

Targeting the tumor stroma in hepatocellular carcinoma

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Abstract

Hepatocellular carcinoma (HCC) is one of the most common and deadly cancers worldwide. In ninety percent of the cases it develops as a result of chronic liver damage and it is thus a typical inflammation-related cancer characterized by the close relation between the tumor microenvironment and tumor cells. The stromal environment consists out of several cell types, including hepatic stellate cells, macrophages and endothelial cells. They are not just active bystanders in the pathogenesis of HCC, but play an important and active role in tumor initiation, progression and metastasis. Furthermore, the tumor itself influences these cells to create a background that is beneficial for sustaining tumor growth. One of the key players is the hepatic stellate cell, which is activated during liver damage and differentiates towards a myofibroblast-

like cell. Activated stellate cells are responsible for the deposition of extracellular matrix, increase the production of angiogenic factors and stimulate the recruitment of macrophages. The increase of angiogenic factors (which are secreted by macrophages, tumor cells and activated stellate cells) will induce the formation of new blood vessels, thereby supplying the tumor with more oxygen and nutrients, thus supporting tumor growth and offering a passageway in the circulatory system. In addition, the secretion of chemokines by the tumor cells leads to the recruitment of tumor associated macrophages. These tumor associated macrophages are key actors of cancer-related inflammation, being the main type of inflammatory cells infiltrating the tumor environment and exerting a tumor promoting effect by secreting growth factors, stimulating angiogenesis and influencing the activation of stellate cells. This complex interplay between the several cell types involved in liver cancer emphasizes the need for targeting the tumor stroma in HCC patients.

Key words: Hepatocellular carcinoma; Stellate cells; Cirrhosis; Angiogenesis; Macrophages; Inflammation

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Core tip: Hepatocellular carcinoma is a primary liver tumor that usually develops in a background of chronic liver disease and fibrosis. It is the underlying chronic inflammation that creates an environment that not only causes but also enhances the formation and growth of tumors. The stromal compartment-including hepatic stellate cells, macrophages and endothelial cells-actively contribute to tumorigenesis, while the tumor itself influences these cells to create a background that is beneficial for tumor growth. This review focuses on the interplay between stroma and tumor cells, as well as therapeutic strategies that aim to target these complex interactions.

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INTRODUCTION

Hepatocellular carcinoma (HCC) is a primary liver tumor that usually develops in a background of chronic liver disease. It is the underlying chronic inflammation that creates an environment that not only causes but also enhances the formation and growth of tumors. Firstly, the continuous state of inflammation as a result of sustained liver damage can lead to hepatocyte cell death as well as compensatory proliferation, which can generate an accumulation of genomic lesions in hepatocytes. Secondly, the initiated cells are surrounded by an inflammatory niche that facilitates their progression towards malignant tumors. For instance, the fibrotic liver is characterized by an increased formation of blood vessels^[1], which will benefit tumor cells for their blood supply as well as facilitating metastasis^[2]. In addition, several factors produced by macrophages and activated stellate cells are known to directly stimulate and enhance tumor growth. Once the cancer has been established, the microenvironment continues to regulate the tumor behavior, influencing the development, progression and even response to therapy. All players within the tumor stroma strongly interact with each other, creating an environment that supports tumor growth (Figure 1). It is therefore not unlikely that future therapies will more and more focus on targeting these complex interactions in the tumor stroma (Figure 2). Ongoing clinical trials are listed in Table 1.

HEPATIC STELLATE CELLS

One major player in the formation of the perfect tumor environment is the activated hepatic stellate cell (HSC)^[3,4]. During liver injury, the stellate cells undergo a transformation from quiescent cells that serve as the liver's resident vitamin-A storing cells, towards "activated" myofibroblast-like cells. These activated HSCs are characterized by increased proliferation and contractility, altered matrix protease activity and the secretion of extracellular matrix (ECM) proteins, as well as tumor growth factors and pro-angiogenic factors.

Several studies have shown that co-culturing HSC with different HCC cell lines induces phenotypic changes in the behavior of the tumor cells^[5,6]. *In vitro* studies show that HSCs can directly influence the tumor cells (through the secretion of growth factors^[7], matrix proteases^[8] and/or ECM proteins^[9]) and there is also evidence from *in vivo* studies that activated HSCs can create an immunosuppressive environment that promotes HCC growth^[10,11]. The

interaction between the tumor cells and HSCs is bidirectional, thereby allowing the tumor to alter the stellate cells (and the overall stromal environment) towards a more pro-tumoral phenotype^[8]. Consistent with these findings, several *in vivo* studies have shown that inducing stellate cell activation increases liver fibrosis and hepatocarcinogenesis^[12-15].

One of the key factors in this HSC-HCC cross talk is transforming growth factor (TGF)- β ^[14,16,17]. Activated HSC are the main source of TGF- β , however most liver cells (including malignant hepatocytes) have the ability to produce TGF- β as well. The TGF- β signaling pathway consists of three distinct ligands, TGF- β 1, TGF- β 2, and TGF- β 3 which all bind to a specific receptor by first engaging with the TGF- β R1, which then heterodimerizes with the TGF- β R2. This causes the phosphorylation of Smad2 and 3, initiating an activation cascade leading to the induction several nuclear transduction proteins. Alternative pathway activation is possible, including the activation of AKT and other intracellular activation proteins. Interestingly, Smad7 antagonizes TGF- β mediated activation of hepatic stellate cells and protects against liver damage^[18]. TGF- β signaling promotes HCC by several distinct mechanisms (reviewed more in detail by Dooley *et al.*^[17]): firstly, through functioning as a growth factor, by which it can act oncogenic or as tumor suppressor depending on the temporal and spatial availability of TGF- β in tumor and stromal cells^[19,20]. And secondly, by transforming HSC to activated myofibroblasts. Interestingly, Inhibitors of TGF- β signaling have been shown to block HCC in different experimental models^[21], leading to the clinical investigation of the TGF- β inhibitor LY2157299 (NCT01246986 and NCT02178358). LY2157299 is a small molecule kinase inhibitor that binds to TGF- β R1 and hence inhibits TGF- β signaling.

The connective tissue growth factor (CTGF) is an extracellular matrix-associated heparin binding protein that is overexpressed in fibrotic lesions, and the overexpression correlates with the severity of fibrosis and can be linked to malignant transformation in patients with chronic hepatitis B^[22]. CTGF is a downstream mediator of some TGF- β effects and it is induced by TGF- β in a SMAD2/3 and stat3 dependent way. Furthermore, IL-13 is able to induce CTGF expression in HSCs by activating TGF- β -independent Smad signaling *via* the Erk-MAPK pathway instead of the canonical JAK/Stat6 pathway^[23]. CTGF expression in HSC leads to increased migration, proliferation, and collagen expression of these cells. In addition, studies have shown that TGF- β can elicit a direct effect on hepatocytes *via* CTGF, thus making it an interesting therapeutic target for multiple cell types involved in the fibrogenesis^[18]. CTGF blocking antibodies have been tested in patients with idiopathic pulmonary fibrosis (NCT00074698) and animal studies have shown that CTGF-inhibition prevents liver fibrosis in

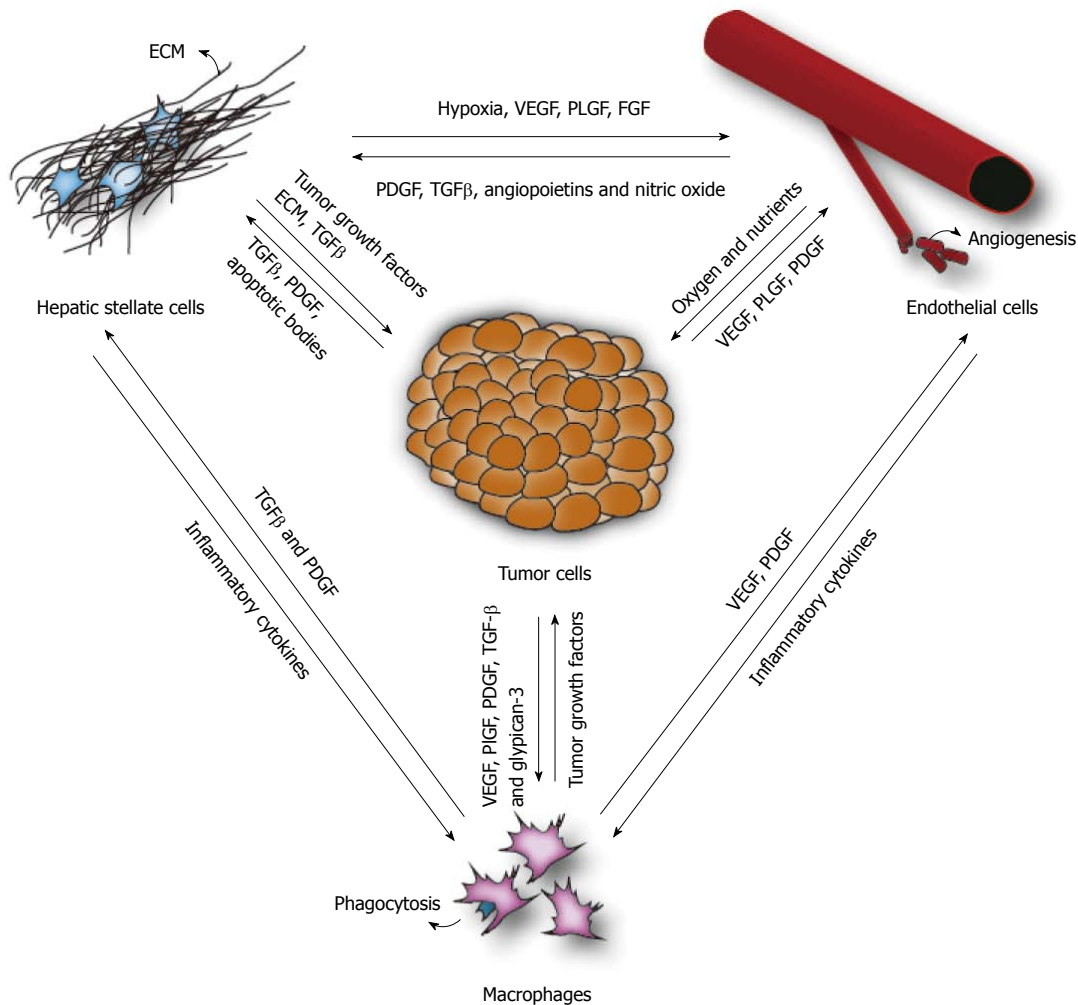


Figure 1 The interaction between tumor stroma and tumor cells in hepatocellular carcinoma. The several actors of the stromal compartment-including hepatic stellate cells, macrophages and endothelial cells-actively contribute to tumorigenesis, while the tumor itself influences these cells to create a background that is beneficial for tumor growth. Tumor cells activate the hepatic stellate cells, leading to the deposition of extracellular matrix (ECM), an increased production of angiogenic factors and the recruitment of macrophages. The increase of angiogenic factors (secreted by macrophages, tumor cells and activated stellate cells) will induce the formation of new blood vessels, thereby supplying the tumor with more oxygen and nutrients, thus supporting tumor growth. The increase of inflammatory cytokines, leads to the recruitment of macrophages, which can exert a pro-tumoral effect by secreting growth factor and influences the activation of stellate cells. This complex interplay between the several cell types involved in liver cancer emphasizes the need for a multi-targeted approach in hepatocellular carcinoma (HCC) patients. VEGF: Vascular endothelial growth factor; PDGF: Platelet-derived growth factor; FGF: Fibroblast growth factor.

rats^[24]. However, no clinical trials on the effect on liver fibrosis have been done.

Another driver of HSC activation are members of the platelet derived growth factor (PDGF) family. PDGFs are potent mitogens for mesenchymal cells and work synergistically with TGF to activate stellate cells. Specific hepatic over-expression of PDGF-C leads to an increase in fibrosis and enhances hepatocarcinogenesis^[12,15]. PDGF-B is also involved in different stages of liver cancer development and is an essential regulator in the development of liver fibrosis^[25]. Hepatic overexpression of PDGF-B accelerates liver cancer, possibly by up regulating TGF- β receptor and by increasing expression of β -catenin as well as VEGF, CD31 and FGF. Several protein tyrosine kinase inhibitors-such as sorafenib, orantinib, sunitinib and SU6668-target PDGFR amongst other targets including VEGFR and FGFR. The protein tyrosine kinase

inhibitor imatinib reduces stromal cell proliferation in this mouse model, which successfully inhibits tumor progression^[13]. Imatinib is currently used to treat gastrointestinal stromal tumors^[26] and could possibly benefit HCC patients.

The deposition of ECM proteins is one of the most characteristic hallmarks of the activated stellate cell. Several of the ECM components such as proteoglycans, laminins, collagens, and fibronectin interact directly and indirectly with HCC cells and the different stroma cell types. This not only changes the tumor phenotype, but also prepares a microenvironment that facilitates tumor growth. Since the ECM acts as a reservoir for growth factors and cytokines, it can rapidly release them to support the tumor's needs.

Heparan sulfate (HS) proteoglycans (PG) are expressed in the ECM and composed of a protein core to which HS is covalently attached as side

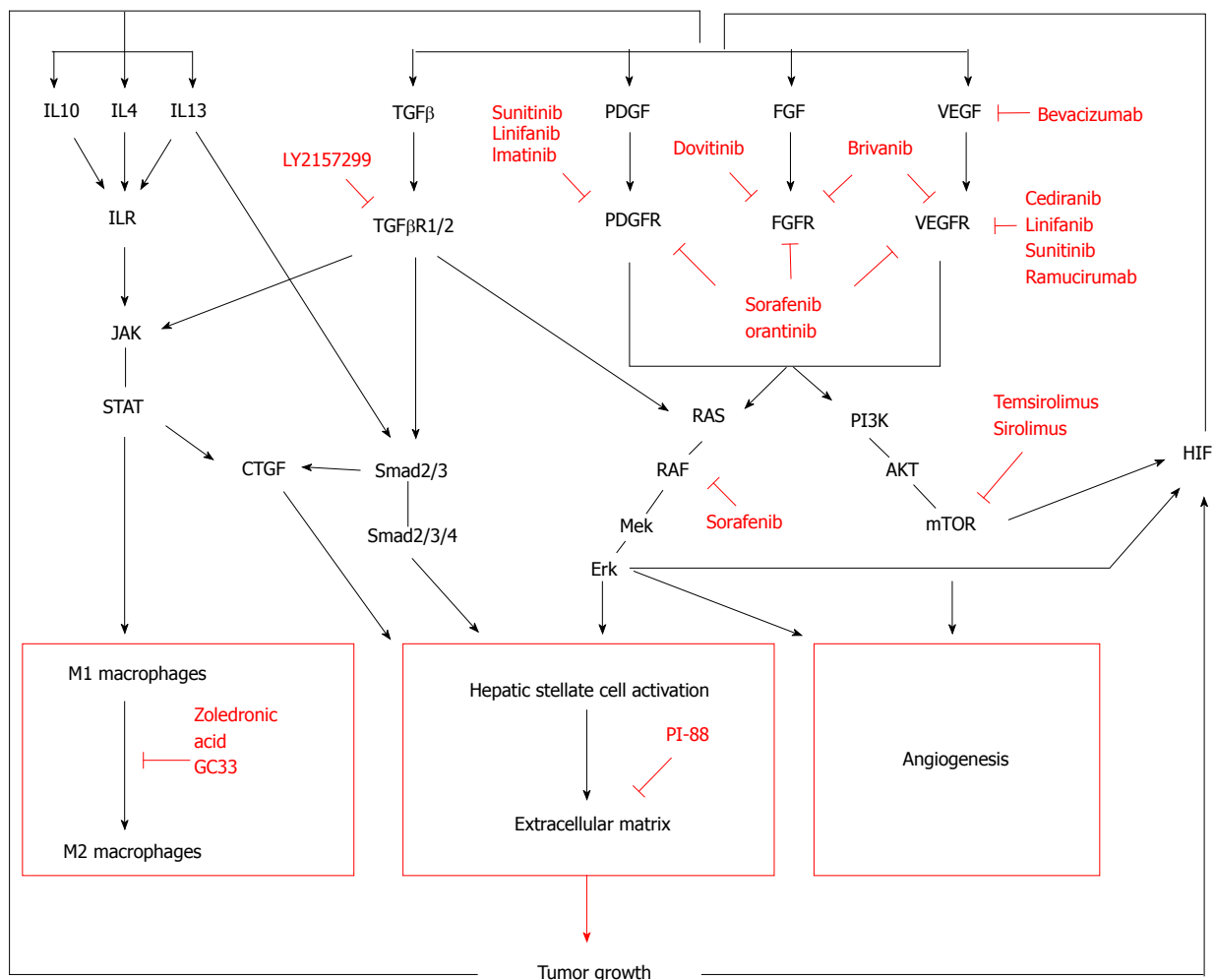


Figure 2 Schematic overview of (simplified) signaling pathways involved in the tumor-stroma interaction, and therapeutic targets that are currently tested in hepatocellular carcinoma. IL: Interleukin; TGF: Transforming growth factor; mTOR: Mammalian target of rapamycin.

chains. They maintain the structural framework of the tissue, store growth factors within the ECM or function as co-receptors. Desulfation of these co-receptor-PG's can abrogate growth factor signaling and inhibit tumor growth^[27,28]. Heparanase cleaves the HS side chains of HSPG, leading to the release of HS-bound proteins, such as growth factors. PI-88 is a heparin sulfate mimic that specifically targets heparanase in cancer, thus preventing the release of growth factors that otherwise would contribute to tumor growth, angiogenesis and metastasis^[29]. The safety and efficiency of PI-88 as an adjuvant therapy for post-operative HCC has been shown in a phase II trial^[30] and a recent follow up study revealed significant clinical benefits for patients with HCC^[31]. Phase III trials are currently ongoing (NCT01402908).

Another important glycoprotein is laminin-5. Laminin-5 is a member of the laminin family, which has been widely reported to be involved in the malignant phenotype of several cancers, including HCC^[32]. Laminin-5 is expressed higher in metastatic HCC and has been shown to stimulate HCC cell migration^[9,32].

Collagen is the major insoluble fibrous protein in the extracellular matrix. Besides its function as a supportive scaffold, collagens can also provoke a cellular response through the integrin family of transmembrane receptors. Several collagen types have been implicated in tumor growth and angiogenesis in different tumors^[33-35] and a recent study has shown that collagen matrix protects malignant hepatocytes from apoptosis^[36]. Antibodies targeting cleaved collagen epitopes have been clinically tested and show promising results in patients with solid tumors^[37,38].

This deposition of ECM leads to an increase in liver stiffness, an important hallmark of the cirrhotic liver, which is also used as a diagnostic tool for patients with CLD^[39]. This change in the mechanical properties of the tumor's surrounding has been associated with a higher risk of developing HCC^[40]. In addition, the increase of ECM and the capillarization of hepatic sinusoids cause a vascular resistance that leads to hypoxia, stimulating the production of pro-angiogenic factors and subsequently inducing angiogenesis^[1,41-43]. The activated HSCs also produce

Table 1 Overview of clinical trials that focus on the tumor environment of hepatocellular carcinoma

	Drug	Targets	Trial	Phase	Status	Ref.	
Tyrosine kinase inhibitors	Sorafenib	PDGFR VEGFR	NCT00105443	III	Completed ¹	[90]	
	Orantinib	RAF/MEK/ERK PDGFR, FGFR VEGFR	NCT02178358	I / II	Completed	[91]	
	Sunitinib	VEGFR	NCT00699374	III	Terminated	[71]	
		PDGFR	NCT00514228	II	Completed	[70]	
		RET	NCT00361309	II	Completed	[69]	
	Linifanib	CSF	NCT00428220	N/A	Ongoing		
		VEGF, PDGF, PDGFR- β , KDR	NCT01009593	III	Terminated		
	Brivanib	CSF	NCT00517920	II	Completed	[68]	
		VEGFR	NCT00858871	III	Completed	[59]	
			FGFR	NCT00908752	III	Ongoing	
				NCT00825955	III	Ongoing	
				NCT01108705	III	Terminated	
				NCT00355238	II	Completed	[92]
				NCT00437424	I	Completed	[93]
		Cediranib	VEGFR	NCT00238394	II		
				NCT00427973	II	Terminated	[54]
		Dovitinib	VEGFR, PDGFR, FGFR	NCT01232296	II	Ongoing	
		Antibodies	Bevacizumab	VEGF	NCT00335829	II	Completed
				NCT00162669	II	Completed	[50]
			NCT00605722	II	Completed	[51]	
			NCT00049322	II	Completed	[52]	
			NCT00280007	II	Terminated		
			NCT01180959	II	Ongoing		
Ramucirumab	VEGFR		NCT00627042	II	Completed	[53]	
			NCT01140347	III	Ongoing		
GC33	Glypican-3		NCT01507168	II	Completed		
			NCT00746317	I	Completed	[84]	
Other kinase inhibitors	Temozolomide	mTOR	NCT00976170	I	Ongoing		
			NCT01008917	I	Ongoing		
	Everolimus	mTOR	NCT01687673	II	Recruiting		
			NCT01035229	III	Completed	[95]	
			NCT01488487	II	Ongoing		
			NCT00516165	I / II	Completed	[66]	
	LY2157299	TGF- β 1		NCT00828594	I	Terminated	
				NCT01246986	II	Recruiting	
	PI-88	Heparanase		NCT02178358	II	Recruiting	
				NCT00568308	III	Terminated	
Zoledronic acid	Macrophages		NCT01402908	III	Ongoing		
			NCT00247728	II	Completed	[30]	
		NCT01259193	II	Ongoing			

¹Sorafenib is currently used as the standard-of-care for advanced hepatocellular carcinoma. CSF: Colony stimulating factor-1-receptor; mTOR: Mammalian target of rapamycin.

angiogenic growth factors, thus enhancing neo-angiogenesis^[8]. This increased vasculature will allow small HCC lesions to progress and eventually metastasize.

ENDOTHELIAL CELLS

The prolonged fibrogenic process leads to an abnormal angioarchitecture distinctive for cirrhosis. Anatomical changes in the cirrhotic liver, such as fibrotic scar tissue compressing portal and central venules are responsible for an increased intrahepatic vascular resistance. In addition, the formation of fibrotic septa, as well as sinusoidal capillarisation, results in an increased resistance to blood flow

and oxygen delivery. This causes hypoxia and the transcription of hypoxia-sensitive pro-angiogenic genes, thus stimulating the formation of new vessels. These new vessels can contribute to the inflammatory response by expressing chemokines and adhesion molecules, thus promoting the recruitment of inflammatory cells, such as macrophages. In addition, hepatic stellate cells are recruited to the angiogenic areas (*via* a number of signaling pathways, including PDGF, TGF- β , angiopoietins and nitric oxide) to contribute in vascular remodeling and stabilization^[44]. Therefore, angiogenesis may contribute to the progression of liver cirrhosis and stimulate the growth of small dysplastic lesions to advanced solid tumors.

HCC is solid tumor that rapidly outgrows its blood

supply and therefore stimulates the formation of new blood vessels to fulfill its high needs in oxygen and nutrients. The malignant hepatocytes, as well as other actors in the microenvironment such as activated stellate cells and macrophages, secrete a number of angiogenic growth factors^[1]. This induces an “angiogenic switch”, which activates endothelial cells and basement membranes to remodel existing vessels, and form new vessels. These new vessels allow the tumor to rapidly expand and offer a passage in the circulatory system, thus facilitating metastasis. Therefore, targeting angiogenesis has become a common cancer therapy to treat solid tumors.

The vascular endothelial growth factor A (VEGF) is one of the key factors regulating angiogenesis. It is secreted by tumor cells, macrophages and stellate cells. VEGF binds to its receptors (VEGFR1 and VEGFR2) on the present endothelial cells, stimulating endothelial cell proliferation and migration into the tumor, which results in vascular sprouting. Elevated VEGF levels are associated with tumor vascularity, metastasis, chemoresistance and poor prognosis^[45-47].

Significant progress on the treatment of advanced HCC has been made possible by sorafenib. Sorafenib is a small molecular inhibitor targeting several tyrosine protein kinases in the Raf/MEK/ERK-pathway (anti-proliferative effect); and PDGF, VEGFR1 and VEGFR2 (anti-angiogenic effect). Sorafenib has become the standard-of-care for patients with advanced HCC and for those progressing after loco-regional therapies^[48]. The success of sorafenib has opened the door for several anti-angiogenic agents to enter clinical studies on HCC^[49]. At the moment, several multikinase inhibitors are being tested in clinical trials, including sunitinib, brivanib, linafinib, cediranib, pazopanib, lenvatinib and axitinib, as well as blocking-antibodies targeting angiogenic pathways.

Bevacizumab, a humanized monoclonal antibody that targets VEGF, has been approved for the treatment of various solid tumors and is currently being investigated as a treatment for HCC. Several phase II trials have been completed and show that bevacizumab is well tolerated in HCC-patients, and could be a promising therapy as a single-agent^[50], in combination with erlotinib^[51] or after loco-regional therapies^[52]. Ramucirumab is a monoclonal antibody targeting VEGFR2 which has been tested as a first line treatment (NCT00627042) for HCC-patients with promising results^[53] and is currently being investigated as second line treatment after sorafenib (NCT01140347)^[53]. Cediranib is a tyrosine kinase inhibitor that targets all VEGF receptors, which has been tested in two clinical trials (NCT00427973 and NCT00238394). Despite some anti-tumor effects, the high toxicity of cediranib makes it an unsuitable drug for HCC-patients HCC^[54,55].

However, targeting VEGF has been shown to

induce therapy escape mechanisms and many patients treated with VEGF-inhibitors or with sorafenib obtain a secondary resistance to therapy. Alternative angiogenic factors, such as the placental growth factor (PlGF), PDGF and fibroblast growth factor (FGF) have been implicated in this acquired tumor resistance and combination therapies could open the door for sustained treatment response^[56]. Additionally, combining sorafenib with conventional chemotherapy could improve outcome and is currently tested in several phase III trials (NCT01015833, NCT01214343)^[57].

Brivanib and dovitinib are tyrosine kinase inhibitors of VEGF and fibroblast growth factor (FGF) signaling pathways, hence anticipating FGF-mediated resistance to anti-VEGF therapy^[58]. Brivanib has been or is being investigated in several phase III trials, including first-line treatment with brivanib vs sorafenib (NCT00858871)^[59], second-line treatment with brivanib after progression on sorafenib treatment (NCT01108705), second-line treatment with brivanib after sorafenib (NCT00825955) and trans-arterial chemoembolization in combination with brivanib (NCT00908752). However, results from the study testing brivanib and sorafenib as first-line therapy in patients with HCC indicate there are no benefits of using brivanib over sorafenib^[59] and study NCT01108705-testing brivanib after sorafenib treatment-has been terminated before completing the trial. Dovitinib trials are still ongoing (NCT01232296).

A drawback of anti-angiogenic therapies is that they aim to deprive the tumor from oxygen, leading to a hypoxic environment that stimulates cancer cells towards a more aggressive phenotype^[60]. Therefore, long-term administration of anti-angiogenic treatment could trigger escape mechanisms and lead to increased metastasis^[61,62].

An interesting way to indirectly target VEGF signaling and the HIF-pathway, is through inhibitors of the mammalian target of rapamycin (mTOR) pathway. mTOR signaling increases VEGF expression by up-regulating hypoxia inducible factor 1 α ^[63]. Furthermore, mTOR-inhibitors can directly influence tumor growth by inhibiting the expression of anti-apoptotic proteins and by inducing autophagy^[64]. Everolimus binds the cyclophilin FKBP-12, which binds the serine-threonine (ST) kinase mTOR when it is associated with raptor and mLST8 to form a complex (mTORC1), and subsequently inhibits downstream signaling, which involves cell cycle regulators and transcription factors such as HIF. mTORC1 lies downstream of phosphatidylinositol 3' kinase (PI3K), which is frequently activated in human cancers. Everolimus has been used in several clinical trials^[65,66], but data from the latest phase III trial (NCT01035229) show no improvement in overall survival^[67]. Temsirolimus is a sirolimus ester, which binds the same receptors. Trials using

temsirolimus are currently ongoing (NCT01008917 and NCT01687673).

The activated stellate cells also play a pivotal role in vascular remodeling, by creating a hypoxic environment, by producing angiogenic factors and also by migrating to angiogenic sites to contribute in the stabilization and maturation of (tumor) blood vessels. Current anti-angiogenic strategies for cancer have mostly focused on endothelial cells. However, combining drugs that target endothelial cells and stellate cells (or pericytes) could work synergistically as a therapy.

Several receptor tyrosine kinase inhibitors target VEGF and PDGF. Linifanib is a potent inhibitor of VEGF, PDGF, PDGFR- β , KDR and colony stimulating factor-1-receptor (CSF). A phase II trial (NCT00517920) showed initial benefits for linifanib in HCC patients^[68], however, the subsequent phase III trial (NCT01009593) had to be terminated for unknown reasons. Sunitinib inhibits receptors for PDGF and VEGF, as well as other receptor tyrosine kinases such as CSF. While several phase II trials (NCT00514228, NCT00361309) have shown promising results^[69,70], it is inferior to sorafenib and the latest phase III trial had to be terminated for safety reasons^[71].

Orantinib is a receptor tyrosine kinase inhibitor that binds and inhibits the autophosphorylation of VEGFR2, PDGF-receptor and fibroblast growth factor receptor (FGFR), thereby inhibiting angiogenesis and cell proliferation. A phase I / II trial has shown a trend towards prolonged progression free survival in patients treated with orantinib after transarterial chemoembolization^[72], and a phase III trial is still ongoing (NCT01465464). Blocking PDGF signaling in mouse models of pancreatic carcinogenesis with orantinib caused regression of blood vessels, as a result of the detachment of pericytes from tumor vessels. The fact that tumor vessels lacking pericytes are more vulnerable suggests that they could be more responsive to other anti-angiogenic drugs^[73,74]. Combining receptor tyrosine kinase inhibitors targeting ECs and pericytes successfully diminished tumor angiogenesis and decreased tumor size compared to a monotherapeutic approach in colon cancer^[75]. Similar effects were seen when PDGF inhibitors were combined with anti-angiogenic treatments^[74]. Thus, targeting stellate cells and endothelial cells may destabilize the existing tumor vasculature more potently than targeting each cell type individually.

MACROPHAGES

After liver damage, the pool of the liver's resident macrophages-Kupffer cells-is rapidly expanded. A harmful incident causes the hepatic macrophages to secrete pro-inflammatory cytokines and chemokines such as IL-1 β , TNF, CCL2, and CCL5, resulting in the activation of protective or apoptotic signaling pathways of hepatocytes and the recruitment of

immune cells that support hepatic injury. There is increasing evidence suggesting that phagocytosis of apoptotic bodies by HSC and by macrophages may directly stimulate fibrogenesis through upregulation of TGF- β ^[76]. Furthermore, these repeated cycles of hepatocyte death and compensatory proliferation provide a mitogenic and mutagenic environment that fuels the development of HCC.

The location of Kupffer cells in the sinusoids allows close interactions with other non-parenchymal liver cells. Firstly, Kupffer cells interact with other immune cells by secreting inflammatory cytokines and chemokines. Secondly, they can activate HSC *via* paracrine mechanisms, likely involving TGF- β and PDGF. These profibrotic functions of Kupffer cells during chronic liver injury possibly contribute to a tumor-stimulating environment in the cirrhotic liver. *In vivo* studies have shown that depleting macrophages reduces angiogenesis and slows down tumor progression in mouse models, and enhances the response to sorafenib^[77].

Macrophages can be classified into two main classes depending on their phenotypic polarization: the M1-phenotype, triggering a Th1 immune response and exerting cytotoxic activity; and the M2-phenotype, which activates a Th2 immune response and promotes angiogenesis, tissue remodeling and tumor progression^[78]. Macrophages can adapt to signals from the microenvironment and change their functional phenotype accordingly^[79]. M1 macrophages are activated as a response to microbial stimuli and interferon gamma, while in a tumor environment the tumour-associated macrophages (TAMs) are mainly polarized towards a M2 phenotype. Increased numbers of M2-macrophages have been associated with angiogenesis, metastasis and poor prognosis.

Tumor associated macrophages are key actors of cancer-related inflammation, being the main type of inflammatory cells infiltrating the tumor environment^[80]. In HCC, tumor cells have been shown to recruit and activate TAMs by the secretion of VEGF, PlGF, PDGF, TGF- β and glypican-3. Glypican-3 is a member of the glypican family of heparin-sulfate proteoglycans linked to the cell surface. It is highly expressed in the majority of HCC cells and is known for its role in the regulation of cell proliferation and apoptosis^[81]. In addition, studies have suggested an involvement in the recruitment of M2-polarized TAM's in human HCC tissues^[82]. Possibly glypican-3 present on the cell surface of malignant cells, binds to CCL5 and CCL3, which are chemokines that attract TAMs. Glypican-3 antibodies could therefore block the recruitment of TAMs *via* CCL5 and CCL3. Antibodies targeting glypican-3 have been tested in several phase I trials for advanced HCC, with promising results. The antibody was well tolerated and preliminary antitumor activity show a threefold prolongation of the median time to progression in patients receiving glypican-3-antibodies compared to

untreated patients^[83,84].

Zoledronic acid (ZA) is a compound widely used to prevent skeletal complications associated with bone metastases. Recent studies have shown a possible direct role as an anti-tumor agent by targeting the TAMs. ZA is taken up by macrophages *via* phagocytosis and leads to apoptosis specifically in TAMs, thus causing a repolarization of the macrophage population^[85,86]. *In vivo* studies of ZA in combination with sorafenib have shown that the latter leads to an increase of M2-macrophages infiltrating the tumor stroma, which can be effectively depleted with ZA. This significantly inhibits angiogenesis, metastasis and tumor progression compared to sorafenib alone^[77]. A phase II study of sorafenib and ZA in advanced HCC has been conducted (NCT01259193), but no results have been published.

DISCUSSION

Several studies have shown that the stroma regulates the malignant transformation, survival, progression and metastasis of hepatocellular carcinoma. Factors derived from the tumor cells in their turn alter the tumor stroma to generate a tumor-permissive microenvironment. This complex interplay between the tumor and the different actors in the stroma establishes a promising axis for therapeutic targets (Figure 2).

VEGF targeting therapies have represented the first success in treating HCC patients in many years, reviving research in this field and leading to an explosion of clinical trials with anti-angiogenic therapies^[49]. However, the success of treatments such as sorafenib needs to be followed by better understanding of the mechanisms that underlie the intrinsic and acquired resistance to anti-angiogenic therapies. Perhaps targeting several actors of the stromal environment and the tumor cells at the same time could be the key for optimal treatment in future therapies.

Sorafenib does not only inhibit angiogenesis, but also alters the inflammatory environment. Sorafenib has been shown to suppress natural killer cells and facilitate tumor growth and metastasis^[87]. Furthermore, multi-tyrosine kinase inhibitors have been shown to increase infiltration of tumor-associated macrophages in the tumor environment which could contribute to the resistance or escape to anti-angiogenic treatment^[77] (although it is important to note that some studies have shown the opposite effect^[88]). Hence the solution could be the use of adjuvant immunotherapy along with tyrosine kinase inhibitors for patients with unresectable HCC in order to obtain long-term response. In fact, one of the first trials to confirm the efficacy of sorafenib in advanced, metastasized renal cell carcinoma, was performed in combination with immunotherapy with IL-2 and interferon-alpha^[89].

Tumor associated macrophages are important actors of cancer-related inflammation, being the main type of inflammatory cells infiltrating the tumor environment. Targeting macrophages as a therapeutic strategy could be done by depleting the overall population of macrophages, or by altering their phenotype from a M2 towards an M1 orientation. Again, macrophages are known to not only interact with the tumor cells and stimulate their growth, they also influence stellate cell activation and angiogenesis.

The stellate cells are one of the key players in the formation of the perfect tumor environment. Not only do they directly affect tumor growth by secreting growth factors^[7], matrix proteases^[8] and/or ECM proteins^[9], they also alter the mechanical properties of the tumor's surrounding. Activated stellate cells are known to stimulate angiogenesis, which allows the tumor cells to grow rapidly and invade in the circulatory system. The deposition of ECM proteins, such as collagens and proteoglycans, serve as a reservoir for growth factors, but also directly provoke a pro-tumoral cellular response. Indeed, the thick layer of ECM in the cirrhotic liver could impair drug delivery and hence decrease response to therapy. Therefore, preventing or reversing the activation of stellate cells could inhibit HCC growth, decrease angiogenesis and increase response to other therapies, such as classic chemotherapy or sorafenib.

As our understanding of the complex interplay between tumor and stroma evolves, the next-generation cancer drugs could target several actors in the tumor-stroma axis and offer a durable treatment for advanced HCC.

REFERENCES

- 1 **Coulon S**, Heindryckx F, Geerts A, Van Steenkiste C, Colle I, Van Vlierberghe H. Angiogenesis in chronic liver disease and its complications. *Liver Int* 2011; **31**: 146-162 [PMID: 21073649 DOI: 10.1111/j.1478-3231.2010.02369.x]
- 2 **Welti J**, Loges S, Dimmeler S, Carmeliet P. Recent molecular discoveries in angiogenesis and antiangiogenic therapies in cancer. *J Clin Invest* 2013; **123**: 3190-3200 [PMID: 23908119 DOI: 10.1172/JCI70212]
- 3 **Puche JE**, Saiman Y, Friedman SL. Hepatic stellate cells and liver fibrosis. *Compr Physiol* 2013; **3**: 1473-1492 [PMID: 24265236 DOI: 10.1002/cphy.c120035]
- 4 **Otranto M**, Sarrazy V, Bonté F, Hinz B, Gabbiani G, Desmoulière A. The role of the myofibroblast in tumor stroma remodeling. *Cell Adh Migr* 2012; **6**: 203-219 [PMID: 22568985 DOI: 10.4161/cam.20377]
- 5 **Geng ZM**, Li QH, Li WZ, Zheng JB, Shah V. Activated human hepatic stellate cells promote growth of human hepatocellular carcinoma in a subcutaneous xenograft nude mouse model. *Cell Biochem Biophys* 2014; **70**: 337-347 [PMID: 24676678 DOI: 10.1007/s12013-014-9918-3]
- 6 **Carloni V**, Luong TV, Rombouts K. Hepatic stellate cells and extracellular matrix in hepatocellular carcinoma: more complicated than ever. *Liver Int* 2014; **34**: 834-843 [PMID: 24397349 DOI: 10.1111/liv.12465]
- 7 **Amann T**, Bataille F, Spruss T, Mühlbauer M, Gäbele E, Schölmerich J, Kiefer P, Bosserhoff AK, Hellerbrand C. Activated hepatic stellate cells promote tumorigenicity of hepatocellular carcinoma. *Cancer Sci* 2009; **100**: 646-653 [PMID: 19175606 DOI:

- 10.1111/j.1349-7006.2009.01087.x]
- 8 **Coulouarn C**, Corlu A, Glaise D, Guénon I, Thorgeirsson SS, Clément B. Hepatocyte-stellate cell cross-talk in the liver engenders a permissive inflammatory microenvironment that drives progression in hepatocellular carcinoma. *Cancer Res* 2012; **72**: 2533-2542 [PMID: 22419664 DOI: 10.1158/0008-5472.CAN-11-3317]
 - 9 **Santamato A**, Fransvea E, Dituri F, Caligiuri A, Quaranta M, Niimi T, Pinzani M, Antonaci S, Giannelli G. Hepatic stellate cells stimulate HCC cell migration via laminin-5 production. *Clin Sci (Lond)* 2011; **121**: 159-168 [PMID: 21413933 DOI: 10.1042/CS20110002]
 - 10 **Zhao W**, Zhang L, Xu Y, Zhang Z, Ren G, Tang K, Kuang P, Zhao B, Yin Z, Wang X. Hepatic stellate cells promote tumor progression by enhancement of immunosuppressive cells in an orthotopic liver tumor mouse model. *Lab Invest* 2014; **94**: 182-191 [PMID: 24296878 DOI: 10.1038/labinvest.2013.139]
 - 11 **Zhao W**, Zhang L, Yin Z, Su W, Ren G, Zhou C, You J, Fan J, Wang X. Activated hepatic stellate cells promote hepatocellular carcinoma development in immunocompetent mice. *Int J Cancer* 2011; **129**: 2651-2661 [PMID: 21213212 DOI: 10.1002/ijc.25920]
 - 12 **Campbell JS**, Hughes SD, Gilbertson DG, Palmer TE, Holdren MS, Haran AC, Odell MM, Bauer RL, Ren HP, Haugen HS, Yeh MM, Fausto N. Platelet-derived growth factor C induces liver fibrosis, steatosis, and hepatocellular carcinoma. *Proc Natl Acad Sci USA* 2005; **102**: 3389-3394 [PMID: 15728360 DOI: 10.1073/pnas.0409722102]
 - 13 **Campbell JS**, Johnson MM, Bauer RL, Hudkins KL, Gilbertson DG, Riehle KJ, Yeh MM, Alpers CE, Fausto N. Targeting stromal cells for the treatment of platelet-derived growth factor C-induced hepatocellular carcinogenesis. *Differentiation* 2007; **75**: 843-852 [PMID: 17999742 DOI: 10.1111/j.1432-0436.2007.00235.x]
 - 14 **Nitta T**, Kim JS, Mohuczy D, Behrns KE. Murine cirrhosis induces hepatocyte epithelial mesenchymal transition and alterations in survival signaling pathways. *Hepatology* 2008; **48**: 909-919 [PMID: 18712785 DOI: 10.1002/hep.22397]
 - 15 **Wright JH**, Johnson MM, Shimizu-Albergine M, Bauer RL, Hayes BJ, Surapisitchat J, Hudkins KL, Riehle KJ, Johnson SC, Yeh MM, Bammler TK, Beyer RP, Gilbertson DG, Alpers CE, Fausto N, Campbell JS. Paracrine activation of hepatic stellate cells in platelet-derived growth factor C transgenic mice: evidence for stromal induction of hepatocellular carcinoma. *Int J Cancer* 2014; **134**: 778-788 [PMID: 23929039 DOI: 10.1002/ijc.28421]
 - 16 **Giannelli G**, Villa E, Lahn M. Transforming growth factor- β as a therapeutic target in hepatocellular carcinoma. *Cancer Res* 2014; **74**: 1890-1894 [PMID: 24638984 DOI: 10.1158/0008-5472.CAN-14-0243]
 - 17 **Dooley S**, Weng H, Mertens PR. Hypotheses on the role of transforming growth factor-beta in the onset and progression of hepatocellular carcinoma. *Dig Dis* 2009; **27**: 93-101 [PMID: 19546546 DOI: 10.1159/000218340]
 - 18 **Dooley S**, Hamzavi J, Ciuculan L, Godoy P, Ilkavets I, Ehrent S, Ueberham E, Gebhardt R, Kanzler S, Geier A, Breitkopf K, Weng H, Mertens PR. Hepatocyte-specific Smad7 expression attenuates TGF-beta-mediated fibrogenesis and protects against liver damage. *Gastroenterology* 2008; **135**: 642-659 [PMID: 18602923 DOI: 10.1053/j.gastro.2008.04.038]
 - 19 **Bogaerts E**, Heindryckx F, Vandewynckel YP, Van Grunsven LA, Van Vlierberghe H. The roles of transforming growth factor- β , Wnt, Notch and hypoxia on liver progenitor cells in primary liver tumours (Review). *Int J Oncol* 2014; **44**: 1015-1022 [PMID: 24504124 DOI: 10.3892/ijo.2014.2286]
 - 20 **Ikushima H**, Miyazono K. TGFbeta signalling: a complex web in cancer progression. *Nat Rev Cancer* 2010; **10**: 415-424 [PMID: 20495575 DOI: 10.1038/nrc2853]
 - 21 **Dituri F**, Mazzocca A, Peidro FJ, Papappicco P, Fabregat I, De Santis F, Paradiso A, Sabba C, Giannelli G. Differential Inhibition of the TGF- β Signaling Pathway in HCC Cells Using the Small Molecule Inhibitor LY2157299 and the D10 Monoclonal Antibody against TGF- β Receptor Type II. *PLoS One* 2013; **8**: e67109 [PMID: 23826206 DOI: 10.1371/journal.pone.0067109]
 - 22 **Gressner OA**, Fang M, Li H, Lu LG, Gressner AM, Gao CF. Connective tissue growth factor (CTGF/CCN2) in serum is an indicator of fibrogenic progression and malignant transformation in patients with chronic hepatitis B infection. *Clin Chim Acta* 2013; **421**: 126-131 [PMID: 23501329 DOI: 10.1016/j.cca.2013.02.029]
 - 23 **Liu Y**, Meyer C, Müller A, Herweck F, Li Q, Müllenbach R, Mertens PR, Dooley S, Weng HL. IL-13 induces connective tissue growth factor in rat hepatic stellate cells via TGF- β -independent Smad signaling. *J Immunol* 2011; **187**: 2814-2823 [PMID: 21804025 DOI: 10.4049/jimmunol.1003260]
 - 24 **Li G**, Xie Q, Shi Y, Li D, Zhang M, Jiang S, Zhou H, Lu H, Jin Y. Inhibition of connective tissue growth factor by siRNA prevents liver fibrosis in rats. *J Gene Med* 2006; **8**: 889-900 [PMID: 16652398 DOI: 10.1002/jgm.894]
 - 25 **Maass T**, Thieringer FR, Mann A, Longerich T, Schirmacher P, Strand D, Hansen T, Galle PR, Teufel A, Kanzler S. Liver specific overexpression of platelet-derived growth factor-B accelerates liver cancer development in chemically induced liver carcinogenesis. *Int J Cancer* 2011; **128**: 1259-1268 [PMID: 20506153 DOI: 10.1002/ijc.25469]
 - 26 **Patel S**. Long-term efficacy of imatinib for treatment of metastatic GIST. *Cancer Chemother Pharmacol* 2013; **72**: 277-286 [PMID: 23503753 DOI: 10.1007/s00280-013-2135-8]
 - 27 **Lai JP**, Sandhu DS, Shire AM, Roberts LR. The tumor suppressor function of human sulfatase 1 (SULF1) in carcinogenesis. *J Gastrointest Cancer* 2008; **39**: 149-158 [PMID: 19373441 DOI: 10.1007/s12029-009-9058-y]
 - 28 **Lai JP**, Thompson JR, Sandhu DS, Roberts LR. Heparin-degrading sulfatases in hepatocellular carcinoma: roles in pathogenesis and therapy targets. *Future Oncol* 2008; **4**: 803-814 [PMID: 19086847 DOI: 10.2217/14796694.4.6.803]
 - 29 **Parish CR**, Freeman C, Brown KJ, Francis DJ, Cowden WB. Identification of sulfated oligosaccharide-based inhibitors of tumor growth and metastasis using novel in vitro assays for angiogenesis and heparanase activity. *Cancer Res* 1999; **59**: 3433-3441 [PMID: 10416607]
 - 30 **Liu CJ**, Lee PH, Lin DY, Wu CC, Jeng LB, Lin PW, Mok KT, Lee WC, Yeh HZ, Ho MC, Yang SS, Lee CC, Yu MC, Hu RH, Peng CY, Lai KL, Chang SS, Chen PJ. Heparanase inhibitor PI-88 as adjuvant therapy for hepatocellular carcinoma after curative resection: a randomized phase II trial for safety and optimal dosage. *J Hepatol* 2009; **50**: 958-968 [PMID: 19303160 DOI: 10.1016/j.jhep.2008.12.023]
 - 31 **Liu CJ**, Chang J, Lee PH, Lin DY, Wu CC, Jeng LB, Lin YJ, Mok KT, Lee WC, Yeh HZ, Ho MC, Yang SS, Yang MD, Yu MC, Hu RH, Peng CY, Lai KL, Chang SS, Chen PJ. Adjuvant heparanase inhibitor PI-88 therapy for hepatocellular carcinoma recurrence. *World J Gastroenterol* 2014; **20**: 11384-11393 [PMID: 25170226 DOI: 10.3748/wjg.v20.i32.11384]
 - 32 **Giannelli G**, Fransvea E, Bergamini C, Marinosci F, Antonaci S. Laminin-5 chains are expressed differentially in metastatic and nonmetastatic hepatocellular carcinoma. *Clin Cancer Res* 2003; **9**: 3684-3691 [PMID: 14506159]
 - 33 **Duan W**, Ma J, Ma Q, Xu Q, Lei J, Han L, Li X, Wang Z, Wu Z, Lv S, Ma Z, Liu M, Wang F, Wu E. The Activation of β 1-integrin by Type I Collagen Coupling with the Hedgehog Pathway Promotes the Epithelial-Mesenchymal Transition in Pancreatic Cancer. *Curr Cancer Drug Targets* 2014 [PMID: 24720337]
 - 34 **Sok JC**, Lee JA, Dasari S, Joyce S, Contrucci SC, Egloff AM, Trevelline BK, Joshi R, Kumari N, Grandis JR, Thomas SM. Collagen type XI α 1 facilitates head and neck squamous cell cancer growth and invasion. *Br J Cancer* 2013; **109**: 3049-3056 [PMID: 24231953 DOI: 10.1038/bjc.2013.624]
 - 35 **Shields MA**, Dangi-Garimella S, Redig AJ, Munshi HG. Biochemical role of the collagen-rich tumour microenvironment in pancreatic cancer progression. *Biochem J* 2012; **441**: 541-552 [PMID: 22187935 DOI: 10.1042/BJ20111240]
 - 36 **Mailloux A**, Epling-Burnette P. Collagen matrix deposition by hepatic stellate cells protects hepatocellular carcinoma from NK-

- mediated cytotoxicity. *J Immunol* 2013; **190**. Available from: http://www.jimmunol.org/cgi/content/meeting_abstract/190/1/MeetingAbstracts/53.7
- 37 **Robert F**, Gordon MS, Rosen LS, Mendelson DS, Mulay M, Adams BJ, Alvarez D, Theuer CP, Leigh BR. Final results from a phase I study of TRC093 (humanized anti-cleaved collagen antibody) in patients with solid cancer. *J Clin Oncol* 2010; **28**: 15
- 38 **Gordon MS**, Rosen LS, Robert F, Mendelson DS, Kleinzweig D, Adams BJ, Theuer CP. A phase I, open-label, dose escalation study of the humanized monoclonal antibody (HuMAb) TRC093, an inhibitor of angiogenesis that binds to cleaved collagen, in patients with locally advanced or metastatic solid tumors. *Ejc Supplements* 2008; **6**: 130-130 [DOI: 10.1016/S1359-6349(08)72348-6]
- 39 **Wong GL**. Transient elastography: Kill two birds with one stone? *World J Hepatol* 2013; **5**: 264-274 [PMID: 23717737 DOI: 10.4254/wjh.v5.i5.264]
- 40 **Wang HM**, Hung CH, Lu SN, Chen CH, Lee CM, Hu TH, Wang JH. Liver stiffness measurement as an alternative to fibrotic stage in risk assessment of hepatocellular carcinoma incidence for chronic hepatitis C patients. *Liver Int* 2013; **33**: 756-761 [PMID: 23405889 DOI: 10.1111/liv.12118]
- 41 **Coulon S**, Legry V, Heindryckx F, Van Steenkiste C, Casteleyn C, Olivier K, Libbrecht L, Carmeliet P, Jonckx B, Stassen JM, Van Vlierberghe H, Leclercq I, Colle I, Geerts A. Role of vascular endothelial growth factor in the pathophysiology of nonalcoholic steatohepatitis in two rodent models. *Hepatology* 2013; **57**: 1793-1805 [PMID: 23299577 DOI: 10.1002/hep.26219]
- 42 **Heindryckx F**, Coulon S, Terrie E, Casteleyn C, Stassen JM, Geerts A, Libbrecht L, Allemeersch J, Carmeliet P, Colle I, Van Vlierberghe H. The placental growth factor as a target against hepatocellular carcinoma in a diethylnitrosamine-induced mouse model. *J Hepatol* 2013; **58**: 319-328 [PMID: 23046674 DOI: 10.1016/j.jhep.2012.09.032]
- 43 **Van Steenkiste C**, Ribera J, Geerts A, Pauta M, Tugues S, Casteleyn C, Libbrecht L, Olivier K, Schroyen B, Reynaert H, van Grunsven LA, Blomme B, Coulon S, Heindryckx F, De Vos M, Stassen JM, Vinckier S, Altamirano J, Bataller R, Carmeliet P, Van Vlierberghe H, Colle I, Morales-Ruiz M. Inhibition of placental growth factor activity reduces the severity of fibrosis, inflammation, and portal hypertension in cirrhotic mice. *Hepatology* 2011; **53**: 1629-1640 [PMID: 21520176 DOI: 10.1002/hep.24238]
- 44 **Hellström M**, Gerhardt H, Kalén M, Li X, Eriksson U, Wolburg H, Betsholtz C. Lack of pericytes leads to endothelial hyperplasia and abnormal vascular morphogenesis. *J Cell Biol* 2001; **153**: 543-553 [PMID: 11331305]
- 45 **Zhan P**, Qian Q, Yu LK. Serum VEGF level is associated with the outcome of patients with hepatocellular carcinoma: a meta-analysis. *Hepatobiliary Surg Nutr* 2013; **2**: 209-215 [PMID: 24570945 DOI: 10.3978/j.issn.2304-3881.2013.06.07]
- 46 **Tsuchiya K**, Asahina Y, Matsuda S, Muraoka M, Nakata T, Suzuki Y, Tamaki N, Yasui Y, Suzuki S, Hosokawa T, Nishimura T, Ueda K, Kuzuya T, Nakanishi H, Itakura J, Takahashi Y, Kurosaki M, Enomoto N, Izumi N. Changes in plasma vascular endothelial growth factor at 8 weeks after sorafenib administration as predictors of survival for advanced hepatocellular carcinoma. *Cancer* 2014; **120**: 229-237 [PMID: 24122122 DOI: 10.1002/cncr.28384]
- 47 **Huang GW**, Yang LY, Lu WQ. Expression of hypoxia-inducible factor 1alpha and vascular endothelial growth factor in hepatocellular carcinoma: Impact on neovascularization and survival. *World J Gastroenterol* 2005; **11**: 1705-1708 [PMID: 15786555]
- 48 **Germano D**, Daniele B. Systemic therapy of hepatocellular carcinoma: current status and future perspectives. *World J Gastroenterol* 2014; **20**: 3087-3099 [PMID: 24696596 DOI: 10.3748/wjg.v20.i12.3087]
- 49 **Pircher A**, Medinger M, Dreves J. Liver cancer: Targeted future options. *World J Hepatol* 2011; **3**: 38-44 [PMID: 21423913 DOI: 10.4254/wjh.v3.i2.38]
- 50 **Boige V**, Malka D, Bourredjem A, Dromain C, Baey C, Jacques N, Pignon JP, Vimond N, Bouvet-Forgeau N, De Baere T, Ducreux M, Farace F. Efficacy, safety, and biomarkers of single-agent bevacizumab therapy in patients with advanced hepatocellular carcinoma. *Oncologist* 2012; **17**: 1063-1072 [PMID: 22707516 DOI: 10.1634/theoncologist.2011-0465]
- 51 **Hsu CH**, Kang YK, Yang TS, Shun CT, Shao YY, Su WC, Sandoval-Tan J, Chiou TJ, Jin K, Hsu C, Cheng AL. Bevacizumab with erlotinib as first-line therapy in Asian patients with advanced hepatocellular carcinoma: a multicenter phase II study. *Oncology* 2013; **85**: 44-52 [PMID: 23838576 DOI: 10.1159/000350841]
- 52 **Britten CD**, Gomes AS, Wainberg ZA, Elashoff D, Amado R, Xin Y, Busuttill RW, Slamon DJ, Finn RS. Transarterial chemoembolization plus or minus intravenous bevacizumab in the treatment of hepatocellular cancer: a pilot study. *BMC Cancer* 2012; **12**: 16 [PMID: 22244160 DOI: 10.1186/1471-2407-12-16]
- 53 **Zhu AX**, Finn RS, Mulcahy M, Gurtler J, Sun W, Schwartz JD, Dalal RP, Joshi A, Hozak RR, Xu Y, Ancukiewicz M, Jain RK, Nugent FW, Duda DG, Stuart K. A phase II and biomarker study of ramucirumab, a human monoclonal antibody targeting the VEGF receptor-2, as first-line monotherapy in patients with advanced hepatocellular cancer. *Clin Cancer Res* 2013; **19**: 6614-6623 [PMID: 24088738 DOI: 10.1158/1078-0432.CCR-13-1442]
- 54 **Zhu AX**, Ancukiewicz M, Supko JG, Sahani DV, Blaszkowsky LS, Meyerhardt JA, Abrams TA, McCleary NJ, Bhargava P, Muzikansky A, Sheehan S, Regan E, Vasudev E, Knowles M, Fuchs CS, Ryan DP, Jain RK, Duda DG. Efficacy, safety, pharmacokinetics, and biomarkers of cediranib monotherapy in advanced hepatocellular carcinoma: a phase II study. *Clin Cancer Res* 2013; **19**: 1557-1566 [PMID: 23362324 DOI: 10.1158/1078-0432.CCR-12-3041]
- 55 **Alberts SR**, Fitch TR, Kim GP, Morlan BW, Dakhlil SR, Gross HM, Nair S. Cediranib (AZD2171) in patients with advanced hepatocellular carcinoma: a phase II North Central Cancer Treatment Group Clinical Trial. *Am J Clin Oncol* 2012; **35**: 329-333 [PMID: 21422991 DOI: 10.1097/COC.0b013e3182118cdf]
- 56 **Lieu CH**, Tran H, Jiang ZQ, Mao M, Overman MJ, Lin E, Eng C, Morris J, Ellis L, Heymach JV, Kopetz S. The association of alternate VEGF ligands with resistance to anti-VEGF therapy in metastatic colorectal cancer. *PLoS One* 2013; **8**: e77117 [PMID: 24143206 DOI: 10.1371/journal.pone.0077117]
- 57 **Miyahara K**, Nouse K, Yamamoto K. Chemotherapy for advanced hepatocellular carcinoma in the sorafenib age. *World J Gastroenterol* 2014; **20**: 4151-4159 [PMID: 24764653 DOI: 10.3748/wjg.v20.i15.4151]
- 58 **Tai WT**, Cheng AL, Shiau CW, Liu CY, Ko CH, Lin MW, Chen PJ, Chen KF. Dovitinib induces apoptosis and overcomes sorafenib resistance in hepatocellular carcinoma through SHP-1-mediated inhibition of STAT3. *Mol Cancer Ther* 2012; **11**: 452-463 [PMID: 22180308 DOI: 10.1158/1535-7163.Mct-11-0412]
- 59 **Johnson PJ**, Qin S, Park JW, Poon RT, Raoul JL, Philip PA, Hsu CH, Hu TH, Heo J, Xu J, Lu L, Chao Y, Boucher E, Han KH, Paik SW, Robles-Aviña J, Kudo M, Yan L, Sobhonlidsuk A, Komov D, Decaens T, Tak WY, Jeng LB, Liu D, Ezzeddine R, Walters I, Cheng AL. Brivanib versus sorafenib as first-line therapy in patients with unresectable, advanced hepatocellular carcinoma: results from the randomized phase III BRISK-FL study. *J Clin Oncol* 2013; **31**: 3517-3524 [PMID: 23980084 DOI: 10.1200/JCO.2012.48.4410]
- 60 **Comito G**, Calvani M, Giannoni E, Bianchini F, Calorini L, Torre E, Migliore C, Giordano S, Chiarugi P. HIF-1 α stabilization by mitochondrial ROS promotes Met-dependent invasive growth and vasculogenic mimicry in melanoma cells. *Free Radic Biol Med* 2011; **51**: 893-904 [PMID: 21703345 DOI: 10.1016/j.freeradbiomed.2011.05.042]
- 61 **Pàez-Ribes M**, Allen E, Hudock J, Takeda T, Okuyama H, Viñals F, Inoue M, Bergers G, Hanahan D, Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell* 2009; **15**: 220-231 [PMID: 19249680 DOI: 10.1016/j.ccr.2009.01.027]
- 62 **Loges S**, Mazzone M, Hohensinner P, Carmeliet P. Silencing or fueling metastasis with VEGF inhibitors: antiangiogenesis revisited. *Cancer Cell* 2009; **15**: 167-170 [PMID: 19249675 DOI: 10.1016/j.ccr.2009.02.007]

- 63 **Zhong H**, Chiles K, Feldser D, Laughner E, Hanrahan C, Georgescu MM, Simons JW, Semenza GL. Modulation of hypoxia-inducible factor 1 α expression by the epidermal growth factor/phosphatidylinositol 3-kinase/PTEN/AKT/FRAP pathway in human prostate cancer cells: implications for tumor angiogenesis and therapeutics. *Cancer Res* 2000; **60**: 1541-1545 [PMID: 10749120]
- 64 **Thomas HE**, Mercer CA, Carnevalli LS, Park J, Andersen JB, Conner EA, Tanaka K, Matsutani T, Iwanami A, Aronow BJ, Manway L, Maira SM, Thorgeirsson SS, Mischel PS, Thomas G, Kozma SC. mTOR inhibitors synergize on regression, reversal of gene expression, and autophagy in hepatocellular carcinoma. *Sci Transl Med* 2012; **4**: 139ra84 [PMID: 22539746 DOI: 10.1126/scitranslmed.3003923]
- 65 **Finn RS**, Poon RT, Yau T, Klumpen HJ, Chen LT, Kang YK, Kim TY, Gomez-Martin C, Rodriguez-Lopez C, Kunz T, Paquet T, Brandt U, Sellami D, Bruix J. Phase I study investigating everolimus combined with sorafenib in patients with advanced hepatocellular carcinoma. *J Hepatol* 2013; **59**: 1271-1277 [PMID: 23928403 DOI: 10.1016/j.jhep.2013.07.029]
- 66 **Zhu AX**, Abrams TA, Miskad R, Blaszkowsky LS, Meyerhardt JA, Zheng H, Muzikansky A, Clark JW, Kwak EL, Schrag D, Jors KR, Fuchs CS, Iafrate AJ, Borger DR, Ryan DP. Phase 1/2 study of everolimus in advanced hepatocellular carcinoma. *Cancer* 2011; **117**: 5094-5102 [PMID: 21538343 DOI: 10.1002/cncr.26165]
- 67 **Zhu AX**, Kudo M, Assenet E, Cattani S, Kang YK, Lim HY, Poon RTP, Blanc JF, Vogel A, Chen CL, Dorval E, Peck-Radosavljevic M, Santoro A, Daniele B, Furuse J, Jappe A, Perraud K, Anak O, Sellami DB, Chen LT. EVOLVE-1: Phase 3 study of everolimus for advanced HCC that progressed during or after sorafenib. *J Clin Oncol* 2014; **32**: 3
- 68 **Toh HC**, Chen PJ, Carr BI, Knox JJ, Gill S, Ansell P, McKeegan EM, Dowell B, Pedersen M, Qin Q, Qian J, Scappaticci FA, Ricker JL, Carlson DM, Yong WP. Phase 2 trial of lenvatinib (ABT-869) in patients with unresectable or metastatic hepatocellular carcinoma. *Cancer* 2013; **119**: 380-387 [PMID: 22833179 DOI: 10.1002/cncr.27758]
- 69 **Sahani DV**, Jiang T, Hayano K, Duda DG, Catalano OA, Ancukiewicz M, Jain RK, Zhu AX. Magnetic resonance imaging biomarkers in hepatocellular carcinoma: association with response and circulating biomarkers after sunitinib therapy. *J Hematol Oncol* 2013; **6**: 51 [PMID: 23842041 DOI: 10.1186/1756-8722-6-51]
- 70 **Koerberle D**, Montemurro M, Samaras P, Majno P, Simcock M, Limacher A, Lerch S, Kovács K, Inauen R, Hess V, Saletti P, Borner M, Roth A, Bodoky G. Continuous Sunitinib treatment in patients with advanced hepatocellular carcinoma: a Swiss Group for Clinical Cancer Research (SAKK) and Swiss Association for the Study of the Liver (SASL) multicenter phase II trial (SAKK 77/06). *Oncologist* 2010; **15**: 285-292 [PMID: 20203173 DOI: 10.1634/theoncologist.2009-0316]
- 71 **Cheng AL**, Kang YK, Lin DY, Park JW, Kudo M, Qin S, Chung HC, Song X, Xu J, Poggi G, Omata M, Pitman Lowenthal S, Lanzalone S, Yang L, Lechuga MJ, Raymond E. Sunitinib versus sorafenib in advanced hepatocellular cancer: results of a randomized phase III trial. *J Clin Oncol* 2013; **31**: 4067-4075 [PMID: 24081937 DOI: 10.1200/JCO.2012.45.8372]
- 72 **Inaba Y**, Kanai F, Aramaki T, Yamamoto T, Tanaka T, Yamakado K, Kaneko S, Kudo M, Imanaka K, Kora S, Nishida N, Kawai N, Seki H, Matsui O, Arioka H, Arai Y. A randomised phase II study of TSU-68 in patients with hepatocellular carcinoma treated by transarterial chemoembolisation. *Eur J Cancer* 2013; **49**: 2832-2840 [PMID: 23764238 DOI: 10.1016/j.ejca.2013.05.011]
- 73 **Bergers G**, Song S, Meyer-Morse N, Bergsland E, Hanahan D. Benefits of targeting both pericytes and endothelial cells in the tumor vasculature with kinase inhibitors. *J Clin Invest* 2003; **111**: 1287-1295 [PMID: 12727920 DOI: 10.1172/JCI117929]
- 74 **Pietras K**, Hanahan D. A multitargeted, metronomic, and maximum-tolerated dose "chemo-switch" regimen is antiangiogenic, producing objective responses and survival benefit in a mouse model of cancer. *J Clin Oncol* 2005; **23**: 939-952 [PMID: 15557593 DOI: 10.1200/JCO.2005.07.093]
- 75 **Shaheen RM**, Tseng WW, Davis DW, Liu W, Reinmuth N, Vellagas R, Wieczorek AA, Ogura Y, McConkey DJ, Drazan KE, Bucana CD, McMahon G, Ellis LM. Tyrosine kinase inhibition of multiple angiogenic growth factor receptors improves survival in mice bearing colon cancer liver metastases by inhibition of endothelial cell survival mechanisms. *Cancer Res* 2001; **61**: 1464-1468 [PMID: 11245452]
- 76 **Canbay A**, Taimr P, Torok N, Higuchi H, Friedman S, Gores GJ. Apoptotic body engulfment by a human stellate cell line is profibrogenic. *Lab Invest* 2003; **83**: 655-663 [PMID: 12746475]
- 77 **Zhang W**, Zhu XD, Sun HC, Xiong YQ, Zhuang PY, Xu HX, Kong LQ, Wang L, Wu WZ, Tang ZY. Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. *Clin Cancer Res* 2010; **16**: 3420-3430 [PMID: 20570927 DOI: 10.1158/1078-0432.CCR-09-2904]
- 78 **Roderfeld M**, Rath T, Lammert F, Dierkes C, Graf J, Roeb E. Innovative immunohistochemistry identifies MMP-9 expressing macrophages at the invasive front of murine HCC. *World J Hepatol* 2010; **2**: 175-179 [PMID: 21160992 DOI: 10.4254/wjh.v2.i5.175]
- 79 **Stout RD**, Jiang C, Matta B, Tietzel I, Watkins SK, Suttles J. Macrophages sequentially change their functional phenotype in response to changes in microenvironmental influences. *J Immunol* 2005; **175**: 342-349 [PMID: 15972667]
- 80 **Capecce D**, Fischietti M, Verzella D, Gaggiano A, Ciccirelli G, Tessitore A, Zazzeroni F, Alesse E. The inflammatory microenvironment in hepatocellular carcinoma: a pivotal role for tumor-associated macrophages. *Biomed Res Int* 2013; **2013**: 187204 [PMID: 23533994 DOI: 10.1155/2013/187204]
- 81 **Yao M**, Wang L, Dong Z, Qian Q, Shi Y, Yu D, Wang S, Zheng W, Yao D. Glypican-3 as an emerging molecular target for hepatocellular carcinoma gene therapy. *Tumour Biol* 2014; **35**: 5857-5868 [PMID: 24633918 DOI: 10.1007/s13277-014-1776-5]
- 82 **Takai H**, Ashihara M, Ishiguro T, Terashima H, Watanabe T, Kato A, Suzuki M. Involvement of glypican-3 in the recruitment of M2-polarized tumor-associated macrophages in hepatocellular carcinoma. *Cancer Biol Ther* 2009; **8**: 2329-2338 [PMID: 19838081]
- 83 **Ikedo M**, Ohkawa S, Okusaka T, Mitsunaga S, Kobayashi S, Morizane C, Suzuki I, Yamamoto S, Furuse J. Japanese phase I study of GC33, a humanized antibody against glypican-3 for advanced hepatocellular carcinoma. *Cancer Sci* 2014; **105**: 455-462 [PMID: 24521523 DOI: 10.1111/cas.12368]
- 84 **Zhu AX**, Gold PJ, El-Khoueiry AB, Abrams TA, Morikawa H, Ohishi N, Ohtomo T, Philip PA. First-in-man phase I study of GC33, a novel recombinant humanized antibody against glypican-3, in patients with advanced hepatocellular carcinoma. *Clin Cancer Res* 2013; **19**: 920-928 [PMID: 23362325 DOI: 10.1158/1078-0432.CCR-12-2616]
- 85 **Rogers TL**, Wind N, Hughes R, Nutter F, Brown HK, Vasiliadou I, Ottewill PD, Hohen I. Macrophages as potential targets for zoledronic acid outside the skeleton-evidence from in vitro and in vivo models. *Cell Oncol (Dordr)* 2013; **36**: 505-514 [PMID: 24177992 DOI: 10.1007/s13402-013-0156-2]
- 86 **Coscia M**, Quaglino E, Iezzi M, Curcio C, Pantaleoni F, Riganti C, Hohen I, Mönkkönen H, Boccadoro M, Forni G, Musiani P, Bosia A, Cavallo F, Massaia M. Zoledronic acid repolarizes tumour-associated macrophages and inhibits mammary carcinogenesis by targeting the mevalonate pathway. *J Cell Mol Med* 2010; **14**: 2803-2815 [PMID: 19818098 DOI: 10.1111/j.1582-4934.2009.00926.x]
- 87 **Zhang QB**, Sun HC, Zhang KZ, Jia QA, Bu Y, Wang M, Chai ZT, Zhang QB, Wang WQ, Kong LQ, Zhu XD, Lu L, Wu WZ, Wang L, Tang ZY. Suppression of natural killer cells by sorafenib contributes to prometastatic effects in hepatocellular carcinoma. *PLoS One* 2013; **8**: e55945 [PMID: 23409093 DOI: 10.1371/journal.pone.0055945]
- 88 **Sprinzl MF**, Reisinger F, Puschnik A, Ringelhan M, Ackermann K, Hartmann D, Schiemann M, Weinmann A, Galle PR, Schuchmann M, Friess H, Otto G, Heikenwalder M, Protzer U. Sorafenib perpetuates cellular anticancer effector functions by modulating the crosstalk between macrophages and natural killer cells. *Hepatology* 2013; **57**: 2358-2368 [PMID: 23424039 DOI: 10.1002/hep.26328]

- 89 **Escudier B**, Lassau N, Angevin E, Soria JC, Chami L, Lamuraglia M, Zafarana E, Landreau V, Schwartz B, Brendel E, Armand JP, Robert C. Phase I trial of sorafenib in combination with IFN alpha-2a in patients with unresectable and/or metastatic renal cell carcinoma or malignant melanoma. *Clin Cancer Res* 2007; **13**: 1801-1809 [PMID: 17363536 DOI: 10.1158/1078-0432.CCR-06-1432]
- 90 **Llovet JM**, Ricci S, Mazzaferro V, Hilgard P, Gane E, Blanc JF, de Oliveira AC, Santoro A, Raoul JL, Forner A, Schwartz M, Porta C, Zeuzem S, Bolondi L, Greten TF, Galle PR, Seitz JF, Borbath I, Häussinger D, Giannaris T, Shan M, Moscovici M, Voliotis D, Bruix J. Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 2008; **359**: 378-390 [PMID: 18650514 DOI: 10.1056/NEJMoa0708857]
- 91 **Kanai F**, Yoshida H, Tateishi R, Sato S, Kawabe T, Obi S, Kondo Y, Taniguchi M, Tagawa K, Ikeda M, Morizane C, Okusaka T, Arioka H, Shiina S, Omata M. A phase I/II trial of the oral antiangiogenic agent TSU-68 in patients with advanced hepatocellular carcinoma. *Cancer Chemother Pharmacol* 2011; **67**: 315-324 [PMID: 20390419 DOI: 10.1007/s00280-010-1320-2]
- 92 **Park JW**, Finn RS, Kim JS, Karwal M, Li RK, Ismail F, Thomas M, Harris R, Baudelet C, Walters I, Raoul JL. Phase II, open-label study of brivanib as first-line therapy in patients with advanced hepatocellular carcinoma. *Clin Cancer Res* 2011; **17**: 1973-1983 [PMID: 21349999 DOI: 10.1158/1078-0432.CCR-10-2011]
- 93 **El-Khoueiry A**, Posey JA, Castillo Ferrando JR, Krishnamurthi SS, Syed S, Kollia G, Walters I, Fischer BS, Masson E. The effects of liver impairment on the pharmacokinetics of brivanib, a dual inhibitor of fibroblast growth factor receptor and vascular endothelial growth factor receptor tyrosine kinases. *Cancer Chemother Pharmacol* 2013; **72**: 53-64 [PMID: 23719718 DOI: 10.1007/s00280-013-2168-z]
- 94 **Buijs M**, Reyes DK, Pawlik TM, Blackford AL, Salem R, Messersmith WA, Weekes CD, Mulcahy M, Kamel IR, Geschwind JF. Phase 2 trial of concurrent bevacizumab and transhepatic arterial chemoembolization in patients with unresectable hepatocellular carcinoma. *Cancer* 2013; **119**: 1042-1049 [PMID: 23132335 DOI: 10.1002/cncr.27859]
- 95 **Zhu AX**, Kudo M, Assenat E, Cattani S, Kang YK, Lim HY, Poon RT, Blanc JF, Vogel A, Chen CL, Dorval E, Peck-Radosavljevic M, Santoro A, Daniele B, Furuse J, Jappe A, Perraud K, Anak O, Sellami DB, Chen LT. Effect of everolimus on survival in advanced hepatocellular carcinoma after failure of sorafenib: the EVOLVE-1 randomized clinical trial. *JAMA* 2014; **312**: 57-67 [PMID: 25058218 DOI: 10.1001/jama.2014.7189]

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