Original Article XPC codon 939 polymorphism is associated with susceptibility to DNA damage induced by aflatoxin B1 exposure

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Abstract: Aflatoxin B1 (AFB1), resulting in the formation of AFB1-DNA adducts, is a known human carcinogen. AFB1-exposure individuals with inherited susceptible carcinogen-repairing genotypes may experience an increased risk of genotoxicity. This study was aimed to investigate whether DNA repair gene xerodermapigmentosum complementation group C codon 939 polymorphism (rs2228001) affected the levels of AFB1-DNA adducts in Guangxi Population (n = 2558), from an AFB1-exposure area. AFB1-DNA adducts were measured by ELISA, and XPC codon 939 genotypes were identified by TaqMan-PCR. We found that longer AFB1-exposure years significantly increased XPC genotypes with codon 939 Gln alleles (namely, XPC-LG and -GG, odds ratios [95% confidence intervals] were 1.37 (1.15-1.63) and 1.99 (1.55-2.55), respectively) was significantly associated with higher levels of AFB1-DNA adducts. Furthermore, there was a positive joint effect between XPC genotypes and long-year AFB1 exposure in the formation of AFB1-DNA adducts. These results suggest that individuals with susceptible genotypes XPC-LG and -GG may experience an increased risk of DNA damage elicited by AFB1 exposure.

Keywords: AFB1, AFB1-DNA adducts, XPC, polymorphism

Introduction

Aflatoxin B1 (AFB1) is an important toxin produced by Aspergillus fungi. This toxin is mainly metabolized by cytochrome P450 into the genotoxic metabolic 8,9-epoxide-AFB1 (AFBO). AFBO can bind to DNA, and cause the formation of AFB1-DNA adducts that may be removed or repaired by DNA repair enzymes [1, 2]. While xerodermapigmentosum complementation group C (XPC) is required for the efficient repair of this DNA adducts [3-5]. The XPC codon 939 polymorphic locus (rs2228001) has been of particular interest in molecular epidemiology studies [6-11]. Increasing evidence has shown this polymorphism may be associated with decreased DNA repair capacity and increased tumor risk [5-8, 12-21]. This suggests that reduced DNA repair capacity may result in the high risk of AFB1-DNA adducts [9, 10]. Therefore, we specifically conducted a study to examine whether XPC codon 939 polymorphism influences the levels of AFB1-DNA adducts among Guangxi population from an AFB1 exposure area.

Materials and methods

Study subjects

A total of 2558 healthy adults (27-78 yrs of age) population who were residence of Guangxi Zhuang Autonomous Regionwere enrolled from affiliated hospitals of Youjiang Medical College for Nationalities and Guangxi Medical University between January 2002 and December 2009. All study subjects, including 1600 individuals previously studied [11], were without any clinical evidence of liver diseases. The characteristic information of all study subjects, including sex, age, ethnicity, hepatitis B virus (HBV) infection, and HCV infection were ascertained as described previously [22]. These having hepatitis B surface antigen (HBsAg)-positive or anti-

Characteristics					
Age, yr					
Mean ± SD	49.29 ± 11.34				
Range	27-78				
Sex					
Male n (%)	1930 (75.4)				
Female n (%)	628 (24.6)				
Ethnicity					
Han n (%)	1175 (45.9)				
Minority n (%)	1383 (54.1)				
Years of AFB1 exposure, yr					
Mean ± SD	40.23 ± 11.51				
Range	5-76				
HBV infection					
HBsAg (-) n (%)	712 (27.8)				
HBsAg (+) n (%)	1846 (72.2)				
HCV infection					
Anti-HCV (-) n (%)	2092 (81.8)				
Anti-HCV (+) n (%)	466 (18.2)				

 Table 1. Characteristics of study population

HCV-positive in their peripheral serum were defined as groups infected HBV and HCV, respectively. Additionally, after informed consent was obtained, each subject donated 4 mL of peripheral blood for AFB1-DNA adducts and XPC codon 939 genotypes analysis. One hundred percent of people asked to participate in this study who did enroll agreed to participate in the investigative study, and no one dropped out. The protocol of this study was approved by the Ethic Committees of the hospitals involved in the study.

DNA isolation

Leukocytes from peripheral venous blood samples were isolated by standard procedures. DNA was then extracted from leukocyte samples by standard phenol-chloroform extraction and ethanol precipitation. DNA was stored at -20°C until additional analysis.

AFB1-exposure years assay

AFB1-exposure years was ascertained by our previously published methods [22]. To analyze, AFB1-exposure years were divided into two groups: short-AFB1 exposure (< 40 yrs) and long-AFB1 exposure (\geq 40 yrs), according to the value of AFB1-exposure years, with one cutoff

points of 40 yrs, the average AFB1-exposure time.

AFB1-DNA adducts assay

AFB1-DNA adducts levels of DNA samples from leucocytes were measured by competitive enzyme-linked immunosorbent assay (ELISA) using monoclonal antibody 6A10 and 50 µg of DNA as described by Hsieh LL, *et al* [23]. The quality control for adduct assays was administered by blank and positive controls. To analyze, AFB1-DNA adduct levels were divided into two groups: low level (\leq 1.00 µmol/mol DNA) and high level (\geq 1.01 µmol/mol DNA), according to the value of AFB1-DNA adduct levels, with one cutoff points of 1.00 µmol/mol DNA, the average adducts levels among study subjects.

XPC genotype analysis

Gene polymorphism analysis of XPC codon 939 was detected by using a previously published TagMan-PCR [11]. Briefly, PCR reactions were run in a 25 µL final volume containing 1 × Premix Ex Taq[™] (TaKaRa, catalog # DRR039A), 0.2 μ M of each probe, 0.2 μ M of each primer, and 50-100 ng of genomic DNA. Cycling conditions were 95°C for 2 min. and 45 cycles of 95°C for 10 s, 60°C for 1 min and 72°C 10 s. Controls were included in each run and repeated genotyping of a random 10% subset yielded 100% identical genotypes. Data analysis for allele discrimination was performed with the iCycleriQ software. The quality control for genotypic assays was administered by blank and positive controls.

Statistical analysis

The association between XPC codon 939 genotypes and the levels of AFB1-DNA adducts was analyzed using *t* test. Logistic regression with an adjustment for age, sex, ethnicity, HBsAg, and anti-HCV was used to estimate the odds ratio (OR) and the 95% confidence interval (Cl) for the XPC codon 939 genotypes. In this analysis, genotype variable was treated as an ordinal variable (XPC-LL coded as 1, XPC-LG as 2, and as 3), and the corresponding risk value was calculated using the additive model. Additionally, the combinative analysis of risk genotypes (XPC-LG + XPC-GG, also called XPC-LG/GG) was accomplished compared with XPC-LL in dominant model. The interactions between XPC

Exposure years	Low add (n = 1	-level High-level lucts adducts 1537) (n = 1021)		-level lucts 1021)		Р		
	n	%	n	%	OR (95% CI) ^a			
Short	871	56.7	488	47.8	Reference			
Long	666	43.3	533	52.2	1.45 (1.23-1.70)	7.00 × 10 ⁻⁷		

 Table 2. Years of AFB1 exposure and the levels of AFB1-DNA adducts

^aAdjusted for age, sex, ethnicity, HBsAg, and anti-HCV.

codon 939 genotypes and modified factors (including race, sex, HBV and HCV infection, and AFB1-exposure years) on the levels of AFB1-DNA adducts were also estimated and tested on a multiplicative scale by combining genotypes and adding a multiplicative term in the logistic regression model. The statistical significance of the term interaction of genotype-modified factors was evaluated through Likelihood ratio test. All tests were two-tailed. A *P*-value of < 0.05 was considered statistically significant in this study. All the analysis was performed by the statistical package for social science (SPSS) version 18.0 (SPSS Institute, Chicago, IL).

Results

Demographic characteristics for subjects

Table 1 showed the demographic data of all study subjects. The mean age study subjects were 49.29 years. While HBV and HCV infective rates were 72.2% and 18.2%, respectively. These results were in accord with our previously published data [11, 22, 24].

AFB1-exposure years increased AFB1-DNA adducts levels

The average AFB1-exposure years were 40.23 (**Table 1**). We also found those individuals featuring long-AFB1 exposure time were likely to have higher levels of AFB1-DNA adducts in their peripheral blood white blood cells (adjusted OR = 1.45, P < 0.01, **Table 2** and **Figure 1A**).

XPC codon 939 polymorphism increased AFB1-DNA adducts levels

To investigate whether the XPC codon 939 polymorphisms were associated with difference in detoxification and DNA repair, which might be reflected in levels of genotoxic damage, we compared this polymorphism with levels of AFB1-DNA adducts (**Figure 1B** and **Table 3**). The data exhibited that the adjusted OR for those individuals carrying the heterozygotes for Lys and Gln allele of XPC codon 939 (XPC-LG) compared with those exhibiting the homozygote for Lys alleles (XPC-LL) was 1.37 (95% Cl, 1.15-1.63), and the corresponding OR for those featuring the homozygote for Gln alleles (XPC-GG) was

1.99 (95% CI, 1.55-2.55), which showed the risk of high AFB1-DNA adduct levels was related with the number of codon 939 GIn alleles. The genotype distributions of XPC codon 939 polymorphisms in the subjects were consistent with Hardy-Weinberg equilibrium.

The XPC codon 939 genotype distribution stratified by age, race, gender, HBV infection, and HCV infection was shown in Table 4. The results demonstrated similar risk estimates of around 1.5-fold increased high-level AFB1-DNA adducts risk with XPC genotypes with codon 939 GIn alleles (XPC-LG/GG, $P_{\text{interaction}} > 0.05$). Interestedly, we found those individuals featuring HBV- or HCV-infection history and carrying XPC-GG had higher risk of increasing AFB1-DNA adducts levels compared to those having XPC-LL (OR = 2.45 for positive-HBsAg status and 2.39 for positive-anti-HCV status). The hepatitis virus infection-gene interactive analysis, however, did not show statistically significant effects on the levels of AFB1-DNA adducts $(P_{\text{interaction}} > 0.05).$

Joint effects of AFB1-exposure years and XPC codon 939 polymorphism on AFB1-DNA adducts levels

We next analyzed the combination effects of AFB1-exposure years and XPC codon 939 polymorphism on the levels of AFB1-DNA adducts (**Table 5**). In this analysis, we used reference the lowest risk group: those who had short AFB1-exposure years and XPC-LL. The results showed those with long AFB1-exposure years and XPC-GG were more likely to have AFB1-DNA adducts in their peripheral blood white blood cells. Additionally, we also evaluated the multiplicatively interactive effects between genotypes and AFB1-exposure years according to the following formula: $OR_{eg} > OR_{eg'} \times OR_{e'g}$ [25]. Some evidence of multiplicatively interaction was observed (2.13 > 1.30 × 1.37).



Figure 1. The effects of AFB1-exposure years and XPC codon 939 polymorphism on AFB1-DNA adduct levels. In this study, AFB1-DNA adducts levels in the peripheral blood leukocytes were evaluated using ELISA. Longer AFB1-exposure years (A) and risk genotypes of XPC codon 939 (B) increased AFB1-DNA adduct amounts. Data were analyzed using *t* test and shown as means ± S.E.

Table 3. XPC genotypes and the levels of AFB1-DNA adducts

XPC genotype	Low-level ad- ducts (n = 1537)		High-level adducts (n = 1021)		OR (95% CI)ª	Р	
	n	%	n	%			
LL	700	45.5	369	36.1	1		
LG	699	43.5	478	46.8	1.37 (1.15-1.63)	3.74 × 10 ⁻⁴	
GG	168	10.9	174	17.0	1.99 (1.55-2.55)	5.62 × 10 ⁻⁸	
LG/GG	837	54.5	654	63.9	1.49 (1.27-1.76)	1.00 × 10 ⁻⁶	

^aAdjusted for age, sex, ethnicity, HBsAg, and anti-HCV. Abbreviations: LL, XPC genotype with codon 939 Lys alleles; LG, XPC genotype with codon 939 Lys and Gln alleles; GG, XPC genotype with codon 939 Gln alleles; LG/GG, the combination of LG and GG genotypes.

Discussion

To the best our knowledge, no studies have investigated the role of DNA-repair gene XPC codon 939 polymorphisms in the risk of the levels of AFB1-DNA adducts, especially from AFB1exposure areas. In this study, we analyzed the association between this polymorphism and the levels of AFB1-DNA adducts among Guangxi population, from an high AFB1-exposure area. The results showed that XPC genotypes with codon 939 Gln alleles were related with higher levels of AFB1-DNA adducts (OR = 1.49, 95% CI = 1.27-1.76). These results may suggest that XPC codon 939 polymorphism may have functional significance in the AFB1-induced DNA damage.

AFB1, an important chemical carcinogen, is mainly metabolized by cytochrome P450 into

the genotoxic metabolic AFBO which can bind to DNA and cause the formation of AFB1guanine adducts. This kind of AFB1-DNA adducts, including 8,9-dihydro-8-(N⁷-guanyl)-9-hydroxy AFB1-DNA (AFB1-N7-Gua-DNA) adduct, and formamidopyridine-AFB1-DNA (AFB1-FAPy-DNA) adduct [2], if not repaired, might induce DNA damage such as base damage and oxidative DNA damage [2, 26, 27].

While XPC gene spans 33kb on chromosome 3p25, and consists of 16 exons and 15 introns. This gene encodes a 940-amino acid protein, an important DNA damage recognition molecule which plays an important role in NER pathway. XPC protein binds tightly with another important NER protein HR23B to form a stable XPC-HR23B complex, the first protein component that recognizes and binds to the DNA damage sites. XPC-HR23B complex can recognize a variety of DNA adducts formed by exogenous carcinogens such as AFB1 and binds to the DNA damage sites. Therefore, it may play a role in the formation process of AFB1-DNA adducts [3-5].

More than one hundred polymorphisms in the XPC gene have been identified and more and more evidence has expressed that the polymorphisms of this gene are associated with the

		Low-level adducts		High-level adducts			
		n	%	n	%	UR (95% CI)	Р
Ethnicity ^a	XPC						
Han	LL	319	45.6	174	36.6	Reference	
	LG	296	42.3	218	45.8	1.35 (1.05-1.74) ^b	0.02
	GG	84	12.0	84	17.6	1.84 (1.29-2.63) ^b	7.70 × 10 ⁻⁴
	LG/GG	380	54.4	302	63.4	1.46 (1.15-1.85) ^b	2.04 × 10 ⁻³
Minority	LL	381	45.5	195	35.8	Reference	
	LG	373	44.5	260	47.7	1.41 (1.11-1.79) ^b	4.86 × 10 ⁻³
	GG	84	10.0	90	16.5	2.10 (1.48-2.97) ^b	3.40 × 10 ⁻⁵
	LG/GG	457	54.5	350	64.2	1.54 (1.23-1.93) ^b	1.94 × 10 ⁻⁴
Age (yrs) ^c	XPC						
≤ 49	LL	366	44.5	194	35.7	Reference	
	LG	369	44.9	254	46.8	1.34 (1.05-1.70) ^d	0.02
	GG	87	10.6	95	17.5	2.12 (1.51-2.99) ^d	1.70 × 10 ⁻⁵
	LG/GG	456	55.5	349	64.3	1.49 (1.16-1.86) ^d	5.93 × 10 ⁻⁴
≥ 50	LL	334	46.7	175	36.6	Reference	
	LG	300	42.0	224	46.9	1.41 (1.10-1.83) ^d	0.01
	GG	81	11.3	79	16.5	1.90 (1.32-2.74) ^d	6.30 × 10 ⁻⁴
	LG/GG	381	53.3	303	63.4	1.51 (1.19-1.93) ^d	7.40 × 10 ⁻⁴
Sex ^e	XPC						
Male	LL	167	43.4	94	25.5	Reference	
	LG	178	46.2	111	30.1	1.07 (0.75-1.53) ^f	0.70
	GG	40	10.4	38	10.3	1.69 (1.01-2.85) ^f	0.04
	LG/GG	218	56.6	275	74.5	1.52 (1.20-1.93) ^f	7.40 × 10 ⁻⁴
Female	LL	533	46.3	275	35.3	Reference	
	LG	491	42.6	367	47.2	1.46 (1.20-1.78) ^f	2.10 × 10 ⁻⁴
	GG	128	11.1	136	17.5	2.10 (1.58-2.79) ^f	3.03 × 10 ⁻⁷
	LG/GG	619	53.7	503	64.7	1.59 (1.32-1.92) ^f	1.00×10^{-6}
HBsAg ^g	XPC						
Positive	LL	196	42.9	96	37.6	Reference	
	LG	195	42.7	120	47.1	1.41 (1.15-1.72) ^h	9.72 × 10 ⁻⁴
	GG	66	14.4	39	15.3	2.45 (1.82-3.30) ^h	3.56 × 10 ⁻⁹
	LG/GG	261	57.1	159	62.4	1.59 (1.32-1.93) ^h	2.00 × 10 ⁻⁶
Negative	LL	504	46.7	273	35.6	Reference	
	LG	474	43.9	358	46.7	1.32 (0.94-1.86) ^h	0.11
	GG	102	9.4	135	17.6	1.13 (0.70-1.83) ^h	0.62
	LG/GG	576	53.3	493	64.4	1.27 (0.92-1.76) ^h	0.15
Anti-HCV ⁱ	XPC						
Positive	LL	562	44.2	296	36.1	Reference	
	LG	567	44.6	382	46.5	1.82 (1.20-2.74) ^j	4.42 × 10 ⁻³
	GG	143	11.2	143	17.4	2.39 (1.29-4.43) ^j	5.67 × 10 ⁻³
	LG/GG	710	55.8	525	63.9	1.93 (1.31-2.85) ^j	8.87 × 10 ⁻⁴
Negative	LL	138	52.1	73	36.3	Reference	
	LG	102	38.5	96	47.8	1.28 (1.05-1.55) ^j	0.01
	GG	25	9.4	32	15.9	1.91 (1.45-2.51) ^j	4.00 × 10 ⁻⁴
	LG/GG	127	47.9	128	63.7	1.40 (1.17-1.68) ^j	2.53 × 10 ⁻⁴

Table 4. XPC polymorphism and associated OR in relation to ethnicity, sex, HBsAg and anti-HCV

^aLikelihood ration test for interaction of the stratified variable (Han and Minority) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.876). ^aAdjusted for age, sex, HBsAg, anti-HCV, and years of AFB1 exposure. ^cLikelihood ration test for interaction of the stratified variable (Age) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.695). ^dAdjusted for race, sex, HBsAg, anti-HCV, and years of AFB1 exposure. ^eLikelihood ration test for interaction of the stratified variable (Male and Female) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.168). ^fAdjusted for age, ethnicity, HBsAg, anti-HCV, and years of AFB1 exposure. ^eLikelihood ration test for interaction of the stratified variable (Male and Female) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.168). ^fAdjusted for age, ethnicity, HBsAg, anti-HCV, and years of AFB1 exposure. ^gLikelihood ration test for interaction of the stratified variable (HBsAg-positive and negative) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.192). ^hAdjusted for age, sex, ethnicity, anti-HCV, and years of AFB1 exposure. ⁱLikelihood ration test for interaction of the stratified variable (Anti-HCV-positive and negative) and XPC genotype was calculated as test for the heterogeneity of ORs across strata (*P*_{interaction} = 0.241). ⁱAdjusted for age, sex, ethnicity, HBsAg, and years of AFB1 exposure. Abbreviations: LL, XPC genotype with codon 939 Lys alleles; LG, XPC genotype with codon 939 Lys and Gln alleles; GG, XPC genotype with codon 939 Gln alleles; LG/GG, the combination of LG and GG genotypes.

AFB1-exposure years	XPC genotype	Low-level adducts		High-level adducts			
		n	%	n	%	OR (95% CI)"	Р
Short	LL	387	25.2	180	17.6	Reference	
	LG	388	25.2	228	22.3	1.26 (0.99-1.61)	0.06
	GG	96	6.2	80	7.8	1.81 (1.28-2.56)	7.89 × 10 ⁻⁴
	LG/GG	484	31.5	308	30.2	1.37 (1.09-1.72)	6.59 × 10 ⁻³
Long	LL	313	20.4	189	18.5	1.30 (1.01-1.68)	0.04
	LG	281	18.3	250	24.5	1.94 (1.52-2.49)	1.29 × 10 ⁻⁷
	GG	72	4.7	94	9.2	2.87 (2.01-4.09)	6.64 × 10 ⁻⁹
	LG/GG	353	23.0	344	33.7	2.13 (1.69-2.69)	1.69 × 10 ⁻¹⁰

^aAdjusted for age, sex, HBsAg, anti-HCV, and race. Abbreviations: LL, XPC genotype with codon 939 Lys alleles; LG, XPC genotype with codon 939 Lys and Gln alleles; GG, XPC genotype with codon 939 Gln alleles; LG/GG, the combination of LG and GG genotypes.

function of DNA repair capacity [6, 7, 11]. In this study, we only analyzed XPC codon 939 polymorphism because this polymorphism changes the amino acids coded, which may be associated with decreased DNA repair capacity [7, 8, 13-18, 20, 21, 28], increased frequency of p53 mutations [29, 30], and increased tumor risk [6, 11, 15]. Recent some studies have shown that low DNA repair capacity resulting from the genetic mutation of XPC codon 939 polymorphism can progress AFB1-induced HCC [31-33], suggesting that XPC codon 939 polymorphism may be important in the repair of AFB1-DNA adducts. Our present data not only supported this hypothesis, but also we found that this polymorphism would be able to interact with AFB1-exposure years, especially long-year AFB1 exposure, in the formation of AFB1-DNA adducts. Possibly, differences in the AFB1exposure years reflect differences in cumulative exposure information. In tissues and cells with longer-years AFB1 exposure, AFB1-DNA adducts are cumulated because of the deficiency of DNA repair ability.

Although some clues of the interactive effects between either HBV or HCV infection and XPC

codon 939 genotypes on the levels of AFB1-DNA adducts were found in this study, the effect seems to be greatest at the genotype of XPC-GG under the conditions of positive infection history. This may be because of low detoxification capacity resulting from chronic liver diseases history, and low DNA repair ability, which results in the formation of AFB1-DNA adducts.

Conclusion

To the best of our knowledge, this is the first report to investigation associations between the polymorphism at the codon 939 of XPC and the levels of AFB1-DNA adducts. We found evidence to suggest that the XPC codon 939 Gln alleles are associated with increased levels of DNA damage that may be due to reduced detoxification and DNA repair function. However, Selection bias might have occurred through the selection of hospital-based subjects. Furthermore, liver disease (resulting from virus infection) itself may affect the metabolism of aflatoxin and modify the levels of aflatoxin-DNA adducts. Additionally, other polymorphisms (such as the polymorphisms of XRCC4) might be able to further modify the effect of XPC polymorphism on AFB1-DNA adducts. Therefore, future studies need to characterize the role of the XPC codon 939 Gln alleles in functional detoxification and DNA repair assays and to test to see whether they affect the levels of other biomarkers of DNA damage. Given that high AFB1-DNA adducts positively associates with liver cancer risk, the finding of a genetic susceptibility (if confirmed) may have implications for cancer screening and prevention.

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Disclosure of conflict of interest

None.

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References

 Guengerich FP, Johnson WW, Shimada T, Ueng YF, Yamazaki H and Langouet S. Activation and detoxication of aflatoxin B1. Mutat Res 1998; 402: 121-128.

- [2] Wang JS and Groopman JD. DNA damage by mycotoxins. Mutat Res 1999; 424: 167-181.
- [3] Araujo SJ and Wood RD. Protein complexes in nucleotide excision repair. Mutat Res 1999; 435: 23-33.
- [4] Batty DP and Wood RD. Damage recognition in nucleotide excision repair of DNA. Gene 2000; 241: 193-204.
- [5] Sugasawa K. XPC: its product and biological roles. Adv Exp Med Biol 2008; 637: 47-56.
- [6] Qiu L, Wang Z, Shi X and Wang Z. Associations between XPC polymorphisms and risk of cancers: A meta-analysis. Eur J Cancer 2008; 44: 2241-2253.
- [7] Zhang D, Chen C, Fu X, Gu S, Mao Y, Xie Y, Huang Y and Li Y. A meta-analysis of DNA repair gene XPC polymorphisms and cancer risk. J Hum Genet 2008; 53: 18-33.
- [8] Liang J, Gu A, Xia Y, Wu B, Lu N, Wang W, Lu C, Zheng Q, Wang S and Wang X. XPC gene polymorphisms and risk of idiopathic azoospermia or oligozoospermia in a Chinese population. Int J Androl 2009; 32: 235-241.
- [9] Xia Q, Huang XY, Xue F, Zhang JJ, Zhai B, Kong DC, Wang C, Huang ZQ and Long XD. Genetic polymorphisms of DNA repair genes and DNA repair capacity related to aflatoxin b1 (AFB1)induced DNA damages. In: Chen C. New Research Directions in DNA Repair. 1. Rijeka: In-Tech; 2013. pp. 377-412.
- [10] Long XD, Yao JG, Zeng Z, Huang CH, Huang ZS, Huang YZ, Ban FZ, Huang XY, Yao LM, Fan LD and Fu GH. DNA repair capacity-related to genetic polymorphisms of DNA repair genes and aflatoxin B1-related hepatocellular carcinoma among Chinese population. In: Kruman I. DNA Repair. Rijeka: InTech; 2011. pp. 505-524.
- [11] Long XD, Ma Y, Zhou YF, Ma AM and Fu GH. Polymorphism in xeroderma pigmentosum complementation group C codon 939 and aflatoxin B1-related hepatocellular carcinoma in the Guangxi population. Hepatology 2010; 52: 1301-1309.
- [12] Caronia D, Patino-Garcia A, Milne RL, Zalacain-Diez M, Pita G, Alonso MR, Moreno LT, Sierrasesumaga-Ariznabarreta L, Benitez J and Gonzalez-Neira A. Common variations in ERCC2 are associated with response to cisplatin chemotherapy and clinical outcome in osteosarcoma patients. Pharmacogenomics J 2009; 9: 347-353.
- [13] De Ruyck K, Szaumkessel M, De Rudder I, Dehoorne A, Vral A, Claes K, Velghe A, Van Meerbeeck J and Thierens H. Polymorphisms in base-excision repair and nucleotide-excision repair genes in relation to lung cancer risk. Mutat Res 2007; 631: 101-110.
- [14] Dong Z, Guo W, Zhou R, Wan L, Li Y, Wang N, Kuang G and Wang S. Polymorphisms of the

DNA repair gene XPA and XPC and its correlation with gastric cardiac adenocarcinoma in a high incidence population in North China. J Clin Gastroenterol 2008; 42: 910-915.

- [15] Francisco G, Menezes PR, Eluf-Neto J and Chammas R. XPC polymorphisms play a role in tissue-specific carcinogenesis: a meta-analysis. Eur J Hum Genet 2008; 16: 724-734.
- [16] Hansen RD, Sorensen M, Tjonneland A, Overvad K, Wallin H, Raaschou-Nielsen O and Vogel U. XPA A23G, XPC Lys939Gln, XPD Lys751Gln and XPD Asp312Asn polymorphisms, interactions with smoking, alcohol and dietary factors, and risk of colorectal cancer. Mutat Res 2007; 619: 68-80.
- [17] Mechanic LE, Millikan RC, Player J, de Cotret AR, Winkel S, Worley K, Heard K, Tse CK and Keku T. Polymorphisms in nucleotide excision repair genes, smoking and breast cancer in African Americans and whites: a populationbased case-control study. Carcinogenesis 2006; 27: 1377-1385.
- [18] Shore RE, Zeleniuch-Jacquotte A, Currie D, Mohrenweiser H, Afanasyeva Y, Koenig KL, Arslan AA, Toniolo P and Wirgin I. Polymorphisms in XPC and ERCC2 genes, smoking and breast cancer risk. Int J Cancer 2008; 122: 2101-2105.
- [19] Takebayashi Y, Nakayama K, Kanzaki A, Miyashita H, Ogura O, Mori S, Mutoh M, Miyazaki K, Fukumoto M and Pommier Y. Loss of heterozygosity of nucleotide excision repair factors in sporadic ovarian, colon and lung carcinomas: implication for their roles of carcinogenesis in human solid tumors. Cancer Lett 2001; 174: 115-125.
- [20] Zhang L, Zhang Z and Yan W. Single nucleotide polymorphisms for DNA repair genes in breast cancer patients. Clin Chim Acta 2005; 359: 150-155.
- [21] Zhou RM, Li Y, Wang N, Zhang XJ, Dong XJ and Guo W. [Correlation of XPC Ala499Val and Lys-939GIn polymorphisms to risks of esophageal squamous cell carcinoma and gastric cardiac adenocarcinoma]. Ai Zheng 2006; 25: 1113-1119.
- [22] Long XD, Ma Y, Wei YP and Deng ZL. The polymorphisms of GSTM1, GSTT1, HYL1*2, and XRCC1, and aflatoxin B1-related hepatocellular carcinoma in Guangxi population, China. Hepatol Res 2006; 36: 48-55.
- [23] Hsieh LL, Hsu SW, Chen DS and Santella RM. Immunological detection of aflatoxin B1-DNA adducts formed in vivo. Cancer Res 1988; 48: 6328-6331.

- [24] Long XD, Ma Y, Qu de Y, Liu YG, Huang ZQ, Huang YZ, Lin ZH, Wei NB and Zhou SC. The polymorphism of XRCC3 codon 241 and AFB1related hepatocellular carcinoma in Guangxi population, China. Ann Epidemiol 2008; 18: 572-578.
- [25] Brennan P. Gene-environment interaction and aetiology of cancer: what does it mean and how can we measure it? Carcinogenesis 2002; 23: 381-387.
- [26] Wood RD. DNA damage recognition during nucleotide excision repair in mammalian cells. Biochimie 1999; 81: 39-44.
- [27] Wilson DM 3rd and Thompson LH. Life without DNA repair. Proc Natl Acad Sci U S A 1997; 94: 12754-12757.
- [28] Laczmanska I, Gil J, Karpinski P, Stembalska A, Trusewicz A, Pesz K, Ramsey D, Schlade-Bartusiak K, Blin N and Sasiadek MM. Polymorphism in nucleotide excision repair gene XPC correlates with bleomycin-induced chromosomal aberrations. Environ Mol Mutagen 2007; 48: 666-671.
- [29] Ryk C, Kumar R, Sanyal S, de Verdier PJ, Hemminki K, Larsson P, Steineck G and Hou SM. Influence of polymorphism in DNA repair and defence genes on p53 mutations in bladder tumours. Cancer Lett 2006; 241: 142-149.
- [30] Sakano S, Matsumoto H, Yamamoto Y, Kawai Y, Eguchi S, Ohmi C, Matsuyama H and Naito K. Association between DNA repair gene polymorphisms and p53 alterations in Japanese patients with muscle-invasive bladder cancer. Pathobiology 2006; 73: 295-303.
- [31] Strom SS, Estey E, Outschoorn UM and Garcia-Manero G. Acute myeloid leukemia outcome: role of nucleotide excision repair polymorphisms in intermediate risk patients. Leuk Lymphoma 2010; 51: 598-605.
- [32] Khan SG, Yamanegi K, Zheng ZM, Boyle J, Imoto K, Oh KS, Baker CC, Gozukara E, Metin A and Kraemer KH. XPC branch-point sequence mutations disrupt U2 snRNP binding, resulting in abnormal pre-mRNA splicing in xeroderma pigmentosum patients. Hum Mutat 2010; 31: 167-175.
- [33] Gangwar R, Mandhani A and Mittal RD. XPC gene variants: a risk factor for recurrence of urothelial bladder carcinoma in patients on BCG immunotherapy. J Cancer Res Clin Oncol 2010; 136: 779-786.