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Polygenic Transmission and Complex Neuro developmental Network for Attention Deficit Hyperactivity Disorder: Genome-Wide Association Study of Both Common and Rare Variants

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Abstract

Attention-deficit hyperactivity disorder (ADHD) is a complex polygenic disorder. This study aimed to discover common and rare DNA variants associated with ADHD in a large homogeneous Han Chinese ADHD case-control sample. The sample comprised 1,040 cases and 963 controls. All cases met DSM-IV ADHD diagnostic criteria. We used the Affymetrix6.0 array to assay both single nucleotide polymorphisms (SNPs) and copy number variants (CNVs). Genome-wide association analyses were performed using PLINK. SNP-heritability and SNP-genetic correlations with ADHD in Caucasians were estimated with genome-wide complex trait analysis (GCTA).

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SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article at the publisher's web-site.

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Pathway analyses were performed using the Interval enRICHment Test (INRICH), the Disease Association Protein–Protein Link Evaluator (DAPPLE), and the Genomic Regions Enrichment of Annotations Tool (GREAT). We did not find genome-wide significance for single SNPs but did find an increased burden of large, rare CNVs in the ADHD sample ($P = 0.038$). SNP-heritability was estimated to be 0.42 (standard error, 0.13, $P = 0.0017$) and the SNP-genetic correlation with European Ancestry ADHD samples was 0.39 (SE 0.15, $P = 0.0072$). The INRICH, DAPPLE, and GREAT analyses implicated several gene ontology cellular components, including neuron projections and synaptic components, which are consistent with a neurodevelopmental pathophysiology for ADHD. This study suggested the genetic architecture of ADHD comprises both common and rare variants. Some common causal variants are likely to be shared between Han Chinese and Caucasians. Complex neurodevelopmental networks may underlie ADHD's etiology.

Keywords

ADHD; GWAS; pathway; neurodevelopment

INTRODUCTION

Attention-deficit hyperactivity disorder (ADHD) is a common behavioral disorder of childhood, affecting 3–6% of school-age children around the world [Faraone et al., 2003]. It has been viewed as a polygenic, multifactorial disorder. Both common and rare DNA variants contribute to its complex etiology [Poelmans et al., 2011; Stergiakouli et al., 2012; Williams et al., 2012].

Genome-wide association studies (GWAS) are hypothesis-free, interrogate all genes and regulatory regions of the genome and have the potential to discover novel risk genes. The first GWAS of ADHD performed by Neale et al. [2008] analyzed 438,784 SNPs in 909 Caucasian ADHD trios. Although none of the SNP association tests achieved genome-wide significance, the top-25 SNPs (based on P -value) implicated some interesting candidate genes, including cytoskeleton-organizer *DCLK1*, extracellular matrix component *SPOCK3*, cell-cell adhesion protein *CDH13*, as well as two potassium-channel regulators *KCNIP1* and *KCNIP4*. Using the same sample set, Lasky-Su et al. [2008] performed a quantitative genome-wide association analysis of ADHD symptoms. A high percentage (30/32, 94%) genes hit by the 58 SNPs with P values less than 10^{-5} were brain-expressed, including five related to transcription factors.

Meanwhile, Lesch et al. [2008] used independent DNA pools from 343 ADHD-affected adults and 304 controls for association analyses of the ADHD diagnostic phenotype. Of the 30 top-hit genes, seven were involved in cell adhesion/migration/neurogenesis (e.g., *CDH13*, *ASTN2*, *CSMD2*, *ITGAE*, *ITGA11*, *CDH23*, *SDK2*), two regulated synaptic plasticity (e.g., *CTNNA2*, *KALRN*), three were transcription factors (*MYT1L*, *TFEB*, *SUPT3H*), and one coded for a potassium channel (*KCNC1*) [Lesch et al., 2008].

Neale et al. [2010a] performed case-control analyses in 896 cases with DSM-IV ADHD and 2,455 controls. A consensus dataset of 1,033,244 SNPs was imputed (using the HapMap

Phase III European CEU and TSI samples as the reference). No genome-wide significant associations were found. The most significant results implicated *PRKG1*, *FLNC*, *TCERGIL*, *PPM1H*, *NXPPI*, ^{CDH13}, *HK1*, and *HKDC1*. Combining data from four ADHD GWAS projects, Neale et al. [2010b] performed a meta-analysis in a sample of 2,064 trios, 896 cases, and 2,455 controls. Even with this much larger sample size, no genome-wide significant associations were found. One reason for this is that the samples were underpowered to estimate effect sizes of common variants of small effect. This has been confirmed by analyses that estimate the variance contributed by common variants all together. Genome-wide complex trait analysis (GCTA) [Yang et al., 2010, 2011; Lee et al., 2011] applied to ADHD samples (4,163 cases and 12,040 controls) from the Psychiatric Genomics Consortium, estimated SNP chip heritability to be 0.28 (SE 0.02; Psychiatric GWAS Consortium ADHD Group. Paper submitted for publication).

Copy number variations (CNVs) have also been implicated in the etiology of ADHD. Elia et al. [2010] found that inherited rare CNVs in an ADHD sample were significantly enriched for genes known to be important for psychological and neurological functions, including learning, behavior, synaptic transmission, and central nervous system development. Williams et al. [Williams et al., 2010, 2012] found an increased burden of large, rare CNVs and reported excess of chromosome 16p13.11 and 15q13.3 duplications and an overlap between CNVs reported for ADHD and autism spectrum disorders. Elia et al. [2012] further showed that CNVs affecting the metabotropic glutamate receptor genes *GRM5*, *GRM7*, *GRM8*, and *GRM1* were enriched across several independent samples.

In summary, although ADHD is acknowledged to be a genetic disorder, GWAS has not revealed any common SNP variants with genome-wide significance. This study used both common and rare variants, using polygenic and pathway analyses, to evaluate the genetic etiology of ADHD in a large homogenous Han Chinese case-control sample.

MATERIALS AND METHODS

Participants

One thousand and forty ADHD cases (876 boys, 84.2%) aged between 6 and 16 years [average (9.7 ± 2.4) years] were recruited from the Child and Adolescent Psychiatric Outpatient Department of the Sixth Hospital, Peking University. All cases met DSM-IV ADHD diagnostic criteria. A clinical diagnosis was first made by a senior child and adolescent psychiatrist based on the parent and teacher completed ADHD Rating Scale-IV (ADHD-RS-IV), and then confirmed by semi-structured interview with the parents and child using the Chinese version of the Clinical Diagnostic Interview Scale [Barkley, 1998; Yang et al., 2004]. Those with major neurological disorders (e.g., epilepsy), schizophrenia, pervasive development disorder, and mental retardation ($IQ < 70$) were excluded. The sample consists of 680 (65.4%) ADHD combined type and 360 (34.6%) inattentive type. The comorbidities included oppositional defiant disorder (ODD) in 380 patients (36.5%), conduct disorder in 58 (5.6%), and tic disorder in 167 (16.1%).

Nine hundred sixty three controls were students from local elementary schools, healthy blood donors from the Blood Center of the First Hospital, Peking University, and healthy

volunteers from our institute. Six hundred and eight were males (63.1%). The average age was (15.4 ± 8.8) years. Parents or adults themselves completed the ADHD Rating Scale-IV (ADHD RS-IV) to exclude ADHD. Major psychiatric disorders, family history of psychosis, severe physical diseases, and substance abuse were also excluded according to a medical history report form. All the cases and controls were of Han Chinese decent.

The study was approved by the Institutional Review Board of the Peking University Health Science Center. After complete description of the study to the subjects, written informed consent was obtained from parents of the ADHD probands.

Genotyping

Both cases and controls were genotyped using the Affymetrix6.0 array at CapitalBio Ltd. (Beijing) using the standard Affymetrix protocol. Samples of cases and controls were added in equal proportion to each chip to avoid batch effects. The Affymetrix 6.0 array included 906,600 SNP probes and 946,000 CNV probes. The SNP genotypes were called with BIRDSEED v2, while CNVs were called with Genotyping Console (GTC) 4.0 using default parameters. A total of 2003 cases and controls passed the first stage sample control with call rates >98%, no first or second-degree relative relationships, and genders consistent with site reports.

Data Quality Control and Statistical Analysis

Data quality control and association analysis were performed using PLINK 1.07 [Purcell et al., 2007, <http://pngu.mgh.harvard.edu/purcell/plink/>]. For inclusion of SNPs we required: call rate >95%, MAF >1%, and HWE P -value $>10^{-6}$. After data cleaning, there were 656,051 SNPs for the association analyses. To examine population stratification, we performed multi-dimensional scaling (MDS). In the pair-wise MDS plot for 10 dimensions, the majority of subjects were tightly clustered, suggesting no substantial population stratification (SF1). We then conducted logistic regression to adjust the association P -value, using the 10 principal components from the MDS procedure as covariates.

CNV calling only included segments larger than 100 kb, spanning at least 10 consecutive, informative SNPs. Quality control for samples excluded 136 individuals (71 cases, 65 controls) who carried more than 40 apparent CNVs. Analysis focused on rare CNVs with frequency <1%. We used the human reference sequence of NCBI Build 36.1 - hg18 to filter known segmental duplications.

Known common CNVs defined by the Genome Structural Variation Consortium (http://projects.tcag.ca/variation/ng42m_cnv.php) and known gaps of at least 200 kb in the SNP array were also filtered. Burden analysis counted the number of total CNVs, deletions and duplications in cases and controls, calculated the CNV rate, as well as percent of cases and controls that carried rare CNVs. The significance of CNV differences between cases and controls was assessed by permutation test with 50,000 replicates.

Polygenic analyses

To investigate the contribution of common SNPs to variation in liability to ADHD, we estimated the SNP-heritability using GCTA [Yang et al., 2011]. A non-zero heritability is estimated if cases are genetically more similar to other cases than they are to controls [Lee et al., 2011]. We removed individuals such that no pair had genetic similarity relationship >0.05 (as this may inflate estimates unfairly), so that 1,010 cases and 917 controls remained. We used Caucasian samples from the Psychiatric Genomics Consortium for ADHD (4,163 ADHD cases and 12,040 controls) and a bivariate model of analysis [Lee et al., 2012b] to estimate the SNP-genetic correlation between Han Chinese and Caucasians for liability to ADHD. Since the SNP frequencies differ between ethnic groups the additive genetic similarities between individuals i and j were estimated as

$$A^{ij} = \frac{1}{L} \sum_{l=1}^L (x_{il} - 2p_{is,l}) \frac{x_{jl} - 2p_{js,l}}{\sqrt{2p_{is,l}q_{is,l}2p_{js,l}q_{js,l}}} \quad (i \neq j)$$

for the L SNPs with minor allele frequency >0.01 and imputation $R^2 > 0.6$ ($L = 917,066$), where i s represent a population that individual i belong to and p and $q = 1 - p$ are allele frequencies of the first and other allele and x_{il} is the number of first alleles for the l th SNP in individual i . The analysis model include sex, cohort and 20 ancestry principal components are covariates.

Pathway analysis

To determine if any neurobiological pathways were implicated by our association signals, we input our top hit intervals from the SNP and CNV association analyses into Interval enrichment Test (INRICH [Lee et al., 2012a]). Associated intervals for SNPs included those with P -values $<10^{-4}$ after correcting for the MDS components. The SNP tagging function in PLINK was used to generate LD independent genomic intervals (tag r^2 : 0.2, tag kb: 1,000). We included CNV intervals that were more prevalent in cases than in controls with at least a trend difference of statistical significance ($P < 0.15$). We used the Gene Ontology (GO) nodes as our target gene sets. After size filtering, 5,237 target gene sets (nodes) each comprising at least three genes were examined. Interval overlap was limited to 20 kb up/downstream of a gene. The number of overlapping genes was recorded as $Real_i$. Ten thousand replicates generated random interval sets each matching to the number of associated intervals. The empirical gene-set P -value equals the percent of replicates with at least $Real_i$ number of random intervals overlapping with genes in a target gene set. Bootstrapping-based re-sampling was used for multiple testing to correct the empirical gene-set P -value over all gene sets.

To explore potential physical interactions among proteins encoded in associated intervals, we used a second method for pathway analysis, that is, Disease Association Protein-Protein Link Evaluator (DAPPLE) [Rossin et al., 2011]. In consideration of the contributions of both common and rare variants to the etiology of ADHD, and that both might separately capture nodes in the ADHD pathogenesis network, we used the same genomic intervals for both SNPs and CNV that we used for INRICH. DAPPLE uses experimentally validated,

protein–protein interaction databases to identify direct and indirect networks from associated proteins and scores network and protein connectivity. We built 10,000 random networks and compared these with the ADHD associated networks to determine if the connectivity of the ADHD networks and each seed protein was greater than expected by chance.

The third pathway analyses we used the Genomic Regions Enrichment of Annotations Tool (GREAT, [McLean et al., 2010] to assess for enrichment of cis-regulatory regions. GREAT examines not only proximal but also distal regulatory regions up to 1 Mb upstream or downstream of transcription start sites. In addition to typical calculation of gene-based P -values for enrichment, GREAT computes a binomial test over genomic regions, which uses the fraction of the genome associated with each ontology term as the probability of selecting the term. This method explicitly accounts for the variability in length of gene regulatory domains, eliminating the bias that leads to false positive enrichments for distal regulatory regions.

RESULTS

Single Variant Analyses

The quantile-quantile (QQ) plot (SF2) for SNPs' association was almost completely diagonal. The lambda statistic (λ) was 1.02. The distribution of observed P -values did not deviate from the distribution expected under the null hypothesis of no association. The corrected Manhattan plot is shown in SF3. The lowest P values were about 10^{-5} to 10^{-6} . The SNPs associated with P values of 10^{-5} or lower are listed in Supplementary Table SI. All hit genes were expressed in brain. Most of them were known to be involved in neurodevelopment (including cell adhesion, neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity: *ITGAI*, *NYAP2*, *ADAM28*, *CNTN2*, *LRFN2*, *NTM*, *GJA1*, *FLRT2*, *PRKG1*, *PICK1*, *CAMK2G*; glutamate receptor and transporter: *GRIK4*, *GRM7*, *SLC38A1*; and related transcription factors: *PAPOLA*, *MED27*, *TAF2*, *ZNF516*).

We included 3,460 rare CNVs (1,817 in cases and 1,643 in controls) in the analyses, with all segments intersecting with one or more genes (hg18). Burden analyses showed a significantly higher rate of rare CNVs (1.875% vs. 1.830%, ratio: 1.02, $P = 0.038$) and proportion of individuals carrying rare CNVs (55.8% vs. 51.2%, ratio: 1.09, $P = 0.026$) for the ADHD group than for controls. Association analyses found six regions nominally associated with ADHD ($P < 0.05$, with 50,000 permutation tests), though none of them survived genome-wide correction (Supplementary Table SII).

Polygenic Analyses

The estimate of the SNP-heritability calculated in the bivariate analysis was 0.42 (SE 0.13) for the Han Chinese sample (h^2_{SNP-HC}). A maximum likelihood ratio test of $H_0: h^2_{SNP} = 0$. In ancestry the bivariate analysis the SNP-heritability for the European sample (h^2_{SNPEA}) was 0.28 (SE 0.02, $P = 0$), in close agreement (as expected) with the univariate estimate PGC Cross Disorder Group, paper in submission. The estimate of the SNP-genetic correlation between Chinese and European samples (r_{g-SNP}) was 0.39 (SE 0.15, $P = 0.0072$).

Pathway Analyses

Interval enrichment tests of the most significantly associated SNPs found 23 pathways enriched for associated signals (Table I). Although none of these achieved significance after correcting for multiple comparisons, many implicated neurobiological functions potentially relevant to ADHD, e.g. neuron projection morphogenesis (*ITGAI*, *GJAI*), neuron migration (*PRKG1*, *GJAI*), endocytic vesicle membrane (*PICK1*, *CAMK2G*), synaptic transmission (*PICK1*, *CAMK2G*, *SLC38A1*, *GRM7*). Pathways related to transcription were observed, that is, transcription initiation from RNA polymerase II promoter (*MED27*, *TAF2*). Interval enrichment tests of CNVs found 9 pathway nominal significant at $P < 0.05$. None achieved significance after correcting for multiple comparisons (Table I). Most were related to transmembrane transport, including water, sodium and potassium ion transport.

DAPPLE identified 16 direct connections among proteins in 152 associated regions (Table II). Compared to 10,000 random networks, the associated network (SF4) is significantly enriched for direct connectivity (16 vs. 9.7, $P = 0.030$). The connected proteins formed six groups. Their functions involved cell adhesion/synaptic formation/plasticity, especially for glutamatergic synaptic plasticity, as well as related transcription factors. For each seed protein, taking the best of the direct and indirect scores and correcting for the number of tests as well as for the number of genes in one locus, we identified seven genes significant for connectivity to be candidate genes for future research: *NCL* ($P = 2 \times 10^{-4}$), *KCNH7* ($P = 8 \times 10^{-4}$), *NXPPI* (1×10^{-3}), *LANCLI* (6×10^{-3}), *CNTNAP2* (9×10^{-3}), *SV2C* (1.2×10^{-2}), and *PICK1* (4×10^{-2}).

Using the same set of associated SNPs and CNVs for GREAT analyses, we found significant enrichment for 6 GO Cellular Component terms after correcting for multiple comparisons (Table III). The six terms were from two GO branches and their child nodes (Fig. 1): synapse (15 genes hit, FDR Q-val: 0.0055; three child nodes were also significant: synapse part, synaptic membrane, and presynaptic membrane) and neuron projection (16 genes hit, FDR Q-val: 0.013; one child node was also significant: axon).

DISCUSSION

This GWAS of ADHD, comprising 1,040 cases and 963 controls, is the first performed in a homogeneous Han Chinese population. Although we did not find any genome-wide significant SNP or CNV variants, we did find significant evidence for a polygenic SNP component and an increased burden of rare CNVs.

The significant SNP-heritability implies that common variants are associated with ADHD, but that our sample is underpowered to detect them at the stringent significance level imposed by the genome-wide burden of multiple testing. The SNP-heritability in Han Chinese was 0.42 (SE 0.15). Although the point estimate is higher than for the larger European ancestry sample from the PGC-ADHD, 0.28 (SE 0.02), its high standard error shows that the estimates are not significantly different. The estimate of the SNP-genetic correlation (r_{g-SNP}) was 0.39 (SE 0.15, $P = 0.0072$), which indicates that common SNP risk variants are shared by the Han Chinese and European Ancestry samples. To our knowledge, this is the first such correlation reported for any disease or disorder. The significant

correlation indicates that ancient common variants associated with ADHD are shared between the ethnic groups.

However, the point estimate of the SNP correlation between Han Chinese and European Ancestry samples is lower than between sub-samples of the European Ancestry cohort.

Specifically, when the PGC-ADHD data was split into two sub-samples, the h^2_{SNP-EA} estimates were 0.21 (SE 0.07) for the first sample and 0.41 (SE 0.03) for the other sample with a genetic correlation of 0.71 (SE 0.17) implying, as expected, more sharing of associated variants and/or higher linkage disequilibrium between causal variants and SNPs within than between ethnic populations.

Despite the fact that no individual SNPs reached association at genome-wide significance, our most significant findings implicated genes participating in neurodevelopmental processes such as cell adhesion, neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity. Similar sets of genes were also suggested by previous ADHD GWAS and a meta-analysis (see Supplementary Table SI). For example, PRKG1 was implicated by Neale et al. [2010a], ITGA1, CAMK2G, CAMK1D were implicated in the meta-analysis by Neale et al. [2010b], and ITGAE and ITGA11 were implicated by Lesch et al. [2008]. Some of our top genes code for glutamate receptors and transporters. The same genes and gene family members (GRM7, GRIK1) were reported in the quantitative GWAS by Lasky-Su et al. [2008], the meta-analysis by Neale et al. [2010b], and the genome-wide CNV study by Elia et al. [2012]. Some genes related to transcription (ZNF544, ZNF385D, ZNF423, ZNF516, ZNF75A, DMRT2, FHIT, FOXP1, and MEIS2) were also implicated by Lasky-Su et al. [2008] and by Neale et al. [2010b]'s meta-analysis.

Although not significant after correcting for multiple comparisons, the pathways revealed by the INRICH analyses of associated SNPs involved neurobiological functions consistent with the prior findings discussed above. For example, neuron projection morphogenesis and neuron migration pathways were implicated by genes encoding adhesion molecules (e.g., GJA1, ITGA1, PRKG1). Neuron migration and axon guidance toward the target in the development of the nervous system involve interactions between molecules on the surface of the axon and those in the extra-cellular matrix [Tsiotra et al., 1993]. The endocytic vesicle membrane and synaptic transmission pathways involve glutamatergic synaptic function. The transcription related pathway is a ubiquitous biological process, if, as our findings suggest, it is implicated in ADHD's pathophysiology, any defects in the implicated transcription network must require other etiologic factors to lead to a pathophysiologic state.

Because the GO "pathways" used by INRICH are based on bibliometric gene annotations rather than experimental data, we also used DAPPLE, which is based only on experimentally documented physical interactions among proteins. Considering the complexity of the genetic basis of ADHD, we hypothesized that both common and rare variants contribute to the disorder and act on similar functional classes of genes [Poelmans et al., 2011; Stergiakouli et al., 2012]. The DAPPLE analyses showed that the proteins implicated by our GWAS were significantly more likely to be interconnected with one another than expected by chance, suggesting that risk variants might exist in suites of genes involved in the underlying biological process of protein-protein interaction networks. The

DAPPLE results are consistent with the INRICH results implicating three pathways: cell adhesion (*NXP1–NRXN1*, *CNTN2–CNTNAP2–ZMIZ1*), glutamate synaptic development (*GRM7–PICK1–GRM3*, *PICK1–EPHA7*), and the transcription pathway (*TAF2–PAPOLA–POLR2F–MED27–MED20*, *POLR2F–NCL–RSL1D1–BYSL*).

Using the regulatory annotation of associated signals, GREAT depicted a clearer outline of associated genes, which encoded proteins comprising neuronal cellular components from the Neuron Projection and Synapse branches of the GO tree. Most of the genes from these pathways were consistent with the INRICH and DAPPLE findings; they encode adhesion molecules, glutamate receptors and proteins involved in axon and synapse development (Supplementary Table SIII).

All the above pathways are consistent with the hypothesis that mis-wiring of the brain during neurodevelopment might cause ADHD. Similar conclusions were drawn by Lesch et al. [2008] and Franke et al. [2009] based on the findings from existing GWAS, which suggested that neuronal spine formation and plasticity might underlie the pathophysiology of ADHD. Consistent with these ideas, a recent integration of ADHD GWAS findings found significant evidence for a neurodevelopmental network of directed neurite outgrowth [Poelmans et al., 2011]. Although our findings are consistent with prior work, they also provide evidence for a more comprehensive network, involving neuron migration, neurite outgrowth, neuronal morphogenesis, and synaptic plasticity, especially glutamatergic synaptic development. The glutamate system is a reasonable candidate for ADHD's pathophysiology as glutamate is the major excitatory neurotransmitter in the central nervous system, and regulates the catecholaminergic activity which has been implicated in ADHD by neurobiological [Scassellati et al., 2012] and treatment [Faraone and Glatt, 2010] studies.

Although our findings are intriguing, we have only captured fragments of the puzzle of ADHD's etiology in this study. We could not paint the full picture. Our work must be considered in the context of its limitations. We had no genome-wide significant findings for any single variant, which might be due to the sample size. However, our bioinformatic and pathway analyses found some interesting genes and neurobiological pathways which implicate complex neurodevelopmental network underlying ADHD. Our finding of a significant polygenic component suggests that there are many common SNP variants with small effect sizes that increase the risk for ADHD. Individually, these SNPs will be difficult to detect with currently available sample sizes.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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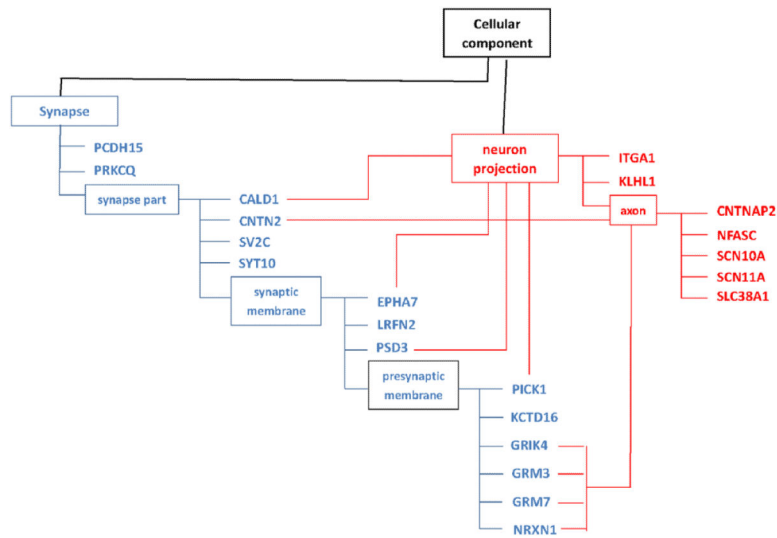


FIG. 1. Neurodevelopmental network predicted by proximal and distal regulatory region among the top hit of genome-wide SNPs and CNVs association.

TABLE I

Pathways Enriched for Associated SNPs and CNVs by INRICH Test

Target	Target size	Interval no.	Emp. P	Cor. P	Gene list
Pathways enriched for associated SNPs ^a					
GO: 0009268 Response to pH	12	2	0.00009999	0.14917	ARSB, GJA1
GO: 0043403 Skeletal muscle tissue regeneration	9	2	0.00019998	0.192162	PLAU, GJA1
GO: 0048812 Neuron projection morphogenesis	18	2	0.00029997	0.231354	ITGA1, GJA1
GO: 0007160 Cell-matrix adhesion	72	3	0.00049995	0.295941	VCL, ITGA1, BCL2L11
GO: 0005916 Fascia adherens	9	2	0.00079992	0.378724	VCL, GJA1
GO: 0006936 Muscle contraction	93	3	0.00109989	0.447111	VCL, ITGA1, GJA1
GO: 0030666 Endocytic vesicle membrane	24	2	0.00269973	0.661668	PICK1, CAMK2G
GO: 0006367 Transcription initiation from RNA polymerase II promoter	67	2	0.00379962	0.727854	MED27, TAF2
GO: 0005741 Mitochondrial outer membrane	85	2	0.00679932	0.847231	GJA1, BCL2L11
GO: 0007229 Integrin-mediated signaling pathway	58	2	0.010399	0.894421	ITGA1, ADAMDEC1
GO: 0005178 Integrin binding	64	2	0.0114989	0.907419	ITGA1, ADAMDEC1
GO: 0007268 Synaptic transmission	266	4	0.0121988	0.915217	PICK1, CAMK2G, SLC38A1, GRM7
GO: 0005764 Lysosome	154	2	0.0157984	0.946011	ARSB, GJA1
GO: 0030165 PDZ domain binding	54	2	0.019898	0.965807	GRM7, GJA1
GO: 0015293 Symporter activity	112	2	0.019898	0.965807	SLC38A1, SLC16A8
GO: 0005624 Membrane fraction	467	4	0.0243976	0.982004	ITGA1, SLC16A8, PSD3, BCL2L11
GO: 0001764 Neuron migration	59	2	0.0255974	0.983003	PRKG1, GJA1
GO: 0045121 Membrane raft	110	2	0.0256974	0.983203	ITGA1, GJA1
GO: 0006814 Sodium ion transport	118	2	0.0265973	0.984003	SLC38A1, SCN9A
GO: 0005654 Nucleoplasm	732	4	0.0214979	0.968806	MED27, CAMK2G, DSSC1, PAPOLA
GO: 0017124 SH3 domain binding	101	2	0.0357964	0.992402	BAIAP2L2, GJA1
GO: 0043234 Protein complex	152	2	0.0437956	0.995601	PICK1, VCL
GO: 0001701 In utero embryonic development	142	2	0.0459954	0.996601	GJA1, BCL2L11
Pathways enriched for associated CNVs ^b					
GO: 0006833 Water transport	36	2	0.00549945	0.508498	AQP9, ADCY8
GO: 0005244 Voltage-gated ion channel activity	149	4	0.0115988	0.734653	KCNH7, KCNQ1, SCN10A, SCN11A
GO: 0055085 Transmembrane transport	619	6	0.0125975	0.752849	KCNH7, SCN10A, KCNQ1, AQP9, ADCY8, SCN11A
GO: 0042493 Response to drug	241	3	0.0164984	0.834233	USP47, CPS1, SCN11A
GO: 0005248 Voltage-gated sodium channel activity	15	2	0.0172983	0.84883	SCN10A, SCN11A
GO: 0001518 Voltage-gated sodium channel complex	12	2	0.0172983	0.84883	SCN10A, SCN11A
GO: 0006814 Sodium ion transport	118	2	0.0258974	0.909018	SCN10A, SCN11A
GO: 0006811 Ion transport	565	5	0.030397	0.928814	KCNH7, KCNQ1, KCNT2, SCN10A, SCN11A

Target	Target size	Interval no.	Emp. P	Cor. P	Gene list
GO: 0006813Potassiumiontransport	153	3	0.0392961	0.956809	KCNH7, KCNQ1, KCNT2

^aWith corrected P -value $<10e-4$.

^bIncluding CNVs more in cases than in controls with $P < 0.15$.

Direct Connections Between Proteins Encoded by Genes Overlapped With Potential Associated SNPs and CNVs

Direct connection	Associated SNP/interval	Gene function ^a	Gene and pathway implicated in other studies of ADHD or psychiatric disorders
	<p><i>NXP1</i>: chr7: 8471930-8567358 <i>NRXN1</i>: chr2: 50226778-50323795</p>	<p>NXP1: The encoded protein forms a very tight complex with alpha neuurexins, a group of proteins that promote adhesion between dendrites and axons NRXN1: The encoded protein functions as cell adhesion molecules and receptors. May play a role in formation or maintenance of synaptic junctions</p>	<p>Both <i>NXP1</i> and <i>NRXN1</i> were among the top hit genes [10^{-5}] in ADHD GWAS by Neale et al. [2010a]. CNVs of <i>NXP1</i> were found in ASD families [Salyakina et al., 2011]. CNVs of <i>NRXN1</i> were also observed in schizophrenia and autism [Doherty et al., 2012] <i>CNTNAP2</i> has been implicated in multiple neurodevelopmental disorders, including Gilles de la Tourette syndrome, schizophrenia, epilepsy, autism, ADHD and mental retardation. <i>CNTNAPS</i> was among the top hit genes in ADHD GWAS by Neale et al. [2010a]. <i>ZMIZ1</i> was among the top hit genes in Neale et al. [2008]</p>
	<p><i>CNTN2</i>: rs2802837 <i>CNTNAP2</i>: chr7: 14563708-14567817 Chr7: 145755346-145813647 <i>ZMIZ1</i>: chr10: 80350172-80476638</p>	<p>CNTN2: It is a neuronal membrane protein that functions as a cell adhesion molecule. It may play a role in the formation of axon connections in the developing nervous system CNTNAP2: This gene encodes a member of the neuexin family which functions as cell adhesion molecules and receptors. This protein is localized at the juxtaparanodes of myelinated axons, and mediates interactions between neurons and glia during nervous system development and is also involved in localization of potassium channels within differentiating axons ZMIZ1: The encoded protein regulates the activity of various transcription factors</p>	<p>Cell adhesion molecule had been reported in previous ADHD GWAS studies [Lasky-Su et al., 2008; Zhou et al., 2008; Neale et al., 2010a]</p>
	<p><i>SCN10A</i>: chr3: 38787799-38889198 <i>GJA1</i>: rs7753979 <i>TNNT2</i>: chr1:199488134-199580414</p>	<p>SCN10A: Voltage-gated sodium channels are integral membrane glycoproteins that are responsible for the initial rising phase of action in most excitable cells GJA1: This gene is a member of the connexin gene family. The encoded protein is a component of gap junctions. Gap channels allow electrical and biochemical coupling between cells and in excitable tissues, such as neurons and heart. TNNT2: The protein encoded by this gene is the troponin-binding subunit of the troponin complex, which is located on the thin filament of striated muscles and regulates muscle contraction in response to alterations in intracellular calcium ion concentration. This gene expresses highest in the heart, but also expresses in the brain</p>	<p>4 SNPs of <i>SCN10A</i> showed nominal association with ADHD in the meta-analysis by Neale et al. [2010b], with the lowest <i>P</i>-value = 0.022 for rs7430438 rs7740467, which is approximately 3 kb upstream of <i>GJA1</i>, was found nominal significant ($P = 0.0204$) in the meta-analysis of ADHD GWAS by Neale et al. [2010b]. 8 SNPs of <i>TNNT2</i> showed nominal association with ADHD in the meta-analysis by Neale et al. [2010b], with the lowest <i>P</i>-value = 0.015 for rs10800775</p>
	<p><i>GRM7</i>: rs13317247 <i>GRM3</i>: chr7: 86132321-86244019 <i>PICK1</i>: rs8142185 <i>EPHA7</i>: chr6: 94194161-94216651</p>	<p>GRM7 and GRM3: <i>L</i>-glutamate is the major excitatory neurotransmitter in the central nervous system, and it activates both ionotropic and metabotropic glutamate receptors. Glutamatergic neurotransmission is involved in most aspects of normal brainfunction and can be perturbed in many neuropathologic conditions. <i>GRM3</i> belongs to the metabotropic glutamate receptors Group II, while <i>GRM7</i> belongs to Group III PICK1: The protein encoded by this gene has been shown to interact with multiple glutamate receptor subtypes, Probable adapter protein that bind to and organize the subcellular localization of a variety of membrane proteins containing some PDZ recognition sequence. Involved in the clustering of various receptors, possibly by acting at</p>	<p>More than 20 SNPs of <i>GRM7</i> showed association with ADHD in the meta-analysis by Neale et al. [2010b], with the lowest <i>P</i> value = 2.96E-3 for rs1532544. CNVs of <i>GRM7</i> were found in a Genome-wide copy number variation study of ADHD [Ella et al., 2012]. rs17051835, rs12491620, rs1450099, and rs3749380 of <i>GRM7</i> were associated with SCZ [Gandia et al., 2009; Ohtsuki et al., 2008; Shibata et al., 2009]. <i>GRM3</i> and <i>EPHA3</i> were identified to be candidate genes for ASD [Casey et al., 2012]. <i>GRM3</i> was associated with SCZ in some candidate gene studies [Cherlyn et al., 2010]. It was also related to psychosis and relapse in bipolar disorder [Dalvie et al., 2010]. <i>PICK1</i> located in the linkage and association region</p>

TABLE III

Significant Enriched Gene Ontology Terms by Genes Associated With Regulatory Regions

Term name	Binom ^a rank	Binom raw P-value	Binom FDR Q-Val	Binom fold enrichment	Binom observed region hits	Binom region set coverage (%)	Hyper ^a rank	Hyper FDR Q-Val	Hyper fold enrichment	Hyper observed gene hits	Hyper total genes	Hyper gene set coverage (%)
Presynaptic membrane	1	2.4669E-11	2.7654E-8	9.4056	16	10.53	1	5.6208E-3	13.5654	6	50	3.85
Synaptic membrane	2	4.2028E-11	2.3557E-8	4.7787	26	17.11	5	1.3568E-2	5.2175	9	195	5.77
Synapse part	4	1.2576E-9	3.5243E-7	3.5619	30	19.74	2	6.9311E-3	4.2597	13	345	8.33
Synapse	5	6.4704E-8	1.4507E-5	2.8497	32	21.05	3	5.4656E-3	3.6862	15	460	9.62
Axon	11	4.2006E-5	4.2808E-3	2.9590	18	11.84	7	1.4342E-2	4.4506	10	254	6.41
Neuron projection	13	1.5147E-4	1.3062E-2	2.0886	28	18.42	4	1.2807E-2	3.1788	16	569	10.26

^a“Binom” represented binomial test over genomic regions; “Hyper” represented hypergeometric test over genes.