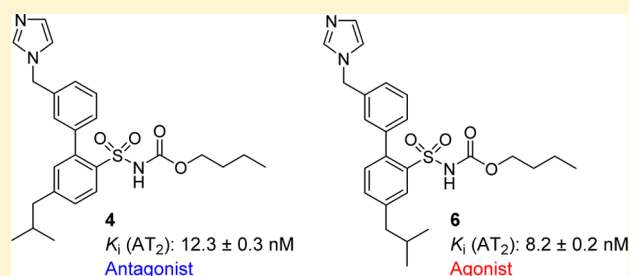


Interconversion of Functional Activity by Minor Structural Alterations in Nonpeptide AT₂ Receptor LigandsCharlotta Wallinder,[†] Christian Sköld,[†] Milad Botros,[‡] Marie-Odile Guimond,[§] Mathias Hallberg,[‡] Nicole Gallo-Payet,[§] Anders Karlén,[†] and Mathias Alterman^{*,†}[†]Organic Pharmaceutical Chemistry, Department of Medicinal Chemistry, BMC, Uppsala University, SE-751 23 Uppsala, Sweden[‡]Beijer Laboratory, Department of Pharmaceutical Biosciences, BMC, Uppsala University SE-751 23 Uppsala, Sweden[§]Service of Endocrinology, Faculty of Medicine and Health Sciences, University of Sherbrooke, Sherbrooke J1H 5N4, Quebec, Canada

Supporting Information

ABSTRACT: Migration of the methylene imidazole side chain in the first reported selective drug-like AT₂ receptor agonist C21/M024 (**1**) delivered the AT₂ receptor antagonist C38/M132 (**2**). We now report that the AT₂ receptor antagonist compound **4**, a biphenyl derivative that is structurally related to **2**, is transformed to the agonist **6** by migration of the isobutyl group. The importance of the relative position of the methylene imidazole and the isobutyl substituent is highlighted herein.

KEYWORDS: AT₂ receptor, nonpeptide ligands, agonist, antagonist, C21/M024, C38/M132



The sartans (angiotensin II receptor blockers, ABRs) have been on the market as antihypertensives for more than two decades and act as selective antagonists at the angiotensin II AT₁ receptor. More recently, the angiotensin AT₂ receptor has emerged as a new target for drug intervention.^{1–3} In 2004, the first selective and drug-like AT₂ receptor agonist C21/M024 (**1**) was disclosed,⁴ and thereafter, a series of structural analogues with agonistic properties have been reported.^{5,6} The activation of the AT₂ receptor is known to render a large number of diverse biological responses.^{7,8} For example, receptor activation stimulates neurite outgrowth in neuronal cells, which is one of the first steps in neuronal differentiation.⁹

In healthy adults the AT₂ receptor expression is low and mainly found in renal, cardiovascular, and brain tissues.^{7,8} Notably, during certain pathological conditions, such as myocardial infarction, vascular injury, brain ischemia, renal failure, and cutaneous wounds, the AT₂ receptor expression is significantly up-regulated.^{7,8}

Recently, several studies conducted in experimental disease models have revealed that the selective potent AT₂ receptor agonist **1** acts as an anti-inflammatory agent and exerts pronounced tissue protective effects in particular in cardiovascular and renal disease. Hence, the compound has had a large impact on the view of the AT₂ receptor as a drug target.^{1,9–14} In addition, the AT₂ receptor is a promising drug target for antagonists. Thus, the AT₂ receptor antagonist EMA401 has demonstrated convincing efficacy data in patients suffering from neuropathic pain (postherpetic neuralgia) in a phase II study.³

We discovered that migration of the methylene imidazole substituent of **1** from the para to the meta substitution pattern delivered a selective AT₂ receptor antagonist, compound

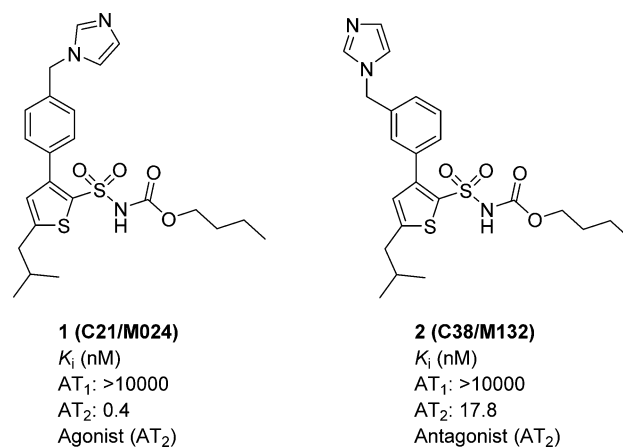


Figure 1. Structures of compounds **1** and **2**.

C38/M132 (**2**) (Figure 1).^{15,16} Thus, a minor structural alteration resulted in the interconversion of a selective AT₂ receptor agonist to a selective AT₂ receptor antagonist.

Herein, we report the synthesis and biological evaluation of four AT₂ receptor ligands, the biphenyl derivatives, and regioisomers, **3**, **4**, **5**, and **6** (Figure 2), and that the relative position of the imidazole and isobutyl substituent determines the functional outcome.

The nonselective Ang II receptor ligand **7** (L-162,782) acts as an AT₁ receptor agonist and **8** (L-162,389) acts as an AT₁

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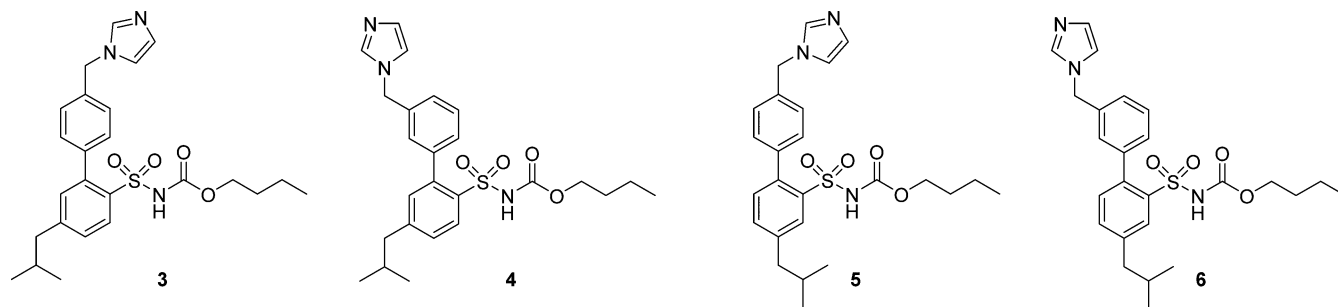


Figure 2. Structures of the regioisomers 3, 4, 5, and 6.

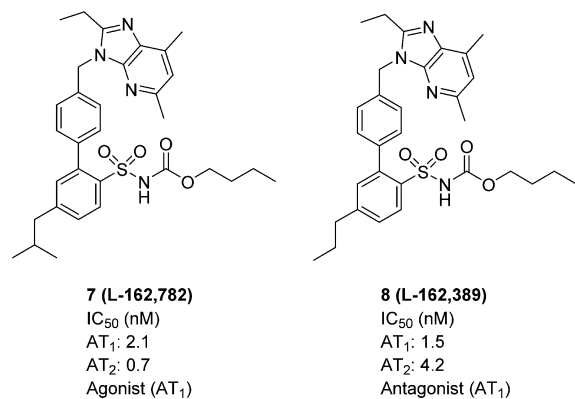


Figure 3. Structures of compounds 7 and 8.

receptor antagonist (Figure 3).¹⁷ Notably, these two structures differ only by a methyl group; the isobutyl group gives rise to agonism, while the *n*-propyl group renders antagonism.¹⁷

Since the only structural difference between the agonist 1 and the antagonist 2 can be found in the position of the methylene imidazole substituent, it can be hypothesized that the change in functionality could be attributed to a reduced ability to stabilize the activated receptor conformation due to a differently positioned imidazole group in 2. However, as an alternative hypothesis the imidazole rings could have the same receptor interaction point, while the isobutyl groups are forced to be positioned differently and cause the difference in functionality (Figure 4A). If so, the topography and location of the lipophilic side chain of ligands binding to the AT₂ receptor is the primary determinant of functionality as was the case at the AT₁ receptor (cf. 7 and 8).

In order to explore this alternative hypothesis and to identify a potential binding mode, a computational analysis was performed. Potential pharmacophore groups, including the imidazole rings, of 1 and 2 were superimposed using the pharmacophore modeling program DISCOtech¹⁸ in Sybyl¹⁹ (see Supporting Information for details). In order to reduce the number of conformations, model structures with methyl instead of butyl substituents on the sulfonylcarbamate group were used in the modeling. From the suggested binding mode model identified, shown in Figure 4A, the isobutyl groups are oriented differently in the two thiophene compounds, which could be of importance for their ability to induce either the agonistic or antagonistic response. On the basis of this model, we hypothesized that it should be possible to convert the meta-substituted antagonist 2 to an agonist if the isobutyl chain could adopt a similar orientation as in the para-substituted agonist 1, illustrated in Figure 4A. To investigate this hypothesis we performed further modeling where we replaced the thiophene

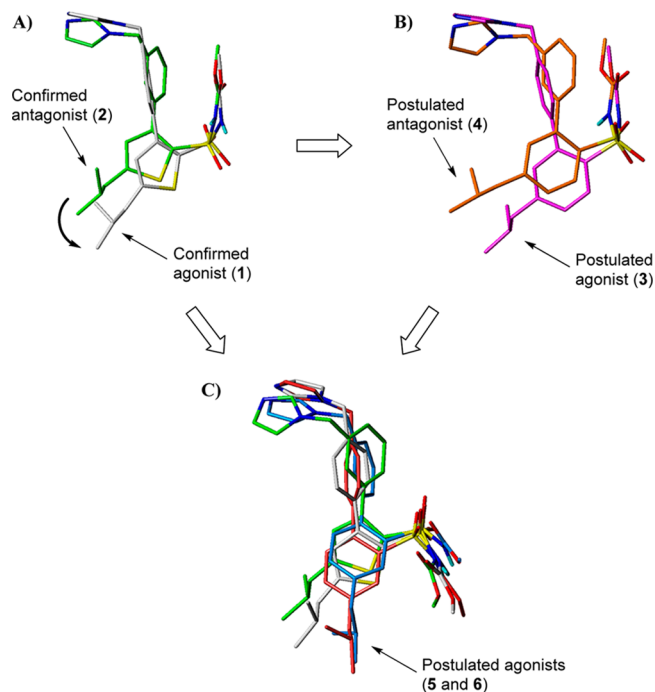


Figure 4. (A) Possible binding mode of two model compounds corresponding to structures 1 (white) and 2 (green). (B) Corresponding hypothesized binding mode for the two model compounds corresponding to the biphenyl analogues, 3 (purple) and 4 (orange), and postulated functions at the AT₂ receptor. (C) Comparison of the suggested binding modes of 1 and 2 with the analogues of 3 and 4, where the isobutyl have been moved to the meta-position, the two model compounds corresponding to compounds 5 (red) and 6 (blue), respectively.

ring with a phenyl group to obtain biphenyl analogues allowing better assessment of the impact of relative positions of the isobutyl side chain (i.e., 3 and 4, see Figure 2). Accordingly, superimposition of 3 and 4 produced a model, depicted in Figure 4B, which was very similar to that shown in Figure 4A. The corresponding analogues with an isobutyl group meta to the sulfonylcarbamate, compounds 5 and 6 (Figure 2), were also modeled and compared with the suggested binding modes of 1 and 2 as seen in Figure 4C. Although the position of the isobutyl group in 6 did not superimpose optimally to the corresponding group in 1, it was clearly closer to this binding mode than that of 2, where the imidazole and the sulfonylcarbamate also could reach the same interactions (Figure 4C). According to the model in Figure 4C, compound 5, although having a meta-methylene imidazole substitution, could adopt a conformation that allows a similar binding mode as 6 and was therefore also hypothesized to act as an agonist. In order to

explore this binding mode hypothesis we set out to synthesize regioisomers 3–6 and determine their AT₂ receptor binding affinity and, in particular, their functional activity at the AT₂ receptor.

The synthesis of 3 has been previously described.⁵ The synthesis of compounds 4–6 are outlined in Scheme 1 and started with the benzenesulfonamides 9 and 10, prepared as previously described by Wallinder et al.²⁰ Compound 13 and 14 were prepared by *N*-alkylation of imidazole by 3- and 4-bromobenzyl bromide in DMF. The crude boronic acids (11 and 12) were coupled with compounds 13 and 14 under Suzuki coupling conditions with Pd(PPh₃)₄ as catalyst and Na₂CO₃ as base using microwave irradiation (150 °C, 5 min) to form compounds 15–17 in good yields (75–87%). Compounds 4–6 were afforded after deprotection of the sulfonamide with BCl₃ and subsequent acylation with *n*-butyl chloroformate in DCM, water, and Na₂CO₃ to yield the final products in 27–51% yield, after purification by preparative HPLC.

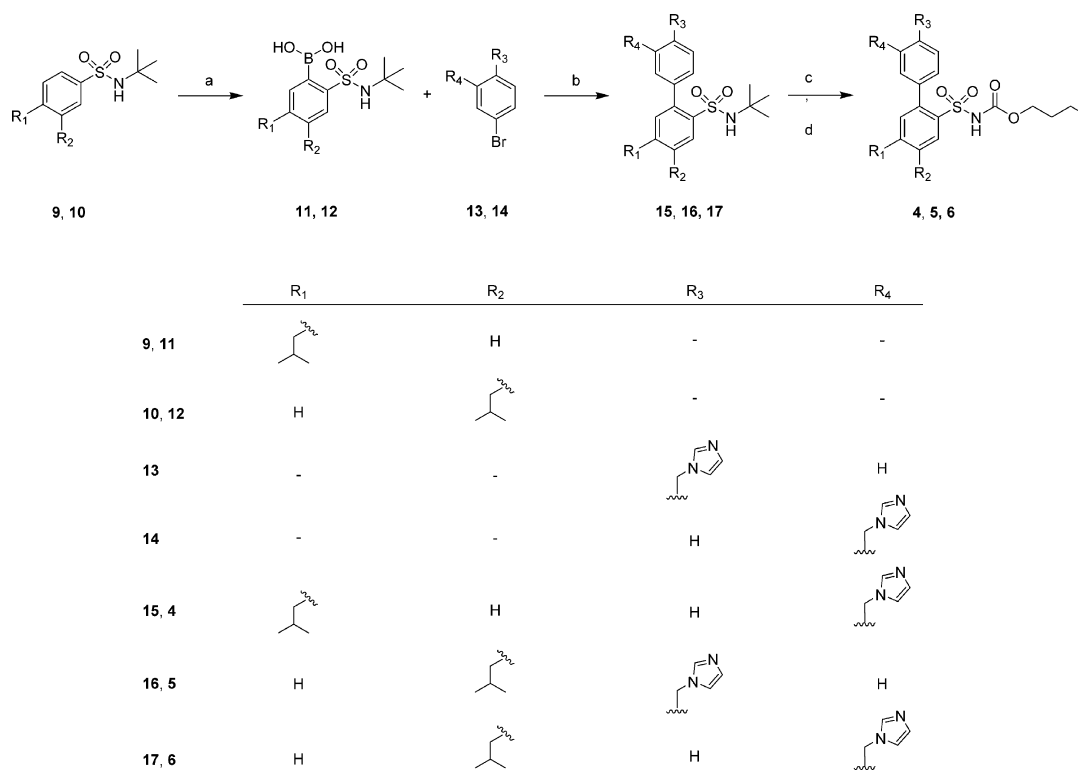
Compounds 3–6 were evaluated for affinity in a radioligand-binding assay by displacement of [¹²⁵I]Ang II from AT₂ receptors in pig uterus membranes as described previously by Nielsen et al.²¹ The natural substrate Ang II and the selective AT₁ receptor antagonist losartan were used as reference substances.²² The results of the binding study are presented in Table 1. Only AT₂ receptor affinity was evaluated since no AT₁ receptor binding have been seen for these methylene imidazoles even upon modifications of the lower part of the structures. This is also verified by compound 3, which previously has been tested for AT₁ receptor affinity ($K_i > 10\,000$ nM).⁵

Table 1. Biological Activities at the AT₂ Receptor

compd	$K_i \pm SD$ (nM)	function (10 nM)
3	0.7 ± 0.1	agonist
4	12.3 ± 0.4	antagonist
5	10.1 ± 0.4	agonist
6	8.2 ± 0.2	agonist

All compounds were evaluated for functionality in a neurite outgrowth assay with NG108-15 cells. The NG108-15 cells express mainly the AT₂ receptor in their undifferentiated state, and a three-day treatment with Ang II or the selective peptidic AT₂ receptor agonist CGP-42112 (*N*_α-nicotinoyl-Tyr-(*N*_ε-Cbz-Arg)-Lys-His-Pro-Ile)^{23,24} induce neurite outgrowth.^{16,25} The signaling pathways involve a sustained increase in Rap1/B-Raf/p42/p44^{mapk} activity and activation of the nitric oxide/guanylyl cyclase/cGMP pathway.^{26,27} Cells were treated in the absence or presence of Ang II, 3, 4, 5, and 6. After 3 days of treatment, cells were examined under a phase-contrast microscope and micrographs were taken. To establish an adequate test concentration, the compounds were tested at various concentrations ranging from 0.1 to 100 nM. No evidence of cell death was observed. All compounds were tested at a concentration of 10 nM, compound 4 was also tested at 100 nM to verify that the lack of neurite outgrowth was not due to too low concentration (data not shown). Coincubation with the selective AT₂ receptor antagonist PD 123,319 reduced neurite outgrowth, verifying that the effect was mediated through the AT₂ receptor. Treatment with PD 123,319 alone did not alter the morphology compared to untreated cells (data not shown), as we have shown previously.¹⁶ The antagonistic effect was verified through coincubation with Ang II resulting in reduced neurite outgrowth.

Scheme 1^a



^aReagents: (a) *n*-BuLi, THF, triisopropyl borate, HCl (aq); (b) Pd(PPh₃)₄, Na₂CO₃, toluene, ethanol, H₂O; (c) BCl₃, DCM; (d) *n*-butyl chloroformate, Na₂CO₃, DCM, H₂O.

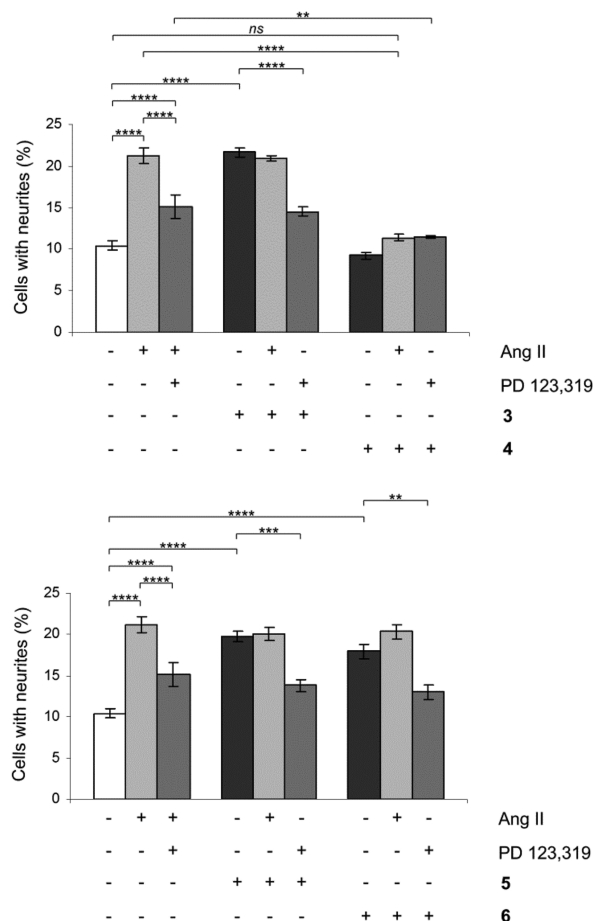


Figure 5. Effect of compounds 3–6 on neurite outgrowth in NG108-15 cells. The cells were plated at a cell density of 3.6×10^4 cells/Petri dish (35 mm) and were cultured for 3 days in the absence or presence of 100 nM Ang II, 10 nM 3, 10 nM 4, 10 nM 5, or 10 nM 6 alone or in combination with 10 μ M PD 123,319 or 100 nM Ang II. Cells with at least one neurite longer than a cell body were counted as positive for neurite outgrowth. The number of cells with neurites was expressed as the percentage of the total number in the micrographs (at least 400 cells according to the experiment). The results are significant according to two-way ANOVA: ****, $p < 0.0001$; ***, $p < 0.001$; **, $p < 0.01$; ns = not significant.

The functionalities are summarized in Table 1 and the neurite outgrowth results are shown in Figure 5.

The biphenyl analogues of 1 and 2, compounds 3 and 4, respectively, exhibited a similar biological response both in affinity to and functional activity at the AT₂ receptor, indicating that the biphenyl scaffold is a bioisoster to the thienylphenyl scaffold in this series of compounds. It is worth noting that the antagonist 4 exhibited a better inhibition of Ang II-induced neurite outgrowth compared to PD 123,319, even though the concentration of 4 was a 1000-fold lower compared to PD 123,319 (10 nM vs 10 μ M), as also seen with 2.^{15,16} Ang II stimulation after pretreatment with 4 resulted in neurite outgrowth no longer significantly different compared to untreated cells, while Ang II stimulation after pretreatment with PD 123,319 showed a neurite outgrowth significantly (****, $p < 0.0001$) higher compared to untreated cells. This indicates that compound 4 is a more efficient AT₂ receptor antagonist than PD 123,319 in the cell assay.

Compound 3 that has been previously synthesized and assessed as an AT₂ receptor ligand⁵ acts as an agonist (Table 1

and Figure 5). When the substitution pattern for the isobutyl chain of 3 was changed to give 5, the affinity dropped approximately 10 times, but the agonistic property was retained (Table 1 and Figure 5). When the same structural modification was performed with 4 to deliver compound 6 the affinity was slightly increased, but more interestingly, the antagonistic property of 4 was interconverted to agonism (Table 1 and Figure 5). Thus, independent of the methylene imidazole position, both an agonist and an antagonist could be obtained by moving the isobutyl side chain. This suggests that the structural feature responsible for functionality is the spatial relationship between the imidazole ring and the isobutyl side chain.

When looking at the peptide-activated GPCRs there seems to be a trend that minor structural alterations in small non-peptidic ligands can interconvert the functional activity, cf. 7 and 8.¹⁷ Small changes in the effector domain of peptides can frequently also have large impact on functionality, as exemplified by [Ile⁸]AngII, which acts as an antagonist at the AT₁ receptor (or partial agonist), while the native [Phe⁸]AngII acts as an agonist.²⁸ Likely it is the same effects on the equilibrium between the active and inactive receptor conformation that we see from these small nonpeptidic ligands.

The presented investigation highlights the importance of the relative position of the methylene imidazole and the isobutyl substituent for functional activity in this class of small molecule AT₂ receptor ligands. Thus, regardless of the position of the methylene imidazole group that previously was proposed to serve as an important determinant for agonism, both agonists and antagonists can be obtained by a short distance migration of the isobutyl group attached to the biphenyl scaffold. This suggests that the structural feature responsible for functionality is the spatial relationship between the imidazole ring and the isobutyl side chain. The data gained from molecular modeling provides a good foundation in the future design of selective AT₂ receptor agonists and antagonists.

■ ASSOCIATED CONTENT

📄 Supporting Information

Synthetic procedure and analytical data of the synthesized compounds, biological evaluation methods, and computational methods. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Author Contributions

The manuscript was written through contributions of all authors.

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Notes

The authors declare no competing financial interest.

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