Pharmacogenomics – how close/far are we to practising individualized medicine for children?

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The translation of pharmacogenomics into clinical practice is a key approach for practising individualized medicine, which aims to maximize drug efficacy and minimize drug toxicity. Since the completion of both the Human Genome Project and the International HapMap project, the development of pharmacogenomics has been greatly facilitated. However, progress in translating pharmacogenomics into clinical practice, especially in paediatric medicine, is unexpectedly slow. Many challenges from different areas remain. This paper discusses the existing applications and the limitations to the implementation of paediatric pharmacogenomics, as well as possible solutions for overcoming these limitations and challenges.

Introduction

Individualized medicine may be described as being the selection of 'the right drug at the right dose for the right patient'. In paediatric therapeutics, as with adults, the age, body size, genome profile, concomitant drug use and organ dysfunction can all affect drug efficacy and safety [1, 2]. Lack of drug efficacy may result in a delay in successful therapeutic treatment, whereas adverse drug reactions (ADRs) may result in poor drug adherence, ultimately leading to a lower quality of life and sometimes life-threatening events.

Genetic factors have been found to account for a significant proportion of the individual variability in drug response [3, 4]. Genetic variation in individuals affects the activities of drug-metabolizing enzymes [cytochrome P450 (CYP) enzymes], hence the pharmacokinetics and pharmacodynamics of drugs. Poor ability to metabolize a drug may lead to accumulation of this drug in the body. Conversely, an individual patient who metabolizes a drug too rapidly may experience instances where the therapeutic level of the drug is not reached, due to enhanced metabolism or 'clearance'. The study and application of pharmacogenomics is thus important in order to optimize drug therapy, hence leading to individualized medicine.

Currently, translating pharmacogenomic knowledge and research into clinical practice remains the first priority in implementing individualized medicine. With advances in genotyping technology and gene mapping, a wealth of pharmacogenomic data and tests are readily available. However, uptake of the use of these resources in clinical practice is unexpectedly slow in all age groups. There are considerable obstacles to the practice of individualized medicine. In children, these obstacles are more challenging due to such factors as various developmental stages and ethical concerns, which make the progress of paediatric pharmacogenomics even slower than for adults. In this article, we discuss the current implementation of paediatric pharmacogenomic knowledge, limitations to this implementation and the possible solutions for overcoming these challenges.

Practising pharmacogenomics

In the last decade, the completion of the Human Genome Project [5], the International HapMap Project [6] and the advances in high-throughput genotyping technologies have facilitated the burgeoning development of pharmacogenomic studies. More than 100 drugs, some of them are commonly used in children, such as thiopurine, warfarin and antipsychotic drugs, are mandated by the US Food and Drug Administration (FDA) to incorporate pharmacogenomic information in drug labelling under sections on Dosage and Administration, Warnings and Precautions and Indications and Usage etc. [7]. Most of the drug labels do not ask for a mandatory genotyping test, but the pharmacogenomic information provides additional guidelines for clinicians to design the drug therapy.

Examples applicable to both adults and children in clinical use

Carbamazepine and human leukocyte antigen (HLA) One of the promising examples of pharmacogenomics in practice is the genotyping test for human leukocyte antigen B (HLA-B) in patients with carbamazepine treatment. Carbamazepine, an anticonvulsant used to treat seizures and nerve pain, can cause severe cutaneous adverse drug reactions, such as Stevens-Johnson Syndrome (SJS) and toxic epidermal necrolysis (TEN). These adverse reactions are commonly found in Asian populations, and studies from China, Hong Kong, Malaysia, Taiwan and Thailand have reported that the HLA-B*1502 variant allele is strongly correlated with carbamazepine-induced SJS/TEN [8-12]. In other populations, such as Caucasians and African-Americans, the prevalence of HLA-B*1502 is almost absent, but study of European patients undergoing carbamazepine treatment has shown that another variant allele, HLA-A*3101, has a mild association with the increased risk of SJS/TEN [13].

In 2007, the US FDA included pharmacogenomic information on the drug label of carbamazepine, which recommended a genetic test for HLA-B*1502 in high-risk patients prior to treatment with carbamazepine. Patients who are positive for HLAB*1502 should not be prescribed with carbamazepine unless the benefits clearly outweigh the risks [14]. The screening for HLA-B*1502 in patients from high-risk ancestry is routinely performed in clinical practice. The successful application of pharmacogenomics in carbamazepine treatment can be attributed to a number of factors. First, the allele variant has a high correlation with the adverse event. In the Taiwanese study, the odds ratio for developing carbamazepine-induced SJS/TEN among patients with a positive result to HLA-B*1502 testing was 2504 (95% confidence interval 126-49 500) [8]. The odds ratios in other studies were also high [9, 10]. Second, the adverse events are severe and life threatening. There is a high mortality rate for SJS/TEN, ranging from 5% in SJS to 30% in TEN [15]. Third, other alternatives to carbamazepine, such as lamotrigine, are available. Therefore, based on the result of the genotyping test, clinicians can consider the best medication for the patient. For these reasons, pharmacogenomic information is very helpful and is widely used in carbamazepine treatment. Certainly, it is routinely used in many areas with a high Chinese population, such as Hong Kong, in both children and adults.

Thiopurine and thiopurine methyltransferase (TPMT) Thiopurine is commonly used to treat acute lymphoblastic leukaemia (ALL), in both adults and children. It is a prodrug that is metabolized by TPMT. To date, over 25 variants in the TPMT gene have been identified, most of these variants being associated with TPMT activity. It has been reported that >90% of ALL patients carry the TPMT*2, TPMT*3A, TPMT*3B or TPMT*3C allele in several populations [16–18]. A functional study showed that these variants enhanced proteolysis of TPMT and resulted in lower catalytic activity [19]. As a consequence, when patients carrying these variants receive standard dosage of thiopurine, cytotoxic molecules will accumulate, and it will subsequently lead to ADRs, such as myelosuppression and fatal bone-marrow toxicity [20–22].

A TPMT genotyping test has been shown to be highly specific and sensitive in ADR prevention. In 1214 German Caucasian patients, performance of genotype tests for the variant TPMT*2 and TPMT*3 alleles (TPMT*3A, TPMT*3B, TPMT*3C and TPMT*3D) achieved a sensitivity of 90% and a specificity of 99% in preventing ADRs [16]. With the observed high sensitivity and specificity, TPMT genotype testing has become commercially available. In 2004, the US FDA approved a TPMT genotype test and revised the drug labelling to recommend a prior TPMT genotype test for thiopurine [7]. The Clinical Pharmacogenetics Implementation Consortium (CPIC) released the guideline for the dose adjustment of thiopurines [23]. In the UK, the standard treatment protocol for ALL (UKALL-2003) includes the pharmacogenomic test for TPMT [24]. Patients with homozygous nonfunctional alleles of TPMT are required to reduce the standard dose by 90%, while a reduction of 30-70% of the standard dose is required for a heterozygous variant of TPMT (with one functional allele, *1) [23, 25]. A prospective study on the TPMT genotype test showed that the dose-adjustment strategy can reduce drug toxicity without compromising efficacy [26]. The cost-effectiveness analysis in four European countries (Germany, Ireland, The Netherlands and the UK) showed that calculated cost per life-year gained by TPMT genotyping in ALL patients was €2100 (~£1820) [27]. This cost is much lower than £30 000 (~€34 700) per qualityadjusted life-year gained, which is the threshold value set up by the UK National Institute for Clinical Excellence (NICE) to indicate a cost-effective outcome [28]. Therefore, pretreatment TPMT genotype testing is a cost-effective approach in clinical practice.

Warfarin, CYP2C9 and VKORC1 Warfarin is the most frequently used medication in the long-term management of anticoagulation therapy in children. Individual dose requirements for warfarin are highly variable and subject to repeated dose adjustments. Age, body size, organ

dysfunction and diet are the main factors that affect the efficacy of warfarin treatment in adults and children [29–33]. Genetic factors that may also influence warfarin dosing have not been investigated thoroughly in children until recently.

A UK study showed that polymorphisms of *CYP2C9* (*2 and *3 alleles) and *VKORC1* (–1639G>A) were significantly associated with required dose adjustments of warfarin in children [31]. The mean warfarin daily dose requirement in children with the *VKORC1* GG genotype was significantly higher than for those with GA or AA genotype. *In vitro* and *ex vivo* studies showed that *CYP2C9*2* and *CYP2C9*3* cause a reduction of warfarin metabolism by 30–40 and 80–90%, respectively, indicating a lower dose requirement for children [34].

In 2007, the US FDA revised the label for warfarin with information pertaining to new pharmacogenomic data. An International Warfarin Pharmacogenetics Consortium algorithm was developed in 2011, and use of this algorithm was suggested to give a better prediction of the appropriate dose of warfarin than the other clinical algorithms or use of a fixed-dose approach [34]. However, this algorithm is based on data obtained from adults and may overestimate the dose for children [31]. Concerning the variability of response to warfarin, guidelines specific for paediatric patients would be preferable.

Examples more applicable to children

Psychotropic drugs and CYP2D6 Cytochrome P450 (CYP) is a well-known superfamily of drug-metabolizing enzymes involved in the metabolism of ~80% of drugs [35]. Cytochrome P450 2D6 is responsible for metabolizing many antipsychotic and antidepressant drugs. Polymorphisms of CYP2D6 have functional significance in drug metabolism and can be classified into the following four categories/drug-metabolizing phenotypes: poor metabolizer (PM); intermediate metabolizer (IM); extensive metabolizer (EM); and ultra-rapid metabolizer (UM) [36].

Atomoxetine, commonly used for treating attentiondeficit/hyperactivity disorder (ADHD) in children, is metabolized by CYP2D6. Pharmacogenomic data have shown that CYP2D6 PM results in 10-fold greater area under the concentration vs. time curve of atomoxetine and higher than normal activity of CYP2D6, which poses risk of ADRs [37]. In US FDA-approved drug labelling, dose adjustment is recommended for CYP2D6 PM, which should be initiated at low dose and increased to the usual target dose if no improvement is observed and the initial dose is well tolerated [38]. A laboratory test for CYP2D6 genotype is currently available. However, due to the lack of clinical utility and clear clinical guideline, a genotyping test is not routinely used by clinicians prior to the treatment of children with ADHD with atomoxetine. Details of limitations will be discussed the section entitled 'To what extent can we practise individualized medicine in children?'.

Cisplatin, TPMT, catechol-O-methyltransferase and ATP-binding cassette transporter C3 Cisplatin is one of the most effective chemotherapeutic agents for treatment of solid tumours. However, the use of cisplatin is limited by the high incidence of ototoxicity in children [39, 40]. This complication causes irreversible, bilateral hearing loss, which seriously hinders language and cognitive development in children [41].

Association studies in children with cisplatin-induced ototoxicity showed that genetic variants in *TPMT* (rs12201199) and catechol-*O*-methyltransferase (*COMT*; rs9332377) are associated with hearing loss [42]. The result was replicated by the same research group using an independent cohort of paediatric patients, and one more genetic variant, in ATP-binding cassette transporter C3 (*ABCC3*; rs1051640), was identified to be associated with hearing loss [43].

In clinical practice, the risk of cisplatin-induced ototoxicity is evaluated by clinical factors such as age, germ-cell tumour, cranial irradiation and vincristine treatment. A predictive model combining the variants of *TPMT*, *COMT* and *ABCC3* with clinical factors showed a significant improvement in predicting the risk of ototoxicity than using clinical risk factors alone [43]. This result illustrates the potential value of incorporating pharmacogenomics into clinical practice.

However, the association between *TPMT*, *COMT* and cisplatin-induced ototoxicity has been questioned because the result could not be replicated in another study and because the laboratory models did not support this association [44]. Therefore, more study should be done to validate the association.

In addition to the aforementioned examples, pharmacogenomic markers for drug efficacy and safety have also been identified for several paediatric antiepileptic drugs and immunotherapy drugs, such as abacavir, clobazam and phenytoin. The pharmacogenomics of these drugs have been discussed in previous review articles [45, 46]. Other common paediatric drugs with available pharmacogenetic tests are shown in Table 1.

Currently, there are only a few paediatric pharmacogenomic tests that are commercially available and used in clinical practice. However, with continuous support from national charity groups, healthcare support groups and regulatory agencies, it is expected that more paediatric pharmacogenomic tests will become available in the near future. In 2005, the US FDA released guidance to the pharmaceutical industry voluntarily to submit pharmacogenomic data for licensing. This guidance is intended to facilitate the use of pharmacogenomic data in drug development. In 2006, the European Medicines Agency (EMA) and the US FDA agreed to have a joint working initiative with respect to the data submission procedure. These efforts have raised awareness of the importance of pharmacogenomic study in drug development in terms of improving efficacy and reducing ADRs. Since

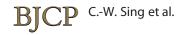


Table 1Selected drugs with available pharmacogenomic tests for children*

Drug	Gene	Therapeutic class or indication	Genetic variant	Comment on genotype	Action recommended by US FDA-approved drug label
Abacavir	HLA-B	Human immunodeficiency virus therapy	HLA-B*5701	Increased risk of hypersensitivity	Abacavir is not recommended for people carrying HLA-B*5701 allele
Atomoxetine	CYP2D6	Attention-deficit/ hyperactivity disorder	CYP2D6*3, CYP2D6*4, CYP2D6*5, CYP2D6*6	Alert to adverse drug effects in poor metabolizer	Start at 0.5 mg kg ⁻¹ day ⁻¹ and only increase to the usual target dose of 1.2 mg kg ⁻¹ day ⁻¹ if symptoms fail to improve after 4 weeks and the initial dose is well tolerated
Cisplatin	TPMT	Cancer	TPMT*3B, TPMT*3C	Increased risk of ototoxicity	Audiometric monitoring should be performed prior to initiation of therapy, prior to each subsequent dose and for several years post-therapy
Clobazam	CYP2C19	Anticonvulsant	CYP2C9*2	Higher concentration of the active metabolite of clobazam	Initial dose should be 5 mg day ⁻¹ and titrated initially to 10–20 mg day ⁻¹ , and may be titrated further to a maximal daily dose of 40 mg if tolerated
Phenytoin	HLA-B	Anticonvulsant	HLA-B*1502	Increased risk of Stevens–Johnson Syndrome and toxic epidermal necrolysis	Consideration should be given to avoiding phenytoin as an alternative for carbamazepine in patients positive for <i>HLA-B*1502</i>
Thioprines (azathioprine, mercaptourine, thioguanine)	TPMT	Acute lymphatic leukaemia	TPMT*2, TPMT*3A, TPMT*3C	Increased risk of myelosuppression	Substantial dosage reduction
Valproic acid	POLG	Anticonvulsant	POLG mutation	Increased risk of liver failure and death in patients with hereditary neurometabolic syndromes	POLG mutation testing should be performed in accordance with current clinical practice
Warfarin	VKORC1, CYP2C9	Anticoagulant	VKORC1 (G-1639A), CYP2C9*2, CYP2C9*3	Increased risk of bleeding	A range of doses based on VKORC1 and CYP2C9 genotypes

^{*}Drugs are selected from the British National Formulary for Children [66] and are listed in the table of the US Food and Drug Administration (US FDA) Pharmacogenomic Biomarkers in Drug Labeling [7].

then, additional clinical trials have incorporated pharmacogenomic tests in phases 2 and 3, and it is expected that pharmacogenomic data will be more commonly listed on drug labels.

To what extent can we practise individualized medicine in children?

A decade after completion of the Human Genome Project, individualized medicine is still in its infancy. Pharmacogenomic tests for children receiving medication are limited, and the progress of translating/incorporating pharmacogenomic data into clinical practice is unexpectedly slow. This poses a question: what obstacles slow down this progress of incorporation?

Gene expression and ontogeny in children

The gene expression profile and ontogeny in children is the biggest challenge for pharmacogenomic study. Most of the pharmacogenomic research identifies variation in the genetic sequence (e.g. single nucleotide polymorphism) as the marker to predict drug response. However, the level of gene expression can vary depending on changes in the environment, such as different developmental stages. The development of children can be subdivided into the stages of infancy, childhood and adolescence. During the developmental process, major organ systems involved in drug biotransformation are developing to reach maturity and, likewise, gene expression, particularly with respect to the drug-metabolizing enzymes, undergoes considerable changes throughout the developmental process [47].

In the study of ontogeny of drug-metabolizing enzymes, variable expression profiles of the enzymes can be observed. While some enzymes are expressed at the highest level in the fetus and become silenced (e.g. flavincontaining monooxygenase 1) or reduced to relatively low levels (e.g. CYP3A7) within a few days to 2 years after birth [48], in contrast, some enzymes (e.g. CYP2D6) are expressed at negligible or low levels in the fetus but increase significantly to high levels within a few weeks to 1 or 2 years after birth [49]. The change of enzyme activity across developmental stages greatly affects the drug response and tolerance in children.

When a genotype–phenotype relationship is identified in children, researchers have to consider the effect of developmental factors. However, the ontogeny of drug biotransformation in children remains unclear, which limits the predictive capacity of pharmacogenomic data for drug efficacy and safety.

Lack of pharmacogenomic information for use in paediatrics

Compared with adults, pharmacogenomic studies in children are scarce, hence pharmacogenomic information for paediatric use on drug labels is limited. Although numerous pharmacogenomic studies have been published, most have been performed in adults. Results from adult may not be applicable to children owing to differences in developmental stage, as discussed in the previous subsection. For example, the kidney and liver are the major organs responsible for drug metabolism and clearance/elimination. Developmental changes in these two organs, such as overall size, which will influence the total mass of drugmetabolizing cells and, in the case of the kidney, the glomerular function, will significantly influence drug metabolism in children. This is often markedly different from that observed in adults [50]. One example is the clearance of voriconazole, an antifungal agent, is ~3-fold higher in children than in adults [51].

With this limitation, paediatricians often remain sceptical and uncertain as how to use data derived from adult studies and apply them to clinical practice in children. In 2009, a survey study performed by The American Society of Pediatric Hematology/Oncology showed that most US paediatric haematologists are familiar with pharmacogenomic data and their application to the use in warfarin treatment. However, they refused to perform the genotype test prior to decision making for drug therapy in children [52]. The clinicians argued that there is a lack of data pertinent to the US paediatric population and most studies performed are retrospective in nature, with a small sample size. A prospective randomized clinical trial on the clinical effectiveness of pharmacogenomic tests in warfarin therapy has been conducted, but children were not included in the trial [53]. They urged a large prospective trial in children to provide a stronger evidence base for the current pharmacogenomic data.

Ethical concerns in paediatrics

The ethical concern of individualized medicine is of great importance, especially in children, and this also partly explains why only a limited number of pharmacogenomic studies have been conducted in children. A recent review discussed the ethical considerations for pharmacogenomic research in the paediatric population and its implementation in the care of children [54]. In paediatric research, acquisition of informed consent is one of the greatest challenges, especially in pharmacogenomic studies. Full disclosure of information is difficult, because

the potential implications of pharmacogenomic data might be unknown at the time of the study. Given this uncertainty, risk-benefit assessment might not always be complete for children and their parents to make an informed decision on their participation in the research trial process.

In clinical practice, genetic discrimination and privacy are major concerns. In addition, pharmacogenomic testing may provide ancillary disease risk information, which might affect the ability of patients to obtain secure life insurance/protection [55]. The psychological burden to both children and parents should also be considered, because ancillary disease risk information implies the potential development of one or more diseases. In addition, privacy is also a concern in pharmacogenomic testing. A survey study conducted in 2007 showed that 66% of subjects had a fear of inadequate protection of their personal data if they were to undergo a pharmacogenomic test [56]. The survey did not involve children, but fear from their parents would probably result in a reluctance to allow any pharmacogenomic test to be performed on their child.

Lack of clinical utility and cost effectiveness

The key for translating pharmacogenomics into clinical practice is clinical utility and cost effectiveness. Currently, only a few pharmacogenomic tests are used routinely by clinicians, because the clinical utility is still being guestioned. Atomoxetine is one of the examples. As discussed in the section of 'Examples more applicable to children', patients with CYP2D6 variants have increased risk of adverse events, and dose adjustment is recommended by the US FDA. Regardless of CYP2D6 phenotype, the normal dose of atomoxetine should be initiated at 0.5 mg kg⁻¹ day⁻¹ and increased after a minimum of 3 days to a target total daily dose of 1.2 mg kg⁻¹. If the patient is CYP2D6 PM, the dose is recommended to be initiated at 0.5 mg kg⁻¹ day⁻¹ and only increased to the usual target dose of 1.2 mg kg⁻¹ day⁻¹ if symptoms fail to improve after 4 weeks and the initial dose is well tolerated [38]. In clinical management, a prescription starting with a low dose and titrating slowly is a basic practice and thus such recommendations seem not to be meaningful. Without knowing the CYP2D6 status, clinicians are still able to dose atomoxetine to a safe and efficient level according to clinical assessment tools [57].

The prevalence of genetic variants among different ethnic populations also limits the clinical utility of some pharmacogenomics tests. For example, CYP2D6 PM are found in 7% of Caucasians but only 1% of Chinese populations [58]. In contrast, *HLA-B*1502* is commonly found in Chinese and other Asian populations but is almost absent in Caucasian and African-American populations. As a result, clinicians would be doubtful of using pharmacogenomic information based on other ethnic populations.

Apart from clinical utility, clinicians also consider the cost effectiveness of pharmacogenomic tests. Advances in genotyping technology will eventually reduce the cost of genotyping; however, performing routine pharmacogenomic testing in patients may still be expensive for individuals and the parent healthcare system. Therefore, both governments and clinicians are concerned about whether the cost of pharmacogenomic tests is justified by the clinical outcomes after performing them. However, there are only a few studies that demonstrate the cost effectiveness of performing pharmacogenomic tests, or show that the test is only cost effective in certain patient groups, such as with the case of testing prior to warfarin therapy [59]. Guidelines exist for economic evaluation of pharmaceutical products in the healthcare system, but they are rarely applied in pharmacogenomic study.

What should be done to facilitate the implementation of pharmacogenomics into clinical practice?

To overcome the challenges and facilitate the implementation of pharmacogenomics into clinical practice in paediatrics, different approaches can be worked on, and examples are given below. These suggestions are not limited to paediatric study and are applicable to all pharmacogenomic research.

Pharmacogenomics network for paediatric study

Drug metabolism in children is complex owing to differences in gene expression during physiological development. Therefore, in order to gain a better understanding of drug metabolism at the different developmental stages, adequate pharmacogenomic studies remain a prerequisite for improving paediatric individualized medicine. If a drug is required to be studied in an agegroup-specific manner, we can envisage that subject recruitment will be a major challenge. In order to have sufficient power to detect any genetic effect/difference, nationwide pharmacogenomic networks for paediatric study and international collaboration are encouraged and essential to allow the collection of a representative sample size and to allow cross-comparisons of the results.

There are pharmacogenomic groups worldwide, but projects specific to paediatrics are limited. The Canadian Pharmacogenomics Network for Drug Safety (CPNDS) is one of the few to have started a nationwide project on paediatric pharmacogenomics, namely the Genetic Approach to Therapy in Children (GATC) [60]. This project establishes a nationwide network for of surveillance ADRs

in children and collects their biological samples for genotyping. They target three ADRs commonly found in children, i.e. cisplatin-induced hearing loss, anthracyclineinduced cardiotoxicity and codeine-induced infant mortality. Thousands of ADR cases in children have been recorded, and pharmacogenetic markers have been successfully identified [42, 61]. The GATC may provide a framework of active pharmacogenomic study in children that can be adopted by other countries. In the USA, there is another paediatric pharmacogenomic project, termed 'PAAR4Kids: Pharmacogenomics of Anticancer Agents Research in Children', launched by National Institutes of Health's Pharmacogenomics Research Network (PGRN). This project studies only the pharmacogenomics of children with ALL and should be extended to other disciplines as well [62].

Guidelines for pharmacogenomics studies in children

Guidelines for conducting good pharmacogenomic studies in children should be developed. These guidelines could provide a framework for future investigators to be able to plan, conduct and evaluate pharmacogenomic analysis in paediatric studies. Several areas should be particularly addressed within these suggested guidelines, namely protocol development, ethical considerations in children, and the documentation of both research methods and results. With such guidelines, the quality and integrity of pharmacogenomic research can be maintained and, consequently, the results of such studies would be more robust.

Pharmacoeconomics in pharmacogenomics

The incorporation of pharmacoeconomic data into pharmacogenomic research will, it is expected, be the catalyst to full implementation of individualized medicine in modern society. Pharmacogenomic tests are not only beneficial to patients, but if they are also shown to be cost effective and to have positive cost-benefit, then their acceptance by physicians will be enhanced. Likewise, the willingness of individual governments/ healthcare sponsors and private insurers to adopt and support pharmacogenomic technologies financially would be encouraged. There are few cost-benefit analyses of pharmaocogenomic studies, but they have adopted different methodologies and, consequently, the consistency of results is poor. A standardized pharmacoeconomic model for pharmacogenomics is favourable, but health economists are uncertain whether existing pharmacoeconomic models are sufficient to evaluate pharmacogenomic tests. Therefore, developing models for pharmcoeconomic analysis in pharmacogenomics should be one of the main focuses in future work on translational pharmacogenomics.



Integration programme for translational pharmacogenomics

In order for clinicians to integrate pharmacogenomics into clinical practice, they need support from various areas, including clinical guidelines, education and computational tools. An integration programme that works on these areas can greatly facilitate the progress of pharmacogenomics in clinical practice, including paediatrics.

An example is the translational pharmcogenomic programme led by St Jude Children's Research Hospital (St Jude). PG4KDS, a project launched by St Jude, aims to evaluate pharmacogenomic test results in children and selectively choose the tests that have strong evidence and closely link to drug response to incorporate into patient medical records. Clinicians will be alerted to any pharmacogenomic information about the patient when prescribing treatment [63]. In addition, St Jude is also developing protocols and computational tools to help clinicians to make prescription decisions based on the pharmacogenomic information [64]. To increase the use of pharmacogenomics, clinicians and pharmacists need to play an active role in decision making on the choice of pharmacogenomic tests and developing clinical protocols so that they have a better understanding of the pharmacogenomic tests and will be better able to use them. In addition, the project works on education by making videos and leaflets for parents and patients, as well as healthcare professionals. These materials are easily accessible in hospitals and online [65].

Conclusion

To date, full implementation of pharmacogenomics into clinical practice, and thus individualized medicine in children, is inadequate. There is a need to have more consortia that focus on paediatric pharmacogenomics and to encourage the reduction in the cost of genotyping in order to understand the ontogeny and gene expression in children. It can be foreseen that individualized medicine in children will be the future of paediatric medicine.

Competing Interests

All authors have completed the Unified Competing Interest form at http://www.icmje.org/coi_disclosure.pdf (available on request from the corresponding author) and declare: ICKW was partly supported by the European Union Seventh Framework Programme FP7/2007–2013 under grant agreement no. 261060, the 'Global Research in Paediatrics Network of Excellence (GRiP)'; no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; no other relationships or activities that could appear to have influenced the submitted work.

REFERENCES

- **1** Marshall JD, Kearns GL. Developmental pharmacodynamics of cyclosporine. Clin Pharmacol Ther 1999; 66: 66–75.
- **2** Biss TT, Kamali F. Warfarin anticoagulation in children: is there a role for a personalized approach to dosing? Pharmacogenomics 2012; 13: 1211–4.
- **3** Kling J. US FDA contemplates collection of pharmacogenomic data. Nat Biotechnol 2003; 21: 590.
- **4** Kalow W, Tang BK, Endrenyi L. Hypothesis: comparisons of inter- and intra-individual variations can substitute for twin studies in drug research. Pharmacogenetics 1998; 8: 283–9.
- **5** Collins FS, Morgan M, Patrinos A. The Human Genome Project: lessons from large-scale biology. Science 2003; 300: 286–90.
- **6** The International HapMap Consortium. The International HapMap Project. Nature 2003; 426: 789–96.
- 7 U.S. FDA. Table of Pharmacogenomic Biomarkers in Drug Labels. US: USFDA, 2013. Available at http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm (updated 06/19/2013; last accessed 1 July 2013).
- **8** Chung WH, Hung SI, Hong HS, Hsih MS, Yang LC, Ho HC, Wu JY, Chen YT. Medical genetics: a marker for Stevens-Johnson syndrome. Nature 2004; 428: 486.
- **9** Man CB, Kwan P, Baum L, Yu E, Lau KM, Cheng AS, Ng MH. Association between HLA-B*1502 allele and antiepileptic drug-induced cutaneous reactions in Han Chinese. Epilepsia 2007: 48: 1015–8.
- 10 Wang Q, Zhou JQ, Zhou LM, Chen ZY, Fang ZY, Chen SD, Yang LB, Cai XD, Dai QL, Hong H, Wang HX. Association between HLA-B*1502 allele and carbamazepine-induced severe cutaneous adverse reactions in Han people of southern China mainland. Seizure 2011; 20: 446–8.
- 11 Locharernkul C, Loplumlert J, Limotai C, Korkij W, Desudchit T, Tongkobpetch S, Kangwanshiratada O, Hirankarn N, Suphapeetiporn K, Shotelersuk V. Carbamazepine and phenytoin induced Stevens-Johnson syndrome is associated with HLA-B*1502 allele in Thai population. Epilepsia 2008; 49: 2087–91.
- **12** Chang CC, Too CL, Murad S, Association HSH. of HLA-B*1502 allele with carbamazepine-induced toxic epidermal necrolysis and Stevens-Johnson syndrome in the multi-ethnic Malaysian population. Int J Dermatol 2011; 50: 221–4.
- 13 McCormack M, Alfirevic A, Bourgeois S, Farrell JJ, Kasperaviciute D, Carrington M, Sills GJ, Marson T, Jia X, de Bakker Pl, Chinthapalli K, Molokhia M, Johnson MR, O'Connor GD, Chaila E, Alhusaini S, Shianna KV, Radtke RA, Heinzen EL, Walley N, Pandolfo M, Pichler W, Park BK, Depondt C, Sisodiya SM, Goldstein DB, Deloukas P, Delanty N, Cavalleri GL, Pirmohamed M. HLA-A*3101 and carbamazepine-induced hypersensitivity reactions in Europeans. N Engl J Med 2011; 364: 1134–43.

- 14 U.S. Food and Drug Administration. Information for Healthcare Professionals: Dangerous Or Even Fatal Skin Reactions – Carbamazepine (Marketed As Carbatrol, Equetro, Tegretol, and Generics). [Internet] Rockville, MD: U.S. Food and Drug Administration, 2007. Available at http://www.fda.gov/Drugs/DrugSafety/PostmarketDrug SafetyInformationforPatientsandProviders/ucm124718.htm (updated 2013 Aug 14; last accessed 10 November 2013).
- 15 Roujeau JC, Kelly JP, Naldi L, Rzany B, Stern RS, Anderson T, Auquier A, Bastuji-Garin S, Correia O, Locati F, Mockenhaupt M, Paoletti C, Shapiro S, Shear N, Schöpf E, Kaufman DW. Medication use and the risk of Stevens-Johnson syndrome or toxic epidermal necrolysis. N Engl J Med 1995; 333: 1600–7.
- **16** Schaeffeler E, Fischer C, Brockmeier D, Wernet D, Moerike K, Eichelbaum M, Zanger UM, Schwab M. Comprehensive analysis of thiopurine S-methyltransferase phenotype-genotype correlation in a large population of German-Caucasians and identification of novel TPMT variants. Pharmacogenetics 2004; 14: 407–17.
- 17 Yates CR, Krynetski EY, Loennechen T, Fessing MY, Tai HL, Pui CH, Relling MV, Evans WE. Molecular diagnosis of thiopurine S-methyltransferase deficiency: genetic basis for azathioprine and mercaptopurine intolerance. Ann Intern Med 1997; 126: 608–14.
- 18 Hon YY, Fessing MY, Pui CH, Relling MV, Krynetski EY, Evans WE. Polymorphism of the thiopurine S-methyltransferase gene in African-Americans. Hum Mol Genet 1999; 8: 371–6.
- **19** Tai HL, Fessing MY, Bonten EJ, Yanishevsky Y, D'Azzo A, Krynetski EY, Evans WE. Enhanced proteasomal degradation of mutant human thiopurine S-methyltransferase (TPMT) in mammalian cells: mechanism for TPMT protein deficiency inherited by TPMT*2, TPMT*3A, TPMT*3B or TPMT*3C. Pharmacogenetics 1999; 9: 641–50.
- **20** Schutz E, Gummert J, Mohr F, Oellerich M. Azathioprine-induced myelosuppression in thiopurine methyltransferase deficient heart transplant recipient. Lancet 1993; 341: 436.
- **21** Wall AM, Rubnitz JE. Pharmacogenomic effects on therapy for acute lymphoblastic leukemia in children. Pharmacogenomics J 2003; 3: 128–35.
- 22 Adam de Beaumais T, Fakhoury M, Medard Y, Azougagh S, Zhang D, Yakouben K, Jacqz-Aigrain E. Determinants of mercaptopurine toxicity in paediatric acute lymphoblastic leukemia maintenance therapy. Br J Clin Pharmacol 2011; 71: 575–84.
- 23 Relling MV, Gardner EE, Sandborn WJ, Schmiegelow K, Pui CH, Yee SW, Stein CM, Carrillo M, Evans WE, Hicks JK, Schwab M, Klein TE. Clinical pharmacogenetics implementation consortium guidelines for thiopurine methyltransferase genotype and thiopurine dosing: 2013 update. Clin Pharmacol Ther 2013; 93: 324–5.
- 24 MRC. UKALL 2003 version 7. 2003. Available at www.ctsu.ox .ac.uk/research/mega-trials/leukaemia-trials/ukall-2003/protocol-version-7 (updated 08/2009; last accessed 1 July 2013).
- 25 Swen JJ, Nijenhuis M, de Boer A, Grandia L, Maitland-van der Zee AH, Mulder H, Rongen GA, van Schaik

- RH, Schalekamp T, Touw DJ, van der Weide J, Wilffert B, Deneer VH, Guchelaar HJ. Pharmacogenetics: from bench to byte an update of guidelines. Clin Pharmacol Ther 2011; 89: 662–73.
- **26** Relling MV, Pui CH, Cheng C, Evans WE. Thiopurine methyltransferase in acute lymphoblastic leukemia. Blood 2006; 107: 843–4.
- 27 van den Akker-van Marle ME, Gurwitz D, Detmar SB, Enzing CM, Hopkins MM, Gutierrez de Mesa E, Ibarreta D. Cost-effectiveness of pharmacogenomics in clinical practice: a case study of thiopurine methyltransferase genotyping in acute lymphoblastic leukemia in Europe. Pharmacogenomics 2006; 7: 783–92.
- **28** McCabe C, Claxton K, Culyer AJ. The NICE cost-effectiveness threshold: what it is and what that means. Pharmacoeconomics 2008: 26: 733–44.
- 29 Streif W, Andrew M, Marzinotto V, Massicotte P, Chan AK, Julian JA, Mitchell L. Analysis of warfarin therapy in pediatric patients: a prospective cohort study of 319 patients. Blood 1999; 94: 3007–14.
- **30** Aithal GP, Day CP, Kesteven PJ, Daly AK. Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. Lancet 1999; 353: 717–9.
- **31** Biss TT, Avery PJ, Brandao LR, Chalmers EA, Williams MD, Grainger JD, Leathart JB, Hanley JP, Daly AK, Kamali F. VKORC1 and CYP2C9 genotype and patient characteristics explain a large proportion of the variability in warfarin dose requirement among children. Blood 2012; 119: 868–73.
- **32** Rogers HL, Bhattaram A, Zineh I, Gobburu J, Mathis M, Laughren TP, Pacanowski M. CYP2D6 genotype information to guide pimozide treatment in adult and pediatric patients: basis for the U.S. Food and Drug Administration's new dosing recommendations. J Clin Psychiatry 2012; 73: 1187–90.
- **33** Khan T, Wynne H, Wood P, Torrance A, Hankey C, Avery P, Kesteven P, Kamali F. Dietary vitamin K influences intra-individual variability in anticoagulant response to warfarin. Br J Haematol 2004; 124: 348–54.
- **34** Johnson JA, Gong L, Whirl-Carrillo M, Gage BF, Scott SA, Stein CM, Anderson JL, Kimmel SE, Lee MT, Pirmohamed M, Wadelius M, Klein TE, Altman RB. Clinical Pharmacogenetics Implementation Consortium Guidelines for CYP2C9 and VKORC1 genotypes and warfarin dosing. Clin Pharmacol Ther 2011; 90: 625–9.
- **35** Wilkinson GR. Drug metabolism and variability among patients in drug response. N Engl J Med 2005; 352: 2211–21.
- **36** Kirchheiner J, Nickchen K, Bauer M, Wong ML, Licinio J, Roots I, Brockmoller J. Pharmacogenetics of antidepressants and antipsychotics: the contribution of allelic variations to the phenotype of drug response. Mol Psychiatry 2004; 9: 442–73.
- **37** Michelson D, Read HA, Ruff DD, Witcher J, Zhang S, McCracken J. CYP2D6 and clinical response to atomoxetine in children and adolescents with ADHD. J Am Acad Child Adolesc Psychiatry 2007; 46: 242–51.

- **38** STRATTERA. 2013. Label Information: STRATTERA. US FDA. Available at http://www.accessdata.fda.gov/drugsatfda_docs/label/2014/021411s044lbl.pdf (last accessed 4 December 2013).
- **39** Li Y, Womer RB, Silber JH. Predicting cisplatin ototoxicity in children: the influence of age and the cumulative dose. Eur J Cancer 2004; 40: 2445–51.
- 40 Brock PR, Bellman SC, Yeomans EC, Pinkerton CR, Pritchard J. Cisplatin ototoxicity in children: a practical grading system. Med Pediatr Oncol 1991; 19: 295–300.
- **41** Knight KR, Kraemer DF, Neuwelt EA. Ototoxicity in children receiving platinum chemotherapy: underestimating a commonly occurring toxicity that may influence academic and social development. J Clin Oncol 2005; 23: 8588–96.
- 42 Ross CJD, Katzov-Eckert H, Dube M-P, Brooks B, Rassekh SR, Barhdadi A, Feroz-Zada Y, Visscher H, Brown AMK, Rieder MJ, Rogers PC, Phillips MS, Carleton BC, Hayden MR. Genetic variants in TPMT and COMT are associated with hearing loss in children receiving cisplatin chemotherapy. Nat Genet 2009; 41: 1345–9.
- 43 Pussegoda K, Ross CJ, Visscher H, Yazdanpanah M, Brooks B, Rassekh SR, Zada YF, Dube MP, Carleton BC, Hayden MR. Replication of TPMT and ABCC3 genetic variants highly associated with cisplatin-induced hearing loss in children. Clin Pharmacol Ther 2013; 94: 243–51.
- **44** Yang JJ, Lim JY, Huang J, Bass J, Wu J, Wang C, Fang J, Stewart E, Harstead EH, E S, Robinson GW, Evans WE, Pappo A, Zuo J, Relling MV, Onar-Thomas A, Gajjar A, Stewart CF. The role of inherited TPMT and COMT genetic variation in cisplatin-induced ototoxicity in children with cancer. Clin Pharmacol Ther 2013; 94: 252–9.
- **45** Lai-Goldman M, Faruki H. Abacavir hypersensitivity: a model system for pharmacogenetic test adoption. Genet Med 2008; 10: 874–8.
- **46** Seo T, Nagata R, Ishitsu T, Murata T, Takaishi C, Hori M, Nakagawa K. Impact of CYP2C19 polymorphisms on the efficacy of clobazam therapy. Pharmacogenomics 2008; 9: 527–37.
- **47** Kearns GL, Abdel-Rahman SM, Alander SW, Blowey DL, Leeder JS, Kauffman RE. Developmental pharmacology–drug disposition, action, and therapy in infants and children. N Engl J Med 2003; 349: 1157–67.
- **48** Hines RN. The ontogeny of drug metabolism enzymes and implications for adverse drug events. Pharmacol Ther 2008; 118: 250–67.
- **49** Stevens JC, Marsh SA, Zaya MJ, Regina KJ, Divakaran K, Le M, Hines RN. Developmental changes in human liver CYP2D6 expression. Drug Metab Dispos 2008; 36: 1587–93.
- 50 Chen N, Aleksa K, Woodland C, Rieder M, Koren G. Ontogeny of drug elimination by the human kidney. Pediatr Nephrol 2006; 21: 160–8.
- **51** Yanni SB, Annaert PP, Augustijns P, Ibrahim JG, Benjamin DK, Jr, Thakker DR. In vitro hepatic metabolism explains higher clearance of voriconazole in children versus adults: role of CYP2C19 and flavin-containing monooxygenase 3. Drug Metab Dispos 2010; 38: 25–31.

- **52** Thornburg CD, Jones E, Bomgaars L, Gage BF. Pediatric warfarin practice and pharmacogenetic testing. Thromb Res 2010; 126: e144–6.
- 53 Anderson JL, Horne BD, Stevens SM, Woller SC, Samuelson KM, Mansfield JW, Robinson M, Barton S, Brunisholz K, Mower CP, Huntinghouse JA, Rollo JS, Siler D, Bair TL, Knight S, Muhlestein JB, Carlquist JF. A randomized and clinical effectiveness trial comparing two pharmacogenetic algorithms and standard care for individualizing warfarin dosing (CoumaGen-II). Circulation 2012; 125: 1997–2005.
- **54** Moran C, Thornburg CD, Barfield RC. Ethical considerations for pharmacogenomic testing in pediatric clinical care and research. Pharmacogenomics 2011; 12: 889–95.
- **55** Henrikson NB, Burke W, Veenstra DL. Ancillary risk information and pharmacogenetic tests: social and policy implications. Pharmacogenomics J 2008; 8: 85–9.
- **56** Voora D, Gage BF. Is primary care ready for pharmacogenetics? Pharmacogenomics 2006; 7: 1–3.
- **57** Trzepacz PT, Williams DW, Feldman PD, Wrishko RE, Witcher JW, Buitelaar JK. CYP2D6 metabolizer status and atomoxetine dosing in children and adolescents with ADHD. Eur Neuropsychopharmacol 2008; 18: 79–86.
- **58** Bertilsson L, Dahl ML, Dalen P, Al-Shurbaji A. Molecular genetics of CYP2D6: clinical relevance with focus on psychotropic drugs. Br J Clin Pharmacol 2002; 53: 111–22.
- **59** Eckman MH, Rosand J, Greenberg SM, Gage BF. Cost-effectiveness of using pharmacogenetic information in warfarin dosing for patients with nonvalvular atrial fibrillation. Ann Intern Med 2009; 150: 73–83.
- **60** Ross CJ, Visscher H, Sistonen J, Brunham LR, Pussegoda K, Loo TT, Rieder MJ, Koren G, Carleton BC, Hayden MR. null. The Canadian Pharmacogenomics Network for Drug Safety: a model for safety pharmacology. Thyroid 2010; 20: 681–7.
- **61** Visscher H, Ross CJ, Rassekh SR, Barhdadi A, Dube MP, Al-Saloos H, Sandor GS, Caron HN, van Dalen EC, Kremer LC, van der Pal HJ, Brown AM, Rogers PC, Phillips MS, Rieder MJ, Carleton BC, Hayden MR, Canadian Pharmacogenomics Network for Drug Safety Consortium. Pharmacogenomic prediction of anthracycline-induced cardiotoxicity in children. J Clin Oncol 2012; 30: 1422–8.
- **62** PGRN. 2010. PAAR4Kids: pharmacogenomics of anticancer agents research in children. Available at http://pharmacogenetics.org/paar4kids/ (last accessed 9 November 2013).
- 63 Crews KR, Cross SJ, McCormick JN, Baker DK, Molinelli AR, Mullins R, Relling MV, Hoffman JM. Development and implementation of a pharmacist-managed clinical pharmacogenetics service. Am J Health Syst Pharm 2011; 68: 143–50.
- 64 Hicks JK, Crews KR, Hoffman JM, Kornegay NM, Wilkinson MR, Lorier R, Stoddard A, Yang W, Smith C, Fernandez CA, Cross SJ, Haidar C, Baker DK, Howard SC, Evans WE, Broeckel U, Relling MV. A clinician-driven automated system for integration of pharmacogenetic interpretations into an electronic medical record. Clin Pharmacol Ther 2012; 92: 563–6.

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- 65 St. Jude Children's Research Hospital. Non-therapeutic Protocol PG4KDS: Clinical Implementation of Pharmacogenetics. USA: St. Jude Children's Research Hospital. 2013. Available at http://www.stjude.org/stjude/v/index.jsp?vgnextoid=28105138e6bdf210VgnVCM1000001 e0215acRCRD&vgnextchannel=4c80bfe82e118010 VgnVCM100000e2015acRCRD (last accessed 20 January 2014).
- **66** Bristish Medicial Association and the Royal Pharmacetuical Society of Great Britian. BNF for Children 2011–2012. London: BMJ Publishing Group, 2011.