

Influence of *Pseudomonas Aeruginosa* on Exacerbation in Patients with Bronchiectasis

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ABSTRACT

Background: A majority of the studies done on the western population have shown that *Pseudomonas aeruginosa* causes many severe infections in patients with bronchiectasis as compared to other pathogens. There is scarcity of similar data from the Asian population. **Materials and Methods:** A prospective study was undertaken to identify the various pathogens isolated from the respiratory samples of 117 patients with bronchiectasis from south India and to compare the clinicomicrobiological profile of infections caused by *P. aeruginosa* and other respiratory pathogens. **Results:** The respiratory pathogens were isolated from 63 (53.8%) patients. *P. aeruginosa* was the most common isolate (46.0%) followed by *Klebsiella pneumoniae* (14.3%) and other pathogenic bacteria. Patients included in the *P. aeruginosa* group had a higher number of exacerbations (p : 0.008), greater number of hospital admissions (p : 0.007), a prolonged hospital stay (p : 0.03), and poor lung function, compared to the patients infected with the non-*Pseudomonas* group. **Conclusion:** It is necessary to investigate the etiology of respiratory tract infections among bronchiectasis patients followed by the prompt management of cases diagnosed with *P. aeruginosa* infections, so as to lower the morbidity and have a better prognosis.

Key words: Bronchiectasis, Morbidity, *Pseudomonas aeruginosa*

INTRODUCTION

Bronchiectasis is a persistent or progressive condition characterized by dilated, thick-walled bronchi.^[1] It is accompanied by chronic productive cough, airway obstruction, and recurrent infections.^[2] A vicious cycle of transmural recurrent infection and subsequent inflammation causes damage primarily to the bronchi and bronchioles. The damaged airways are susceptible to infection usually with colonizing, but severely damaging bacterial and fungal microbes.^[3] The symptoms of bronchiectasis vary from intermittent episodes of expectoration to persistent daily expectoration often of large volumes of purulent sputum and may be associated with other non-specific respiratory symptoms including dyspnea, chest pain, and hemoptysis, and may progress to respiratory failure and corpulmonale.^[1]

The main bacterial pathogens that are commonly isolated in bronchiectasis are *Haemophilus influenzae* and *Pseudomonas aeruginosa*. Other microorganisms encountered include *Streptococcus pneumoniae*, *Haemophilus parainfluenzae*, *Staphylococcus aureus*, *Moraxella catarrhalis*, nontuberculous mycobacteria, and *Aspergillus spp.*^[4] About one-third of the patients with bronchiectasis are chronically colonized with *P. aeruginosa*. Patients with *P. aeruginosa* experience an accelerated decline in lung function and more frequent exacerbations. Patients with no pathogens isolated from their sputum have the mildest disease.^[5] It is commonly recommended that microbiological identification of the pathogen and characterization of its antimicrobial susceptibility pattern would aid in decisions regarding antibiotic therapy, should it become needed.^[4] Considering that infections with *P. aeruginosa* have been associated with higher morbidity compared with other bacterial infections, and also the paucity of microbiological studies on bronchiectasis patients in the Asian population, a prospective study was performed to analyze the clinicomicrobiological profile in patients with bronchiectasis.

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Quick Response Code:



Website:
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DOI:
10.4103/0974-777X.150885

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MATERIALS AND METHODS

A prospective study was conducted for a period of one year from January to December 2012. This study was approved by our Institutional Ethics Committee. Following informed consent at enrollment, patients clinically diagnosed with acute exacerbation of bronchiectasis were included. Those patients with prior antibiotic therapy in the previous two weeks were excluded from the study. An infective exacerbation was defined as a change in one or more of the common symptoms of bronchiectasis (increasing sputum volume or purulence, worsening dyspnea, increased cough, declining lung function, increased fatigue/malaise) or the appearance of new symptoms (fever, pleurisy, hemoptysis, requirement for antibiotic treatment).^[1] Lower respiratory tract specimens including sputum and bronchoalveolar lavage fluid were collected at the time of an infective exacerbation, prior to the commencement of antibiotic treatment. The specimens were transported within two hours after collection to the Microbiology Laboratory for further processing. The quality of specimens was evaluated based on gram-stain findings, followed by culture and susceptibility testing. All sputum gram stains were read under an oil immersion objective (x100) and evaluated according to the Bartlett criteria. The specimens were scored 0, +1, or +2 according to the number of leukocytes seen per field and 0, -1, and -2 according to the number of squamous epithelial cells seen per field. Specimens with total scores of zero or less were considered inadequate and heavily contaminated with oropharyngeal flora. Those containing greater than 25 leucocytes and fewer than 10 squamous epithelial cells per field were optimal specimens and processed further.^[6] The bronchoalveolar lavage fluid was processed by a semi-quantitative culture with a positive threshold of 10⁴ CFU/mL.^[7] The specimens were cultured on Blood agar, Chocolate agar, and MacConkey agar plates and incubated at 37°C for 18-24 hours. The Blood agar and Chocolate agar plates were incubated in 5% CO₂ (capnophilic atmosphere). Identification of the bacterial isolates was done following standard bacteriological techniques.^[8] The antibiotic sensitivity of the isolates was determined by the Kirby Bauer's disk diffusion method on Mueller-Hinton agar (BD) following the Clinical Laboratory Standard Institute (CLSI) guidelines.^[9] These strains were then checked for extended spectrum beta-lactamase production (ESBL) using the double disk approximation method.^[10] For quality control, *E. coli* ATCC 25922, *P. aeruginosa* ATCC 27853, and *K. pneumoniae* ATCC 700603 were used. Duplicate isolates from the same patient were excluded from the analysis.

The isolates were grouped as *Pseudomonas* and non-*Pseudomonas* groups for analysis of the demographic characteristics; clinical severity in terms of the number of infective exacerbations in the previous year; high resolution computed tomography findings (HRCT); lung function tests, and antibiotic susceptibility profile. To assess lung function, the spirometry values of forced vital capacity (FVC), forced expiratory volume in one second (FEV1), and FEV1/FVC ratio was obtained for all patients. The HRCT thorax was assessed for the number of lobes involved (the lingula was considered as a separate lobe). Each lobe of both lungs was graded for bronchiectatic changes on a 0-3 scale, giving a maximum of 18 points: 0: No bronchiectasis; 1: one or less than one bronchopulmonary segment involved; 2: more than one bronchopulmonary segment involved; 3: gross cystic bronchiectasis.^[11]

RESULTS

A total of 117 patients with bronchiectasis were studied. The mean age of the study group was 52.9 years and the range was 7-86 years. Sixty-six were female (56.4%) and 51 (43.6%) were male. Normal oropharyngeal flora was grown in culture from 54 (46.2%) lower respiratory tract specimens. The respiratory pathogens were isolated from the rest of the 63 (53.8%) patients. *P. aeruginosa* was the most common isolate (46.0%) followed by *K. pneumoniae* and other pathogenic bacteria, accounting for the rest of the isolates (54%) [Table 1].

The clinical details of the studied patients [Table 2] showed that cough with expectoration was the presenting symptom in all the patients (100%) followed by hemoptysis in 26.5% of the cases. A past history of pulmonary tuberculosis was present in 34 (29%) patients, whereas, diabetes was noticed in 16 (13.7%) cases. Wheeze and crackles were present in 27.4 and 20.5% of the patients, respectively. Patients

Table 1: Frequency of isolation of pathogens from Bronchiectasis patients

Isolates	Frequency (%)
<i>Pseudomonas aeruginosa</i>	29 (46.0)
<i>Klebsiella pneumoniae</i>	9 (14.3)
<i>Acinetobacter spp.</i>	8 (12.7)
<i>Haemophilus influenzae</i>	4 (6.3)
<i>Streptococcus pneumoniae</i>	3 (4.8)
<i>Moraxella catarrhalis</i>	3 (4.8)
<i>Escherichia coli</i>	2 (3.2)
<i>Nocardia spp.</i>	2 (3.2)
<i>Stenotrophomonas maltophilia</i>	2 (3.2)
<i>Staphylococcus aureus</i>	1 (2.8)
Total	63

infected with *P. aeruginosa* had a significantly higher number of exacerbations (p : 0.008), greater number of hospital admissions (p : 0.007), a prolonged hospital stay (p : 0.03), poor lung function (p : < 0.05), and increased severity of the disease (p : 0.001) compared to the patients infected with other pathogenic bacteria [Table 3].

The *P. aeruginosa* isolates were largely sensitive to most of the antibiotics tested, with 93.1% of them being sensitive to meropenem, piperacillin, and cefepime [Table 4]. Higher rates of drug resistance were noted among the Enterobacteriaceae isolates and *Acinetobacter spp.* Meropenem was found to be the most effective antibiotic against *K. pneumoniae* (66.7%). All isolates of *K. pneumoniae*,

E. coli, and *Acinetobacter spp.* were found to be sensitive to colistin and tigecycline (100%). Among the gram-positive bacterial isolates, *S. pneumoniae* and *Nocardia sp.* were the sensitive strains, whereas, the isolate of *S. aureus* was found to be methicillin-resistant *S. aureus*.

DISCUSSION

Correlating with the earlier evidence,^[12] the present study has also observed a higher incidence of bronchiectasis in females as compared to males. Bronchiectasis often occurs in patients who have systemic diseases or other underlying associated conditions. The two basic pathogenic factors are airway obstruction and bacterial infection in the bronchial tree, leading to bacterial colonization of the bronchial mucosa and subsequent progressive lung damage.^[13] A history of previous severe lower respiratory tract infections due to bacterial and viral pneumonia, pertussis or tuberculosis should be sought in all patients with bronchiectasis. Where possible, the temporal relationship of the identified infections with the onset of chronic respiratory symptoms should be determined.^[1] Thirty-four (29%) subjects in the present study had a past history of pulmonary tuberculosis.

It has been suggested that all children and adults with bronchiectasis should have an assessment of lower respiratory tract microbiology.^[1] Understanding the local spectrum of lower respiratory bacteriology among patients with bronchiectasis will help in choosing the appropriate empirical therapy, pending culture results. *P. aeruginosa* was the most common isolate (46%) in our study, a finding also noted in other studies.^[11,13,14] Other studies have shown *H. influenzae* to be the most commonly isolated pathogen.^[15,16] The differences could be due to the varied distribution of organisms in different geographical locations. Patients infected with *P. aeruginosa* are known to experience a more accelerated decline in lung function and more frequent exacerbations than those infected with other organisms.^[5] A similar observation was made in the present study with patients infected with *P. aeruginosa* having a higher number of exacerbations (p : 0.008) and a prolonged hospital stay (p : 0.03). Spirometry (FVC and FEV1) also demonstrated a significant difference (p : < 0.05) in the *Pseudomonas* versus non-*pseudomonas* group. HRCT is considered to be the best investigation for bronchiectasis patients to determine the involvement of different lobes of the lung with precision. In the present study, HRCT has revealed the increased severity of the disease based on the lobes of the lung involved in patients having *P. aeruginosa* infection (p : 0.001), as compared to the non-*pseudomonas* group.

Table 2: Frequency of clinical signs and symptoms among the study group

Clinical parameter	No. of patients (%)
Symptoms	
Productive cough	117 (100)
Hemoptysis	31 (26.5)
Chest pain	19 (16.2)
Past h/o tuberculosis	34 (29)
Signs	
Wheeze	32 (27.4)
Crackles	24 (20.5)
Clubbing	19 (16.2)

Table 3: Comparison of demographic factors and the clinical profile of bronchiectasis patients infected with *Pseudomonas aeruginosa* versus non-*Pseudomonas aeruginosa* bacteria

Characteristics	<i>Pseudomonas</i> group (n = 29)	Non- <i>Pseudomonas</i> group (n = 34)	p-value
Demographic details			
Gender (M:F)	12:17	20:14	
Mean age	56	51.6	0.14
Important Symptoms and Signs			
Mean duration of cough (days)	17.5	19.06	0.78
Mean number of exacerbations	31.19	4.6	0.008
Hemoptysis	06	13	0.92
Chest pain	08	07	0.81
Wheeze	10	11	0.37
Clubbing	04	09	0.23
History of smoking	05	07	0.83
Past h/o pulmonary tuberculosis	16	18	0.23
Effect on lung functions			
Mean FVC	50.9	58.5	0.016
Mean FEV1	41.5	49.8	0.05
FEV1/FVC	80.2	85.1	0.329
Mean HRCT score	7.8	3.2	0.001
Effect on disease outcome			
Mean duration of hospital stay	29.29	9.5	0.03
Mean number of hospital admissions	2.46	1.76	0.007

Table 4: Antimicrobial susceptibility profile of pathogens from bronchiectasis patients (in percentage)

Gram-negative bacteria																	
Organisms (No.)	AC	PIP	AMC	CFX	FRX	CAZ	FEP	AZT	COT	AMK	GEN	NET	TOB	CIP	TZP	TIM	MEM
<i>Pseudomonas aeruginosa</i> (29)	NT	93.1	NT	NT	NT	89.6	93.1	NT	NT	86.2	79.3	86.2	86.2	86.2	93.1	NT	93.1
<i>Klebsiella pneumoniae</i> (9)	0	NT	22.2	33.3	33.3	NT	44.4	33.3	33.3	66.7	44.4	55.5	NT	44.4	55.5	44.4	66.7
<i>Acinetobacter spp.</i> (8)	0	NT	0	12.5	0	NT	62.5	0	62.5	62.5	75	75	NT	75	62.5	62.5	75
<i>Haemophilus influenzae</i> (4)	75	NT	75	NT	75	NT	NT	NT	NT	NT	NT	NT	NT	100	NT	NT	NT
<i>Moraxella catarrhalis</i> (3)	66.7	NT	100	NT	100	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT	NT
<i>Escherichia coli</i> (2)	0	NT	0	0	0	NT	0	0	50	0	0	0	NT	0	0	0	100
<i>Stenotrophomonas maltophilia</i> (2)	NT	NT	NT	NT	NT	NT	NT	NT	100	NT	NT	NT	NT	100	NT	NT	NT
Gram-positive bacteria																	
Organisms (No.)	AC	AMC	FRX	CIP	COT	ERY	GEN	TET	CHL	LNZ	TEC	VAN	RIF				
<i>Streptococcus pneumoniae</i> (3)	NT	NT	100	100	66.7	0	NT	33.3	NT	NT	NT	NT	NT				
<i>Nocardia spp.</i> (2)	100	100	100	100	0	50	0	50	NT	NT	NT	NT	NT				
<i>Staphylococcus aureus</i> (1)	0	0	0	0	0	0	0	100	100	100	100	100	100				

AC: Amoxicillin (10 µg), AMC: Amoxicillin-clavulanic acid (20/10 µg), AMK: Amikacin (30 µg), AZT: Aztreonam (30 µg), CAZ: Ceftazidime (30 µg), CFX: Cefuroxime (30 µg), CHL: Chloramphenicol (30 µg), CIP: Ciprofloxacin (5 µg), COT: Trimethoprim-sulfamethoxazole (23.75/1.25 µg), ERY: Erythromycin (15 µg), FEP: Cefepime (30 µg), FRX: Ceftriaxone (30 µg), GEN: Gentamicin (10 µg), LNZ: Linezolid (30 µg), Net: Netilmicin (30 µg), MEM: Meropenem (10 µg), PIP: Piperacillin (100 µg), RIF: Rifampicin (5 µg), TEC: Teicoplanin (30 µg), TET: Tetracycline (30 µg), TIM: Ticarcillin-clavulanic acid (75/10 µg), TOB: Tobramycin (10 µg), TZP: Piperacillin-tazobactam (100/10 µg), VAN: Vancomycin (30 µg), NT: not tested

Although the isolates of *P. aeruginosa* were found to be sensitive to the antibiotics tested, most of them were mucoid strains. Once acquired, *P. aeruginosa* (especially the mucoid type) is difficult to eradicate from bronchiectasis and cystic fibrosis patients.^[14] Prompt eradication treatment at the very onset of infection prior to its transition to a mucoid variant would seem advantageous.^[17] Adequate antibiotic therapy, including a combination dosage, modes of delivery, and duration of therapy should be given due consideration. Addition of macrolide antibiotics to the treatment regimen are effective in reducing the number of exacerbations in patients with bronchiectasis, by modulation of the inflammatory response, and their ability to impede biofilm formation.^[18]

Even though *P. aeruginosa* and *H. influenzae* are the most common bacteria isolated from non-cystic fibrosis bronchiectasis patients, other pathogens including *K. pneumoniae*, *Acinetobacter spp.*, and *S. maltophilia* were isolated. These gram-negative bacilli were found to be multidrug-resistant strains. Previous exposure to anti-microbial agents and repeated contact with the healthcare system could lead to colonization and infection with these multidrug-resistant bacteria. In this study, only one isolate of *S. aureus* was obtained. Persistent isolation of *S. aureus* should lead to consideration of underlying allergic bronchopulmonary aspergillosis or cystic fibrosis.^[1,18]

The study stresses the need to investigate the etiology of respiratory tract infections among bronchiectasis patients and prompt management of cases diagnosed with *P. aeruginosa* infections so as to lower the morbidity and have a better prognosis.

REFERENCES

- Pasteur MC, Bilton D, Hill AT. British Thoracic Society guideline for non-CF bronchiectasis. *Thorax* 2010;65:i1-58.
- Baydarian M, Walter RN. Bronchiectasis: Introduction, Etiology, and Clinical Features. *Dis Mon* 2008;54:516-26.
- Moulton BC, Barker AF. Pathogenesis of Bronchiectasis. *Clin Chest Med* 2012;33:211-7.
- Feldman C. Bronchiectasis: New Approaches to Diagnosis and Management. *Clin Chest Med* 2011;32:535-46.
- O'Donnell AE. Bronchiectasis. *Chest* 2008;134:815-23.
- Lentino JR, Lucks DA. Nonvalue of Sputum Culture in the Management of Lower Respiratory Tract Infections. *J Clin Microbiol* 1987;25:758-62.
- Chastre J, Combes A, Luyt CE. The Invasive (Quantitative) Diagnosis of Ventilator-Associated Pneumonia. *Respir Care* 2005;50:797-807.
- Collee JG, Miles RS, Watt B. Tests for identification of bacteria. In: Collee JG, Fraser AG, Marmion BP, Simmons A, editors. *Mackie and McCartney Practical Medical Microbiology*. 14th ed. New York: Churchill Livingstone; 1996. p. 131-49.
- Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-First Informational Supplement. CLSI document M100-S21. Wayne, Pennsylvania USA: Clinical and Laboratory Standards Institute; 2011.
- Tofteland S, Haldorsen B, Dahl KH, Simonsen GS, Steinbakk M, Walsh TR, et al. Effects of phenotype and genotype on methods for detection of extended-spectrum-beta-lactamase-producing clinical isolates of *Escherichia coli* and *Klebsiella pneumoniae* in Norway. *J Clin Microbiol* 2007;45:199-205.
- Wilson CB, Jones PW, O'Leary CJ, Hansell DM, Cole PJ, Wilson R. Effect of sputum bacteriology on the quality of life of patients with bronchiectasis. *Eur Respir J* 1997;10:1754-60.
- Morrissey BM, Harper RW. Bronchiectasis: Sex and gender considerations. *Clin Chest Med* 2004;25:361-72.
- Palvatwichai A, Chaoprasong C, Vattanatham A, Wongs A, Jatakanon A. Clinical, laboratory findings and microbiologic characterization of bronchiectasis in Thai patients. *Respirology* 2002;7:63-6.
- Hla SW, Hui KP, Tan WC, Ho B. Genome macro restriction analysis of sequential *Pseudomonas aeruginosa* isolates from bronchiectasis patients without cystic fibrosis. *J Clin Microbiol* 1996;34:575-8.
- Li AM, Sonnappa S, Lex C, Wong E, Zacharasiewicz A, Bush A, et al. Non-CF bronchiectasis: Does knowing the aetiology lead to changes in management? *Eur Respir J* 2005;26:8-14.
- King PT, Holdsworth SR, Freezer NJ, Villanueva E, Holmes PW. Microbiologic follow-up study in adult bronchiectasis. *Respir Med*

2007;101:1633-8.

17. Jones AM. Eradication therapy for early *Pseudomonas aeruginosa* infection in CF: Many questions still unanswered. Eur Respir J 2005;26:373-5.
18. Amorima A, Gamboab F, Azevedoc P. New advances in the therapy of non-cystic fibrosis bronchiectasis. Rev Port Pneumol 2013;19:266-75.

How to cite this article: Chawla K, Vishwanath S, ManuM, Lazer B. Influence of *Pseudomonas aeruginosa* on exacerbation in patients with bronchiectasis. J Global Infect Dis 2015;7:18-22.

Source of Support: Nil. **Conflict of Interest:** None declared.

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