

Prospective MEG Biomarkers in ASD: Pre-Clinical Evidence and Clinical Promise of Electrophysiological Signatures

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Autism spectrum disorders (ASD†) are characterized by social impairments and restricted/stereotyped behaviors and currently affect an estimated 1 in 68 children aged 8 years old. While there has been substantial recent focus on ASD in research, both the biological pathology and, perhaps consequently, a fully effective treatment have yet to be realized. What has remained throughout is the hypothesis that ASD has neurobiological underpinnings and the observation that both the phenotypic expression and likely the underlying etiology is highly heterogeneous. Given the neurodevelopmental basis of ASD, a biologically based marker (biomarker) could prove useful not only for diagnostic and prognostic purposes, but also for stratification and response indices for pharmaceutical development. In this review, we examine the current state of the field for MEG-related biomarkers in ASD. We describe several potential biomarkers (middle latency delays [M50/M100], mismatch negativity latency, gamma-band oscillatory activity), and investigate their relation to symptomatology, core domains of dysfunction (e.g., language impairment), and putative biological underpinnings.

INTRODUCTION

Autism spectrum disorders (ASD) are defined by restricted/stereotyped behaviors and social impairments [1] and have a massive impact on both the individuals with ASD and society itself. Currently, an estimated 1 in 68 children aged 8 years old suffer from an ASD [2]. Neither the biological pathology nor, perhaps consequently, a fully effective treatment have yet to be realized for ASD, even after substantial focus on ASD in research. In fact, the list of possible pathogenic mechanisms has dramatically increased, including multiple genetic mutations, combinations thereof and gene-environment interactions

[3], manifesting through a variety of mechanisms, including imbalance of excitation and inhibition [4] as well as hypo/hyper-connectivity [5]. What seems to be of consensus is that ASD have neurobiological underpinnings and, moreover, phenotypic expression, and likely the underlying etiology is highly heterogeneous. As such, a biologically based marker (biomarker), rooted in one or more of the various etiologies, could prove useful for both diagnostic and prognostic purposes. Furthermore, a valuable role of such a biomarker lies in subject/patient stratification in both clinical trials (e.g., by population enrichment through biologically based inclusion criteria)

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†Abbreviations: ASD, autism spectrum disorders; biomarker, biologically based marker; EEG, electroencephalography; MEG, magnetoencephalography; MRI, magnetic resonance imaging; fMRI, functional magnetic resonance imaging; PET, positron emission tomography; ERP, event-related potential; ERF, event-related field; EMG, electromyography; DTI, diffusion tensor imaging; FA, fractional anisotropy; MMF, magnetic mismatch field; MMN, mismatch negativity; Gamma, 30-80 Hz oscillatory activity; GABA, γ -Aminobutyric acid; mRNA, messenger RNA; EAAT, excitatory amino acid transporters; AMPA, α -Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; GAD65/67, glutamic acid decarboxylase; MRS, magnetic resonance spectroscopy; NLGN3, neuroligin-3; PV, parvalbumin; PEERS, Program for the Education and Enrichment of Relational Skills.

Keywords: ASD, biomarker, MEG, signature, translational, latency delay, Gamma

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Table 1. Electrophysiological signatures of ASD and their potential for biomarker use.

Biomarker	Simple to Implement	Sensitive	Specific	Scale	Responsive to Treatment	Biological Basis	Predictive
M50/100 Latency Delay	✓	✓	✓	✓	✓ (pre-clinical)	✓ (hypothesized)	untested
MMF/N	✓	✓	to RDoC domain but not clinical label	✓	untested	less clear	untested
Gamma Band Dysfunction	✓	✓	to RDoC domains, but not clinical label	✓	✓	✓	✓

and subsequently matching/selecting treatment to patient. In such a way, the heterogeneous ASD population would be biologically divided — thus made at least somewhat less heterogeneous — into good candidates for a particular treatment (a sub-population that would be targeted from inclusion) versus a less favorable sub-population. Additionally, it would provide biologically targeted objective indices of drug target engagement and activity for dose finding and as early signals of possible efficacy. Particularly since recruitment would be targeted toward that subset of the heterogeneous autism spectrum that exhibit this particular endophenotype, it should be expected to respond (perhaps, normalize) if indeed it is biologically tuned to the mode of action of the drug being evaluated. Exploring the biological basis of ASD in the pursuit of such biomarkers will also likely continue to identify substrates and neurobiological mechanisms underpinning the symptoms of ASD, informing our neurobiological understanding of the disorder and the basis of its comorbidities (e.g., attention deficit hyperactivity disorder and seizure disorders, both associated with neurotransmitter imbalance) as well as providing targets for further development of interventions.

In general, for a biomarker to be effective, it must be sensitive and specific to the disorder/dysfunctional domain, scale with severity, be simple to implement, and demonstrate robustness. An additional opportunity emerges in use of the biomarker to bridge the gap between clinical and pre-clinical environments, essentially providing a degree of validity to pre-clinical models that a biologically relevant phenomenon or trait is, in fact, being recapitulated [6]. As the label itself implies, a “bio” marker owes a plausible biological mechanism (or, at least, hypothesis), commonly rooted in both clinical and pre-clinical findings. Several emerging biomarkers have been proposed for adoption in ASD, ranging from blood-based assays [7], magnetic resonance imaging (MRI) [8], positron emission tomography (PET) imaging [9] and electrophysiology, and electroencephalography (EEG) or magnetoencephalography (MEG) [6,10-12]. In addition, other markers not as firmly rooted in biology have been also suggested for adoption in ASD, such as behavioral and eye-tracking performance [13]. This review focuses

on electrophysiological biomarkers, as they satisfy the above measurement requirements and offer plausible biological hypotheses for their basis. Specifically, while both EEG and MEG methodologies have been used to probe ASD, in this review, we will focus on MEG.

CURRENT ELECTROPHYSIOLOGICAL SIGNATURES OF ASD: NON-INVASIVE, PRECISE, AND SENSITIVE TO NEURONAL MASS FIRING

Of the various classes of candidate biomarkers, electrophysiological signatures of ASD hold particular promise in the study and treatment of ASD because of the non-invasive nature of recordings for a population traditionally challenged by imaging procedure compliance demands (Table 1). Passive paradigms, along with recording of early, obligate responses offer strategies to defend against confounding influences of attention, performance, and movement. Over the last several decades, researchers have explored numerous electrophysiological signatures and have found varying relationships of these measures to ASD symptom severity and specificity [14]. While this association of observational findings and symptomatology and the non-invasive nature of recordings are not unique to electrophysiological methods (e.g., other imaging methods such as fMRI/MRI/PET show such profiles [15,16]), electrophysiology has the important distinction of being exquisitely sensitive to the temporal dimension, and, thus, unlike MRI, fMRI, and PET, electrophysiological methodologies allow investigations and characterization of the timing, synchrony, and connectivity of actual neural activity rather than simply spatial identification of secondary associations (e.g., hemodynamic response through neurovascular coupling). While exhibiting sensitivity to similar neuronal electrical activity as EEG, MEG extends the capabilities of EEG with a more precise spatial resolution than its electrical measuring cousin. In contrast to EEG, MEG’s resolution is in the range of millimeters [17], similar to the range of fMRI. In addition, MEG is relatively immune (via the use of spatial filtering-based source localization) from contaminating EMG artifact in high frequency bands arising from scalp and facial muscle [18] as

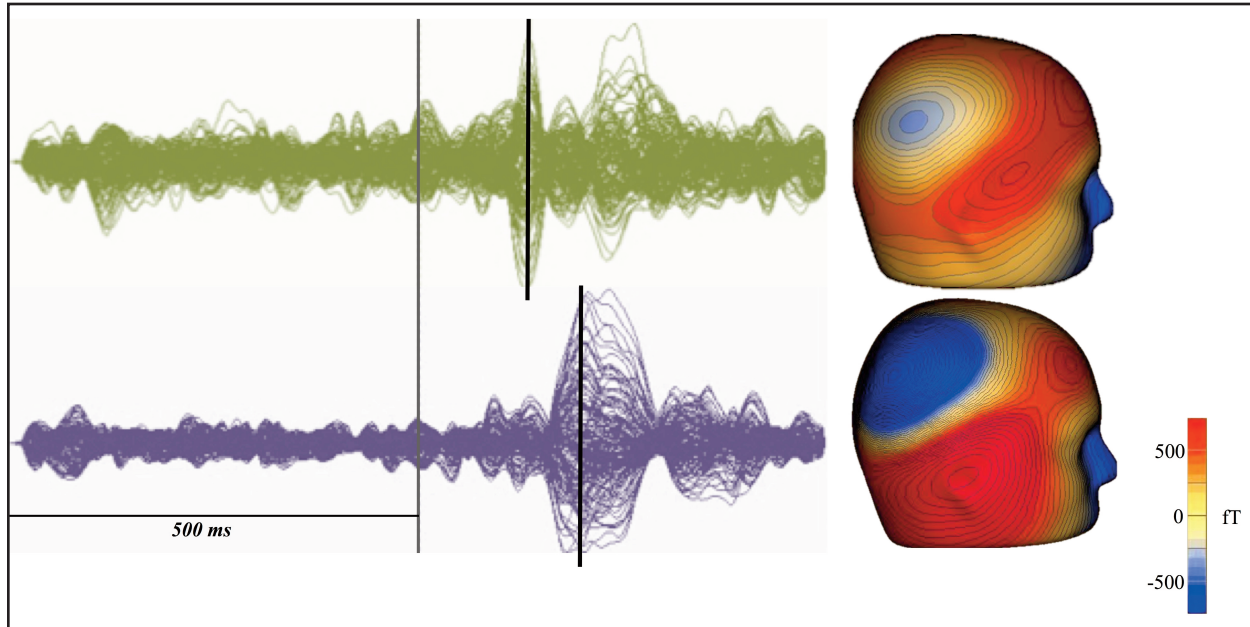


Figure 1. Delayed M100 response latency in ASD, when compared to age matched typically developing controls. ASD (bottom) demonstrate a delay of the M100 response compared to typically developing controls (top) as detected by MEG. At left, ERP of typically developing (green) and children with ASD (purple), with stimulus marked in gray and M100 denoted by black bar. At right, 3D topographic representations of the M100 responses demonstrate that both populations M100 arise from the region of auditory cortex (top: TD; bottom: ASD).

well as microsaccadic eye movements [19]. Signatures derived from electrophysiological measures, and especially MEG, technically offer the potential of increased sensitivity as well as specificity for neurophysiological abnormalities that may underlie symptom domains of ASD and thus present opportunities for evaluation as potential biomarkers.

M50 AND M100 LATENCY DELAYS: PREDICTIVE WITH A PLAUSIBLE BIOLOGICAL BASIS

One of the most replicable electrophysiological findings is the latency delay of middle-latency M50/M100 responses in ASD as compared to typically developing children. First demonstrated by Gage and colleagues, right hemisphere M100 latencies exhibit both altered maturation [20] and reduced representational dynamic range [21]. These findings went against some concurrent findings that resolved no latency prolongation of the M100 response [22], though this study has exclusively low functioning subjects with ASD for the experimental group, and methodological differences such as choice of stimuli (1000Hz/1200Hz standard/deviant) may account for the lack of findings. Furthermore, the M100 response was defined as the “first power maximum following the 50-msec latency,” which has been shown to be either the M50 or M100, depending on the age — and potentially, diagnostic status — of the subject [23]. While it is posited that both M50 and M100 are delayed in ASD, to detect M50 latency prolongation, larger cohort sizes have been needed [24]; importantly, distinction should be made between responses that are “late M50s” versus

“early or typical M100s” as potential overlap in latency range can obscure group findings; topographic representations of surface magnetic fields can readily aid this resolution. In addition to this potential misattribution, M100 response latencies, even in response to simple sinusoidal tones, are not stationary. Middle latency-evoked potentials can differ depending on stimulus properties such as frequency [25], intensity [26], and other features [27]. As such, it is important to standardize all stimulus properties and delivery methodology. For instance, we the authors always conduct experiments with stimuli at 45 dB above sensation level, well above the plateau of intensity effects of the stimuli on latency. Also, during each session, multiple (200, 300, 500, and 1000Hz tones) stimuli are used to both fully characterize responses and allow comparison to other institutions that use different frequencies. Direct comparisons of M100 latencies recorded at different sites occurred in a recent multi-center collaboration [28]. Here, great care was undertaken to standardize all aspects of the stimuli presentation, although a persistent up-to-5 ms delay continued to exist between sites, which may either reflect technical precision limits or imperfect subject matching between sites or a combination thereof.

Delays in the M100 latency were recapitulated later in a larger cohort and survived co-varying for age and language ability [29]. Generally, these delays are of the order of ~10ms (Figure 1) and thus, in themselves, quite demanding of measurement precision. M50 latency delays were also observed in a similarly large-sized sample [24]. While M50 responses are larger in amplitude as well as later in children than adults [23], both the M50 and M100 latency delays seen in ASD are ~10 percent of the typical

Table 2. Summary of MEG based M50/M100 findings in ASD, and their relation to structural and behavioral measures.

Study	M50	M100	Correlate to Age	Correlate to IQ	Correlate to Language Impairment	Correlate to White Matter Microstructure
Gage et al., 2003 [21]	Not Reported	Reduced dynamic range of M100 latency to different frequencies in the right hemisphere	Within-subject-normalized M100 latencies were not modulated by age (ANOVA)	Not Tested	Not Tested	Not Tested
Gage et al., 2003 [20]	Not Tested	M100 latency is prolonged in ASD	Altered maturation of M100 in right hemisphere	Not Tested	Not Tested	Not Tested
Tecchio et al., 2003 [22]	Not Tested	No M100 differences exhibited between TD and ASD	M100 decreased as a function of age (only 1 hemisphere reported)	Not Tested	Not Tested	Not Tested
Oram Cardy et al., 2004 [23]	No differences exhibited between TD and ASD	No differences exhibited between TD and ASD	Both TD and ASD children have delayed M50/M100 latencies compare to healthy adults	Not Tested	Not Tested	Not Tested
Roberts et al., 2010 [29]	No resolvable difference in latency or strength	Delayed M100 in right hemisphere of ASD	Age correlated to M100 latency in TD only	No relationship between cognitive ability and M100 latency	No relationship between language ability and M100 latency	Not Tested
Roberts et al., 2013 [24]	10% delayed M50 when averaged across hemisphere	Not Reported	ASD and TD both show maturation, with no difference in slope of fit, but with different intercepts	No relationship between cognitive ability and M100 latency	No relationship between language ability and M100 latency	FA and M50 latency negatively correlate in TD only
Edgar et al., 2013 [62]	Not Tested	10% delay in ASD in right hemisphere	Age predicts M100 latency	Not Tested	Not Tested	Not Tested
Edgar et al., 2014 [30]	Delayed M50 for both ASD+/-LI (Language Impairment)	M100 detected less often in ASD +LI than TD in younger children (6-10yrs old). In older subject (11-15 years old) groups, ASD-LI has more responses than ASD+LI. M100 delayed in ASD	Age predicts M50/100 latency	No relationship between cognitive ability and M100 latency	No relationship between language ability and M100 latency	Not Tested
Oram Cardy et al., 2008 [31]	Not Reported	Not Reported	Left hemisphere M50/M100 predicted age	No relationship between cognitive ability and M100 latency	Right hemisphere M50 and to less extent M100 predicts language ability, especially true for receptive language	Not Tested
Roberts et al., 2012 [32]	Not Tested	M100 not delayed in SLI	Not Tested	No relationship between cognitive ability and M100 latency	Not Tested	Not Tested

latency (M50 [24], M100 [29]). Thus, M50 delays (~5ms) are even more demanding to resolve, so less statistical power may be needed to resolve M100 delays (given similar measurement precision), explaining the predominance of focus on this later component. In both cases, large sample studies are necessary to avoid misrepresenting the broad distribution of responses across the spectrum. Nonetheless, M50 latency delays in ASD have also been replicated in later analyses [30]. Maturational changes in both M100 (especially right hemisphere) and M50 (bilaterally resolved) suggest that although there is a pattern of latency shortening with increasing age — in fact, at a rate similar to that of typical development — a persistent shift or prolongation of latency exists in ASD compared to controls at any given age, thereby defining an atypical developmental trajectory, despite potentially similar rate [20,24].

Confounding these results are the observations that M100 responses are detectable less often in young children with ASD and comorbid language impairment than typically developing children [30], potentially skewing the distribution of samples in whom responses are indeed obtained. Another yet unresolved aspect of M50/M100 latency delays are their association with ultimate clinical language impairment. M50 latency and, to a lesser extent, M100 latencies are reported to predict oral language ability [31], though this has failed to be replicated in all cohorts [29]. Recent evidence suggests an association of M50/M100 latencies with language ability is better seen with more specific intermediate-level language phenotyping (e.g., phonological processing using the CTOPP), although this has not been fully evaluated. Interestingly, M100 delays seem specific to ASD and are not evident in specific language impairment (SLI) [32]. Thus, currently, it appears that the M50/M100 delays may indicate atypical auditory processing and may indeed underlie clinical language impairments, but a direct and striking correlational mapping between simple M50/M100 latency and ultimate high-level behavioral language performance is not present.

Beyond a diagnostic and potentially prognostic role, M50/M100 latency delays also show the translational promise of signaling treatment efficacy in pre-clinical models. Prenatal valproic acid insult in rodents can recapitulate key aspects of ASD in the offspring, including several auditory related alterations such as auditory cortex (N1) latency delays [6]. In this rodent model, behavioral training can recover otherwise delayed auditory P1 (analogous to M50) latencies [33]. Similar changes have been found in children with ASD, with behavioral training recovering N1 (EEG equivalent of M100) latencies in children with ASD, though care should be taken due to the extremely small number of participants [34]. Partial support for this finding arises from studies on other sensory modalities, where recovery of typical event-related potential (ERP) latencies through the Early Start Denver Model (a high intensity, applied behavioral analysis-based intervention focusing on social and emotion domains

shown to be effective in a randomized clinical trial [35]) was concurrent with behavioral improvements, though in a larger sample cohort [36]. Interestingly, recent pre-clinical work now suggests a link between N1 latency and sociability in several studies [37,38], though similar associations of M100 and behavioral measures of ASD have yet to be thoroughly studied clinically (Table 2).

Finally, in fulfillment of a biomarker's plausible biological basis, early event-related potential/field (ERP/ERF) latency delays could, in principle, arise from multiple mechanisms. Currently, the best experimentally supported hypotheses are either white matter alterations leading to poor signal conduction [24,39-41] or synaptic dysfunction [42-46] leading to poor signal transduction. Additional mechanisms and combinations can, of course, also be considered. White matter alterations not directly visible to electrophysiologic techniques can be probed using diffusion-sensitive MRI called diffusion tensor imaging (DTI). Such methods also afford a quantitative description of properties of the white matter microstructure, often represented in the terms "mean diffusivity (MD)" and "fractional anisotropy (FA)." These are then used to draw inferences about white matter developmental changes. While DTI indices such as FA are sometimes interpreted as measures of microstructural "integrity," it should be borne in mind that there are several underlying metrics and their combination in summary measures such as FA can obscure findings. For example, although the thalamocortical white matter projections of children with ASD exhibit similar mean FA microstructural metrics to typically developing controls, the age-FA relationship evident in typical development [39] was not present in ASD. When compared directly to the M50 response latency, increased FA correlated with decreased M50 latency, suggesting a biophysical hypothesis linking white matter conduction velocity and cortical response timing; such a relationship was not observed in ASD [24]. However, a more in-depth investigation in Roberts et al. [24] showed that white matter maturation in ASD might still be occurring (as resolved in changing mean diffusivity or the eigenvalue components thereof), despite being unresolved in FA, but importantly through an atypical mechanism driven by atypical maturational trajectories of axial and radial diffusivity components [24].

As such, DTI supports the notion of ongoing, but atypical, maturation of the thalamocortical white matter, perhaps accounting for M50/M100 maturational trajectories showing age-related latency shortening, but nonetheless persistent delays with respect to typically developing controls. Further insight to the biological underpinnings of early ERP-related latency delays in ASD comes from subjects with 16p11.2 abnormalities. These subjects are at greater risk for ASD (15 percent of 16p11.2 alterations also demonstrating ASD [47]). A multi-site collaboration recently observed M100 latency delays in subjects with 16p11.2 deletions. Such delays were again observed in conjunction with white matter abnormalities [48], though the exact interpretation of such microstructure alterations

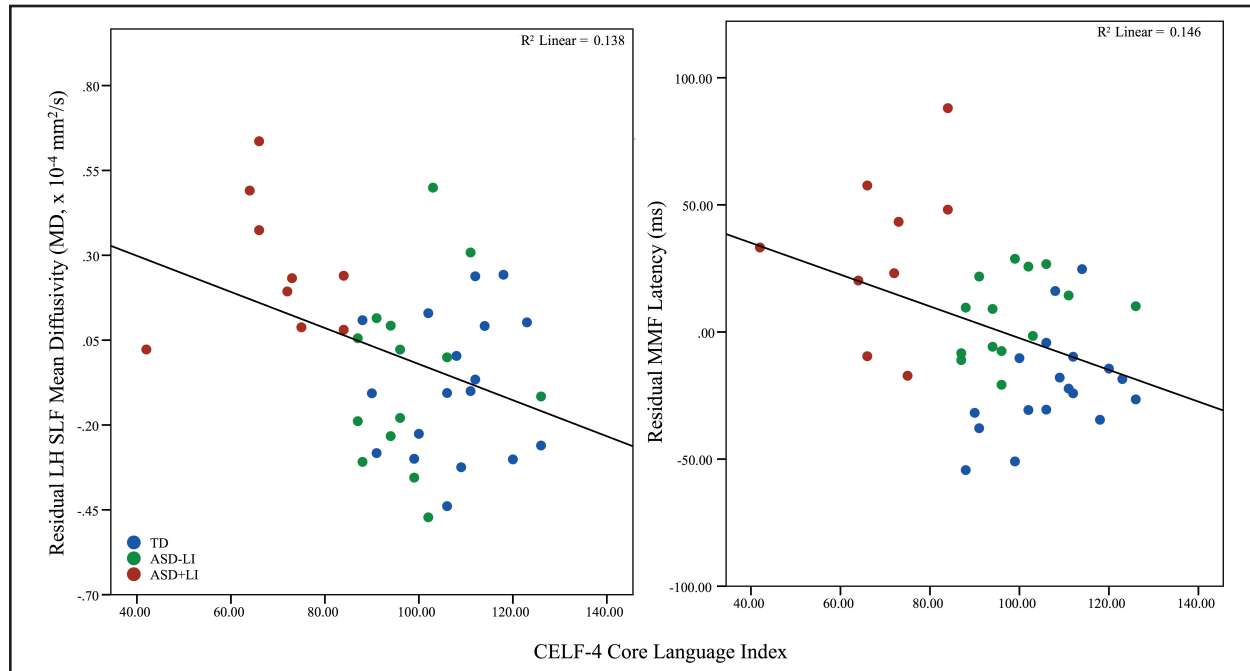


Figure 2. Mismatch fields and microstructure of arcuate fasciculus demonstrate qualitatively similar relationships to language ability. Age corrected mean diffusivity of the arcuate fasciculus (left) and age corrected mismatch field latency (right) both show negative correlations to language ability as indexed via the CELF-4 core language index. No hypothesis currently directly links the latency of mismatch fields to the white matter microstructure of the arcuate fasciculus, in contrast to the M50/100 relationship to thalamocortical white matter microstructure. Nonetheless, a clear analogy is offered.

is unclear, due to increases in the size of several anatomic structures and white matter in general. Interestingly, pre-clinical work with rodent models that recapitulates key aspects of ASD suggest that M100 latency delays are actually due to delays in anterior auditory field (AAF) rather than primary auditory cortex responses [46], spatial scales that are unresolvable by MEG [17]. This last finding may suggest that synaptic dysfunction may partially account for prolonged latencies, due to improper integration in auditory cortex.

MISMATCH NEGATIVITY/FIELD (MMN/F): PREDICTS LANGUAGE IMPAIRMENT AND ASD, THOUGH LESS-DIRECT BIOLOGICAL UNDERPINNING

Similar to the M50/M100 latency delays in response to simple auditory presentation of sinusoidal tones seen in ASD, children with ASD can also exhibit a latency delay of the auditory magnetic mismatch field [49]. The magnetic mismatch field (MMF) and its EEG counterpart, mismatch negativity (MMN), refer to the resultant ERP derived from subtracting a standard or frequent stimulus' ERP from that of a rare or novel stimulus [50] and essentially representing the “difference wave” as an index of “change detection.” To perform such an experiment, a stream of standard stimuli are presented, with irregular interruption by the deviant stimuli (e.g., /a//a//a//u//a/). Responses for the standard stimuli and deviant stimuli are separately averaged into

ERPs, and then the standard stimulus ERP is subtracted from the deviant stimulus ERP. The MMF/N measure is sensitive to changing stimulus parameters, for example, a phoneme category boundary being crossed between two stimuli [50]. This signature exhibits face validity, since change detection is critical for the accurate processing of speech, and, moreover, language impairment is a commonly observed feature of ASD [51]. The MMF/N latency deficit is not, however, dependent on the use of language stimuli (i.e., vowels); delays are also observed to sinusoidal tones [49], implicating an auditory processing sensitivity. Furthermore, the MMF delay [38] is in addition to any intrinsic ASD-related delay in the earlier M100 component.

Analogous alterations to MMF latencies have been replicated using different paradigms [52], though the exact nature of the alteration is not stationary; for example, the MMF is sometimes missing in ASD populations [22]. Similar results have been exhibited via EEG, though they are not always consistent [53]. For instance, Gomot and colleagues have found shortened MMF latencies using EEG, although it is not clear if the difference is due to the use of higher frequency stimuli than aforementioned reported studies with findings of prolongation of latencies or small sample sizes [54,55]. Others have also found no differences in MMF latency when using multiple stimuli types, but in small sample populations [56].

Interestingly, from the view of a biomarker, MMF latency delays scales within the ASD cohort with clinical measures of language ability and is a sensitive and spe-

cific predictor of language impairment [10]. When compared to another MEG signature (e.g., M100 latency delay) or a structural imaging marker, MMF timing alterations have a better accuracy for both predicting both ASD and also language impairment within ASD [57]. The specificity of MMF to ASD-related language impairment is in question, though, as selective MMF delays also exist in children who demonstrate language impairment but are not on the autism spectrum [32].

When considering MMN/F alterations, it is necessary to keep in mind results from a similar paradigm: rapid temporal processing. This paradigm consists of two rapidly presented tones (e.g., 150ms apart), and studies have demonstrated that the earlier components of the second tone are less likely to be exhibited by language-impaired children regardless of ASD diagnosis as compared to children with no language perturbations (typically developing and ASD) [58]. The specificity of this may also be called into question with dyslexic adults demonstrating weaker amplitude responses [59], although its sensitivity for impaired language function is supported. As such, it may be that both rapid temporal processing measures and MMF/N are best considered markers of language domains and not specific for clinical labels.

What remains unknown is the biological basis for the MMF/N and how this might be related to the biological perturbations frequently exhibited by those with ASD. Addressing this indirectly, however, there is emerging evidence from DTI studies that white matter microstructure of the left hemisphere arcuate fasciculus (a key part of the language pathway) also exhibits atypicalities in ASD and indeed as a function of language impairment. Visual comparison of quantitative associations between MMF latency and clinical language assessment on the one hand and white matter microstructural metrics versus clinical language assessment on the other are strikingly analogous (Figure 2). A direct hypothesized mechanism for the biological coupling of the white matter and cortical measures is not yet available, in contrast to the M50/M100 and thalamocortical white matter association discussed above.

GAMMA-BAND OSCILLATORY ACTIVITY: PERVASIVE, REPLICABLE, PREDICTIVE WITH A SPECIFIC PUTATIVE BIOLOGICAL BASIS

A recent topic of increased study for ASD is Gamma-band oscillatory activity (Gamma [30-80 Hz]). While not specific to ASD [60], Gamma dysfunction (“oscillopathy”) may relate to certain core domains of symptomatology, with the ultimately impacted system/domain dependent on the time of critical dysfunction or region involved [61]. Importantly, auditory response-related Gamma has been repeatedly demonstrated to be altered in ASD [6,62-66]. Evoked (phase-locked) Gamma power for both transient responses to simple sinusoidal tones and driven by 40Hz auditory steady state stimuli is reduced in ASD [65,66]. Concomitant with such phase-locked

deficits, increased induced (non-phase locked activity) activity [64] has been observed, though this does not consistently achieve statistical significance [6]. Interestingly, during spoken word recognition, a reverse profile of Gamma dysfunction occurs compared to simpler stimuli; evoked Gamma is increased, and total (both phase and non-phase locked) Gamma power, which is driven by induced power changes, responses are diminished [63]. As such, the exact Gamma dysfunction exhibited seems dependent on stimulus type.

For both types of stimuli (simple/complex), relationships to symptomatology and/or core functional domains can be found. The total Gamma power elicited by the spoken word recognition task was positively correlated to figurative language abilities in controls, with qualitatively the opposite association exhibited in ASD. Separately, the transient and steady state Gamma responses that result from simpler stimuli both correlate to more basic functionality such as communication and symptom severity rating of ASD [62,65]. The relationship of Gamma in response to simpler stimuli and more basic functioning has an intuitive basis, since this Gamma dysfunction is in a lower level of sensory processing.

Gamma deficits are not specific to the auditory system; replicable high frequency oscillatory alterations have been found elsewhere (e.g., visual system [67,68], and at rest [69-71]), with some finding correlations of these Gamma perturbations to symptom severity [71]. Similar to the alterations seen at rest, pre-stimulus measures of Gamma are increased in ASD (sometimes considered evidence of a “noisy” brain), with elevated pre-stimulus Gamma corresponding to language abilities in ASD [62]. Together, such findings demonstrate an emerging coherent picture of the pervasiveness and importance of Gamma oscillopathy with regard to symptomatology and core domain dysfunction in ASD and its potential suitability as a biomarker. Moreover, in fulfillment of this status, Gamma functioning is also predictive of later outcome; infants’ frontal region Gamma not only correlates to current [72] but also future [73] cognitive and language abilities, both of which are core domains associated with ASD. Also during infancy, Gamma metrics are discriminative for risk status (low versus high) for ASD; two separate studies have demonstrated differential Gamma profiles in low risk (no sibling with ASD) versus high risk (at least one sibling with ASD) for autism [74,75]. Furthermore, after Tierney et al. removed all subjects that finally met criteria for ASD, difference still remained significant, suggesting that Gamma may be an endophenotype of ASD [59]. This is supported by work by Rojas and colleagues, where two studies have identified Gamma dysfunction in first-degree relatives of ASD [64,65]. While the identification of Gamma dysfunction in high-risk infants or first-degree relative of people with ASD may paradoxically undermine the use of Gamma as a diagnostic biomarker (since it is also altered in “unaffected”/undiagnosed relatives), a broad autism phenotype (BAP) has long been associated

within first-degree relatives of people with ASD [76], suggesting a possible biological sensitivity that exceeds clinical resolution and categorization.

Lastly, Gamma-related metrics also hold the potential for measuring treatment efficacy, not only for synaptically targeted pharmaceuticals but also for behavioral interventions, as a reversal of Gamma perturbations coincided and correlated with effective intervention using Program for the Education and Enrichment of Relational Skills (PEERs) [77], a behavioral intervention utilizing parental assistance for the teaching of social functioning to teens with ASD, shown to be effective in a randomized controlled study [78].

Biological Evidence for Human Oscillopathies

Gamma is posited to depend on numerous constituents, including potassium channel subtypes, spike conductance trajectories, and a strong role for both glutamatergic and γ -Aminobutyric acid (GABA)-ergic signaling [79]. There are two widely accepted models for the mechanism of Gamma generation: the I-I model and E-I model. In both models, inhibitory feedback onto specific cells (interneurons and pyramidal cells, respectively, for each model) cause a temporary quiescence in population firing [79]. Therefore, Gamma is thought to arise from local circuit interactions that depend heavily on GABAergic/glutamatergic receptor-based kinetics [79]. Supporting this putative GABA-Gamma interaction are findings that relative cortical GABA levels correlate to Gamma in motor and visual cortex [80,81], and initial work by our lab and others suggest this is true for auditory cortex as well. Such emerging correlations are not without controversy, with at least one other laboratory showing no correlation between relative cortical GABA and Gamma [82]. The possibility of regional variability in this coupling also cannot be excluded in accounting for varying observations in the literature, along with the usual suspects of measurement precision and sample size.

With this link between GABA and the mechanistic underpinning of Gamma, and the clear Gamma alterations in ASD, it is not surprising that the balance of excitation and inhibition has been posed as a pathogenic mechanism for ASD [4]. In addition, there has been substantial data from human studies to suggest the existence of such an imbalance [83]. The high comorbidity of seizure disorders such as epilepsy in ASD also points to hyper-excitability and an imbalance in GABA and glutamate [84]. Indeed, the high co-occurrence of seizure disorders in ASD and the known glutamatergic/gabaergic basis of some, if not all, seizures may have led in considerable part to Rubenstein and Merzenich's proposed model of E/I imbalance as a basis for ASD [4]. With respect to glutamate-related alterations, a GluR6 receptor subunit has been tied to ASD inheritance [85-87]. Subjects with ASD can also demonstrate increased expression of glutamate transmission-related messenger RNA (mRNA) and proteins, for example, Excitatory Amino Acid Transporters (EAAT) and α -Amino-3-hydroxy-5-

methyl-4-isoxazolepropionic acid (AMPA) isotypes [88]. Additionally, Magnetic Resonance Spectroscopy (MRS) has demonstrated possible increased glutamate in the cortex of ASD individuals [89], although this last finding is not unambiguously interpreted because of the role of glutamate in other intracellular processes [90]. For inhibition, the 15q11 locus, which encodes many GABA signaling-related proteins, is deleted in as much as 3 percent of the ASD population [91]. Also, both GABA production enzymes (glutamic acid decarboxylase [GAD] 65 and GAD 67 and receptors [GABAA and GABAB]) have been found to be decreased in postmortem studies of subjects with ASD [92-94]. This is supported by several studies in which subjects with ASD exhibited decreased cortical GABA concentrations [42-44]. As such, Gamma activity in general, and oscillopathies in particular, seem not only supported with a biological mechanism, but this mechanism appears to be altered in ASD, enhancing the optimism of Gamma-based measures as candidate biomarkers.

MOUSE MODEL VERIFICATION OF THE BIOLOGICAL MECHANISMS OF INTERMEDIATE PHENOTYPES: CLINICAL AND BASIC RESEARCH INFORMING EACH OTHER

Recent work has shown that mouse models that recapitulate key aspects of ASD, whether due to prenatal environmental insult [6], pharmacological treatment [95], or genetic manipulation [37,96,97], demonstrate analogous perturbations to their auditory electrophysiological responses with both delayed early/middle latencies and aberrant oscillatory profile. Such alterations can be related to the synaptic/circuitry alterations exhibited by these mice. In mice treated prenatally with valproic acid, Gamma deficits are correlated to neuroligin-3 (NLGN3), a protein integral in the building and maintenance of synapses [6]. This same prenatal treatment also causes a hyper-connectivity and hyper-excitability within circuit functioning [98]. Lastly, when studied in the aforementioned mouse models, altered GABAergic cell expression is found. Mice treated prenatally with valproic acid display reductions in Parvalbumin containing (PV+) interneuron cells counts in [99], cells thought to be crucial in Gamma activity [100]. Such PV+ modulation is also observed in another animal model exhibiting stimulus-related Gamma activity deficits, with down-regulated PV+ cell count and protein expression in NR1neo^{-/-} mice [97]. Interestingly, the NR1 neo^{-/-} mouse model's hippocampal pyramidal cells also exhibit intrinsic hyper-excitability, further demonstrating altered E/I balance [97]. These alterations to synaptic transmission and circuit parameters could lead not only to alterations to oscillatory activity, but also delays in ERP components. Figure 3 draws some analogies between human studies of ASD and various pre-clinical models probing aspects of ASD neurobiology.

Such pre-clinical work has also informed ongoing human studies. Several mouse models exhibit negative

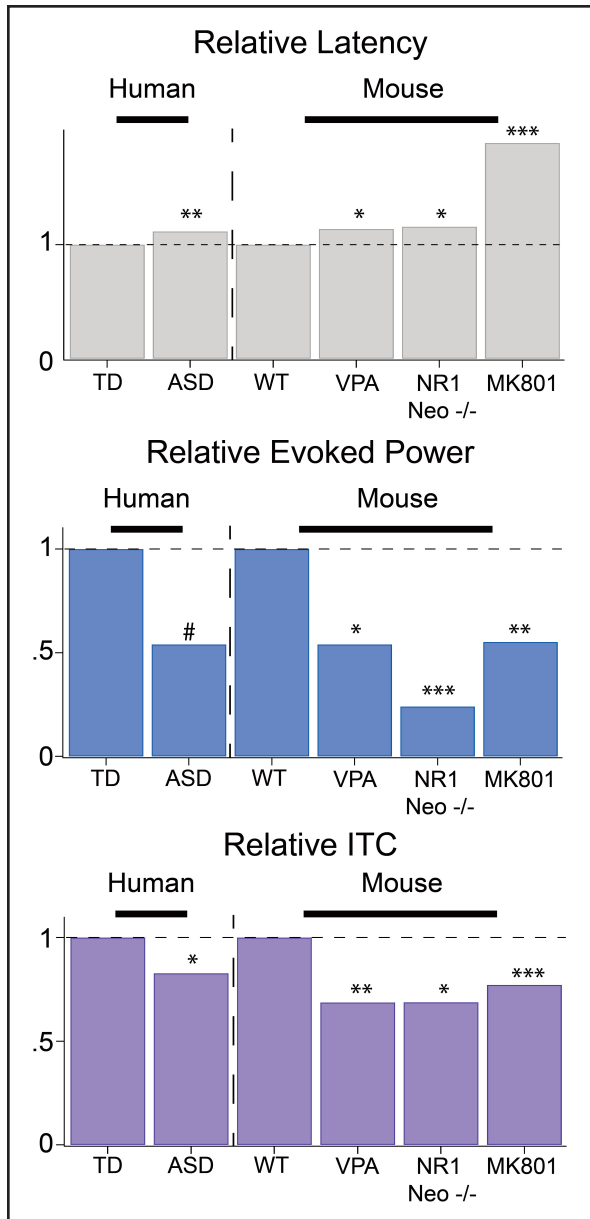


Figure 3. Pre-clinical work mirrors that of clinical research for multiple biomarkers. M100 latency (top) delays seen in humans are mirrored in several mouse models derived from different mechanisms. The same mice also exhibit decreases in evoked GAMMA responses (middle) and GAMMA inter-trial coherence (ITC) (bottom). (VPA, prenatal insult; NR1, Neo^{-/-}, genetic insult; MK801, pharmacological challenge). Adapted from Port et al. [61]. ***P < 0.001, **P < 0.01, *P < 0.05, #P < 0.1

correlations between sociability and baseline Gamma-band activity [38,97]. This prompted work that showed similar results in children with ASD [62]. A putative link between baseline Gamma and sociability was demonstrated using optogenetics by Yizhar and colleagues, who demonstrated increased baseline Gamma-band activity due to increased excitatory drive, which coincided with a decrease in sociability; additionally, such a deficit could be

reversed by recovering E/I balance [101]. Separately, others have demonstrated that pharmacological reversal of Gamma signal to noise deficits in a genetic insult model of ASD with GABA-B agonist treatment reversed behavior impairments [97]. Such recovery of symptomology coinciding with reversal of Gamma abnormalities is relevant and reflected in humans; for example, as mentioned above, Gamma-band function is partially recovered with PEERS intervention [77].

CONCLUSION AND OUTLOOK

MEG-based biomarkers display great potential for use in ASD. Not only are biomarkers presented here grounded in biology, each also have substantial support from the literature for their use. These MEG biomarkers may, in fact, be deemed more suitable for ASD than more invasive assays (i.e., blood-based), since the subject is only mildly inconvenienced. Moreover, compared to other imaging techniques, MEG is favorable due to the use of non-claustrophobia/anxiety inducing setups for which MRI-based techniques suffer due to the requirement of a magnetic bores and loud machine noise and minimal sensory contact as opposed to applying EEG head caps.

Although the current state of electrophysiological biomarkers in ASD is promising, there are still unresolved issues regarding the sensitivity and specificity of each candidate signature. Therefore, at the moment, the greatest promise may be toward a multivariate combination of such biomarkers [57]. The unresolved concerns for clinical specificity of such biomarkers may, in fact, reflect the nature of biological dysfunction and offer opportunities to investigate comorbidities and potentially redefine clinical categories. What these biomarkers may actually detect are core domain alterations and not ASD per se. As such, this would explain the occurrence of similar abnormalities in other disorders (e.g., Gamma alterations in bipolar [102] and schizophrenia [103]). This is not unprecedented; the new focus from the National Institute of Mental Health on Research Domain Criteria encourages testing the relationship between biomarkers and specific cognitive and sensory domains even if the biomarkers are not selective for traditionally defined disorders. While further study is needed to perfect these biomarkers to the point of clinical use, promise is nevertheless exhibited even in these early stages. Indeed, this approach may be more clinically relevant in disorders as heterogeneous as ASD as roles extend beyond diagnosis and prognosis toward stratification (sub-population definition) and monitoring of treatment response. For instance, if a biomarker could stratify subjects with ASD based on certain biological alterations (i.e., poor synaptic function as revealed by Gamma alterations versus poor signal transmission as revealed by white matter alterations), new sub-types of ASD (those with/without Gamma and/or white matter alterations) could be defined. Then treatments targeted to those alterations could be implemented and monitored for efficacy.

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