

Research Article

Diarrheagenic *Escherichia coli* Phylogroups Are Associated with Antibiotic Resistance and Duration of Diarrheal Episode

Susan Mosquito,¹ Maria J. Pons,^{2,3} Maribel Riveros,¹
Joaquim Ruiz,² and Theresa J. Ochoa^{1,4}

¹Instituto de Medicina Tropical Alexander von Humboldt, Universidad Peruana Cayetano Heredia, Lima 031, Peru

²ISGlobal, Barcelona Centre for International Health Research (CRESIB), Hospital Clínic, Universitat de Barcelona, 08036 Barcelona, Spain

³Universidad Peruana de Ciencias Aplicadas (UPC), Lima 09, Peru

⁴School of Public Health, University of Texas, Houston, TX 77030, USA

Correspondence should be addressed to Theresa J. Ochoa; theresa.j.ochoa@uth.tmc.edu

Received 28 July 2014; Revised 7 February 2015; Accepted 11 February 2015

Academic Editor: Boris Martinac

Copyright © 2015 Susan Mosquito et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Conventionally, in *Escherichia coli*, phylogenetic groups A and B1 are associated with commensal strains while B2 and D are associated with extraintestinal strains. The aim of this study was to evaluate diarrheagenic (DEC) and commensal *E. coli* phylogeny and its association with antibiotic resistance and clinical characteristics of the diarrheal episode. Phylogenetic groups and antibiotic resistance of 369 *E. coli* strains (commensal strains and DEC from children with or without diarrhea) isolated from Peruvian children <1 year of age were determined by a Clermont triplex PCR and Kirby-Bauer method, respectively. The distribution of the 369 *E. coli* strains among the 4 phylogenetic groups was A (40%), D (31%), B1 (21%), and B2 (8%). DEC-control strains were more associated with group A while DEC-diarrhea strains were more associated with group D ($P < 0.05$). There was a tendency ($P = 0.06$) for higher proportion of persistent diarrhea (≥ 14 days) among severe groups (B2 and D) in comparison with nonsevere groups (A and B1). Strains belonging to group D presented significantly higher percentages of multidrug resistance than the rest of the groups ($P > 0.01$). In summary, DEC-diarrhea strains were more associated with group D than strains from healthy controls.

1. Introduction

Conventionally, *Escherichia coli*, a common isolate in clinical laboratories, is classified into two major groups: commensal and pathogenic. Additionally pathogenic isolates may produce different diseases, being then subdivided in diarrheagenic and extraintestinal *E. coli*. Human infections caused by extraintestinal *E. coli* include meningitis, urinary tract infections, sepsis, pneumonia, surgical site infections, and infections in other extraintestinal locations [1]. However, when classified into subtypes, *E. coli* mainly fall into four phylogenetic groups: A, B1, B2, and D [1]. Previous studies have shown that commensal *E. coli* strains tend to be associated within phylogenetic groups A and B1 [1, 2], whereas the extraintestinal pathotypes fall within phylogenetic groups B2 and D [3, 4]. Regarding uropathogenic *E. coli* (UPEC) strains determinants including phylogroups markers are well

established. However, for diarrheagenic *E. coli* (DEC), the scenario remains unclear. There is no information about the association between the phylogenetic group and the clinical data of the diarrheal episode [5].

Previous reports describe the emerging antibiotic resistance in commensal and diarrheagenic *E. coli* in Peru [6–10]. However, there is no sufficient data in the correlation of phylogeny and antibiotic resistance [11]. Therefore we conducted this study to determine the association between the phylogenetic group and antibiotic resistance in a large number of *E. coli* (commensal and diarrheagenic) strains from Peruvian infants.

2. Materials and Methods

2.1. Samples. Commensal and diarrheagenic *E. coli* strains were isolated during a previous passive surveillance diarrhea

study [12]. In this study 1032 children were followed from 2 to 12 months of life, obtaining a total of 1079 *E. coli* strains that were analyzed by a real time multiplex PCR to determine DEC pathotypes [13]. A total of 369 isolates from this study were randomly selected and analyzed, including 74 commensal *E. coli*, 94 DEC from asymptomatic children (DEC-control), and 201 DEC isolated from children with diarrhea (DEC-diarrhea). DEC pathotypes included in this study were enteropathogenic *E. coli* (EPEC), enterotoxigenic *E. coli* (ETEC), enteroaggregative *E. coli* (EAEC), and diffusely adherent *E. coli* pathotypes (DAEC); Shiga toxin-producing *E. coli* (STEC) and enteroinvasive *E. coli* (EIEC) strains were not included due to their low prevalence [12].

2.2. Phylogenetic Group Determination. The phylogenetic groups were determined as previously described [14]. In all cases the bacteria DNA was extracted boiling.

2.3. Clinical Data of Diarrheal Episodes. Variables such as episode duration (days), maximum number of stools per day, total number of stools per episode, and a modified Vesikari modified score [15] were analyzed and associated with phylogenetic groups in those cases in which no coinfections were previously reported [12].

2.4. Antibiotic Resistance. Antibiotic susceptibility to ampicillin (10 µg), trimethoprim-sulfamethoxazole (23.75/1.25 µg), chloramphenicol (30 µg), nalidixic acid (30 µg), and tetracycline (30 µg) was determined by disk diffusion in accordance with the CLSI guidelines [16]. Multiresistance was defined as resistance to three or more unrelated antibiotic families.

2.5. Statistical Analysis. Vesikari severity score was expressed by mean ± standard deviation and median (range) values were given for duration of the episode and number of stools. The comparisons between groups were made using chi-squared or Fisher's exact test. Student's *t*-test was used for the comparison of Vesikari severity scores between groups.

3. Results

3.1. Phylogenetic Group Frequency. The *E. coli* strains (DEC and commensals) were distributed in the four phylogenetic groups: A (147 isolates, 40%), D (116 isolates, 31%), B1 (76 isolates, 21%), and B2 (30 isolates, 8%). No significant difference in the prevalence of phylogenetic groups was found within each pathotype when analyzed by control/diarrhea. In total were analyzed 87 EPEC (38 DEC-control, 49 DEC-diarrhea), 83 ETEC (26 DEC-control, 57 DEC-diarrhea), 94 EAEC (24 DEC-control, 70 DEC-diarrhea), 31 DAEC (6 DEC-control; 25 DEC-diarrhea), and 74 commensal isolates.

The phylogroup A was the most common, in both groups, control and diarrhea, in EPEC (45 isolates, 52%) and ETEC (44, 53%), while mostly DAEC isolates (27 isolates, 87%) belong to phylogroup D. Regarding EAEC, differences were

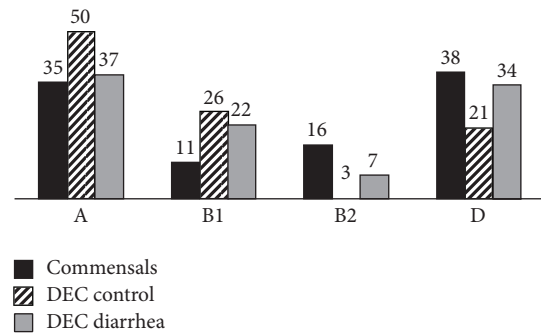


FIGURE 1: Percentage of phylogenetic groups in commensal *E. coli* strains ($n = 74$), diarrheagenic *E. coli* from healthy controls (DEC-control) ($n = 94$), and DEC from children with diarrhea (DEC-diarrhea) ($n = 201$).

found between those isolates causing diarrhea and those recovered from healthy children. Thus EAEC isolates causing diarrhea were mostly classified as phylogroup D (29 isolates, 41%), while those recovered from healthy children predominantly belong to the phylogroup A (10 isolates, 42%) (Table 1). Analyzing together the DEC isolates, those classified as DEC-control strains were more associated with A group (50%) while the DEC-diarrhea strains were more associated with D group (34%) ($P < 0.05$). Meanwhile, commensal *E. coli* ($n = 74$) were more associated with A (26 isolates, 35%) and D (28 isolates, 38%) phylogroups. The commensal group also had a high prevalence of B2 group (12 isolates, 16%) unlike both DEC groups ($P < 0.05$). Both DEC-control (24 isolates, 26%) and DEC-diarrhea (54 isolates, 27%) groups were more associated with the phylogroup B1 than commensals strains (7 isolates, 14%) ($P < 0.05$) (Figure 1).

3.2. Clinical Data of Diarrheal Episodes. From the 201 DEC-diarrhea isolates analyzed in the study, 127 strains were isolated from diarrhea episodes in which no other pathogen was detected. In these 127 patients, no significant differences were found for the studied variables among the four phylogenetic groups. In general, the episode duration was 5 days (1–25), the maximum number of stools/day was 5 (3–11), the total number of stools/episode was 20 (3–128), and the Vesikari score was 6 ± 2.6 . We found a higher proportion ($P = 0.06$) of persistent diarrhea (14 or more days) among B2–D groups (23.9%) compared to among A–B1 groups (9.88%). No differences were found for either acute or prolonged diarrhea (7–14 days) between severe (B2–D) and nonsevere (A–B1) groups.

3.3. Antibiotic Resistance. Resistance to trimethoprim-sulfamethoxazol, tetracycline, chloramphenicol, and nalidixic acid and multiresistance were significantly different among the four phylogenetic groups ($P < 0.05$) (Figure 2). In general, B2 and D groups presented higher percentage of antibiotic resistance than A and B1 groups. In the case of multiresistance D group presented significantly higher percentages than the rest of the groups ($P < 0.01$) (Figure 2).

TABLE 1

Phylogroup	Commensal		Diarrheagenic pathotypes																	
	EAEc (94)		EPEC (87)		DAEC (31)		ETEC (83)													
	N	%	Control	Diarrhea	Control	Diarrhea	Control	Diarrhea	Control	Diarrhea	Control	Diarrhea								
A	26	35.1	10	41.7*	20	28.6*	21	55.3	24	49.0	0	0	0	0	2	8.0	16	61.5	28	49.1
B1	8	10.8	4	16.7	14	20.0	12	31.6	11	22.4	0	0	0	0	0	0	8	30.8	19	33.3
B2	12	16.2	2	8.3	7	10.0	0	0	5	10.2	1	16.7	1	4.0	1	4.0	0	0	2	3.5
D	28	37.8	8	33.3	29	41.4	5	13.1	9	18.4	5	83.3	22	88.0	2	7.7	2	7.7	8	14.0
Total	74	100	24	100	70	100	38	100	49	100	6	100	25	100	26	100	57	100	57	100

* P < 0.05.

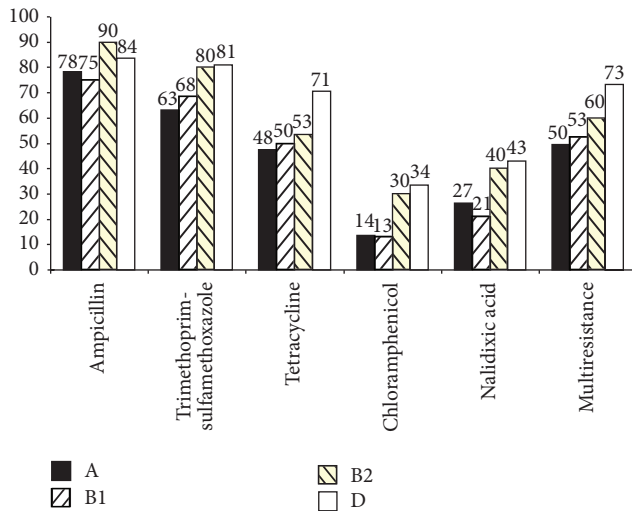


FIGURE 2: Percentages of multiresistance (resistance to 3 or more different antibiotics families) and antibiotic resistance among the four phylogenetic groups A ($n = 147$), B1 ($n = 76$), B2 ($n = 30$), and D ($n = 116$).

4. Discussion

The *E. coli* phylogroups differ in their ecological niches, life-history characteristics, and propensity to cause disease. In this manner B2 and D groups are less frequently isolated from the environment [17]. Regarding human illness, *E. coli* isolates recovered from extraintestinal body sites are more likely to belong to B2 or D phylotypes than to A or B1 [18, 19]. However, differences in the prevalence of the different phylogenetic groups among virulent extraintestinal *E. coli* have been observed in previous studies [20]. Despite the fact that the two phylogenetic groups most frequently related with virulence in the extraintestinal *E. coli* are the aforementioned B2 and D, some reports have shown a high frequency of group A (46%) among *E. coli* causing urinary infections [20]. Alternatively, some reports showed that gut commensal *E. coli* were mostly related with group A [14]. In addition, in a previous study in children in Costa Rica, commensal *E. coli* were related to phylogenetic groups A and D (36%), while the studied DEC belong to B1 (35%), A (29%), B2 (23%), and D (14%) [19]. Another study analyzing DEC (EAEC, EPEC, and STEC) isolated from children in Rumania showed that 51% of the strains belonged to group A, followed by 23% of the strains that belonged to group B2 [21]. In the present study, commensal strains were more associated with A and D groups as has been previously reported [19].

Phylogenetic groups B2 and D have been related with virulence factors that cause infections at an extraintestinal level. Additionally, the B2 strains have been shown to persist for longer periods in infants than other *E. coli* strains [22].

Previous studies have tried to relate the clinical data of infection with the phylogenetic groups [23]. However, these types of studies have not addressed intestinal pathogenic strains that cause diarrhea. In this report, no significant differences were found for the clinical variables analyzed among the four phylogenetic groups. However, we found

a greater tendency of persistent diarrhea in phylogenetic groups B2 and D (24%) than in groups A and B1 (10%). Regarding isolates belonging to the B2 group, this long persistence has been previously observed [22], while no data has been found regarding other phylogroups. In this line an association between the diarrheagenic pathotype DAEC and persistent diarrhea was also observed, in accordance with what has been previously reported [24]. Interestingly, DAEC isolates belong largely to the D phylogroup.

When we analyzed the relation between phylogenetic group and antibiotic resistance we found that the isolates belonging to the group D were more related with multiresistance than those belonging to other groups. Although previous reports in extraintestinal strains showed that virulence-related phylogenetic groups, especially B2, were associated with low levels of antibiotic resistance [11] our data showed a different scenario, in which B2 and D isolates were those exhibiting high levels of antibiotic resistance. A possibility to take into account is the possibility that as phylogroup D results in more prolonged diarrhea, the use of antibacterial agents may be needed, and then these isolates may be under a more intense antibiotic pressure which may facilitate the acquisition of antibiotic resistance mechanisms.

DEC accounts for more than 120,000 deaths/year, being involved, together with rotavirus in around 40% of all diarrhea related children deaths [25]. Recent studies have showed that both EPEC and ETEC isolates are related with an increased mortality [26]; in fact, it is considered that ETEC isolates account for more than 40,000 deaths each year. In this sense it is of interest to note that in the present study most of the ETEC or EPEC isolates belong to the phylogroup A, classically considered as a low-virulent phylogroup. Studies on extraintestinal *E. coli* have showed that the development of antibiotic resistance may be correlated with a decreased virulence [27]. Thus, although no clear reason may be stated to explain the high percentage of DEC isolates belonging to low-virulent phylogroups, the high levels of antibiotic resistance present in the area and these described inverse relations between virulence and antibiotic resistance may be suggested as a potential explanation.

One study limitation is the fact that specific virulence factors encoding genes were not studied; therefore, a direct relationship between virulence and resistance was not possible to evaluate. Furthermore, multilocus sequence typing (MLST) method is more specific than the Clermont triplex PCR method used in this study [28]. However, the Clermont triplex PCR method is a rapid and cost-effective method that has been used extensively. Moreover, this method is able to be implemented in resource constraining sites. A recent study indicates that strains belonging to cryptic lineages of *Escherichia* are the more related to failure by this triplex method; however, in human fecal samples these lineages are unlikely to be found (2-3% frequency) [29].

5. Conclusion

The present data show the relationships between *E. coli* phylogenetic lineages, diarrheagenic character, and severity

of illness. Moreover, an association between phylogroup D and prolonged disease duration has been found. Interestingly, the isolates belonging to this phylogroup also presented the higher levels of multiresistance, showing that those isolates able to cause a more prolonged disease also possess higher levels of antibiotic resistance, probably because they usually required antibiotic treatment and then are under a more intense antibiotic pressure.

Further investigations to elucidate the relationship between phylogeny, specific virulence factors, and mechanisms of resistance are needed in order to better understand DEC and commensal *E. coli* strains.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This study was partially supported by the Agencia Española de Cooperación Internacional (AECID), Spain, Programa de Cooperación Interuniversitaria e Investigación Científica con Iberoamérica (D/019499/08, D/024648/09, and D/030509/10) (TJ and JR). Theresa J. Ochoa is supported by the National Institute of Health, USA, Public Health Service award RO1-HDO67694-01A1. Joaquim Ruiz is supported by I3 program, of the Ministerio de Economía y Competitividad, Spain (Grant no. CES11/012). Maria J. Pons has a postdoctoral fellowship from CONCYTEC/FONDECYT.

References

- [1] J. B. Kaper, "Pathogenic *Escherichia coli*," *International Journal of Medical Microbiology*, vol. 295, no. 6-7, pp. 355-356, 2005.
- [2] U. Dobrindt, "(Patho-)genomics of *Escherichia coli*," *International Journal of Medical Microbiology*, vol. 295, no. 6-7, pp. 357-371, 2005.
- [3] O. Clermont, C. Cordevant, S. Bonacorsi, A. Marecat, M. Lange, and E. Bingen, "Automated ribotyping provides rapid phylogenetic subgroup affiliation of clinical extraintestinal pathogenic *Escherichia coli* strains," *Journal of Clinical Microbiology*, vol. 39, no. 12, pp. 4549-4553, 2001.
- [4] P. Escobar-Páramo, O. Clermont, A.-B. Blanc-Potard, H. Bui, C. Le Bouguéneq, and E. Denamur, "A specific genetic background is required for acquisition and expression of virulence factors in *Escherichia coli*," *Molecular Biology and Evolution*, vol. 21, no. 6, pp. 1085-1094, 2004.
- [5] A. H. Regua-Mangia, K. Irino, R. da Silva Pacheco, R. M. Pimentel Bezerra, A. R. Santos Périssé, and L. M. Teixeira, "Molecular characterization of uropathogenic and diarrheagenic *Escherichia coli* pathotypes," *Journal of Basic Microbiology*, vol. 50, supplement 1, pp. S107-S115, 2010.
- [6] A. Bartoloni, L. Pallecchi, C. Fiorelli et al., "Increasing resistance in commensal *Escherichia coli*, Bolivia and Peru," *Emerging Infectious Diseases*, vol. 14, no. 2, pp. 338-340, 2008.
- [7] C. Gomes, L. Ruiz, M. J. Pons, T. J. Ochoa, and J. Ruiz, "Relevant role of efflux pumps in high levels of rifaximin resistance in *Escherichia coli* clinical isolates," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 107, no. 9, pp. 545-549, 2013.
- [8] S. Mosquito, J. Ruiz, M. J. Pons, D. Durand, F. Barletta, and T. J. Ochoa, "Molecular mechanisms of antibiotic resistance in diarrhoeagenic *Escherichia coli* isolated from children," *International Journal of Antimicrobial Agents*, vol. 40, no. 6, pp. 544-548, 2012.
- [9] T. J. Ochoa, J. Ruiz, M. Molina et al., "High frequency of antimicrobial drug resistance of diarrheagenic *Escherichia coli* in infants in Peru," *The American Journal of Tropical Medicine and Hygiene*, vol. 81, no. 2, pp. 296-301, 2009.
- [10] M. J. Pons, S. Mosquito, T. J. Ochoa et al., "Niveles de resistencia a quinolonas y otros antimicrobianos en cepas de *Escherichia coli* comensales en niños de la zona periurbana de Lima, Perú," *Revista Peruana de Medicina Experimental y Salud Publica*, vol. 29, no. 1, pp. 82-86, 2012.
- [11] E. L. Hannah, J. R. Johnson, F. Angulo, B. Haddadin, J. Williamson, and M. H. Samore, "Molecular analysis of antimicrobial-susceptible and -resistant *Escherichia coli* from retail meats and human stool and clinical specimens in a rural community setting," *Foodborne Pathogens and Disease*, vol. 6, no. 3, pp. 285-295, 2009.
- [12] T. J. Ochoa, L. Ecker, F. Barletta et al., "Age-related susceptibility to infection with diarrheagenic *Escherichia coli* among infants from periurban areas in Lima, Peru," *Clinical Infectious Diseases*, vol. 49, no. 11, pp. 1694-1702, 2009.
- [13] C. E. Guion, T. J. Ochoa, C. M. Walker, F. Barletta, and T. G. Cleary, "Detection of diarrheagenic *Escherichia coli* by use of melting-curve analysis and real-time multiplex PCR," *Journal of Clinical Microbiology*, vol. 46, no. 5, pp. 1752-1757, 2008.
- [14] O. Clermont, S. Bonacorsi, and E. Bingen, "Rapid and simple determination of the *Escherichia coli* phylogenetic group," *Applied and Environmental Microbiology*, vol. 66, no. 10, pp. 4555-4558, 2000.
- [15] C. A. Contreras, T. J. Ochoa, J. Ruiz et al., "Genetic diversity of locus of enterocyte effacement genes of enteropathogenic *Escherichia coli* isolated from Peruvian children," *Journal of Medical Microbiology*, vol. 61, no. 8, pp. 1114-1120, 2012.
- [16] Clinical and Laboratory Standards Institute, "Performance standards for antimicrobial susceptibility testing," Nineteenth Informational Supplement M100-S21, CLSI, Wayne, Pa, USA, 2011.
- [17] S. T. Walk, E. W. Alm, L. M. Calhoun, J. M. Mladonicky, and T. S. Whittam, "Genetic diversity and population structure of *Escherichia coli* isolated from freshwater beaches," *Environmental Microbiology*, vol. 9, no. 9, pp. 2274-2288, 2007.
- [18] D. M. Gordon, *The Influence of Ecological Factors on the Distribution and Genetic Structure of Escherichia Coli*, ASM Press, Washington, DC, USA, 2004, <http://www.asmscience.org/content/journal/ecosalplus/10.1128/ecosalplus.6.4.1>.
- [19] C. Pérez, O. G. Gómez-Duarte, and M. L. Arias, "Diarrheagenic *Escherichia coli* in children from Costa Rica," *The American Journal of Tropical Medicine and Hygiene*, vol. 83, no. 2, pp. 292-297, 2010.
- [20] N. Adib, R. Ghanbarpour, H. Solatzadeh, and H. Alizade, "Antibiotic resistance profile and virulence genes of uropathogenic *Escherichia coli* isolates in relation to phylogeny," *Tropical Biomedicine*, vol. 31, no. 1, pp. 17-25, 2014.
- [21] C.-R. Usein, D. Tatu-Chitoiu, S. Ciontea, M. Condei, and M. Damian, "*Escherichia coli* pathotypes associated with diarrhea in Romanian children younger than 5 years of age," *Japanese Journal of Infectious Diseases*, vol. 62, no. 4, pp. 289-293, 2009.

- [22] F. L. Nowrouzian, I. Adlerberth, and A. E. Wold, "Enhanced persistence in the colonic microbiota of *Escherichia coli* strains belonging to phylogenetic group B2: role of virulence factors and adherence to colonic cells," *Microbes and Infection*, vol. 8, no. 3, pp. 834–840, 2006.
- [23] J. P. Horcajada, S. Soto, A. Gajewski et al., "Quinolone-resistant uropathogenic *Escherichia coli* strains from phylogenetic group B2 have fewer virulence factors than their susceptible counterparts," *Journal of Clinical Microbiology*, vol. 43, no. 6, pp. 2962–2964, 2005.
- [24] S. M. Soto, J. Bosch, M. T. Jimenez de Anta, and J. Vila, "Comparative study of virulence traits of *Escherichia coli* clinical isolates causing early and late neonatal sepsis," *Journal of Clinical Microbiology*, vol. 46, no. 3, pp. 1123–1125, 2008.
- [25] C. F. Lanata, C. L. Fischer-Walker, A. C. Olascoaga, C. X. Torres, M. J. Aryee, and R. E. Black, "Global causes of diarrheal disease mortality in children <5 years of age: a systematic review," *PLoS ONE*, vol. 8, no. 9, Article ID e72788, 2013.
- [26] K. L. Kotloff, J. P. Nataro, W. C. Blackwelder et al., "Burden and aetiology of diarrhoeal disease in infants and young children in developing countries (the Global Enteric Multicenter Study, GEMS): a prospective, case-control study," *The Lancet*, vol. 382, no. 9888, pp. 209–222, 2013.
- [27] J. Vila, K. Simon, J. Ruiz et al., "Are quinolone-resistant uropathogenic *Escherichia coli* less virulent?" *Journal of Infectious Diseases*, vol. 186, no. 7, pp. 1039–1042, 2002.
- [28] D. M. Gordon, O. Clermont, H. Tolley, and E. Denamur, "Assigning *Escherichia coli* strains to phylogenetic groups: multi-locus sequence typing versus the PCR triplex method," *Environmental Microbiology*, vol. 10, no. 10, pp. 2484–2496, 2008.
- [29] O. Clermont, D. M. Gordon, S. Brisse, S. T. Walk, and E. Denamur, "Characterization of the cryptic *Escherichia* lineages: rapid identification and prevalence," *Environmental Microbiology*, vol. 13, no. 9, pp. 2468–2477, 2011.