–620. <http://dx.doi.org/10.3390/agronomy3040595>.
11. Boetzer M, Henkel CV, Jansen HJ, Butler D, Pirovano W. 2011. Scaffolding pre-assembled contigs using SSPACE. *Bioinformatics* 27: 578–579. <http://dx.doi.org/10.1093/bioinformatics/btq683>.
12. Boetzer M, Pirovano W. 2012. Toward almost closed genomes with GapFiller. *Genome Biol* 13:R56. <http://dx.doi.org/10.1186/gb-2012-13-6-r56>.
13. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.