

# Metagenomic Evaluation of Bacterial and Archaeal Diversity in the Geothermal Hot Springs of Manikaran, India

Sonu Bhatia,<sup>a</sup> Navneet Batra,<sup>a</sup> Ashish Pathak,<sup>b</sup> Stefan J. Green,<sup>c</sup> Amit Joshi,<sup>d</sup> Ashvini Chauhan<sup>b</sup>

Department of Biotechnology, GGSDS College, Chandigarh, India<sup>a</sup>; School of the Environment, Florida A&M University, Tallahassee, Florida, USA<sup>b</sup>; DNA Services Facility, University of Illinois at Chicago, Chicago, Illinois, USA<sup>c</sup>; Department of Biotechnology, SGGGS College, Chandigarh, India<sup>d</sup>

**Bacterial and archaeal diversity in geothermal spring water were investigated using 16S rRNA gene amplicon metagenomic sequencing. This revealed the dominance of Firmicutes, Aquificae, and the Deinococcus-Thermus group in this thermophilic environment. A number of sequences remained taxonomically unresolved, indicating the presence of potentially novel microbes in this unique habitat.**

Received 23 December 2014 Accepted 29 December 2014 Published 19 February 2015

**Citation** Bhatia S, Batra N, Pathak A, Green SJ, Joshi A, Chauhan A. 2015. Metagenomic evaluation of bacterial and archaeal diversity in the geothermal hot springs of Manikaran, India. *Genome Announc* 3(1):e01544-14. doi:10.1128/genomeA.01544-14.

**Copyright** © 2015 Bhatia et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Navneet Batra, [batranavneet@gmail.com](mailto:batranavneet@gmail.com).

North India is home to several geothermal springs, presenting an opportunity to study their microbial ecology (1, 2). Of particular interest are a series of hot water springs located close to Manikaran (32°02'N, 77°21'E; elevation 1,760 m) in the north-western Himalayas (3). This geothermal field lies in the Parbati Valley and extends in a linear zone of 1.5 km, where, sporadically, thermal springs emerge as spouts with temperatures of up to 96°C (3–5).

Previous microbial ecology studies on Manikaran springs mainly utilized culture-dependent approaches (1, 2); thus, our objective was to use metagenomics so that a comprehensive understanding of bacterial and archaeal diversity in these springs can be obtained. Replicate water samples were collected in sterilized containers from three sites at the Manikaran springs and filtered through 0.2- $\mu$ m pore-size sterilized filters by vacuum filtration. The filters were stored on ice and transported to GGSDS College, Chandigarh, where genomic DNA was extracted using the Powersoil DNA Isolation Kit (MoBio Laboratories, Inc., Carlsbad, CA, USA). DNA was then amplified using a two-stage PCR approach, similar to that described previously (6). Briefly, 515F and 806R primers (7, 8), targeting the V4 variable region of bacterial and archaeal small subunit rRNA genes, were used for amplification and sequencing through adapter and bar code incorporation via a second, 8-cycle PCR employing the AccessArray Barcode Library for Illumina (Fluidigm, South San Francisco, CA, USA). Cycling conditions for the first reaction were 95°C for 5 min, followed by 28 cycles of 95°C for 30 sec, 55°C for 45 sec, and 68°C for 30 sec, with a 7-min elongation step at 68°C. The second reaction was at 95°C for 5 min, followed by 8 cycles of 95°C for 30 sec, 60°C for 45 sec, and 68°C for 30 sec, with a 7-min elongation step at 68°C. Samples were pooled and purified using solid-phase reversible immobilization implemented with AMPure XP beads. Sequencing was performed using an Illumina MiSeq Microbiology kit, with primers CS1\_515F and CS2\_806R and the CS2rc primer (Fluidigm) for the index read. Raw sequences were merged using CLC genomics version 7.5.1 (CLC bio, Qiagen, Boston, MA, USA)

and quality trimmed (Q20) to obtain a total of 56.7 Mb of sequence data, which were analyzed using MG-RAST (9).

The Gram-positive, endospore-forming Firmicutes (28 to 84%) were dominant in the spring water, followed by Aquificae (2 to 64%) and the Deinococcus-Thermus group (1 to 18%). These phyla are mainly thermophilic and found in other extreme environments (10, 11). *Bacillus megaterium*, *Bacillus sporothermodurans*, *Hydrogenobacter* sp. GV4-1, *Thermus thermophiles*, and *Thermus brockianus* were the main bacterial species in the spring water.

Crenarchaeota (0.04–3%) was the main archaeal phylum, with *Pyrobaculum aerophilum* and *P. cladifontis* predominating because they are hyperthermophilic and metabolically versatile. Unlike many archaea, *Pyrobaculum* can thrive in microaerophilic environments by growing chemolithoautotrophically by sulfur reduction or organotrophically by sulfur respiration and fermentation, as shown by recent genome studies (12, 13).

Additionally, several bacterial and archaeal sequences remained taxonomically unresolved, indicating potentially novel microorganisms in this geothermal ecosystem. Additional metagenomics of this habitat will facilitate identification of microorganisms possessing industrially relevant traits, such as enzymes (14) and other compounds.

**Nucleotide sequence accession number.** The DNA sequences from this metagenomic project were deposited in the Sequence Read Archive under the accession number [SRX792272](https://www.ncbi.nlm.nih.gov/sra/SRX792272).

## ACKNOWLEDGMENTS

Partial funding for this study was provided by the U.S. Department of Defense grants W911NF-10-1-0146 and W911NF-10-R-0006.

## REFERENCES

1. Kumar M, Yadav AN, Tiwari R, Prasanna R, Saxena AK. 2014. Deciphering the diversity of culturable thermotolerant bacteria from Manikaran hot springs. *Ann Microbiol* 64:741–751. <http://dx.doi.org/10.1007/s13213-013-0709-7>.
2. Verma A, Gupta M, Shrikot P. 2014. Isolation and characterization of

- thermophilic bacteria in natural hot water springs of Himachal Pradesh (India). *Bioscan* 9:947–952.
3. Cinti D, Pizzino L, Voltattorni N, Quattrocchi F, Walia V. 2009. Geochemistry of thermal waters along fault segments in the Beas and Parvati valleys (north-west Himalaya, Himachal Pradesh) and in the Sohna town (Haryana), India. *Geochem J* 43:65–76. <http://dx.doi.org/10.2343/geochemj.1.0011>.
  4. Pandey OP, Negi JG. 1995. Geothermal fields of India: a latest update, vol 1, p 163–171. *In Proc. World Geothermal Congress International Geothermal Association, Bochum, Germany*.
  5. Chandrasekharam D, Alam MA, Minissale A. 2005. Thermal discharges at Manikaran, Himachal Pradesh, India, p 1–4. *In Proc. World Geothermal Congress. International Geothermal Association, Bochum, Germany*.
  6. Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. 2014. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. *MBio* 5:e01283-14. <http://dx.doi.org/10.1128/mBio.01283-14>.
  7. Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Huntley J, Fierer N, Owens SM, Betley J, Fraser L, Bauer M, Gormley N, Gilbert JA, Smith G, Knight R. 2012. Ultra-high-throughput microbial community analysis on the Illumina HiSeq and MiSeq platforms. *ISME J* 6:1621–1624. <http://dx.doi.org/10.1038/ismej.2012.8>.
  8. Moonsamy PV, Williams T, Bonella P, Holcomb CL, Höglund BN, Hillman G, Goodridge D, Turenchalk GS, Blake LA, Daigle DA, Simen BB, Hamilton A, May AP, Erlich HA. 2013. High throughput HLA genotyping using 454 sequencing and the Fluidigm Access Array system for simplified amplicon library preparation. *Tissue Antigens* 81:141–149. <http://dx.doi.org/10.1111/tan.12071>.
  9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
  10. Hall JR, Mitchell KR, Jackson-Weaver O, Kooser AS, Cron BR, Crosse LJ, Takacs-Vesbach CD. 2008. Molecular characterization of the diversity and distribution of a thermal spring microbial community by using rRNA and metabolic genes. *Appl Environ Microbiol* 74:4910–4922. <http://dx.doi.org/10.1128/AEM.00233-08>.
  11. Pagaling E, Grant WD, Cowan DA, Jones BE, Ma Y, Ventosa A, Heaphy S. 2012. Bacterial and archaeal diversity in two hot spring microbial mats from the geothermal region of Tengchong, China. *Extremophiles* 16:607–618. <http://dx.doi.org/10.1007/s00792-012-0460-1>.
  12. Mardanov AV, Gumerov VM, Slobodkina GB, Beletsky AV, Bonch-Osmolovskaya EA, Ravin NV, Skryabin KG. 2012. Complete genome sequence of strain 1860, a crenarchaeon of the genus *Pyrobaculum* able to grow with various electron acceptors. *J Bacteriol* 194:727–728. <http://dx.doi.org/10.1128/JB.06465-11>.
  13. Bernick DL, Karplus K, Lui LM, Coker JK, Murphy JN, Chan PP, Cozen AE, Lowe TM. 2012. Complete genome sequence of *Pyrobaculum oguniense*. *Stand Genomic Sci* 6:336–345.
  14. Batra N, Singh J, Joshi A, Bhatia S. 2011. Applications of  $\beta$ -Gal-III isozyme from *Bacillus coagulans* RCS3, in lactose hydrolysis. *Int J Biol Macromol* 49:879–884. <http://dx.doi.org/10.1016/j.ijbmac.2011.08.004>.