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Stereological Analysis of the Rat and Monkey Amygdala

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Abstract

The amygdala is part of a neural network that contributes to the regulation of emotional behaviors. Rodents, especially rats, are used extensively as model organisms to decipher the functions of specific amygdala nuclei, in particular in relation to fear and emotional learning. Analysis of the role of the nonhuman primate amygdala in these functions has lagged work in the rodent but provides evidence for conservation of basic functions across species. Here we provide quantitative information regarding the morphological characteristics of the main amygdala nuclei in rats and monkeys, including neuron and glial cell numbers, neuronal soma size, and individual nuclei volumes. The volumes of the lateral, basal, and accessory basal nuclei were, respectively, 32, 39, and 39 times larger in monkeys than in rats. In contrast, the central and medial nuclei were only 8 and 4 times larger in monkeys than in rats. The numbers of neurons in the lateral, basal, and accessory basal nuclei were 14, 11, and 16 times greater in monkeys than in rats, whereas the numbers of neurons in the central and medial nuclei were only 2.3 and 1.5 times greater in monkeys than in rats. Neuron density was between 2.4 and 3.7 times lower in monkeys than in rats, whereas glial density was only between 1.1 and 1.7 times lower in monkeys than in rats. We compare our data in rats and monkeys with those previously published in humans and discuss the theoretical and functional implications that derive from our quantitative structural findings.

INDEXING TERMS

amygdaloid complex; neuron; astrocyte; oligodendrocyte; neuropil; human

The amygdala, also known as the amygdaloid complex, is a brain region located in the rostral portion of the human medial temporal lobe (Freese and Amaral, 2009). In rodents and primates including humans the amygdala consists of 13 nuclei and cortical areas demarcated into several subdivisions (Table 1) (Amaral et al., 1992; Sorvari et al., 1996; Pitkäen, 2000). Amygdala function has long been associated with emotional behavior, particularly fear (Davis, 2000; LeDoux, 2003; Feinstein et al., 2011), stress (Roozendaal et al., 2009), anxiety

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(Rauch et al., 2003), and appetitive and aversive conditioning (Shabel and Janak, 2009). The amygdala has also been linked to the neural systems that underlie normal social behavior (Adolphs, 2003), essentially via the modulation of fear (Emery et al., 2001; Prather et al., 2001; Amaral et al., 2003; Bauman et al., 2004). In addition, the amygdala has been shown to be important for the modulation of memory processes, particularly under arousing or emotionally salient conditions (McGaugh, 2004; Roozendaal et al., 2009).

Rodents, especially rats, are used extensively as model organisms to study the structural organization of the amygdala and to decipher the functions of specific amygdala nuclei (Pitkänen et al., 1997; Balleine and Killcross, 2006), in particular with respect to fear and emotional learning (Sigurdsson et al., 2007). In contrast, primates have been the model of choice for studies of the role of the amygdala in social behavior (Amaral et al., 2003; Adolphs, 2010). Several neuroanatomical studies, however, have already pointed out striking differences in the organization of the amygdaloid complex in rats, monkeys, and humans at the cellular level. Differences include, for example, the distribution of calcium-binding proteins (McDonald, 1994a; Pitkänen and Kemppainen, 2002), chondroitin sulfate proteoglycans (Pantazopoulos et al., 2008), GABA_A receptors (McDonald and Mascagni, 1996), corticosteroid receptors (Pryce, 2008), and glutamate receptor subunits (McDonald, 1994b). Similarly, a relatively comprehensive description of the fundamental morphological characteristics and connectivity of the amygdaloid complex in rats, cats, and monkeys (Price et al., 1987) also summarized and suggested a number of potentially important species differences.

Stephan and Andy (1977) performed an informative comparative study of the volume of the amygdala in a large number of species. However, there was only limited information regarding the connectivity and neurochemical characteristics of the amygdaloid complex at that time and the nomenclature used in that study did not correspond to our current understanding of the structural organization of the amygdala (Table 1) (Amaral et al., 1992; Sorvari et al., 1996; Pitkänen, 2000). Moreover, it is now clear that although a first step, the simple measure of the volume of a brain structure is insufficient, by itself, to make clear interpretations regarding species differences in brain structure and function (Williams and Herrup, 1988). Indeed, there is not necessarily a direct link between the volume of a structure and the number of neurons in that structure, especially between species that might be only distantly related (Herculano-Houzel et al., 2007). It is therefore critically important to obtain reliable, quantitative information regarding the number of neurons, which are the fundamental functional units of the nervous system, in order to make inferences about the structural organization and putative functional processes subserved by defined nuclei of the amygdaloid complex in different species.

Although a cursory examination of Nissl-stained, coronal brain sections is sufficient to recognize that the amygdala differs between rodents and primates, little reliable quantitative information defining the number of neurons and glial cells, neuronal soma size, and individual nuclei volumes exists. The aim of this study was to provide these normative data in rats and monkeys. We therefore implemented modern, design-based stereological techniques to count neuron and glia numbers, measure neuronal soma size and the volume of the main amygdala nuclei (lateral, basal, paralaminar (in monkeys), accessory basal, medial

and central nuclei) in mature rats (at 7 months of age) and monkeys (at 5–9 years of age). We further compared our quantitative data with previously acquired stereological data from the human amygdala (Schumann and Amaral, 2005), in order to provide a critical perspective on the relative development and structural organization of the main amygdala nuclei in two model organisms widely used to decipher the basic principles of amygdala functions in humans.

MATERIALS AND METHODS

Experimental animals

Monkeys—Four rhesus monkeys, *Macaca mulatta*, (two males: 5.3 and 9.4 years of age; two females: 7.7 and 9.3 years of age) were used for this study. Monkeys were born from multiparous mothers and raised at the California National Primate Research Center (CNPRC). They were maternally reared in 2,000 m² outdoor enclosures and lived in large social groups until they were killed. These monkeys were the same animals used in quantitative studies of the monkey hippocampal formation (Jabès et al., 2010, 2011).

Rats—Four Sprague-Dawley rats, *Rattus norvegicus*, two males and two females at 7 months of age, were used for this study. All rats were retired breeders obtained from a commercial supplier (Charles River, San Diego, CA). Animals were perfused, as described below, within 24 hours of their arrival in our laboratory. In the meantime they were housed singly in standard laboratory cages with food and water ad libitum.

All experimental procedures were approved by either the Fribourg Veterinary Commission or the Institutional Animal Care and Use Committee of the University of California, Davis, and were conducted in accordance with the National Institutes of Health guidelines for the use of animals in research.

Brain acquisition

Monkeys—Each monkey was injected with the cell-division marker, 5'-bromo-2-deoxyuridine (BrdU, 150 mg/kg intraperitoneally; Boehringer Mannheim, Germany), 4 weeks prior to death in the context of another experiment (Jabès et al., 2010). Monkeys were deeply anesthetized with an intravenous injection of sodium pentobarbital (50 mg/kg, Fatal-Plus, Vortech Pharmaceuticals, Dearborn, MI) and perfused transcardially with 1% and then 4% paraformaldehyde (PFA) in 0.1 M phosphate buffer (PB; pH 7.4) following protocols previously described (Lavenex et al., 2009a). Coronal sections were cut using a freezing, sliding microtome in six series at 30 µm and one series at 60 µm (Microm HM 450, Microm International, Germany). The 60-µm sections were collected in 10% formaldehyde solution in 0.1 M PB (pH 7.4) and postfixed at 4°C for 4 weeks prior to Nissl staining with thionin. All other series were collected in tissue collection solution and kept at –70°C until further processing (Lavenex et al., 2009a).

Rats—Animals were deeply anesthetized with pentobarbital (Nembutal, 50 mg/kg i.v.) and perfused transcardially with 1% and 4% PFA in 0.1 M PB (pH 7.4). The brains were postfixed for 6 hours in the same fixative, cryoprotected in 10% and 20% glycerol solutions

in 0.1 M PB (pH 7.4; for 24 and 72 hours, respectively), frozen in powdered dry ice, and stored at -70°C until sectioning. Coronal sections were cut using a freezing, sliding microtome in four series at $40\ \mu\text{m}$ (Microm HM 450, Microm International, Germany). One series was collected in 10% formaldehyde solution in 0.1 M PB (pH 7.4) and postfixed at 4°C for 4 weeks prior to Nissl staining with thionin. All other series were collected in tissue collection solution and kept at -70°C until further processing.

Histological processing

The procedure for Nissl-stained sections followed our standard laboratory protocol described previously (Lavenex et al., 2009). Briefly, sections were taken out of the 10% formaldehyde solution, thoroughly washed, mounted on gelatin-coated slides, and air-dried overnight at 37°C . Sections were then defatted 2×2 hours for monkeys and 2×1 hour for rats in a mixture of chloroform/ethanol (1:1, vol), partially rehydrated, and air-dried overnight at 37°C . Sections were then fully rehydrated and stained 40 seconds for rats and 20 seconds for monkeys in a 0.25% thionin solution (Fisher Scientific, Waltham, MA, cat. no. T-409), dehydrated, and coverslipped with DPX (BDH Laboratories, Poole, UK).

Anatomical boundaries of the amygdala

Rats—The cytoarchitectonic organization of the rat amygdala has been described in detail previously (Krettek and Price, 1978; McDonald, 1982; Cassell et al., 1986; Price et al., 1987; Pitkänen et al., 1995; Savander et al., 1995, 1996; Jolkkonen and Pitkänen, 1998), and a nomenclature congruent with that used for monkeys and humans has been provided by Pitkänen (2000) for defining the rat amygdala nuclei (Table 1). We thus delineated the five main nuclei of the rat amygdala (i.e., lateral, basal, accessory basal, central, and medial) according to these descriptions for all subsequent analyses (Fig. 1).

Monkeys—The nomenclature and the basic description of the morphological characteristics of the amygdala nuclei have been described in detail previously by Price et al. (1987), Amaral et al. (1992), and Pitkänen and Amaral (1998) for the cynomolgus monkey (*Macaca fascicularis*). We used these descriptions to determine the boundaries of the six main nuclei (i.e., lateral, basal, accessory basal, paralaminar, central, and medial) of the *Macaca mulatta* amygdala (Table 1; Fig. 2).

Stereological analyses

Volume measurements and neuron and glia counts were performed with StereoInvestigator 7.0 (MicroBright-Field, Williston, VT). We estimated the volume of the brain (telencephalon and diencephalon), the volume of the whole amygdala, the volume of the main amygdala nuclei (lateral, basal, paralaminar (in monkeys), accessory basal, central, and medial) according to the Cavalieri principle on Nissl-stained sections cut at $60\ \mu\text{m}$ in monkeys and $40\ \mu\text{m}$ in rats (Gundersen and Jensen, 1987; West and Gundersen, 1990; Lavenex et al., 2000a,b). An average of 15 sections per animal for monkeys ($480\ \mu\text{m}$ apart) and an average of 28 sections per animal for rats ($160\ \mu\text{m}$ apart) were used to measure the volume of the whole amygdala. We estimated total amygdala volumes in the left and the right hemisphere and did not find any lateralization in rats ($t_{(3)} = -0.606$, $P = 0.587$) or in

monkeys ($t_{(3)} = -1.222, P = 0.309$). We therefore performed estimates unilaterally for the other parameters (see below).

About 29 sections per animal for monkeys (240 μm apart) and about 21 sections per animal for rats (160 μm apart) were used for volume measurements of the main amygdala nuclei. As there was no lateralization for overall amygdala size, we estimated the volumes of individual amygdala nuclei in the left hemisphere for half of the animals, and in the right hemisphere for the other half. Brain volume refers to the volume of the telencephalon and diencephalon bilaterally. Thirty-four to 38 sections per animal for monkeys (1,920 μm apart), with the first section selected randomly within the first seven sections through the brain, and 29 to 32 sections per animal for rats (480 μm apart), with the first section selected randomly within the first three sections through the brain, were used for brain volume measurements. As for brain volume measurements, we used the section cutting thickness (60 μm in monkeys, 40 μm in rats) in order to calculate the volume of individual amygdala nuclei.

The total number of neurons in the main amygdaloid nuclei was determined using the optical fractionator method (Gundersen 1986; West et al., 1991). This design-based method enables the estimation of the number of cells that is independent of volume estimates. Neuron number was estimated in the right or in the left amygdala only, as for volume measurements. About 11 sections per animal for monkeys (480 μm apart), with the first section selected randomly within the first two sections through the nucleus of interest, and about 13 sections per animal for rats (160 μm apart) were used for neuron counts (Supporting Information 1). We used a 100 \times PlanFluor oil objective (N.A. 1.30) on a Nikon Eclipse 80i microscope (Nikon Instruments, Melville, NY) linked to PC-based StereoInvestigator 7.0. The sampling scheme was established to obtain individual estimates of neuron number with estimated coefficients of error (CE) around 0.10 (CE average (neurons) = 0.107). Section thickness was measured at each counting site in rats and at every other counting site in monkeys (average thickness: 11.89 μm in rats and 13.20 μm in monkeys). The thickness of processed sections was thus 30% that of the cutting thickness in rats and 22% that of the cutting thickness in monkeys, due to the shrinkage induced by the different steps of tissue processing including different defatting times for rats and monkeys.

The volume of neuronal somas was determined using the Nucleator method (Gundersen, 1988). We measured an average of 260 neurons per nucleus, sampled at every counting site during the optical fractionator analysis. Briefly, the nucleator can be used to estimate the mean cross-sectional area and volume of cells. A set of rays emanating from a point randomly chosen within the nucleus is drawn and oriented randomly. The length of the intercept from the point to the cell boundary (l) is measured and the cell volume is obtained by $V = (4/3) \times 3.1416 \times (\text{mean } l)^3$. Essentially, this is the formula used to determine the volume of a sphere with a known radius. Note that the nucleator method provides accurate estimates of neuron size when isotropic-uniform-random sectioning of brain structures is employed (Gundersen, 1988). In our study all brains were cut in the coronal plane. Estimates of cell size might therefore be impacted by the nonrandom orientation of neurons in the different amygdala nuclei, which could lead to a systematic over- or underestimation of cell size in any given structure.

The total number of glial cells in the amygdala nuclei was determined using the optical fractionator method during the neuron counting. Thus, the same sampling scheme as for neuron counts was used for glial cells counts (CE average (glia) = 0.132). In rats and monkeys we differentiated between neurons, glial, and endothelial cells (Konopaske et al., 2007; Morris et al., 2008). In monkeys we were able to further distinguish oligodendrocytes and astrocytes based on morphological criteria identifiable in Nissl preparations (Fig. 3). We refer the reader to the original publications by (Palackal et al., 1993; Grady et al., 2003; Hamidi et al., 2004; Fitting et al., 2008) for detailed descriptions. Neurons are heavily stained with a large and single nucleolus (Fig. 3). Astrocytes display relatively smaller size and pale staining of the nucleus with a thin rim of Nissl-stained cytoplasm immediately adjacent to it (Fig. 3). Oligodendrocytes are smaller than astrocytes and can be identified by round, darker nuclei more densely packed with chromatin (Fig. 3). Microglia were not counted but were characterized in rats and monkeys by their smallest nucleus, darkest, irregular shape, sometimes rod-shaped, oval, or bent (Morris et al., 2008). The differentiation between oligodendrocytes and astrocytes was not sufficiently reliable in rats due to their overall smaller cell size and the lack of clear morphological criteria in Nissl preparations.

No gender difference was found for any of the estimated parameters, so data from both genders were combined for presentation. We also evaluated both left and right amygdaloid nuclei in a systematic manner (as described above) and no lateralization was found (data not shown). Thus, our findings and all subsequent considerations are valid for, and can be generalized to, both left and right amygdala in both males and females.

3D reconstructions

We performed 3D reconstructions of the amygdala nuclei in order to better appreciate the morphological and topological differences between rats, monkeys and humans (Fig. 4; Supporting Information 2 and 3). We used every Nissl-stained, coronal brain section, 160 μm apart for rats, 240 μm apart for monkeys to trace the main amygdala nuclei with StereoInvestigator 7.0. Tracings of the human amygdala nuclei were obtained with StereoInvestigator 5.0 (Schumann and Amaral, 2005). We then imported the tracings into the Reconstruct software (<http://synapses.clm.utexas.edu/>) (Fiala, 2005), and aligned the serial sections manually by using the contour of the brain as a reference between individual sections. The software generated a surface reconstruction of the brain and main amygdala nuclei using the Boissonnat (1988) method. Finally, 3D reconstruction images were rendered in Blender software v. 2.49 (open-source free-ware, <http://www.blender.org>).

Photomicrographic production

Low-magnification photomicrographs were taken with a Leica DFC420 digital camera on a Leica MZ9.5 stereo-microscope (Leica Microsystems, Wetzlar, Germany). High-magnification photomicrographs were taken with a Leica DFC490 digital camera on a Nikon Eclipse 80i microscope (Nikon Instruments, Tokyo, Japan). Artifacts located outside of the sections were removed and gray levels were adjusted in Adobe Photoshop CS4, v. 11.0 (Adobe, San Jose, CA).

RESULTS

Volumes

We used the Cavalieri method to estimate the volumes of the brain, the whole amygdala, and the main amygdala nuclei in rats and monkeys (Table 2). We also present human data previously published by Schumann and Amaral (2005) to enable a comparison of the relative size of individual nuclei in the three species. Estimates of the volume of the brain (telencephalon and diencephalon bilaterally) revealed that the monkey brain is about 53 times larger than the rat brain. In comparison, the volume of the whole amygdala (unilaterally) was only 18 times larger in monkeys than in rats. Importantly, this size difference was not homogeneous among amygdala nuclei, and distinct amygdala nuclei exhibited different scaling factors between rats and monkeys (Table 2; Fig. 5A). The lateral, basal, and accessory basal nuclei were, respectively, 32, 39, and 39 times larger in monkeys than in rats. In contrast, the central and medial nuclei were only 8 and 4 times larger, respectively, in monkeys than in rats. The combined volume of the remaining amygdala nuclei (i.e., nuclei other than those described above) was 10 times larger in monkeys than in rats (see Table 2).

By way of comparison, the human brain is ≈ 10 times larger than the monkey brain and the human amygdala is 7 times larger than that of the monkey. Again, however, distinct nuclei exhibited different scaling factors between monkeys and humans (Table 2; Fig. 5A). In particular, the lateral nucleus was 12 times larger in humans than in monkeys. In contrast, the volumes of the basal, accessory basal and central nuclei were respectively 6, 6, and 4 times larger in humans. The combined volume of the remaining amygdala nuclei was 6 times larger in humans than in monkeys.

Altogether, these volumetric data revealed important species differences in the relative size of the main amygdala nuclei in rats, monkeys, and humans. Most interestingly, the lateral, basal, and accessory basal nuclei, which are highly interconnected with the neocortex, were dramatically more developed in monkeys and humans than in rats. Indeed, the summed volume of the lateral, basal (including the paralamina nucleus in monkeys and humans), and accessory basal nuclei represented 62% of total amygdala volume in monkeys and 69% in humans, whereas it represented only 28% of total amygdala volume in rats. In contrast, the central and medial nuclei exhibited relatively smaller size differences between rats and monkeys. These differences can easily be appreciated in 3D reconstructions of the amygdala in the three species (Fig. 4 and Supporting Information 2 and 3).

Neuron and glial cell numbers

We used the optical fractionator method to estimate the number of neurons and glial cells (astrocytes and oligodendrocytes) in distinct nuclei of the rat and monkey amygdala (Table 3), and compared our results with the number of neurons found in the human amygdala (Schumann and Amaral, 2005). We found that the numbers of neurons in the lateral, basal, and accessory basal nuclei were, respectively, 14, 11, and 16 times greater in monkeys than in rats (Table 3; Fig. 5B). In contrast, the numbers of neurons in the central and medial nuclei were only 2.3 and 1.5 times greater in monkeys than in rats, respectively. Thus,

species differences in neuron numbers are substantial but not as striking as species differences in the volume of individual amygdaloid nuclei. This suggests that differences in neuron numbers are accompanied by even more important differences in other cellular components between rats and monkeys.

Interestingly, we found that the differences in the number of glial cells between rats and monkeys were about twice as large as differences in neuron number. We found that the numbers of glial cells in the monkey lateral, basal, and accessory basal nuclei were, respectively, 27, 24, and 36 times larger than in the rat. In the central and medial nuclei, there were, respectively, 6 and 3 times more glial cells in monkeys than in rats. Consequently, the ratio between the number of glial cells and the number of neurons was about two times greater in monkeys than in rats; this ratio was relatively constant across all amygdala nuclei (Table 3). Unfortunately, we were unable to reliably distinguish between astrocytes and oligodendrocytes in the rat (see Materials and Methods section). Thus, we could not determine experimentally if one type of glial cells underlies this species difference or if both astrocytes and oligodendrocytes contribute to this difference between rats and monkeys. No data were available for the human amygdala.

Neuronal soma size

We used the nucleator technique to estimate the volume of neuronal somas in the amygdala nuclei of rats and monkeys; Schumann and Amaral (2006) estimated neuronal somas in human cases with the same method (Table 4). Altogether, these data indicate that the average size of amygdala neurons is progressively larger from rats to monkeys to humans. Across the main amygdala nuclei, neuronal somas were about two times larger in monkeys than in rats and about two times larger in humans than in monkeys (Table 4; Fig. 5C). In rats and monkeys, neurons exhibited distinct size distributions in different amygdala nuclei (Fig. 6). In monkeys, the greater variation in neuronal soma size was clearly indicative of different neuron populations within the basal and accessory basal nuclei. The basal nucleus is well known to have a range of neuronal sizes that goes from larger dorsally to smaller ventrally (Freese and Amaral, 2009). The variation was less obvious in the lateral nucleus, even though this nucleus clearly contains different populations of neurons in both rats (Price et al., 1987) and monkeys (Pitkänen and Amaral, 1998). In general, amygdala neuron populations were more heterogeneous in monkeys than in rats, suggesting a potentially greater diversity of different cell types in primates.

Cell density and neuropil volume

Neuron density and neuropil volume provide additional information about the structural organization of a brain region that neuron number or overall volume measurements alone do not necessarily provide. We therefore calculated neuron density for the main amygdala nuclei (Table 4; Fig. 5D), based on our previous measures of volumes and neuron numbers (Tables 2, 3). Neuron density was between 2.36 (lateral nucleus) and 3.73 (basal nucleus) times lower in monkeys than in rats (average: 3.01), and between 3.00 (basal + paralaminar nuclei) and 4.67 (lateral nucleus) times lower in humans than in monkeys (average: 3.84). In the lateral nucleus, for example, there were on the order of 9,000 neurons per cubic millimeter in the human, 42,000 neurons per cubic millimeter in the monkey, and 99,000

neurons per cubic millimeter in the rat. These data suggest that the volume of the neuropil becomes progressively larger from rats, to monkeys, to humans. We therefore calculated the percentage of the volume occupied by neuronal somas in order to obtain an estimate of neuropil volume in individual amygdala nuclei (Table 4). On average, neuronal somas occupied $\approx 10\%$ of the volume of the amygdala in rats and $\approx 6\%$ in monkeys (Fig. 5E). Using data from Schumann and Amaral (2005, 2006), we estimated that the volume occupied by neuronal somas is $\approx 3\%$ in the human amygdala. Interestingly, the percentage of the volume of individual amygdala nuclei occupied by neuronal somas was inversely related to the size of the neuronal somas (Fig. 5C,E). The increase in neuropil volume was thus proportionally larger than the increase in neuronal soma size. Indeed, the percentage of amygdala volume occupied by the neuropil was progressively larger from rats (90%), to monkeys (94%), to humans (97%). In addition, neuron density in the different amygdala nuclei across the three different species was inversely related to the volume of neuronal somas (Fig. 7; $\log_{10}(\text{neuron density}) = 10.116 - 1.745 \times \log_{10}(\text{soma size})$; $R^2 = 0.85$; $F(1.59) = 342.444$; $P < 0.001$). Interestingly, glial density was also slightly lower in monkeys than in rats (Table 4), but this difference was more limited than that observed for neuron density (monkey/rat ratios: neuron density, 0.27–0.42; glia density, 0.60–0.93). Altogether, these results suggest that species differences in neuropil volume are likely due to differences in the dendritic and axonal arborizations of amygdala neurons, which might reflect an increased level of connectivity in primates.

DISCUSSION

Our stereological data in rats and monkeys provide fundamental, quantitative information regarding the morphological characteristics of the main amygdala nuclei in two species used as model organisms to understand the basic principles of human amygdala functions. We found: 1) that the lateral, basal, and accessory basal nuclei were dramatically more developed in monkeys than in rats, whereas the volume and neuron numbers in the central and medial nuclei differed far less between rats and monkeys; 2) a larger volume of the amygdala is made up of neuropil in monkeys than in rats. The higher percentage of neuropil volume implies a) a greater neuronal arborization and b) a greater number of glial cells relative to neuron number in primates (Fig. 8).

In this discussion, we first consider some methodological issues that might impact quantitative analyses of cell numbers and morphological features of the brain. We then evaluate our results in light of previous studies and conclude that our data provide a reliable reference regarding the morphological characteristics of the main amygdala nuclei in rats and monkeys. Finally, we discuss the theoretical and functional implications that derive from our quantitative structural findings.

Methodological considerations

There are a number of methodological issues that must be taken into consideration when evaluating the data we have presented. First, the rats and monkeys used in this study were reared in different environmental conditions. The monkeys were reared by their mothers in 2,000 m² outdoor naturalistic enclosures and lived in large social groups. In contrast, the rats

were retired breeders housed in standard laboratory cages. It is quite possible that different rearing conditions and life experiences might have influenced the development of the amygdala (Joseph, 1999; Okuda et al., 2009). However, the relatively small variations that might be induced by rearing conditions are very unlikely to account for the major differences in the morphological features, such as main amygdala nuclei volumes and neuron and glial cell numbers, observed between species. Moreover, the rearing conditions described here correspond to those typically employed in functional studies of the amygdala in rats (Pitkänen et al., 1997; LeDoux, 2000; Balleine and Killcross, 2006; Sigurdsson et al., 2007) and monkeys (Emery et al., 2001; Prather et al., 2001; Amaral et al., 2003; Bauman et al., 2004).

Second, data on the volume of individual nuclei were not corrected for potential differences in shrinkage of brain tissue during fixation. The rats and monkeys used in the current study were all perfused with the same fixative (4% PFA) following the same protocol used routinely in our laboratory. In contrast, the human brains used previously (Schumann and Amaral, 2005) were fixed by immersion in 10% formalin. We have previously shown, using monkey brains (Lavenex et al., 2009), that different methods of fixation (i.e., perfusion vs. immersion) using the same fixative solution (4% PFA in PB) have a significant impact on the neuroanatomical characteristics of the brain. In brief, neuropil volume is larger in immersion-fixed brains as compared to perfusion-fixed brains. In contrast, neuronal soma volume is smaller in immersion-fixed brains than in perfusion-fixed brains. Moreover, the differential shrinkage observed between immersion-fixed and perfusion-fixed tissue varies considerably between different cell types (Lavenex et al., 2009), so that it is impossible to determine any single correction factor that would enable direct comparison of these parameters for all cell populations. However, this issue is unlikely to affect the conclusions of our study, as we focused our comparisons on the relative rather than absolute volumes of the different amygdala nuclei in the different species. In contrast, the measurements of neuronal soma size in humans (Schumann and Amaral, 2006) might have underestimated true neuronal size, as compared to the perfusion-fixation condition that was used for rats and monkeys. If this hypothesis is true, then the actual differences in neuronal soma size between humans and the other two species might be even greater than that reported in the current study. The determination of the total number of neurons is not sensitive to fixation artifacts and potential differences in shrinkage, as the optical fractionator method is independent of the measure of the volume of the structure (West et al., 1991). Thus, the comparison of total neuron numbers in the main amygdala nuclei of rats and monkeys is valid and reliable.

Interlaboratory differences—Considering the important differences between laboratories that can sometimes outweigh differences between experimental conditions (Scorcioni et al., 2004; Altemus et al., 2005; see also discussion below), it is particularly important to consider that our quantitative data were derived from Nissl-stained, coronal brain sections processed following the same protocol in the laboratories of the two senior investigators (P.L.: rats and two monkeys; D.G.A.: two monkeys). The first author (L.J.C.) performed all quantitative analyses with the same microscope and stereological system in P.L.'s laboratory at the University of Fribourg. We compared the current data in rats and

monkeys with those obtained in humans and previously published by Schumann and Amaral (2005). The stereological analysis of human cases was performed by another experimenter (C.M.S.) in D.G.A.'s laboratory at the University of California, Davis. We have thus reduced the potential influence of an experimenter bias to a minimum in order to make sound species comparisons. Moreover, comparison of our comprehensive dataset with those of other studies performed in different laboratories further supports the reliability and generalization of our normative data (see discussion below and Supporting Information 4).

Normalization procedures—Some researchers have considered the need to normalize morphometric data in order to compare different parameters of brain structure that might vary between species or experimental conditions. However, a major hurdle in performing a sound normalization is to find an adequate reference that does not falsely impact the parameter under study (Lavenex et al., 2002, 2000b). For example, it would be easy to conclude that the amygdala is less developed in monkeys, as compared to rats, if one defines the size of the amygdala in relation to the size of the brain. This conclusion is obviously incorrect considering that 1) the actual volume and the numbers of neurons and glial cells are indeed greater in the monkey amygdala, as compared to rats; and 2) we do not know what factors might impact the development of the rest of the brain in different species. This is especially true for species that are only distantly related (Herculano-Houzel et al., 2007). Consequently, we did not use any normalization procedure in the current study. Instead, we compared directly the various parameters that we estimated and considered the implications that could be inferred from defined species differences in our specific estimates.

Comparison with previous studies

Rats—Previous studies of neuron number and nuclei volume in the rat amygdala have reported variable results (Supporting Information 4). Data from two separate studies, which considered the volume and neuron number of at least three of the main amygdala nuclei (Tuunanen and Pitkänen, 2000; Pêgo et al., 2008), differed significantly from each other but in a consistent manner as compared to our own results in the six main amygdala nuclei. The neuron numbers in the lateral, basal, and accessory basal nuclei of 6-month-old Sprague-Dawley rats reported by Tuunanen and Pitkänen (2000) were about 50% lower than our estimates. Similarly, the results on volume and neuron number in the lateral, basal, and central nuclei in 4-month-old Wistar rats reported by Pêgo et al. (2008) were consistently 20–30% below our estimates. As Pêgo et al. (2008) used different criteria to delineate the accessory basal nucleus, it is difficult to compare our estimate to theirs.

In contrast, our estimates of neuron number in the central and medial nuclei were consistent with the results reported by Fan et al. (2008). Our estimate of the volume of the central nucleus was also consistent with their results, but the volume of the medial nucleus that they reported was smaller than our own estimate; this is reasonable, however, since Fan et al. excluded the cell free layer I in their volume measurements. Our estimate of the number of neurons in the basal nucleus is consistent with previous estimates by Rubinow and Juraska (2008). Similarly, our estimates of neuronal soma size are consistent with those reported by Cooke et al. (2000) in the medial nucleus and by Berdel et al. (1997) in the lateral nucleus.

Altogether, the consistency of our results with those of a number of studies performed in different laboratories, together with the reliability of our stereological analysis system established in the monkey amygdala (see below) and hippocampal formation (Jabès et al., 2010, 2011), support the reliability of our stereological data on the volumes and neuron numbers in the main nuclei of the rat amygdala. Our data can therefore be considered a reliable base on which to build models and perform interspecies comparisons of amygdala structure and putative functional circuits.

Monkeys—Our results on nuclei volume and neuron number in the monkey amygdala are consistent with recent data reported by Carlo et al. (2010) (Supporting Information 4). Considering the volume of individual nuclei, the estimates of the paralamina nucleus are the only ones that differed by more than 20% between the two studies (29%). Considering the number of neurons, only the estimates of the central and medial nuclei differed by more than 10% between the two studies (central: 27%, medial: 17%). Since these differences are relatively small, they do not significantly impact the comparisons with the rat or human amygdala. Indeed, such small differences in the estimations of volume and neuron number performed by different experimenters in different laboratories, using different materials and methods, provide a strong validation of the current results. To our knowledge, there are no other data on neuron size and glia number in the rhesus monkey amygdala.

Humans—The stereological data on the human amygdala reported by Schumann and Amaral (2005) were also supported by those of another study, which found similar results in the lateral nucleus of the human amygdala (Kreczmanski et al., 2007) (Supporting Information 4). Despite differences in the delimitation of individual nuclei, Vereecken et al. (1994) found a total number of neurons in the whole amygdala that was close to that found by Schumann and Amaral (2005). The data on neuronal soma size were consistent with those of Aliashkevich et al. (2003) in the lateral and basal nuclei (Supporting Information 4). Data on neuronal soma size in humans from Berretta et al. (2007) were also consistent for the basal and accessory basal nuclei but not in the lateral nucleus, where the data reported by Berretta were 28% below those reported by Schumann and Amaral (2005). Bezchlibnyk et al. (2007) reported neuronal soma sizes twice as big as those found by Schumann and Amaral (2005). These differences could be due to the use and calibration of different analysis systems, or the use of paraffin-embedded brains by Bezchlibnyk et al. (2007) and the use of frozen sections by Schumann and Amaral (2005). Nevertheless, the data previously reported by Schumann and Amaral (2005) are largely consistent with the results of other studies and can thus be used as a reliable reference to establish sound comparisons with our current data in rats and monkeys.

Cell density and glia/neuron ratio

Our data suggested that the greater glia/neuron ratio in the primate amygdala (i.e., in both monkeys and humans) is mainly due to a lower density of neurons in primates as compared to rats. The amygdala glia/neuron ratio (total number of glia divided by the total number of neurons) is greater in humans (5.36, derived from the data on glia density published by Hamidi et al. (2004) and the volume measurements of Schumann and Amaral (2005)) than in monkeys (1.50) or rats (0.68). An increase in the glia/neuron ratio with increased

complexity of the brain between species has previously been reported in the cerebral cortex (Friede and Van Houten, 1962), as well as more specifically in frontal cortical areas within primates (Sherwood et al., 2006). In the human amygdala, Hamidi et al. (2004) estimated a glia density (oligodendrocytes + astrocytes; 50,254 cells/mm³) that is close to what we calculated in the monkey amygdala (53,591 cells/mm³). Although our study found glial density to be higher in the rat (78,819 cells/mm³), what is important is that the differences in glial density between species (1.55 times higher in rats than monkeys, and 1.55 times higher in rats than humans) are not nearly as pronounced as the differences in neuron density (3.2 times higher in rats than monkeys, and 12 times higher in rats than humans). These results indicate that, in contrast to neuron density, glial density is relatively constant across species.

The glia/neuron ratio that we determined in the monkey amygdala (1.50) was much higher than previous estimates of glia/neuron ratio (including microglia) from several other regions of the *Macaca mulatta* rhesus monkeys brain. Previously reported glia/neuron ratios were: 1.0 in the prefrontal cortex (Dombrowski et al., 2001), 0.46 in the visual cortex (O’Kusky and Colonnier, 1982), 0.56 in various frontal, parietal, temporal, and occipital regions (Christensen et al., 2007), and 0.82 across all cortical areas (Lidow and Song, 2001). As glia density is relatively constant across the brain, the higher glia/neuron ratio observed in the amygdala is more likely linked to a lower neuron density (34,000 neurons/mm³) as compared to neocortical areas (prefrontal cortex: 38,569–58,708 neurons/mm³ (Dombrowski et al., 2001); visual cortex: 120,000 neurons/mm³ (O’Kusky and Colonnier, 1982)). Similarly, neuron density in the human amygdala (9,000 neurons/mm³) appears to be markedly lower than neuron density in the human posterior orbitofrontal (area 13; 30,000 neurons/mm³) (Semendeferi et al., 1998) and prefrontal (area 10; 34,000 neurons/mm³) (Semendeferi et al., 2001) cortices. These differences in glia/neuron ratios reveal important structural and cellular variations in the organization of the different gray matter areas in the primate brain. The low neuron density of the amygdala, as compared to neocortical areas, could reflect a higher degree of intrinsic and extrinsic interconnectivity, and thus the functional specificity of this structure.

Neuropil

The lower neuronal density in the primate amygdala, as compared to rodents, is related to a larger neuropil volume, which is likely associated with greater complexity in the dendritic and axonal arborizations of individual amygdala neurons. This likely suggests that primates have a greater capacity to integrate information in specific amygdala circuits than do rats. A larger volume of neuropil in humans compared to monkeys has been shown in several cortical regions: Broca’s area (Schenker et al., 2008), the face area of the primary motor cortex (Sherwood et al., 2003), area 10 of the prefrontal cortex (Semendeferi et al., 2001), and the visual cortex (Zilles et al., 1982). Accordingly, a smaller neuronal density in humans as compared to monkeys has been shown in the posterior orbitofrontal cortex (area 13) (Semendeferi et al., 1998) and prefrontal cortex (area 10) (Semendeferi et al., 2001). In the rat amygdala, dendrites and axons represent more than 80% of the neuropil volume, glial cells 14% and synapses only 2% (Cooke et al., 2007). Although largely similar, glial density is slightly lower in monkeys and humans than in rats. The proportion of neuropil occupied by glial cells might thus be somehow compensated for by the greater size of glial cells in

humans than in monkeys, and in monkeys than in rats (Oberheim et al., 2006, 2009). Nevertheless, we also found in the stratum radiatum of the CA1 region of the adult monkey hippocampus that glial processes occupy less than 10% of the neuropil (Lavenex et al., 2011). The larger neuropil volume observed in primates therefore most likely reflects an increase in dendrites and axons. Accordingly, the complexity of the dendritic arborization of amygdala neurons is greater in monkeys than in rats (Rai et al., 2005; Mohandas Rao et al., 2009). Indeed, assumption-based 2D Scholl analyses of amygdala neurons reported between 22 and 29 intersections (between dendrites and the concentric analysis circles centered on the soma) for neurons randomly selected in the rat amygdala (Rai et al., 2005; Mohandas Rao et al., 2009), and between 125 to 210 dendritic intersections for neurons randomly selected in the monkey amygdala (Herzog, 1982; see Altemus et al., 2005 for data on the dendritic arborization of pyramidal neurons in the CA1 field of the rat and monkey hippocampus). Taken together, these results suggest a greater complexity of amygdala networks that might contribute to a greater integration of inputs reaching the amygdala in primates. This hypothesis is supported by the higher convergence of information through the cascade of intrinsic amygdala circuits.

Input convergence and integration of information in amygdala circuits

One obvious interpretation of our data is that the differences in the relative size and neuron numbers of the main amygdala nuclei observed between rats and monkeys are linked to their degree of connectivity with other brain structures. This hypothesis is in agreement with the theory that brain structures with major anatomical and functional links evolve together independently of evolutionary changes in other unrelated structures (Barton and Harvey, 2000). The lateral, basal, and accessory basal nuclei are more developed in primates than in rodents, and parallel the greater development of the neocortical areas with which these nuclei are interconnected in primates. As discussed above, the larger neuropil and greater neuronal soma size observed in the primate amygdala is associated with a greater complexity of the dendritic and local axonal arborizations, which could subsequently have a direct impact on the integration capacity of the functional network that includes this brain region. Here we consider the major connections of the main amygdala nuclei (Freese and Amaral, 2009), in rats and monkeys (the connectivity of the human amygdala remains largely unknown), in order to evaluate the hypothesis of a higher convergence and integration of information in the primate amygdala.

As discussed above, the relative development of the main amygdala nuclei might be influenced by their interconnections with other brain structures, i.e., their afferent and efferent connections (Stephan et al., 1987). In rats and monkeys the lateral, basal, and accessory basal nuclei are the main recipients and originators of neocortical-amygdala connections (Pitkänen, 2000; Freese and Amaral, 2009). Cortical projections to the amygdala are organized similarly in rats and monkeys (McDonald, 1998) but these connections are far more substantial in monkeys than in rats due to the dramatic expansion of the neocortex in primates (Super and Uylings, 2001). Similarly, the efferent projections from the amygdala to the neocortex are more substantial in primates. For example, the amygdala projects to primary sensory areas in monkeys (Amaral and Price, 1984; Iwai and Yukie, 1987; Amaral et al., 1992; Yukie, 2002) and in humans (Catani et al., 2003), but

these connections are apparently absent in rats (Krettek and Price, 1977). Consequently, and in contrast to rodents, the primate amygdala could influence the sensory processing at primary cortical stages (Freese and Amaral, 2009). The expansion of cortical areas and the greater complexity of cortical information reaching the amygdala are thus associated with a greater development of the amygdala nuclei interconnected with the neocortex.

In contrast, the central nucleus is connected mainly to visceral and autonomic systems (Pitkänen, 2000; Freese and Amaral, 2009). These systems are presumed to have been conserved throughout the course of evolution, thus leading to more limited species differences in the size and neuron number of the central nucleus. Similarly, the relatively small species differences in the volume and neuron number of the medial nucleus is paralleled by the highly conserved organization of the olfactory cortex with which it is interconnected (Crosby and Humphrey, 1944). Projections from the olfactory cortical areas are essentially the same in rats and monkeys, except that it represents a greater fraction of the total sensory inputs in rats than in monkeys, in which other sensory modalities have become dominant (Price, 1973; Carmichael et al., 1994).

The main amygdala nuclei, including the lateral, basal, accessory basal, central, and medial nuclei, are also characterized by their unique intraamygdala connections (see Freese and Amaral, 2009, for a comprehensive description). Within the amygdala circuitry, the connections between these different nuclei are highly unidirectional. The lateral nucleus projects to the basal, accessory basal, medial, and central nuclei, whereas the return projections are either very meager or totally nonexistent. The basal nucleus projects to the accessory basal, medial, and central nuclei, but the return projections are either meager or nonexistent. The accessory basal nucleus projects most strongly to the central nucleus and sends moderate projections to the medial nucleus, whereas it returns only light projections to the lateral and basal nuclei. The medial nucleus receives its strongest afferents from the lateral nucleus and moderate projections from the accessory basal nucleus. The medial nucleus projects most heavily to the central nucleus and returns only light projections to the basal and accessory basal nuclei, but not to the lateral nucleus. Finally, the central nucleus is the site of convergence of projections originating in all the other main amygdala nuclei. In contrast, the central nucleus projects only lightly to the lateral, basal, and accessory basal nuclei.

Interestingly, we found the largest species differences in neuron numbers at the earliest stages of the intrinsic amygdala circuitry, that is, within the lateral, basal, and accessory basal nuclei. Indeed, there were about 13 times more neurons in these three nuclei combined in monkeys (3.73 million) than in rats (290 thousand). In contrast, the central nucleus, upon which inputs from the other amygdaloid nuclei converge, exhibit relatively small species differences. There were only 2.3 times more neurons in monkeys (297 thousand) than in rats (127 thousand). Although fundamental amygdala functions are likely conserved across species, the relative importance of the functional processes carried out by distinct amygdalar circuits might differ between rodents and primates. Specifically, the functions subserved by the interconnections of the lateral, basal, and accessory basal nuclei with the neocortex in primates might be relatively preponderant compared to the functions subserved by the circuitry involving the central nucleus. Cortical information reaching the primate amygdala

via the lateral, basal, and accessory basal nuclei can be processed within these nuclei and sent back to the neocortex to modulate cortical activity and cognitive processes. Intrinsic amygdala projections can further process these inputs and forward highly integrated information in order to influence the function of the central nucleus. In addition, direct projections from the lateral, basal, and accessory basal nuclei can also directly contribute to the modulation of neuronal activity in various subcortical structures (see Freese and Amaral, 2009, for details regarding these connections). In sum, the basic functions carried out by the amygdala are likely conserved between species, even though the processes carried out by the central and medial nuclei might be under greater influence by cortical inputs in primates, via projections from the lateral, basal, and accessory basal nuclei. Thus, to illustrate the functional impact of these morphometric differences between species, let us consider that the amygdala serves essentially as a danger detector, in both rodents and primates, thus facilitating the survival of the individual animal. Although this survival function is essentially conserved in primates, because the primate amygdala can integrate additional contextual information, this additional information can further lead to the regulation of more complex behaviors such as the modulation of social interactions (Amaral et al., 2003). Going one step farther, as in the case of the human, where integration may be even greater, the amygdala is also activated when subjects are presented with untrustworthy faces (Adolphs et al., 1998; Winston et al., 2002), positive words (Hamann and Mao, 2002), happy faces (Canli et al., 2002), amusement-inducing movies (Aalto et al., 2002), and other complex social stimuli that might not have an equivalent in rodents. Thus, the highly developed lateral nucleus of the monkey and human amygdala has greater computational resources to carry out learning and template matching for a wide range of environmental stimuli.

CONCLUSION

This quantitative study has revealed important differences in the structural organization and cellular components of the main amygdala nuclei in rats, monkeys, and humans. First, the primate amygdala is characterized by a lower neuron density, larger neuropil volume, and greater glia/neuron ratio than the rat amygdala. These fundamental differences in the structural organization of the amygdala could be linked to a greater complexity and integration capacity of sensory information within the intrinsic amygdala circuits in primates. Second, the lateral, basal, and accessory basal nuclei were dramatically more developed in monkeys than in rats, whereas the volume and neuron numbers in the central and medial nuclei differed relatively little between rats and monkeys. This suggests that although the fundamental function of the amygdala, to regulate fear and emotional learning, is conserved across species, amygdala function might be under greater influence of cortical activity in primates, and therefore integrate additional contextual information that influences the regulation of more complex behaviors such as social interactions.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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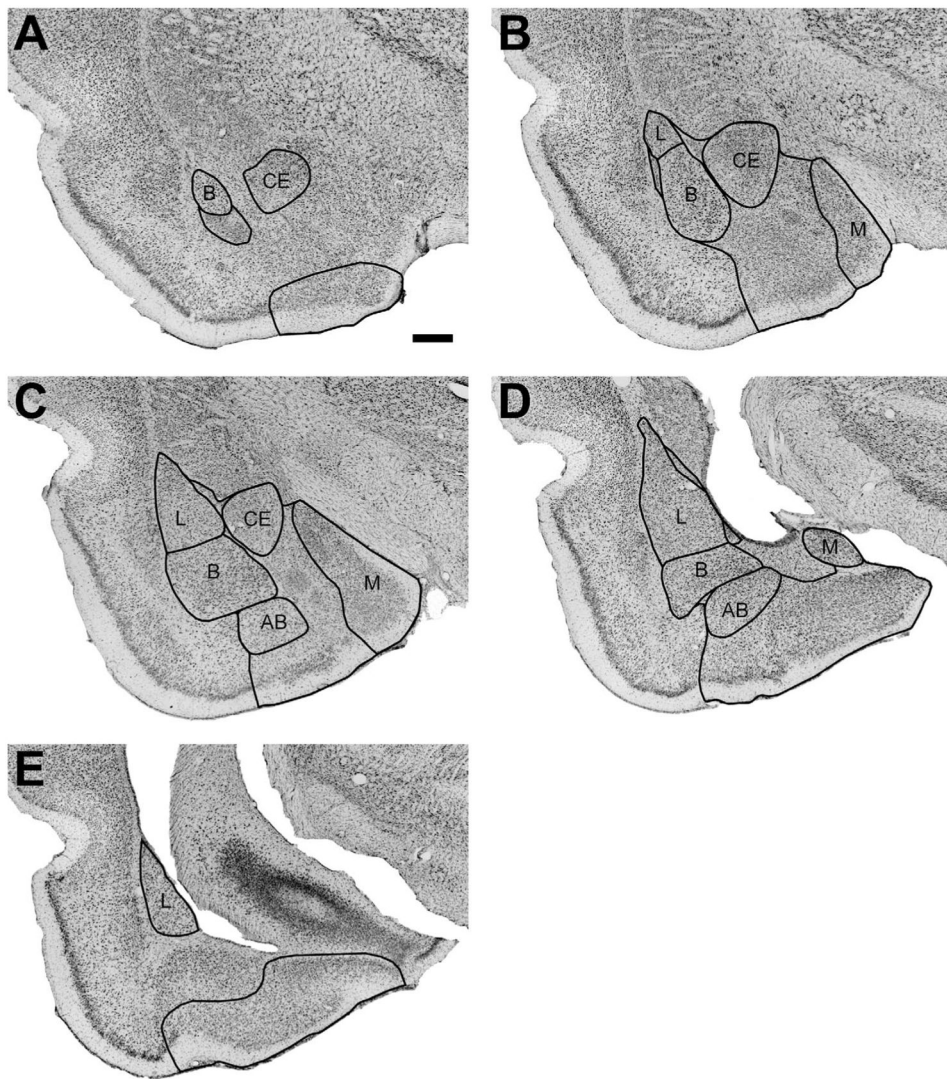


Figure 1. Low-magnification photomicrographs of representative coronal sections through the rat amygdala illustrating the locations of the main nuclei. L, lateral; B, basal; AB, accessory basal; CE, central; M, medial. Nonlabeled areas represent remaining nuclei of the amygdala; see list in Table 1. Scale bar = 500 μ m in A (applies to all panels).

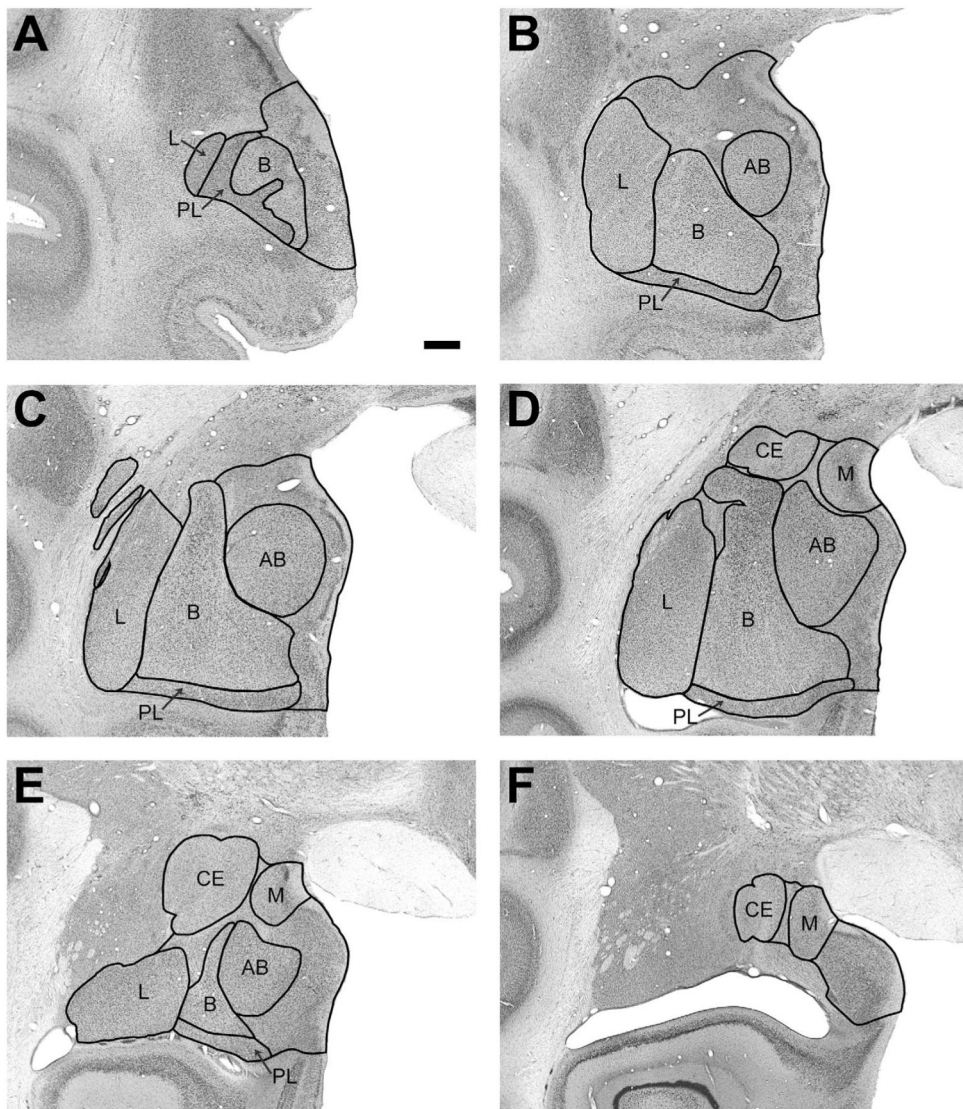


Figure 2. Low-magnification photomicrographs of representative coronal sections through the monkey amygdala illustrating the locations of the main nuclei. L, lateral; B, basal; PL, paralamina; AB, accessory basal; CE, central; M, medial. Nonlabeled areas represent remaining nuclei of the amygdala; see list in Table 1. Scale bar = 1 mm in A (applies to all panels).

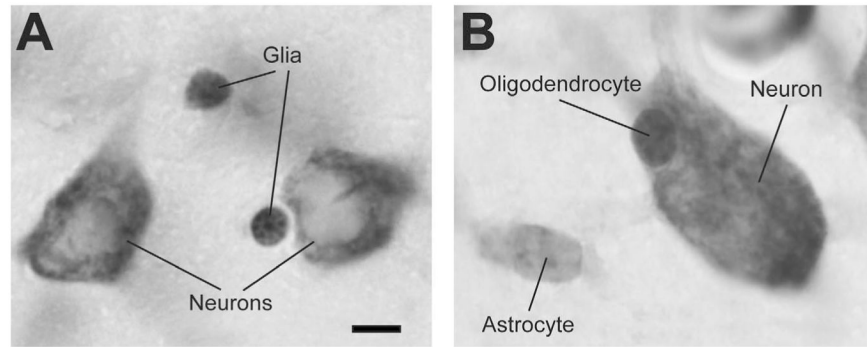


Figure 3.

Classification and identification of different cell types in rats and monkeys. **A:** Typical neuron and glial cells viewed with a 100 \times objective in the rat amygdala basal nucleus as revealed in Nissl-stained sections cut at 40 μm . **B:** Typical neuron and glial cells viewed with a 100 \times objective in the monkey amygdala basal nucleus as revealed in Nissl-stained sections cut at 60 μm . Note that we were able to reliably distinguish between astrocytes and oligodendrocytes in monkeys, but not in rats, based on morphological criteria (see main text for details). Scale bar = 5 μm in A (applies to B).

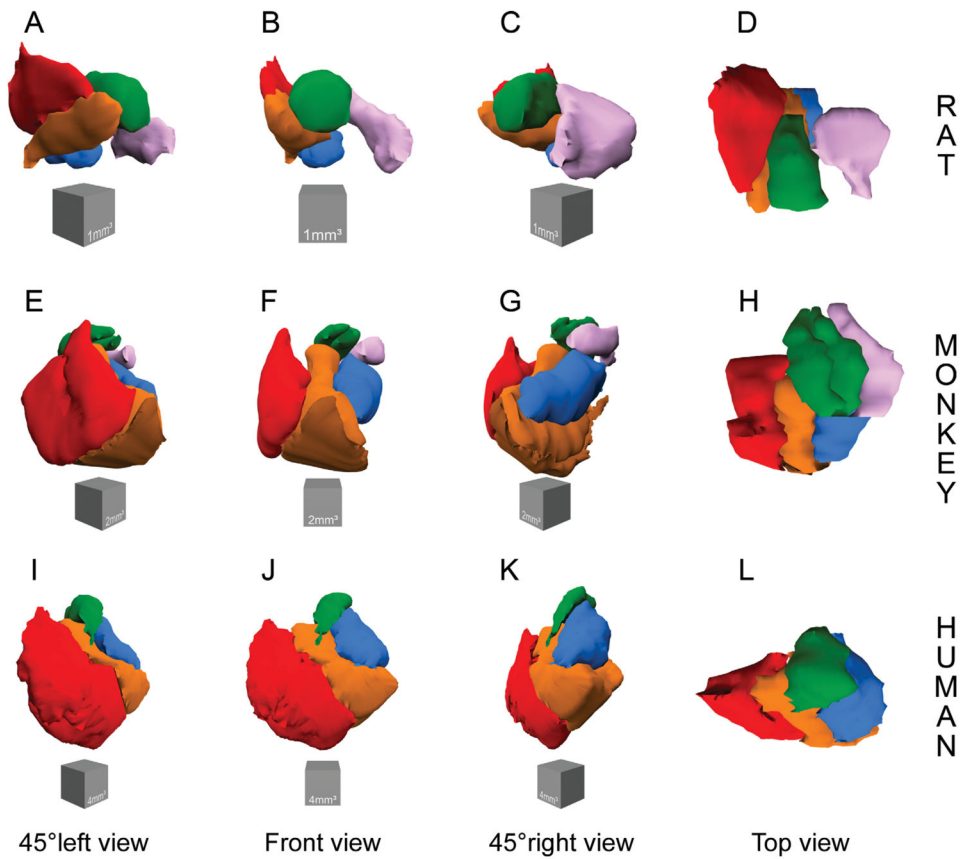
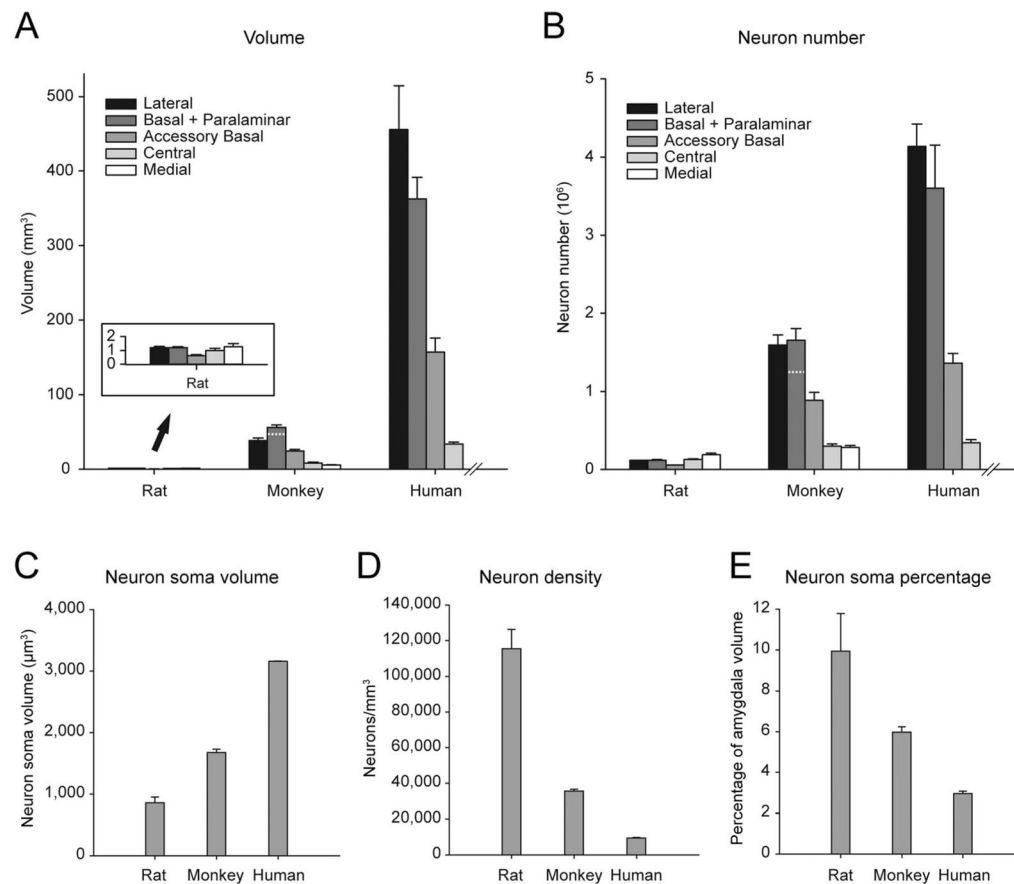


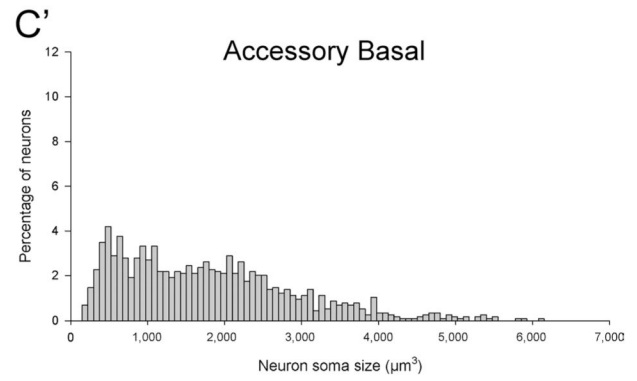
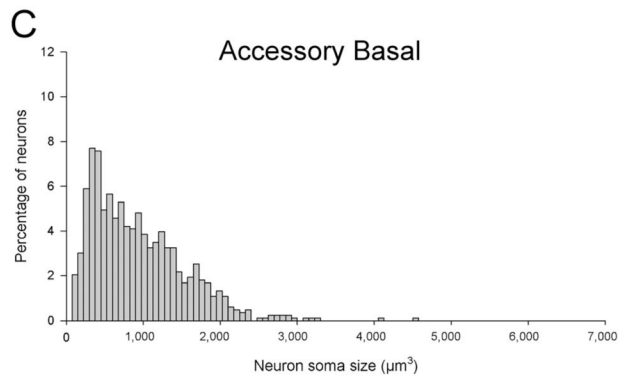
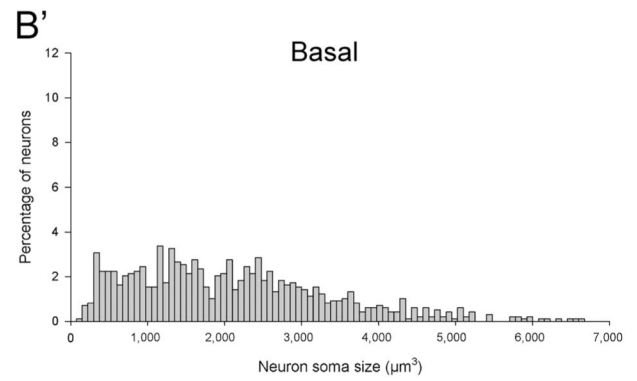
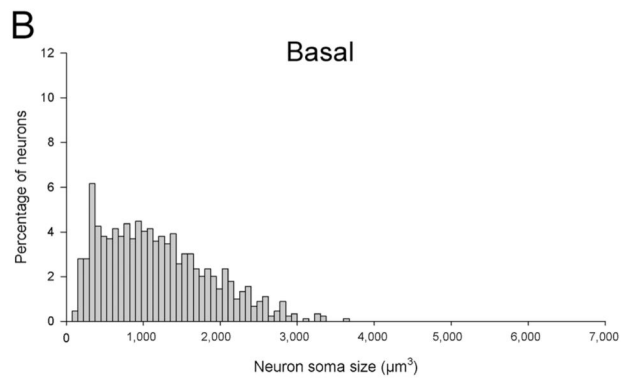
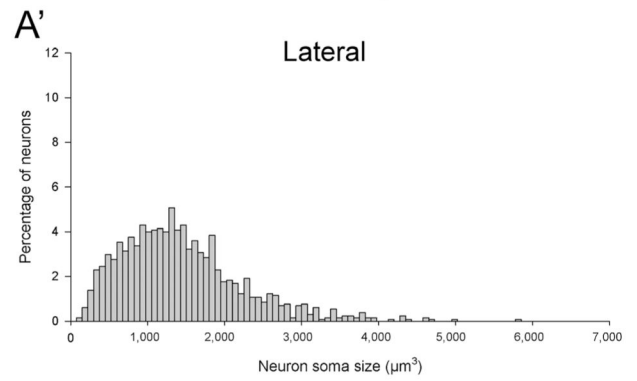
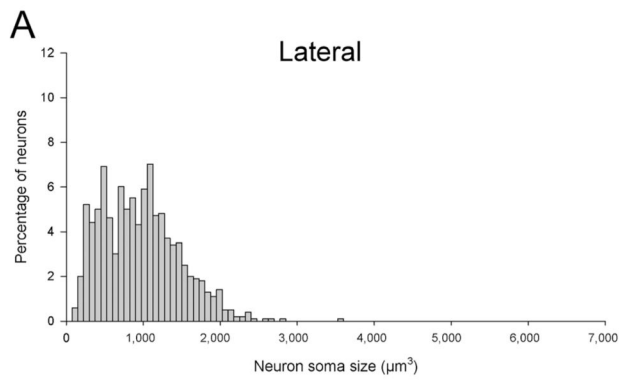
Figure 4. Three-dimensional reconstruction of the main amygdala nuclei in rat (A–D), monkey (E–H), and human (I–L). Lateral nucleus is in red, basal nucleus is in orange, paralaminar nucleus is in dark orange (in monkeys only), accessory basal nucleus is in blue, central nucleus is in green, and medial nucleus is in pink (not represented in humans). Scales: rat, gray cube is 1 mm³; monkey, gray cube is 2 mm³; human, gray cube is 4 mm³. 3D reconstructions are available in movie format as Supporting Information 2 and 3 online.

**Figure 5.**

A: Volumes of the main amygdala nuclei in rats, monkeys, and humans. The deep nuclei (lateral, basal, accessory basal) are relatively more developed in monkeys and humans than in rats. Note that the volume of the human medial nucleus was not estimated by Schumann and Amaral (2005). A dashed line in the bar representing the basal nucleus in monkeys indicates the volume of the paralaminar nucleus (upper part of the bar), and is included to enable the comparison with the human data (which included the paralaminar nucleus in the basal nucleus). **B:** Neuron numbers in the main amygdala nuclei of rats, monkeys, and humans. Note that neuron number in the human medial nucleus was not estimated by Schumann and Amaral (2005). A dashed line in bar representing the basal nucleus in monkeys indicates the number of neurons of the paralaminar nucleus (upper part of the bar), and is included to enable the comparison with the human data. **C:** Average neuronal soma size in the amygdala of the rat, monkey, and human. **D:** Average neuron density in the amygdala of the rat, monkey, and human. **E:** Percentage of total amygdala volume occupied by neuronal somas in the amygdala of the rat, monkey, and human. A lower percentage is associated with greater development of the neuropil. Error bars \pm SD.

Rat

Monkey



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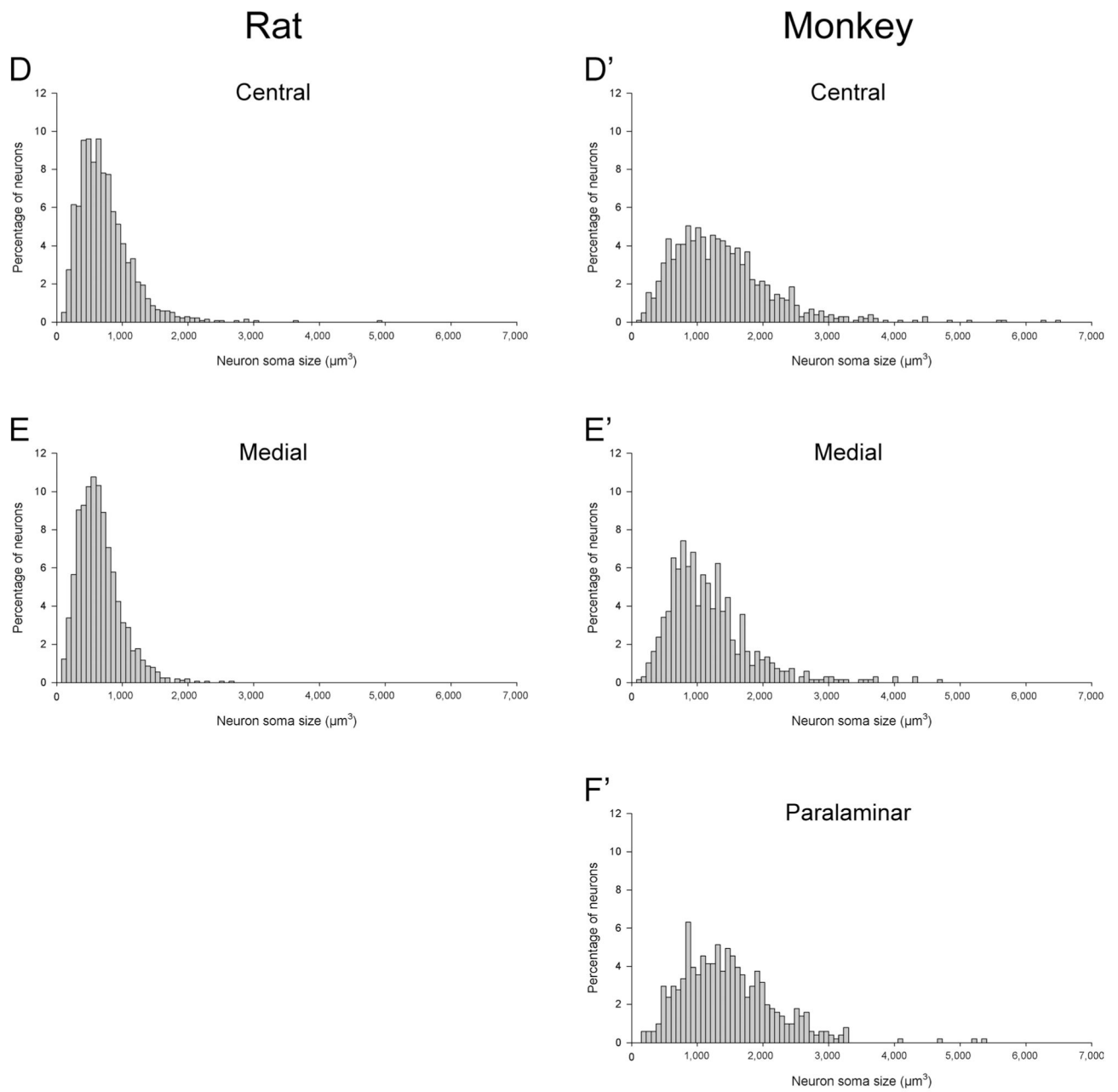


Figure 6.

Distributions of neuronal soma size in the main amygdaloid nuclei of rats (A–E) and monkeys (A'–F'). A, A': Lateral nucleus; B, B': basal nucleus; C, C': accessory basal nucleus; D, D': central nucleus; E, E': medial nucleus; F': paralamniar nucleus (only in monkeys).

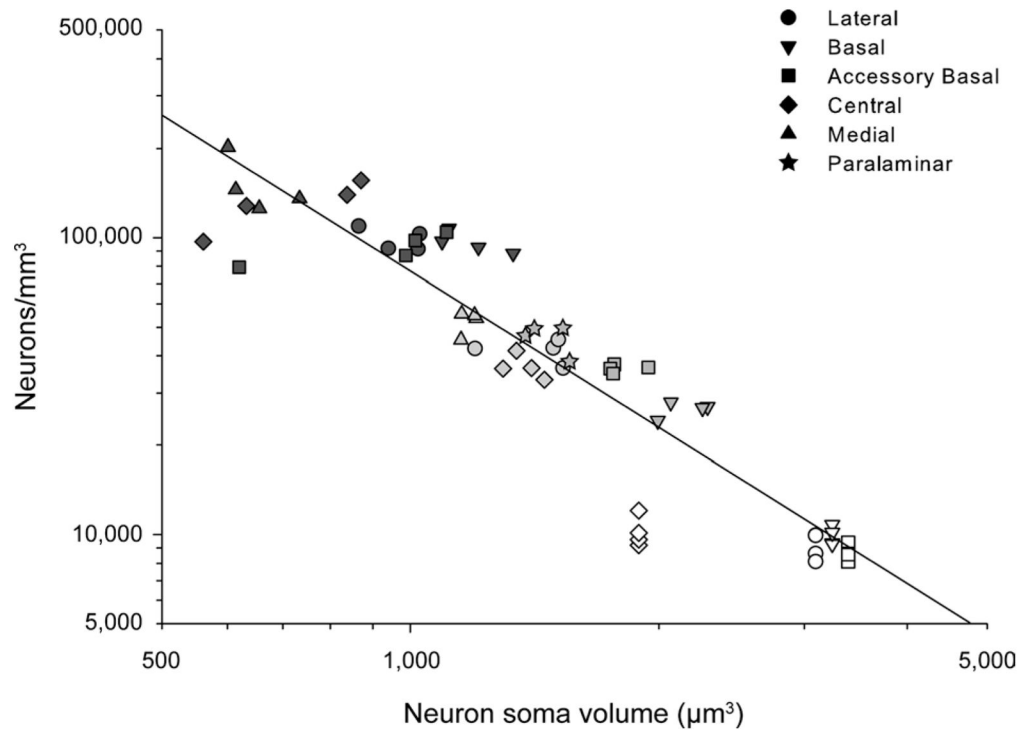


Figure 7.

Relationship between neuronal density and neuronal soma size in the main nuclei of the rat, monkey, and human amygdala. Logarithmic regression of neuron density and neuron soma size in the amygdala ($\log_{10}(\text{neuron density}) = 10.116 - 1.745 \times \log_{10}(\text{soma size})$; $R^2 = 0.85$; $F(1.59) = 342.44$; $P < 0.001$). Rat, dark gray; monkey, gray; human, white. Lateral nucleus, circles; basal nucleus, upside-down triangles; accessory basal nucleus, squares; central nucleus, diamonds; medial nucleus, triangles; paralaminar (only in monkeys), stars.

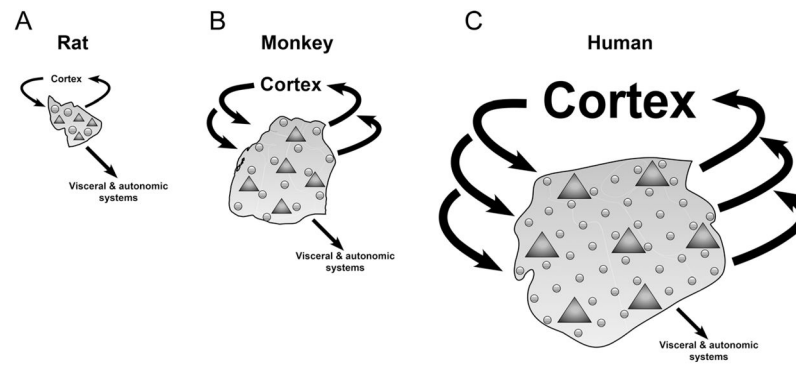


Figure 8.

Summary representation of the amygdala in rats (A), monkeys (B), and humans (C). Glial cells, circles; neurons, triangles: rats < monkeys < humans. Neuron density: rats > monkeys > humans. Glia density: rats = monkeys = humans. Glia/neuron ratio: rats < monkeys < humans. Connectivity with visceral and autonomic systems (mainly via the central nucleus): rats = monkeys = humans. Connectivity with cortical systems (between the neocortex and the lateral, basal, and accessory basal nuclei): rats < monkeys < humans. The proportions of the different parameters are not precisely scaled.

TABLE 1

Nomenclature of the Different Subdivisions of the Amygdaloid Complex in the Rat (*Rattus norvegicus*), Monkey (*Macaca mulatta*), and Human (*Homo sapiens*)

Rat	Monkey	Human
lateral nucleus (L)	lateral nucleus (L)	lateral nucleus (L)
dorsolateral division (Ldl)	dorsal division (Ld)	lateral division (Ll)
ventrolateral division (Lvl)	dorsal intermediate division (Ldi)	medial division (Lm)
medial division (Lm)	ventral intermediate division (Lvi)	
	ventral division (Lv)	
basal nucleus (B)	basal nucleus (B)	basal nucleus (B)
magnocellular division (Bmc)	magnocellular division (Bmc)	magnocellular division (Bmc)
intermediate division (Bi)	intermediate division (Bi)	intermediate division (Bi)
parvicellular division (Bpc)	parvicellular division (Bpc)	parvicellular division (Bpc)
accessory basal nucleus (AB)	accessory basal nucleus (AB)	accessory basal nucleus (AB)
magnocellular division (Abmc)	magnocellular division (ABmc)	magnocellular division (ABmc)
parvicellular division (ABpc)	parvicellular division (ABpc)	parvicellular division (ABpc)
	ventromedial division (ABvm)	ventromedial division (ABvm)
central nucleus (CE)	central nucleus (CE)	central nucleus (CE)
capsular division (CEc)	medial division (CEm)	medial division (CEm)
lateral division (CEl)	lateral division (CEl)	lateral division (CEl)
intermediate division (CEi)		
medial division (CEm)		
medial nucleus (M)	medial nucleus (M)	medial nucleus (M)
rostral division (Mr)		
central division		
dorsal part (Mcd)		
ventral part (Mcv)		
caudal division (Mc)		
–	paralaminar nucleus (PL)	paralaminar nucleus (PL)
intercalated nuclei (I)	intercalated nuclei (I)	intercalated nuclei (I)
anterior amygdaloid area (AAA)	anterior amygdaloid area (AAA)	anterior amygdaloid area (AAA)
nucleus of the lateral olfactory tract (NLOT)	nucleus of the lateral olfactory tract (NLOT)	nucleus of the lateral olfactory tract (NLOT)
anterior cortical nucleus (COa)	anterior cortical nucleus (COa)	anterior cortical nucleus (COa)
periamygdaloid cortex (PAC)	periamygdaloid cortex (PAC)	periamygdaloid cortex (PAC)
medial division (PACm)	(PAC2)	(PAC1)
sulcal division (PACs)	(PAC3)	(PAC3)
	sulcal division (PACs)	sulcal division (PACs)
	oral portion (PACo)	oral portion (PACo)
posterior cortical nucleus (COp)	posterior cortical nucleus (COp)	posterior cortical nucleus (COp)
amygdalo-hippocampal area (AHA)	amygdalo-hippocampal area (AHA)	amygdalo-hippocampal area (AHA)
medial division (AHAm)	ventral division (AHA _v)	
lateral division (AHA _l)	dorsal division (AHA _d)	

Rat	Monkey	Human
bed nucleus of the accessory olfactory tract (BAOT)	–	–
Pitkänen (2000)	Amaral et al. (1992)	Sorvari et al. (1996)

TABLE 2
Volume of the Brain, Amygdala, and Main Amygdala Nuclei in the Rat, Monkey, and Human

	Mean (mm ³)			Relative ² (%)			Ratio		
	Rat	Monkey	Human ¹	R	M	H	M/R	H/M	H/R
Brain ³	994	52,360	—	—	—	—	53	—	—
SD	110	4,875	—	—	—	—	—	—	—
Amygdala	10.62	192.60	1,416.20	100	100	100	18	7	133
SD	1.02	6.26	132.49	—	—	—	—	—	—
Lateral	1.19	38.40	455.63	11	20	32	32	12	382
SD	0.10	3.57	58.62	—	—	—	—	—	—
Basal	1.20	47.15	—	11	24	—	39	—	—
SD	0.06	2.67	—	—	—	—	—	—	—
Basal + Paralaminar	—	55.98	362.33	—	29	26	—	6	302
SD	—	3.36	29.22	—	—	—	—	—	—
Accessory Basal	0.63	24.38	157.00	6	13	11	39	6	251
SD	0.07	2.14	18.77	—	—	—	—	—	—
Central	1.00	8.15	33.73	9	4	2	8	4	34
SD	0.14	1.44	2.84	—	—	—	—	—	—
Medial	1.28	5.42	—	12	3	—	4	—	—
SD	0.23	0.50	—	—	—	—	—	—	—
Paralaminar	—	8.84	—	—	5	—	—	—	—
SD	—	0.90	—	—	—	—	—	—	—
Remaining ⁴	6.60	65.70	407.53	62	34	29	10	6	62
SD	0.90	14.17	40.89	—	—	—	—	—	—

H: human ; M: monkey ; R: rat.

¹ Data reported from Schumann and Amaral (2005).

² Refers to the relative percentage of volume occupied by each nucleus in the amygdala. Amygdala volume equals to 100%.

³ Refers to the volume of the telencephalon and diencephalon bilaterally.

⁴ Remaining contains the medial nucleus, intercalated nuclei, anterior amygdaloid area, nucleus of the lateral olfactory tract, anterior cortical nucleus, periamygdaloid cortex, posterior cortical nucleus, amygdalo-hippocampal area and bed nucleus of the accessory olfactory tract (in rat).

TABLE 3

Cell Numbers in the Main Nuclei of the Rat, Monkey, and Human Amygdala

		Mean			Ratio		
		Rat	Monkey	Human ¹	M/R	H/M	H/R
Lateral	Neuron	117,486	1,592,284	4,136,500	13.6	2.6	35.2
	SD	1,229	128,381	286,150			
	Glia	80,744	2,156,678	—	26.7	—	—
	SD	7,393	189,246	—			
	Oligodendrocytes	—	1,112,480	—	—	—	—
	SD	—	131,622	—	—	—	—
	Astrocytes	—	1,044,198	—	—	—	—
	SD	—	64,819	—	—	—	—
Basal	Glia/neuron ratio	0.7	1.4	—	2.0	—	—
	Neuron	115,774	1,247,181	—	10.8	—	—
	SD	13,793	140,356	—	—	—	—
	*Neuron (B + PL)	—	1,655,232	3,601,500	—	2.2	31.1
	SD	—	148,516	552,180	—	—	—
	Glia	106,521	2,513,417	—	23.6	—	—
	SD	15,942	175,853	—	—	—	—
	Oligodendrocytes	—	1,326,864	—	—	—	—
	SD	—	94,497	—	—	—	—
	Astrocytes	—	1,186,553	—	—	—	—
	SD	—	82,050	—	—	—	—
	Glia/neuron ratio	0.9	2.0	—	2.2	—	—
Accessory basal	Neuron	57,017	885,352	1,357,250	15.5	1.5	23.8
	SD	1,800	101,469	125,680	—	—	—
	Glia	35,780	1,273,935	—	35.6	—	—
	SD	7,010	89,538	—	—	—	—
	Oligodendrocytes	—	572,196	—	—	—	—
	SD	—	82,235	—	—	—	—
	Astrocytes	—	701,739	—	—	—	—
	SD	—	—	—	—	—	—

		Mean			Ratio		
		Rat	Monkey	Human ¹	M/R	H/M	H/R
	SD		23,552				
Central	Glia/neuron ratio	0.6	1.4	—	2.3	—	—
	Neuron	127,174	297,079	344,500	2.3	1.2	2.7
	SD	10,434	30,889	39,602			
	Glia	86,691	486,762	—	5.6	—	—
	SD	11,387	39,116				
	Oligodendrocytes	—	210,656	—	—	—	—
	SD		28,544				
	Astrocytes	—	276,106	—	—	—	—
	SD		15,546				
	Glia/neuron ratio	0.7	1.6	—	2.4	—	—
Medial	Neuron	188,742	282,622	—	1.5	—	—
	SD	19,688	25,053				
	Glia	103,936	338,080	—	3.3	—	—
	SD	9,036	26,695				
	Oligodendrocytes	—	102,661	—	—	—	—
	SD		13,889				
	Astrocytes	—	235,419	—	—	—	—
	SD		17,978				
	Glia/neuron ratio	0.6	1.2	—	2.2	—	—
	Paralaminar	Neuron	—	408,051	—	—	—
SD			49,453				
Glia		—	288,184	—	—	—	—
SD			21,186				
Oligodendrocytes		—	96,422	—	—	—	—
SD			23,397				
Astrocytes		—	191,762	—	—	—	—
SD			28,505				
Glia/neuron ratio		—	0.7	—	—	—	—

H: human ; M: monkey ; R: rat.

Data reported from Schumann and Amaral (2005).

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TABLE 4

Neuron Soma Volume and Cell Density in the Main Nuclei of the Rat, Monkey, and Human Amygdala

	Mean			Ratio		
	Rat	Monkey	Human ¹	M/R	H/M	H/R
Lateral						
Neuron soma volume ²	963	1,433	3,099	1.49	2.16	3.22
SD	76	158	62			
Neuron density ³	99,000	42,000	9,000	0.42	0.21	0.09
SD	8,400	3,700	900			
Glial density ³	68,000	57,000	—	0.84	—	—
SD	10,200	8,700	—			
% of nuclei volume ⁴	9.55	5.96	2.84	0.62	0.48	0.30
SD	1.26	0.79	0.29			
Neuron soma volume	1,187	2,154	3,245	1.81	1.51	2.73
SD	109	144	88			
Neuron density	97,000	26,000	—	0.27	—	—
SD	13,200	1,600	—			
Neuron density (B + PL)	—	30,000	10,000	—	0.33	0.10
SD	—	1,200	700			
Glial density	89,000	53,000	—	0.60	—	—
SD	11,900	4,800	—			
% of nuclei volume	11.44	5.69	3.21	0.50	0.56	0.28
SD	1.39	0.61	0.23			
Neuron soma volume	932	1,804	3,393	1.94	1.88	3.64
SD	214	92	72			
Neuron density	92,000	36,000	9,000	0.39	0.25	0.10
SD	11,100	1,000	500			
Glial density	57,000	53,000	—	0.93	—	—
SD	7,400	5,200	—			
% of nuclei volume	8.74	6.54	2.94	0.75	0.45	0.34
SD	2.82	0.41	0.19			
Neuron soma volume	726	1,374	1,893	1.89	—	—
SD	—	—	—			
Central						
Neuron soma volume	726	1,374	1,893	1.89	—	—
SD	—	—	—			

	Mean			Ratio		
	Rat	Monkey	Human ¹	M/R	H/M	H/R
SD	152	69	44			
Neuron density	130,000	37,000	10,000	0.28	0.27	0.08
SD	24,900	3,500	1,200			
Glia density	89,000	61,000	—	0.69	—	—
SD	21,500	6,200	—			
% of nuclei volume	9.70	5.06	1.94	0.52	0.38	0.20
SD	3.64	0.40	0.24			
Neuron soma volume	651	1,176	—	1.81	—	—
SD	60	27	—			
Neuron density	150,000	52,000	—	0.35	—	—
SD	19,300	4,700	—			
Glia density	85,000	63,000	—	0.74	—	—
SD	24,400	10,800	—			
% of nuclei volume	9.81	6.16	—	0.63	—	—
SD	1.71	0.63	—			
Neuron soma volume	—	1,470	—	—	—	—
SD	—	87	—			
Neuron density	—	46,000	—	—	—	—
SD	—	5,300	—			
Glia density	—	33,000	—	—	—	—
SD	—	5,100	—			
% of nuclei volume	—	6.80	—	—	—	—
SD	—	0.70	—			

H: human ; M: monkey ; R: rat.

¹ Data reported from Schumann and Amaral (2005) and (2006).

² Neuron soma volume in cubic micrometer.

³ Cell number per cubic millimeter.

⁴ Refers to the fraction of the amygdala nuclei volume occupied by neuronal somas.