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Integrins and cancer: regulators of cancer stemness, metastasis, and drug resistance

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Summary

Interactions between cancer cells and their surroundings can trigger essential signaling cues that determine cell fate and influence the evolution of the malignant phenotype. As the primary receptors involved in cell-matrix adhesion, integrins present on the surface of tumor and stromal cells have a profound impact on the ability to survive in specific locations, but in some cases these receptors can also function in the absence of ligand binding to promote stemness and survival in the presence of environmental and therapeutic stresses. Understanding how integrin expression and function is regulated in this context will enable the development of new therapeutic approaches to sensitize tumors to therapy and suppress their metastatic phenotype.

Keywords

Integrins; cancer; drug resistance; stemness

Integrin heterodimers and ligand specificity in cancer

When the extracellular matrix (ECM) is proteolytically degraded or deformed by mechanical forces, cells are prompted to undergo responsive changes that influence remodeling during physiological and pathological events. Integrins are a family of heterodimeric cell surface receptors that sense these changes and trigger a range of cellular responses by forming a physical connection between the inside and outside of a cell to allow the bi-directional “integration” of signals to control cell adhesion, migration, proliferation, survival, and differentiation [1]. While integrins regulate processes important for a range of physiological functions, these receptors also play a crucial role in promoting a more malignant tumor cell phenotype in the setting of cancer [2].

The ability of integrins to dictate cellular responses to a variety of inputs stems from their capacity to differentially recognize distinct environments. To allow for this flexibility, integrins are comprised of 18 α subunits and 8 β subunits that pair to form at least 24

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different functional heterodimeric receptors that each bind to one or more ECM ligands. This specificity allows integrin-ligand binding events to enforce distinct niches or boundaries, so that cells expressing only certain integrin heterodimers can pass within an extracellular matrix containing specific components such as laminin, collagen, vitronectin, or fibronectin. Since a given integrin can bind to multiple ligands, and a single ligand can recognize multiple integrin heterodimers, spatio-temporal patterns of integrin vs. ligand expression ultimately determine how a cell senses and responds to its environment. Integrin control of matrix metalloproteinase (MMPs) on the surface of cells is also a determining factor for invasive behavior [3]. In the context of cancer, this cell adhesion-dependent aspect of integrin function plays a critical role in determining a cell's ability to break through a defined tumor margin in order to locally invade and ultimately metastasize. Ligand binding also controls whether a certain tumor cell can disseminate to a particular metastatic niche, such as bone, brain, or lung environments characterized by distinct ECM signatures.

Upon encountering a specific ligand, integrins undergo a conformational change that switches them from an inactive low avidity state to a high avidity state [4]. This change is based in part on the ability of ligated integrins to cluster in the plane of the membrane leading to “outside-in” signaling via a physical linkage to the actin cytoskeleton (Box 1). Alternatively, intracellular signaling can also activate “inside-out” signals that affect integrin affinity/avidity for extracellular ligands [5]. By selectively recruiting adapter or scaffolding proteins such as CAS, Shc, and Grb2, integrins play an important role in potentiating the activity of receptor tyrosine kinases, including receptors for growth factors such as VEGF, FGF, or EGF [1, 6-9]. Although these “canonical” integrin signaling pathways have been extensively characterized, new specificities and signaling components are still being discovered [10].

Similar to growth factor receptors, integrin clustering within the plasma membrane is regulated by numerous direct or indirect integrin binding partners, and serves to amplify signal-generating capacity (Figure 1). Accordingly, integrin clustering could represent an Achilles' heel for integrin function that could be exploited therapeutically. Galectins, a family of β -galactoside-binding lectins recently associated with metastasis [11], influence tumor cell behavior by binding to carbohydrates on the extracellular domain of integrins and regulate clustering. Several galectins have recently been identified to play a role in tumor progression including Galectin-1, which promotes lung cancer metastasis by potentiating integrin $\alpha_6\beta_4$ and Notch1/Jagged2 signaling [12], and Galectin-3, which induces integrin β_3 -mediated anchorage-independence and drug resistance [13]. Tetraspanins also play key roles in clustering integrins by regulating their trafficking and function [14, 15]. The tetraspanin CD151 in particular shows promise as a diagnostic marker as well as a therapeutic target [16]. Thus, through their effects on integrin clustering, galectins and tetraspanins could provide a means to control integrin signaling independent of ligand binding and promote tumor cell dissemination and metastasis.

Not only can integrin function be suppressed by competitively blocking integrin-ligand binding events, but it is also possible to suppress ligand-independent integrin clustering by manipulating the function of proteins such as galectins or tetraspanins that can cluster and promote integrin activity in the absence of cell adhesion in a permissive microenvironment.

In fact, combining these strategies could provide therapeutic opportunities to short circuit the ability of integrins to generate signals across distinct environments. Developing such an approach will require a better understanding of the cues and responses that are spatially and temporally distinct during the course of cancer progression. This review is therefore focused on highlighting newly appreciated roles of integrins in driving a stem phenotype, drug resistance, and metastasis.

Dissecting integrin-dependent regulation of stem cells

Although epithelial stem cells play a critical role in the physiological development, maintenance, and remodeling of organs and tissues [17], their properties are also associated with the initiation and progression of carcinomas [18]. Since the stem cell niche is tightly regulated by signals from the local microenvironment including the ECM, certain integrins may be critical for the ability of stem cells to sense and respond to these cues in both normal tissues and cancer. Indeed, a number of integrins have recently been highlighted as important markers and functional regulators of stem cells, suggesting that additional insight into how integrins contribute to the stem cell phenotype will allow the development of therapeutic approaches to modulate stemness in aggressive cancers.

Integrin regulation of stem cells during development and physiological remodeling

Recent studies have identified specific integrins that are enriched in epithelial stem cells and critical for their behavior. Integrin $\beta 1$ (CD29) is highly expressed in normal stem cells and regulates their biology in various organs. Stem cells are typically associated with a particular local microenvironment or niche that provides critical signals to direct their self-renewal and pluripotency. For example, ECM proteins such as periostin and tenascin-C are found in stem cell niches [19, 20]. Increasing evidence demonstrates that integrin $\beta 1$ maintains the stem cell niche, preserves a stable stem cell population, and controls the balance between stem cell renewal and differentiation [21]. In the epidermis, the hair follicle bulge creates a smooth muscle cell niche by expressing integrin $\alpha 8\beta 1$ [22]. Integrin signaling is also involved in proliferation and differentiation of cutaneous epithelial stem cells where integrin $\beta 1$ promotes keratinocyte adhesion and integrin $\alpha v\beta 6$ activates transforming growth factor- β (TGF- β) [23]. In addition, MT1-MMP activates a $\beta 1$ -integrin/RhoGTPase signaling cascade that regulates stem cell shape by controlling skeletal stem cell lineage commitment [24]. In intestinal epithelial stem cells, integrin $\beta 1$ deletion increases epithelial proliferation suggesting that integrin expression can limit adult stem cell proliferation [25]. Indeed, integrin dependent adhesion to the basement membrane induces cell intrinsic polarity, resulting in asymmetric division, which ultimately allows the continuing maintenance of stem cells and the generation of differentiated cells [26].

In the mammary gland, the microenvironment provides cues to control the behavior of epithelial stem and progenitor cells [27], and integrins are important markers for identifying these cell types, as separate epithelial lineages arising from the same precursors in the breast can be distinguished on the basis of their integrin profiles. Luminal cells express low levels of the integrin $\alpha 6$ (CD49f) and $\beta 1$ subunits, while cells in the basal layer including mammary stem cells express higher levels [28-30]. Additionally, functional studies have

demonstrated that the integrin $\beta 1$ subunit is essential for the regenerative potential of the adult mammary gland [21].

Expression levels of integrin $\beta 3$ (CD61) can distinguish mammary luminal progenitors ($\beta 3^{-}$) from mature, differentiated luminal ($\beta 3^{-}$) cells [31]. There is also evidence that an $\alpha v\beta 3$ -mediated stemness pathway is important for physiological remodeling events mediated by adult stem cells, as TGF $\beta 2$ induces $\alpha v\beta 3$ expression on adult mammary stem cells during mid-pregnancy to promote mammary stem cell expansion, clonogenicity, and expression of the master stem cell regulator Slug [32]. Whereas virgin mice lacking integrin $\alpha v\beta 3$ develop normal mammary glands, mammary remodeling during pregnancy is defective [32]. This pathway can be usurped during breast cancer, as $\alpha v\beta 3$ contributes to cancer stem cell properties including tumorsphere formation, tumor initiation [32] and metastasis [33]. These examples illustrate the capacity of specific integrins to influence stem cell behavior during maintenance or remodeling of normal tissues, roles that may be conserved in “stem-like” cancer cells.

Integrin regulation of cancer stem cells during tumor initiation

Accumulating evidence suggests that a relatively small number of tumor initiating cells (TICs) within a given tumor represents the subpopulation capable of self-renewal, tumorigenesis, and generation of the heterogeneous tumor cell populations observed in many cancer types. Also known as cancer stem cells (CSCs), TIC have been established as drivers of tumor progression, drug resistance, and disease relapse [18]. Although integrins are well-known for their contributions to tumor progression, it is unclear whether these effects are related to their role in CSCs. Many of the same integrins that are enriched on normal adult stem and progenitor cells are also markers of CSCs, including integrin subunits $\beta 1$, $\alpha 6$, and $\beta 3$ [34]. Among these, $\alpha 6$ is the most widely observed, enriching for CSCs in breast [35], prostate [36], squamous cell carcinoma [37] and colorectal [38] cancers. Recent studies have also characterized integrin $\beta 4$ as a new CSC marker in lung cancer, where it is involved in self-renewal, tumor propagation, and chemotherapy resistance [39]. Therefore, differential surface expression of specific integrins may identify a small sub-population of the most aggressive and dangerous tumor cells.

In addition to serving as cancer stem *markers*, there is recent evidence that integrins also potentiate cancer stem cell *function*. In glioblastoma, disrupting integrin $\alpha 6$ function suppresses the cancer stem cell phenotype, highlighting the role of this integrin for the maintenance of glioblastoma stem cells [40]. Integrin $\alpha 6$ also contributes to breast cancer initiation by regulating a FAK-mediated induction of the Polycomb complex protein BMI-1, necessary for cancer cell self-renewal [41]. In fact, two alternative splice variants of integrin $\alpha 6$ drive opposite phenotypes. Whereas the $\alpha 6A$ variant promotes an epithelial phenotype, the $\alpha 6B$ variant is essential for establishing a stem-like mesenchymal phenotype [42]. Interestingly, these variants differ only in the cytoplasmic tail of integrin $\alpha 6$. Simply disabling the signaling domain of integrin $\beta 4$ abrogates the CSC capabilities of prostate cancer cells, without impacting the development of the prostate [43]. Moreover, integrin $\beta 3$ is necessary and sufficient for the CSC phenotype in breast [32, 44], pancreas [13], and lung [13] cancers. Thus, targeting the ability of specific integrins to modulate cell adhesion

events important for cancer stem cells by manipulating integrin/ECM interactions may represent a therapeutic strategy to suppress the function of cancer stem cells.

Ligand independent functions

Several studies now show that integrins may influence cancer stem cells independent from their capacity to interact with the ECM. In lung and pancreatic cancers, integrin $\alpha v \beta 3$ forms cluster on the surface of cells growing in suspension, without any physical link to ECM ligands. This clustering is mediated by a Galectin-3 interaction with $\alpha v \beta 3$ independent of its ligand binding domain, which in turn promotes recruitment of activated KRAS and ultimately leads to a “stemness” phenotype [13]. Treating cells with integrin antagonists that compete for ligand binding or expressing mutant integrin constructs with impaired ligand binding ability do not compromise the ability of $\alpha v \beta 3$ to drive this stem phenotype, suggesting that it is ligation independent [13]. These recent examples suggest that tumor cells may utilize certain integrins independent from their role as adhesion receptors to promote survival within non-permissive environments, such as those encountered during metastatic dissemination or tumor initiation (Figure 2).

For the case of $\alpha v \beta 3$, its adhesion-independent functions trigger pathways distinct from the typical RAS signaling cascade and cytoskeletal links. Expression of $\alpha v \beta 3$ is necessary and sufficient to drive tumor cell metastasis by virtue of $\beta 3$ -mediated the recruitment and activation of Src kinase in a manner that is not dependent on FAK or MEK/ERK signaling [33]. Furthermore, only unligated $\alpha v \beta 3$ can form a complex with KRAS to recruit RALB and TBK1 and promote NF κ B activity [13]. Therefore, while the activation of RAS family members is triggered downstream of multiple integrins, it is becoming clear that integrin-mediated signaling pathways are diverse and context-dependent, with certain integrins capable of directing specific stemness-related reprogramming. While this mode of integrin function provides an important role during development, its usurpation by cancer stem cells likely allows their survival and transit into inappropriate locations that exacerbates metastasis and tumor progression, and renders tumors highly resistant to therapies that fail to target this tumor subpopulation.

Connecting integrins and drug resistance

Despite advances in cancer treatment, many cancer therapies are limited by the development of resistance that results from a variety of factors, including alterations in the drug target, activation of pro-survival pathways, and ineffective induction of cell death. Resistance to anticancer therapeutics can be divided into two categories: intrinsic resistance derived from genetic or environmental factors pre-existing in the tumor, or acquired resistance resulting from adaptive responses, activation of alternative pathways, and selection of resistant subpopulations.

Cell adhesion-mediated drug resistance, a pro-survival and anti-apoptotic program, is dictated by integrins/ECM interactions [45]. This evasion strategy can select for cells already expressing certain integrins and/or cells capable of inducing integrin gene expression. For example, integrin $\beta 1$ has been implicated as a driver of resistance to radiotherapy in head and neck cancer [46], lapatinib and trastuzumab resistance in breast

cancer [47], and erlotinib resistance in lung cancer [48]. In lung cancer, erlotinib increased expression of $\beta 1$, $\alpha 2$, and $\alpha 5$ and enhanced cell adhesion, while silencing integrin $\beta 1$ restored erlotinib sensitivity by reducing Src and Akt activity, implicating the $\beta 1$ /Src signaling pathway as a key mediator of acquired resistance to EGFR targeted therapies [48]. It is also likely that multiple integrins cooperatively contribute to drug resistance. Indeed, matrix-attached ovarian carcinoma cells tolerate dual PI3K/mTOR inhibition by inducing an adaptive pro-survival response, which can be blocked by simultaneous inhibition integrin $\beta 1$, integrin $\beta 4$, ILK, and FAK [49]. There is also evidence that the ECM can directly modulate cell sensitivity to treatment therapies. In ovarian cancer, the matrix-associated growth factor TGF- β , sensitizes cells to paclitaxel-induced cell death by mediating FAK/Rho signaling pathway through preferential ligation to integrin $\alpha \nu \beta 3$ [50].

The involvement of integrins in the promotion of a CSC phenotype is likely an additional contributor to chemoresistance and tumor relapse. In breast cancer, an integrin $\alpha 6$ -CSC population is enriched after taxol treatment [42]. Although several studies suggest that CSCs are enriched after cancer therapy, this phenotype has not been confirmed using spontaneous tumorigenesis mouse models or in human cancers. A recent study demonstrates that CSC expressing integrin $\beta 4$ are enriched after cisplatin treatment in the $Kras^{G12D}; Trp53^{fl/fl}$ spontaneous lung cancer model [39]. $\beta 3$ is also involved in intrinsic and acquired resistance to erlotinib and lapatinib, as this integrin is highly expressed after acquired resistance to EGFR inhibitors where it drives an NF κ B signaling pathway leading to erlotinib resistance [13]. Together, these findings suggest that several integrins play a role in cancer drug resistance, possibly through controlling CSC behavior.

Integrin involvement in drug resistance may also depend on modulation of the immune response. A variety of drugs induce a DNA damage response that enhances the expression of integrin $\alpha \nu \beta 3$ on tumor cells as they acquire drug resistance, and these therapy-resistant tumor cells are more readily phagocytized by dendritic cells to suppress the immune response [51]. In contrast, monoclonal antibodies that block integrin adhesion may also act to enhance immune response. For example, when tumors highly express $\beta 3$ integrin, systemic therapy with the anti- $\alpha \nu \beta 3$ antibody LM609 invokes host defense mechanisms and triggers antibody-dependent cellular cytotoxicity [52]. Understanding how tumor cells are recognized by the immune system can provide novel therapeutic strategies to combat drug resistance.

Critical roles for integrins during the metastatic cascade

Metastasis is a multistep process that requires a cancer cell to escape from the primary tumor, survive in the circulation, colonize distant sites, and proliferate. Since integrin function can enable and enhance many facets of these steps, it is not surprising that increased expression of certain integrins within the primary tumor are associated with poor prognosis and enhanced metastasis in a variety of cancers [1] (Figure 3). This increased expression may reflect higher numbers of CSCs, with their enriched expression of certain integrins [34] and may explain why some tumors progress, while others do not. Additionally, integrin expression in both CSCs and other tumor cells can be induced by cues

from the microenvironment [53, 54] resulting in increased integrin signaling that promotes the various steps responsible for metastatic progression.

Local invasion

The initial phase of tumor dissemination from a primary site involves a variety of signals that can be modulated by the function of multiple integrins [1, 55]. During local invasion, a cancer cell uses migration mechanisms similar to those in non-neoplastic cells during physiological processes such as embryonic morphogenesis, e.g., a cell must acquire a capacity to spread within the tissue and invade the matrix extracellular. To migrate, the cancer cell modifies its shape to interact with the surrounding tissue. This initial step requires a phenotypic conversion known as the epithelial-to-mesenchymal transition (EMT). This process is critical during development, but is also frequently triggered during metastasis. EMT is characterized by the transition to a mesenchymal phenotype, involving the disassembly of cell-cell contacts, cytoskeletal reorganization, and acquisition of mesenchymal markers and migratory properties [56]. It is not surprising that the mechanisms critical for EMT, stemness, and drug resistance demonstrate significant overlap [57], and it is likely that integrins play a critical role in allowing tumor cells to become more aggressive and therapy-resistant.

Anchorage-independent survival in the bloodstream

Non-transformed cells depend on integrins to relay cues from the ECM to maintain organ integrity and prevent cells from inappropriately wandering to other tissues. While tumor cells display some degree of anchorage-independence, their detachment from the ECM can promote cell death. Once tumor cells escape from a primary site and intravasate into blood or lymphatic vessels, they must therefore adapt to survive in the absence of adhesion to ECM. Growing evidence supports a central role for integrins in controlling growth and survival under anchorage-independent conditions, a property critical for hematogenous metastasis. For example, integrin $\beta 1$ promotes anchorage-independent growth in prostate [58] and breast cancer cells by activating a FAK/PAK/MAPK signaling cascade [59], while integrin $\beta 3$ interacts with c-Src independently of FAK signaling to drive increased anchorage-independence and lymph node metastasis in pancreatic and breast cancer tumor models [32]. The paradigm of integrin-mediated anchorage-independent growth may be explained in part by the ability of specific integrins to form cell surface clusters in response to growth factors or oncogenes and drive downstream signaling [13, 60].

Colonization in the metastatic niche

Metastatic colonization of distant organs requires the cell survival and expansion of cancer cells at each secondary site. This process is successfully accomplished by only a minority of cancer cells that reach the distant organ, as seeding a “metastatic niche” requires specific recognition between cancer cells and their surrounding ECM. Since the integrin repertoire expressed by a given tumor cell may dictate that cell’s ability to respond to a particular niche and initiate metastatic colonization, integrins may be critical for the “homing” of tumor cells to organ environments that promote metastasis.

The propensity for metastasis has recently been linked to the accumulation of certain ECM proteins within a particular metastatic niche. Tenascin C is a ligand for $\beta 1$ and $\beta 3$ integrins that is produced within the lung metastatic niche and correlates with poor outcome for breast cancer [61]. Periostin is a ligand for $\alpha v\beta 3$ and $\alpha v\beta 5$ that is enriched in breast cancer lymph node metastases [62]. Expressed on breast cancer cells, the $\alpha v\beta 3$ ligand L1-CAM is required for breast cancer metastasis to the lungs, allowing tumor cells to bind and extravasate through the lung endothelium [63]. VCAM-1 drives the metastatic spread of tumor cells to lymph nodes where $\alpha 4\beta 1$ is expressed on lymph node endothelial cells. VEGF-C/PI3K α -driven remodeling of lymph nodes activates integrin $\alpha 4\beta 1$ on lymph node lymphatic endothelium, which in turn serves as an adhesive ligand for VCAM– tumor cells [64]. Since binding of $\alpha 4\beta 1$ to VCAM-1 also contributes to the vascular arrest and extravasation of melanoma or lymphoma cells to the lung or spleen, this integrin-mediated binding event is gaining interest as a possible target for cancer therapy [65]. Similarly, integrins $\alpha v\beta 3$, $\alpha 2\beta 1$, and $\alpha 4\beta 1$ play key roles in bone metastasis, as their collective ligands represent ECM proteins normally expressed by bone-associated cells: $\alpha v\beta 3$ binds osteopontin, while $\alpha 2\beta 1$ and $\alpha 4\beta 1$ bind type I collagen on the bone matrix and VCAM-1 on bone endothelial cells [66].

These examples portray the complexity of integrin-ligand binding events that may govern how and where circulating tumor cells travel to form new micrometastatic colonies. Once entrenched within an appropriate metastatic niche, tumor cells must adapt again to allow their survival and proliferation. Integrin $\beta 1$ -mediated filopodium-like protrusions that support the initial interactions between the extravasated cancer cells and ECM components of the host help these cells to trigger adhesion-dependent signaling events including FAK activation, which then phosphorylates and activates ERKs leading to rapid proliferation of these cancer cells within the host tissue [67, 68]. It is also likely that integrins that provide anchorage-independent growth advantages for circulating tumor cells will have less influence over the later steps of metastasis when adhesion-dependent growth may again prevail. However, it is important to point out that a single integrin can regulate very different aspects of tumor progression. For example, unligated integrin $\alpha v\beta 3$ promotes tumor cell reprogramming to a stem-like cell fate, while ligated $\alpha v\beta 3$ can produce distinct cues derived from the ECM driving cell invasion and proliferation (Figure 2).

Concluding remarks

Recent findings have demonstrated that integrins participate in the regulation of stem-cell and cancer stem-cell biology and are required for cancer progression and drug resistance (Figure 3). Further understanding of which specific integrins are required for these events, whether these integrins are interchangeable or specifically required, whether these integrins define a subset of cells that expand in response to changes in the microenvironment or whether a dynamic program allows the cells to turn on and off their expressions as well as the complete integrin “signalosome” will open up new avenues for cancer treatment (Box 2). The development of new agents to target integrins and their signals should consider the context dependency of integrin function, including 2D, 3D, and in vivo assays that better reflect the tumor microenvironment or different locales encountered during the metastatic cascade.

Box 1**The canonical integrin signaling cascade**

Integrins are the major cell surface receptors for extracellular matrix molecules, which play critical roles in a variety of biological processes. Focal adhesion kinase (FAK) is a key component of the signal transduction pathways triggered by integrins. When integrins interact with their specific ligands, they recruit FAK through their β subunit. FAK undergoes autophosphorylation that leads to its association with Src, resulting in activation of both kinases. Then, the active FAK/Src complex recruits p130CAS and paxillin that in turn recruit Crk leading to activation of RAC1, p21-activated kinase (PAK), Jun amino-terminal kinase (JNK), and nuclear factor κ B (NF κ B). Alternatively, the FAK/Src complex can recruit and activate RAP1, which in turn activates extracellular signal-regulated kinase (ERK) and mitogen-activated protein kinase (MAPK) through BRAF. The FAK/Src complex may also lead to its association with growth factor receptor-bound protein 2 (GRB2), which in turn can activate RAS leading to the activation of the RAF-MEK-ERK pathway, and PI 3-kinase (PI3K) has also been shown to bind FAK leading to activation of PI3K and its downstream effectors.

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Box 2**Outstanding questions**

- Which integrins are required to promote stemness, drug resistance, and metastasis?
- Are integrins interchangeable or specifically required?
- Do integrins define a subset of cells with stemness properties?
- Do cancer cells can turn on and off integrin expression when required?
- What is the complete integrin signalosome?

Highlights

- Integrins contribute to cancer progression via adhesion-dependent and -independent pathways.
- Specific integrins not only represent stem cell markers, but also dictate stem cell behavior.
- Integrins drive therapeutic resistance through canonical and non-canonical mechanisms.
- Integrin expression contributes to multiple steps of the metastatic cascade.

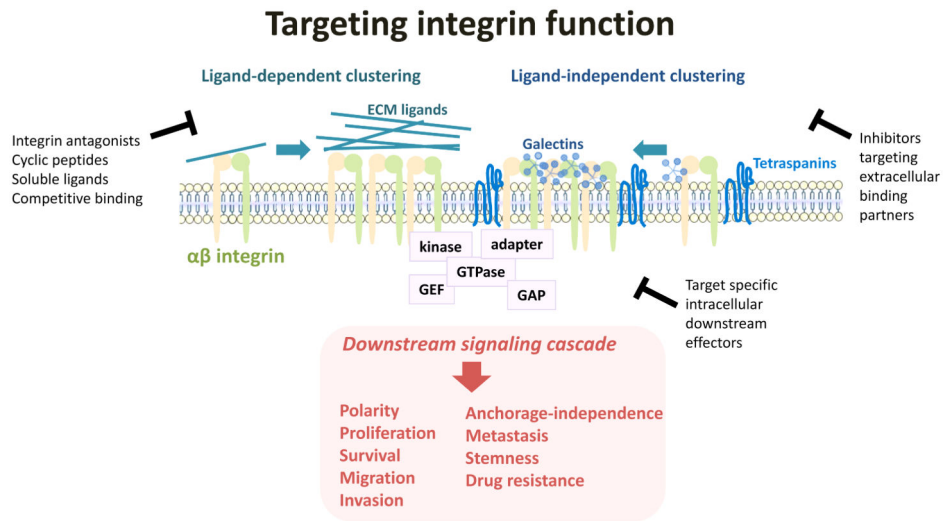


Figure 1. Integrin clustering is critical for generation of downstream signals
 Integrin function can be blocked upstream by preventing ligand binding or ligand-independent clustering, or by targeting specific downstream integrin effectors.

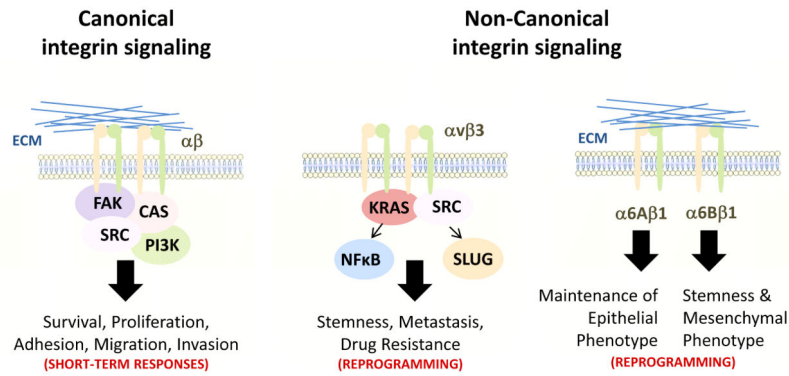


Figure 2. Canonical and non-canonical role of integrin $\alpha v\beta 3$

Integrin signaling generated by binding to extracellular matrix ligands occurs via focal adhesion complexes leading to physical changes in cellular movement and activity. In the absence of ligand binding, $\alpha v\beta 3$ integrin instead recruits KRAS and Src to drive cellular reprogramming events that lead to phenotypic changes that promote stemness, metastasis, and drug resistance. The 6A and 6B splice variants of $\alpha 6\beta 1$ integrin serve as a phenotypic switch between epithelial vs. mesenchymal states.

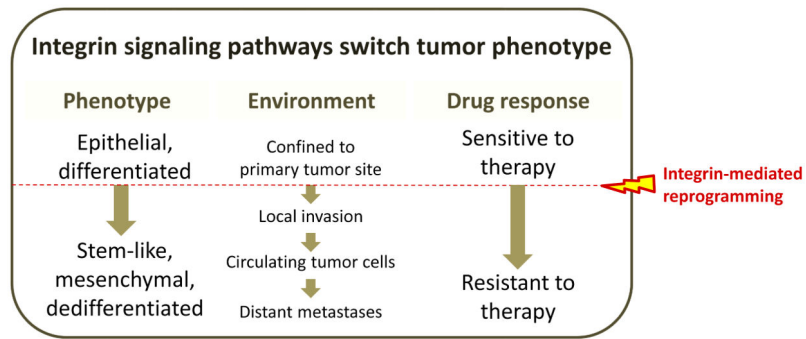


Figure 3. Reprogramming of cancer cells by integrin signaling pathways
 Integrin signaling is capable of reprogramming tumor cells to promote invasion, hematogeneous dissemination, and seeding of distant metastatic sites. Similarly, stemness and drug resistance can be triggered by changes in integrin expression and function. Understanding these events offers new therapeutic opportunities for cancer.