Submit a Manuscript: http://www.wjgnet.com/esps/ Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx DOI: 10.4254/wjh.v7.i3.600 World J Hepatol 2015 March 27; 7(3): 600-606 ISSN 1948-5182 (online) © 2015 Baishideng Publishing Group Inc. All rights reserved.

MINIREVIEWS

Occult hepatitis B virus infection and blood transfusion

Dong Hee Seo, Dong Hee Whang, Eun Young Song, Kyou Sup Han

Dong Hee Seo, Labgenomics Clinical Laboratories, Seongnamsi 463-400, South Korea

Dong Hee Whang, Department of Laboratory Medicine, Inje University College of Medicine, Seoul Paik Hospital, Seoul 100-032, South Korea

Eun Young Song, Kyou Sup Han, Department of Laboratory Medicine, Seoul National University College of Medicine, Seoul 110-744, South Korea

Author contributions: All authors contributed to this paper. Conflict-of-interest: There are no conflicts of interests.

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/4.0/

Correspondence to: Dong Hee Seo, MD, PhD, Labgenomics Clinical Laboratories, B-6F, 700, Daewangpangyo-ro, Bundang-gu, Seongnam-si 463-400, Gyeonggi-do,

South Korea. seo2023@nate.com Telephone: +82-31-6280730 Fax: +82-31-6280701 Received: August 23, 2014

Peer-review started: August 24, 2014 First decision: November 3, 2014 Revised: November 29, 2014 Accepted: December 16, 2014 Article in press: December 16, 2014 Published online: March 27, 2015

Abstract

Transfusion-transmitted infections including hepatitis B virus (HBV) have been a major concern in transfusion medicine. Implementation of HBV nucleic acid testing (NAT) has revealed occult HBV infection (OBI) in blood donors. In the mid-1980s, hepatitis B core antibody (HBc) testing was introduced to screen blood donors in HBV non-endemic countries to prevent transmission of non-A and non-B hepatitis. That test remains in use for preventing of potential transmission of HBV from hepatitis B surface antigen (HBsAg)-negative blood donors, even though anti-hepatitis C virus tests

have been introduced. Studies of anti-HBc-positive donors have revealed an HBV DNA positivity rate of 0%-15%. As of 2012, 30 countries have implemented HBV NAT. The prevalence of OBI in blood donors was estimated to be 8.55 per 1 million donations, according to a 2008 international survey. OBI is transmissible by blood transfusion. The clinical outcome of occult HBV transmission primarily depends on recipient immune status and the number of HBV DNA copies present in the blood products. The presence of donor anti-HBs reduces the risk of HBV infection by approximately five-fold. The risk of HBV transmission may be lower in endemic areas than in non-endemic areas, because most recipients have already been exposed to HBV. Blood safety for HBV, including OBI, has substantially improved, but the possibility for OBI transmission remains.

Key words: Occult hepatitis B infection; Transfusion; Antihepatitis B core antibody; Nucleic acid testing; Blood service

© **The Author(s) 2015.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Hepatitis B surface antigen negative but hepatitis B virus (HBV) DNA positive blood products can evoke hepatitis in blood recipients. Anti-hepatitis B core and HBV nucleic acid testing screening tests are necessary to prevent occult HBV infection transmission by transfusion. Anti-HBs antibody in donors and recipients can protect against hepatitis B infection.

Seo DH, Whang DH, Song EY, Han KS. Occult hepatitis B virus infection and blood transfusion. *World J Hepatol* 2015; 7(3): 600-606 Available from: URL: http://www.wjgnet.com/1948-5182/full/v7/i3/600.htm DOI: http://dx.doi.org/10.4254/wjh.v7.i3.600

INTRODUCTION

Hepatitis B virus (HBV) infection via blood transfusion



WJH | www.wjgnet.com 600 March 27, 2015 | Volume 7 | Issue 3 |

is a major concern in transfusion medicine^[1-4]. Screening tests for hepatitis B surface antigens (HBsAg) and anti-hepatitis B core (HBc) antibodies detect HBV transmissible blood and prevent recipient HBV infection. After the introduction of HBV nucleic acid tests (NAT) in blood donor screening, the residual risk of HBV infection by transfusion decreased^[5,6]. Implementation of this test revealed occult hepatitis B virus infection (OBI) in blood donors. OBI is defined as the presence of HBV DNA in the liver (with detectable or undetectable HBV DNA in the serum) of individuals who tested negative for HBsAg^[7]. The amount of viral DNA in the serum is typically very low in cases of true OBI. Because testing liver tissue is not always practical or possible, OBI is often diagnosed through serum HBV DNA and viral marker tests^[8,9].

A positive OBI test may be found in blood donors as a result of various clinical conditions, including: (1) the incubation period of acute infections; (2) the tail-end stage of chronic hepatitis B; (3) low-level viral replication after recovery from hepatitis; and (4) escape mutants not detected by current HBsAg tests^[10,11]. HBV transmission by blood transfusion from an OBI donor was first reported in 1978^[12]. An increasing number of studies on OBI infectivity of blood products have recently been published^[13,14]. In this review, we summarize the role of blood screening tests for HBV infections and update the known risks of OBI transfusion transmission.

ANTI-HBC ANTIBODY

Hepatitis B core antigen (HBcAg) appears in hepatocytes within 2 wk after HBV infection; infectious viremia including HBsAg and polymerase are present in the blood after 3 wk. Anti-HBc IgG forms during the recovery phase of infection and is persistent for life, thus, the presence of this antibody in blood indicates past HBV infection[15]. The analytic sensitivity of HBsAg tests in the 1980s was lower than that of current assays. In 1983, Nath et al[16] found that 1 of 16 samples with anti-HBc in the absence of anti-HBs was found to have HBsAg when tested with a more sensitive test. Therefore, additional screening for HBV and surrogate tests for non-A, non-B hepatitis were necessary in the 1980s until the anti-hepatitis C virus (HCV) antibody test became available^[17]. The anti-HBc test was introduced in the mid-1980s for screening of blood donors in HBV non-endemic countries, such as the United States. Even after the introduction of the anti-HCV test in the early 1990s, the anti-HBc test continues to be used for donor screening in many countries to prevent potential transmission of HBV from HBsAg-negative donors^[18,19]. Several studies have reported effective screening of blood for anti-HBc[20,21]. However, HBV endemic countries were unable to implement anti-HBc screening because many blood products would be discarded due to positive screening tests even though most of the blood would be safe for transfusion. Cases of posttransfusion hepatitis B from positive carrier blood and posttransfusion fulminant hepatitis B from blood containing precoredefective HBV mutants have been reported in Norway and Japan, respectively, countries that did not screen donors for anti-HBc $^{[22,23]}$. In 1989, Japan introduced anti-HBc testing with a modified algorithm in which anti-HBc-reactive blood with titers < 1:32 or \geq 1:32 with anti-HBs \geq 200 mIU/mL were used for transfusion $^{[24]}$.

Anti-HBc prevalence is related to regional hepatitis B prevalence, and both are typically proportional to one another. The prevalence rates of anti-HBc in blood donors in the United States are 0.23%^[25]; United Kingdom, $0.56\%^{[21]}$; Denmark, $0.70\%^{[26]}$; Japan, $1.1\%^{[27]}$; Germany, $1.88\%^{[28]}$; Italy, $4.85\%^{[29]}$; India, $10.82\%^{[30]}$; South Korea, $13.5\%^{[31]}$; Egypt, $14.2\%^{[32]}$; Greece, $14.9\%^{[33]}$; and Pakistan, $17.28\%^{[34]}$ (Figure 1). O'Brien et al^[35] reported 5585 (1.13%) anti-HBc repeat-reactive blood donors among 493344 blood donors in Canada, of which 29 (0.52%) were HBsAgnegative but HBV DNA-positive^[35]. The anti-HBc test lacks specificity and reactivity of the test reagents varies by manufacturer. Therefore, comparison of anti-HBc positivity should be conducted with caution; it is better to describe general features rather than directly comparing studies. Efforts to improve test specificity have included the addition of reductants such as dithiothreitol and cysteine[36]. High donor HBsAg antibody (anti-HBs) levels (> 100 mIU/mL) are assumed to be putatively protective against the transfusion transmission of HBV^[37]. Anti-HBc only or isolated anti-HBc are defined as anti-HBc positive without HBsAg and anti-HBs. An HBV DNA positivity of 0%-15% among those donors positive for anti-HBc only was reported in studies performed in Greece, China, Japan, and Germany^[10]. Therefore, the presence of anti-HBc only does not necessarily indicate active viral replication and transmission potential.

NAT IN BLOOD SERVICE

Improved molecular methods for the detection of viral nucleic acids made it possible to introduce NAT for donor screening in the late 1990s^[38]. By 1997, several countries in Europe had initiated voluntary screening of pooled plasma donations using NAT, and a directive was issued by the EU requiring HCV RNA testing for all plasma intended for fractionation in Europe by July 1, 1999. Routine NAT for HBV was first introduced in German blood transfusion services in January of 1997^[39]. The United States began screening source plasma pools for HCV and human immunodeficiency virus (HIV)-1 RNA in early 1998 under the Food and Drug Administration's Investigational New Drug (IND) program. Currently, more than 90% of whole blood and nearly all source plasma are screened for HCV and HIV by NAT. NAT for blood screening is typically conducted using a multiplex polymerase chain reaction

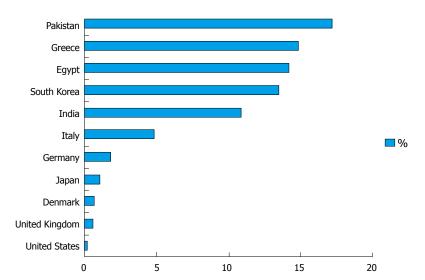


Figure 1 Prevalence of anti-hepatitis B core in blood donors.

method that simultaneously detects the presence of HIV, HCV and HBV^[40]. And a multiplex reaction positive case requires discrimination testing^[41]. NAT can be performed on pooled samples to reduce running costs^[42]. In pooling test system, primary NAT positive cases require a secondary confirmation procedure for identification of positivity of individual donation samples. The American Red Cross implemented automated triplex NAT for HIV, HCV, and HBV in June of 2009^[25], Japan implemented NAT in November 1997 and all donations have been screened since October 1999. Japanese Red Cross Blood Centers serologically screened 500 samples that were pooled and tested for HBV, HCV and HIV-1. To increase HBV detection sensitivity in Japan, the sample pooling size was reduced to 50 in 2000, and 20 in 2004^[43].

As of 2012, 30 countries have implemented HBV NAT^[44,45], as shown in Table 1. According to international survey results, there were a total of 170 HBV NAT-only positive donors among 19887649 blood donations in 2008, and the prevalence of OBI among blood donors was an estimated 8.55 per 1 million donations^[44]. In Taiwan, 8 (0.13%) HBV NATonly positive donors were identified among 5,973 random donor samples^[46]. In China, 22 among 165371 HBsAg-negative plasma samples were identified as OBI-positive; their alanine aminotransferase levels were normal and their viral loads low, with a median of 14 IU/mL^[47]. In Iran, 4% of 1000 healthy blood donors were anti-HBc- and OBI-positive, and 8.23% of 11,240 volunteer donors were anti-HBc positive and OBI-positive in Mexico^[48,49]. To detect OBI, the HBV DNA test is substantial, and minipool NAT can be used to reduce the cost of testing in developing countries. However, in HBV endemic areas, minipool NAT can result in many primary positive cases, which means that many blood products must be retained until the positivity of individual sample is resolved through a secondary confirmation test. Therefore, it is better to implement NAT for individual-donations rather than minipools for NAT in HBV endemic areas^[50-53]. HBV replicates more slowly than HIV or HCV, and

its doubling time during the ramp-up phase was estimated to be 2.56 d by Biswas $et\ a^{[54]}$. Therefore, minipool NAT for HBV detection is less effective than for HIV or HCV detection. Each country should develop its own blood screening strategy based on HBV prevalence, yields of infectious units by different screening methods and cost-effectiveness of testing methods.

OBI TRANSMISSION BY BLOOD TRANSFUSION

Occult HBV is transmissible by blood transfusion, although the transmission rate is considered to be very low. The clinical outcome of OBI transmission mainly depends on the immune status and copies of HBV DNA in blood products of the recipient. A lookback program by the Japanese Red Cross showed that window period-derived blood components evoked 50% (11/22) seroconversion in recipients, but tailend chronic hepatitis B infections caused only 3% (1/33) seroconversion in recipients^[55]. Satake et al^[55] concluded that the blood infectivity rate during the window period was 10-fold higher than the transmission rates from occult carriers with low-titer anti-HBc. In Canada, a look-back study identified 9.7% anti-HBcpositive recipients and 4 HBV DNA-positive, HBsAgnegative, anti-HBc-positive donors[35]. However, hepatitis cases were reported to have occurred as the result of transfusion of anti-HBc-positive, anti-HBspositive (12 IU/L), HBV DNA-positive (180 IU/mL) blood product^[56].

A study on blood component infectivity by Allain *et al*^[14] reported that the presence of anti-HBs in donors reduces the risk of HBV infection by approximately five-fold, while therapeutic fresh frozen plasma over platelet concentrate increases the risk by approximately three-fold by logistic regression analysis^[14]. A case of OBI transmission by plasma has been reported, but not by red blood cells^[57,58]. Because the amount of plasma containing viral particles is

Table 1 Introduction of hepatitis B virus nucleic acid testing in donor screening

Year	Country
1997	Germany
1999	Austria, Japan
2004	Singapore, Spain
2005	Poland, France (OT + army), South Africa
2006	Greece, Italy, Portugal, Thailand
2007	Hong Kong, Kuwait, Malaysia, New Zealand, Slovenia,
	Switzerland
2008	Finland, Israel, Latvia, Netherlands, Taiwan
2009	United States, Denmark, Ireland, United Kingdom
	(England and Wales)
2010	Australia, United Kingdom (Scotland), Canada
2012	South Korea

very small in RBCs, they are considered to be less infective than plasma products. In a look-back study in Taiwan, Su *et al*^[59] identified 12 (0.11%) OBI-positive donors among 10824 repository samples; they also identified no post-transfusion hepatitis cases among the recipients. They suggested that the risk of HBV transmission is lower in hyperendemic areas such as Taiwan than in non-endemic areas, because most recipients have already experienced an HBV infection. In a look-back study in Hyogo-prefecture, one of 12 recipients was diagnosed with post-transfusion hepatitis B. Of the remaining 11 recipients, 7 were lost to follow-up, and 4 were negative for HBV^[60].

CONCLUSION

The life-saving role of blood transfusion makes it an essential component of modern medical practice. However, blood used for transfusion is not always free of transmissible diseases. According to the 2008 World Health Organization Global Database on Blood Safety, approximately 92 million blood donations are collected worldwide each year. Of these donations, 48% are collected in high-income countries. Blood products from these countries are screened for HBV, HCV, and HIV. However, 39 countries do not routinely test blood donations for transfusion-transmissible infections^[61]. HBsAg-positive blood products, including OBI, could be transfused to patients in these counties more frequently than in the countries that screen blood. Although the prevalence of OBI in blood donors differs by country, a 2008 international survey estimated it to be 8.55 per 1 million donations. To reduce the risk of HBV transmission, HBsAg testing was introduced, followed by anti-HBc and HBV NAT in countries where additional testing was feasible. These approaches were also effective for the prevention of transfusiontransmission of OBI. A modified algorithm for anti-HBc screening of blood donors was implemented in intermediate HBV endemic areas, such as Japan, where the test alone could not be introduced because of the resultant high blood discard rates. Unlike anti-HBc screening, HBV NAT can detect infections during

window periods. Therefore, despite higher costs, HBV NAT is more suitable for screening in areas with endemic HBV.

Although OBI infectivity depends on recipient immunity and blood product type, it is transmissible by transfusion. One study observed higher genetic diversity in occult HBV genotype B and C strains from South East Asian blood donors^[62]. Epigenetic factors have also been identified in HBV cccDNA molecules, and studies to understand OBI immunopathogenesis are currently underway^[63-68]. Even with HBV NAT screening, there is a risk of false-negative results. NAT using sample pooling systems in particular cannot detect low-level viremia. Pathogen inactivation technologies that destroy viral DNA or RNA in blood products using chemical agents and ultraviolet illumination have been introduced as alternatives in blood services^[69,70]. HBV vaccination of potential recipients is also an important preventive measure. In conclusion, following the implementation of anti-HBc and HBV NAT screening, blood safety for HBV including OBI has improved substantially, but the potential for OBI transmission remains.

REFERENCES

- Korelitz JJ, Busch MP, Kleinman SH, Williams AE, Gilcher RO, Ownby HE, Schreiber GB. A method for estimating hepatitis B virus incidence rates in volunteer blood donors. National Heart, Lung, and Blood Institute Retrovirus Epidemiology Donor Study. *Transfusion* 1997; 37: 634-640 [PMID: 9191825]
- Wang JT, Lee CZ, Chen PJ, Wang TH, Chen DS. Transfusion-transmitted HBV infection in an endemic area: the necessity of more sensitive screening for HBV carriers. *Transfusion* 2002; 42: 1592-1597 [PMID: 12473140]
- 3 Candotti D, Allain JP. Transfusion-transmitted hepatitis B virus infection. *J Hepatol* 2009; 51: 798-809 [PMID: 19615780 DOI: 10.1016/j.jhep.2009.05.020]
- Perkins HA, Busch MP. Transfusion-associated infections: 50 years of relentless challenges and remarkable progress. Transfusion 2010; 50: 2080-2099 [PMID: 20738828 DOI: 10.1111/j.1537-2995.2010.02851.x]
- 5 Roth WK, Weber M, Petersen D, Drosten C, Buhr S, Sireis W, Weichert W, Hedges D, Seifried E. NAT for HBV and anti-HBc testing increase blood safety. *Transfusion* 2002; 42: 869-875 [PMID: 12375659]
- 6 Cable R, Lelie N, Bird A. Reduction of the risk of transfusion-transmitted viral infection by nucleic acid amplification testing in the Western Cape of South Africa: a 5-year review. Vox Sang 2013; 104: 93-99 [PMID: 22924987 DOI: 10.1111/j.1423-0410.2012.01640.x]
- Raimondo G, Allain JP, Brunetto MR, Buendia MA, Chen DS, Colombo M, Craxì A, Donato F, Ferrari C, Gaeta GB, Gerlich WH, Levrero M, Locarnini S, Michalak T, Mondelli MU, Pawlotsky JM, Pollicino T, Prati D, Puoti M, Samuel D, Shouval D, Smedile A, Squadrito G, Trépo C, Villa E, Will H, Zanetti AR, Zoulim F. Statements from the Taormina expert meeting on occult hepatitis B virus infection. *J Hepatol* 2008; 49: 652-657 [PMID: 18715666 DOI: 10.1016/j.jhep.2008.07.014]
- Brojer E, Grabarczyk P, Liszewski G, Mikulska M, Allain JP, Letowska M. Characterization of HBV DNA+/HBsAg- blood donors in Poland identified by triplex NAT. *Hepatology* 2006; 44: 1666-1674 [PMID: 17133474 DOI: 10.1002/hep.21413]
- 9 Hollinger FB. Hepatitis B virus infection and transfusion medicine: science and the occult. *Transfusion* 2008; 48: 1001-1026 [PMID:



- 18454738 DOI: 10.1111/j.1537-2995.2008.01701.x]
- 10 Allain JP. Occult hepatitis B virus infection: implications in transfusion. Vox Sang 2004; 86: 83-91 [PMID: 15023176]
- 11 Gerlich WH, Wagner FF, Chudy M, Harritshoj LH, Lattermann A, Wienzek S, Glebe D, Saniewski M, Schüttler CG, Wend UC, Willems WR, Bauerfeind U, Jork C, Bein G, Platz P, Ullum H, Dickmeiss E. HBsAg non-reactive HBV infection in blood donors: transmission and pathogenicity. *J Med Virol* 2007; 79: S32-S36 [DOI: 10.1002/jmv.20963]
- Hoofnagle JH, Seeff LB, Bales ZB, Zimmerman HJ. Type B hepatitis after transfusion with blood containing antibody to hepatitis B core antigen. N Engl J Med 1978; 298: 1379-1383 [PMID: 652005 DOI: 10.1056/NEJM197806222982502]
- 13 Raimondo G, Caccamo G, Filomia R, Pollicino T. Occult HBV infection. *Semin Immunopathol* 2013; 35: 39-52 [PMID: 22829332 DOI: 10.1007/s00281-012-0327-7]
- 14 Allain JP, Mihaljevic I, Gonzalez-Fraile MI, Gubbe K, Holm-Harritshøj L, Garcia JM, Brojer E, Erikstrup C, Saniewski M, Wernish L, Bianco L, Ullum H, Candotti D, Lelie N, Gerlich WH, Chudy M. Infectivity of blood products from donors with occult hepatitis B virus infection. *Transfusion* 2013; 53: 1405-1415 [PMID: 23362802 DOI: 10.1111/trf.12096]
- 15 Koziel MJ, Siddiqui A. Hepatitis B virus and hepatitis delta virus. In: Mandell, Douglas, and Bennett's principles and practice of infectious diseases. 6th ed. Philadelphia: Elsevier, 2005: 1864-1890
- 16 Nath N, Pielech M, Dodd RY. Hepatitis-associated markers in the American Red Cross blood donor population. V. Prevalence of antibodies to core antigen in three blood services regions. *Vox Sang* 1983; 44: 312-318 [PMID: 6858079]
- 17 Koziol DE, Holland PV, Alling DW, Melpolder JC, Solomon RE, Purcell RH, Hudson LM, Shoup FJ, Krakauer H, Alter HJ. Antibody to hepatitis B core antigen as a paradoxical marker for non-A, non-B hepatitis agents in donated blood. *Ann Intern Med* 1986; 104: 488-495 [PMID: 3006567]
- 18 Kuhns MC, Kleinman SH, McNamara AL, Rawal B, Glynn S, Busch MP. Lack of correlation between HBsAg and HBV DNA levels in blood donors who test positive for HBsAg and anti-HBc: implications for future HBV screening policy. *Transfusion* 2004; 44: 1332-1339 [PMID: 15318857]
- 19 O'Brien SF, Yi QL, Fan W, Scalia V, Kleinman SH, Vamvakas EC. Current incidence and estimated residual risk of transfusion-transmitted infections in donations made to Canadian Blood Services. *Transfusion* 2007; 47: 316-325 [PMID: 17302779 DOI: 10.1111/j.1537-2995.2007.01108.x]
- 20 Mosley JW, Stevens CE, Aach RD, Hollinger FB, Mimms LT, Solomon LR, Barbosa LH, Nemo GJ. Donor screening for antibody to hepatitis B core antigen and hepatitis B virus infection in transfusion recipients. *Transfusion* 1995; 35: 5-12 [PMID: 7998069]
- 21 Allain JP, Hewitt PE, Tedder RS, Williamson LM. Evidence that anti-HBc but not HBV DNA testing may prevent some HBV transmission by transfusion. *Br J Haematol* 1999; 107: 186-195 [PMID: 10520040]
- 22 Larsen J, Hetland G, Skaug K. Posttransfusion hepatitis B transmitted by blood from a hepatitis B surface antigen-negative hepatitis B virus carrier. *Transfusion* 1990; 30: 431-432 [PMID: 2360234]
- 23 Kojima M, Shimizu M, Tsuchimochi T, Koyasu M, Tanaka S, Iizuka H, Tanaka T, Okamoto H, Tsuda F, Miyakawa Y. Posttransfusion fulminant hepatitis B associated with precore-defective HBV mutants. Vox Sang 1991; 60: 34-39 [PMID: 2053327]
- 24 Tani Y, Aso H, Matsukura H, Tadokoro K, Tamori A, Nishiguchi S, Yoshizawa H, Shibata H. Significant background rates of HBV and HCV infections in patients and risks of blood transfusion from donors with low anti-HBc titres or high anti-HBc titres with high anti-HBs titres in Japan: a prospective, individual NAT study of transfusion-transmitted HBV, HCV and HIV infections. *Vox Sang* 2012; 102: 285-293 [PMID: 22082342 DOI: 10.1111/j.1423-0410.2011.01561.x]
- 25 Stramer SL, Zou S, Notari EP, Foster GA, Krysztof DE, Musavi

- F, Dodd RY. Blood donation screening for hepatitis B virus markers in the era of nucleic acid testing: are all tests of value? *Transfusion* 2012; **52**: 440-446 [PMID: 21810101 DOI: 10.1111/j.1537-2995.2011.0328.x]
- 26 Christensen PB, Titlestad IL, Homburg KM, Georgsen J, Kristensen T. Hepatitis B core antibodies in Danish blood donors: a surrogate marker of risk behaviour. Vox Sang 2001; 81: 222-227 [PMID: 11903997]
- Yotsuyanagi H, Yasuda K, Moriya K, Shintani Y, Fujie H, Tsutsumi T, Nojiri N, Juji T, Hoshino H, Shimoda K, Hino K, Kimura S, Iino S, Koike K. Frequent presence of HBV in the sera of HBsAg-negative, anti-HBc-positive blood donors. *Transfusion* 2001; 41: 1093-1099 [PMID: 11552064]
- Schmidt M, Nübling CM, Scheiblauer H, Chudy M, Walch LA, Seifried E, Roth WK, Hourfar MK. Anti-HBc screening of blood donors: a comparison of nine anti-HBc tests. *Vox Sang* 2006; 91: 237-243 [PMID: 16958836 DOI: 10.1111/j.1423-0410.2006.00818.x]
- 29 Manzini P, Girotto M, Borsotti R, Giachino O, Guaschino R, Lanteri M, Testa D, Ghiazza P, Vacchini M, Danielle F, Pizzi A, Valpreda C, Castagno F, Curti F, Magistroni P, Abate ML, Smedile A, Rizzetto M. Italian blood donors with anti-HBc and occult hepatitis B virus infection. *Haematologica* 2007; 92: 1664-1670 [PMID: 18055990 DOI: 10.3324/haematol.11224]
- 30 Chaudhuri V, Nanu A, Panda SK, Chand P. Evaluation of serologic screening of blood donors in India reveals a lack of correlation between anti-HBc titer and PCR-amplified HBV DNA. *Transfusion* 2003; 43: 1442-1448 [PMID: 14507277]
- 31 Seo DH, Whang DH, Song EY, Kim HS, Park Q. Prevalence of antibodies to hepatitis B core antigen and occult hepatitis B virus infections in Korean blood donors. *Transfusion* 2011; 51: 1840-1846 [PMID: 21332731 DOI: 10.1111/j.1537-2995.2010.03056.x]
- 32 Said ZN, Sayed MH, Salama II, Aboel-Magd EK, Mahmoud MH, Setouhy ME, Mouftah F, Azzab MB, Goubran H, Bassili A, Esmat GE. Occult hepatitis B virus infection among Egyptian blood donors. World J Hepatol 2013; 5: 64-73 [PMID: 23646231 DOI: 10.4254/wjh.v5.i2.64]
- 33 Zervou EK, Dalekos GN, Boumba DS, Tsianos EV. Value of anti-HBc screening of blood donors for prevention of HBV infection: results of a 3-year prospective study in Northwestern Greece. *Transfusion* 2001; 41: 652-658 [PMID: 11346702]
- 34 Bhatti FA, Ullah Z, Salamat N, Ayub M, Ghani E. Anti-hepatits B core antigen testing, viral markers, and occult hepatitis B virus infection in Pakistani blood donors: implications for transfusion practice. *Transfusion* 2007; 47: 74-79 [PMID: 17207233 DOI: 10.1111/j.1537-2995.2007.01066.x]
- O'Brien SF, Fearon MA, Yi QL, Fan W, Scalia V, Muntz IR, Vamvakas EC. Hepatitis B virus DNA-positive, hepatitis B surface antigen-negative blood donations intercepted by anti-hepatitis B core antigen testing: the Canadian Blood Services experience. *Transfusion* 2007; 47: 1809-1815 [PMID: 17880605 DOI: 10.1111/j.1537-2995.2007.01396.x]
- 36 Cheng Y, Dubovoy N, Hayes-Rogers ME, Stewart J, Shah D. Detection of IgM to hepatitis B core antigen in a reductant containing, chemiluminescence assay. *J Immunol Methods* 1999; 230: 29-35 [PMID: 10594351]
- 37 Reesink HW, Allain JP. Management of donors and blood products reactive for hepatitis B virus DNA. *Vox Sang* 2006; 91: 281 [PMID: 17105601 DOI: 10.1111/j.1423-0410.2006.00843.x]
- 38 Candotti D, Allain JP. Molecular virology in transfusion medicine laboratory. *Blood Transfus* 2013; 11: 203-216 [PMID: 23356973 DOI: 10.2450/2012.0219-12]
- Roth WK, Weber M, Seifried E. Feasibility and efficacy of routine PCR screening of blood donations for hepatitis C virus, hepatitis B virus, and HIV-1 in a blood-bank setting. *Lancet* 1999; 353: 359-363 [PMID: 9950441]
- 40 Margaritis AR, Brown SM, Seed CR, Kiely P, D'Agostino B, Keller AJ. Comparison of two automated nucleic acid testing systems for simultaneous detection of human immunodeficiency virus and hepatitis C virus RNA and hepatitis B virus DNA. Transfusion 2007; 47: 1783-1793 [PMID: 17880602 DOI: 10.1111/



- j.1537-2995.2007.01343.x]
- 41 Charlewood R, Flanagan P. Ultrio and Ultrio Plus non-discriminating reactives: false reactives or not? *Vox Sang* 2013; 104: 7-11 [PMID: 22765287 DOI: 10.1111/j.1423-0410.2012.01624.x]
- 42 Kleinman SH, Strong DM, Tegtmeier GG, Holland PV, Gorlin JB, Cousins C, Chiacchierini RP, Pietrelli LA. Hepatitis B virus (HBV) DNA screening of blood donations in minipools with the COBAS AmpliScreen HBV test. *Transfusion* 2005; 45: 1247-1257 [PMID: 16078909 DOI: 10.1111/j.1537-2995.2005.00198.x]
- 43 Reesink HW, Engelfriet CP, Henn G, Mayr WR, Delage G, Bernier F, Krusius T, Assal A, Gallian P, Corbi C, Morel P, David B, De Micco P, Murokawa H, Yugi H, Hino S, Tadokoro K, Flesland O, Brojer E, Letowska M, Olim G, Nascimento F, Gonçalves H, Castro L, Morais M, Stezinar SL, Alvarez M, Sauleda S, González R, Niederhauser C, Stolz M, Allain JP, Owusu-Ofori S, Eglin R, Stramer S, Busch M, Strong DM, Epstein J, Biswas R. Occult hepatitis B infection in blood donors. Vox Sang 2008; 94: 153-166 [PMID: 18205672 DOI: 10.1111/j.1423-0410.2008.01017.x]
- Roth WK, Busch MP, Schuller A, Ismay S, Cheng A, Seed CR, Jungbauer C, Minsk PM, Sondag-Thull D, Wendel S, Levi JE, Fearon M, Delage G, Xie Y, Jukic I, Turek P, Ullum H, Tefanova V, Tilk M, Reimal R, Castren J, Naukkarinen M, Assal A, Jork C, Hourfar MK, Michel P, Offergeld R, Pichl L, Schmidt M, Schottstedt V, Seifried E, Wagner F, Weber-Schehl M, Politis C, Lin CK, Tsoi WC, O'Riordan J, Gottreich A, Shinar E, Yahalom V, Velati C, Satake M, Sanad N, Sisene I, Bon AH, Koppelmann M, Flanagan P, Flesland O, Brojer E, Letowska M, Nascimento F, Zhiburt E, Chua SS, Teo D, Stezinar SL, Vermeulen M, Reddy R, Park Q, Castro E, Eiras A, Gonzales Fraile I, Torres P, Ekermo B, Niederhauser C, Chen H, Oota S, Brant LJ, Eglin R, Jarvis L, Mohabir L, Brodsky J, Foster G, Jennings C, Notari E, Stramer S, Kessler D, Hillyer C, Kamel H, Katz L, Taylor C, Panzer S, Reesink HW. International survey on NAT testing of blood donations: expanding implementation and yield from 1999 to 2009. Vox Sang 2012; 102: 82-90 [PMID: 21933190 DOI: 10.1111/ j.1423-0410.2011.01506.x]
- 45 Kang JW, Kwon SY, Seo YI, Lee MK, Huh K, Park SJ, Kim MH, Cho NS. HBV NAT implementation in Korea: 1 year experience. Vox Sang 2014; 107: 150
- 46 Lin KT, Chang CL, Tsai MH, Lin KS, Saldanha J, Hung CM. Detection and identification of occult HBV in blood donors in Taiwan using a commercial, multiplex, multi-dye nucleic acid amplification technology screening test. *Vox Sang* 2014; 106: 103-110 [PMID: 23909571 DOI: 10.1111/vox.12075]
- 47 Zheng X, Ye X, Zhang L, Wang W, Shuai L, Wang A, Zeng J, Candotti D, Allain JP, Li C. Characterization of occult hepatitis B virus infection from blood donors in China. *J Clin Microbiol* 2011; 49: 1730-1737 [PMID: 21411575 DOI: 10.1128/JCM.00145-11]
- 48 Vaezjalali M, Rashidpour S, Rezaee H, Hajibeigi B, Zeidi M, Gachkar L, Aghamohamad S, Najafi R, Goudarzi H. Hepatitis B viral DNA among HBs antigen negative healthy blood donors. Hepat Mon 2013; 13: e6590 [PMID: 23675384 DOI: 10.5812/hepatmon.6590]
- 49 García-Montalvo BM, Farfán-Ale JA, Acosta-Viana KY, Puerto-Manzano FI. Hepatitis B virus DNA in blood donors with anti-HBc as a possible indicator of active hepatitis B virus infection in Yucatan, Mexico. *Transfus Med* 2005; 15: 371-378 [PMID: 16202051 DOI: 10.1111/j.1365-3148.2005.00610.x]
- 50 Assal A, Barlet V, Deschaseaux M, Dupont I, Gallian P, Guitton C, Morel P, David B, De Micco P. Comparison of the analytical and operational performance of two viral nucleic acid test blood screening systems: Procleix Tigris and cobas s 201. Transfusion 2009; 49: 289-300 [PMID: 19000230 DOI: 10.1111/j.1537-2995.2008.01965.x]
- 51 Vermeulen M, Lelie N, Sykes W, Crookes R, Swanevelder J, Gaggia L, Le Roux M, Kuun E, Gulube S, Reddy R. Impact of individual-donation nucleic acid testing on risk of human immunodeficiency virus, hepatitis B virus, and hepatitis C virus transmission by blood transfusion in South Africa. *Transfusion* 2009; 49: 1115-1125 [PMID: 19309474 DOI: 10.1111/j.1537-2995.2009.02110.x]

- Yang MH, Li L, Hung YS, Hung CS, Allain JP, Lin KS, Tsai SJ. The efficacy of individual-donation and minipool testing to detect low-level hepatitis B virus DNA in Taiwan. *Transfusion* 2010; 50: 65-74 [PMID: 19709394 DOI: 10.1111/j.1537-2995.2009.02357.x]
- Arora S, Doda V, Kirtania T. Sensitivity of individual donor nucleic acid testing (NAT) for the detection of hepatitis B infection by studying diluted NAT yield samples. *Blood Transfus* 2014 Oct 23; Epub ahead of print [PMID: 25369612 DOI: 10.2450/2014.0048-14]
- 54 Biswas R, Tabor E, Hsia CC, Wright DJ, Laycock ME, Fiebig EW, Peddada L, Smith R, Schreiber GB, Epstein JS, Nemo GJ, Busch MP. Comparative sensitivity of HBV NATs and HBsAg assays for detection of acute HBV infection. *Transfusion* 2003; 43: 788-798 [PMID: 12757531]
- 55 Satake M, Taira R, Yugi H, Hino S, Kanemitsu K, Ikeda H, Tadokoro K. Infectivity of blood components with low hepatitis B virus DNA levels identified in a lookback program. Transfusion 2007; 47: 1197-1205 [PMID: 17581154 DOI: 10.1111/j.1537-2995.2007.01276.x]
- 56 Levicnik-Stezinar S, Rahne-Potokar U, Candotti D, Lelie N, Allain JP. Anti-HBs positive occult hepatitis B virus carrier blood infectious in two transfusion recipients. *J Hepatol* 2008; 48: 1022-1025 [PMID: 18436328 DOI: 10.1016/j.jhep.2008.02.016]
- 57 Coppola N, Loquercio G, Tonziello G, Azzaro R, Pisaturo M, Di Costanzo G, Starace M, Pasquale G, Cacciapuoti C, Petruzziello A. HBV transmission from an occult carrier with five mutations in the major hydrophilic region of HBsAg to an immunosuppressed plasma recipient. *J Clin Virol* 2013; 58: 315-317 [PMID: 23856167 DOI: 10.1016/j.jcv.2013.06.020]
- Furuta RA, Kondo Y, Saito T, Tomita M, Oka K, Kishimoto Y, Tani Y, Shibata T. Transfusions of red blood cells from an occult hepatitis B virus carrier without apparent signs of transfusion-transmitted hepatitis B infection. *Transfus Med* 2008; 18: 379-381 [PMID: 19140823 DOI: 10.1111/j.1365-3148.2008.00898.x]
- 59 Su TH, Chen PJ, Chen TC, Cheng HR, Li L, Lin KS, Kao JH, Chen DS, Liu CJ. The clinical significance of occult hepatitis B transfusion in Taiwan--a look-back study. *Transfus Med* 2011; 21: 33-41 [PMID: 20726954 DOI: 10.1111/j.1365-3148.2010.01036.x]
- 60 Bouike Y, Imoto S, Mabuchi O, Kokubunji A, Kai S, Okada M, Taniguchi R, Momose S, Uchida S, Nishio H. Infectivity of HBV DNA positive donations identified in look-back studies in Hyogo-Prefecture, Japan. *Transfus Med* 2011; 21: 107-115 [PMID: 21118317 DOI: 10.1111/j.1365-3148.2010.01057.x]
- 61 World Health Organization. Blood safety: Key global fact and figures in 2011. Available from: URL: http://www.who.int/ worldblooddonorday/media/who_blood_safety_factsheet_2011.pdf
- 62 Candotti D, Lin CK, Belkhiri D, Sakuldamrongpanich T, Biswas S, Lin S, Teo D, Ayob Y, Allain JP. Occult hepatitis B infection in blood donors from South East Asia: molecular characterisation and potential mechanisms of occurrence. *Gut* 2012; 61: 1744-1753 [PMID: 22267593 DOI: 10.1136/gutjnl-2011-301281]
- 63 Mason AL, Xu L, Guo L, Kuhns M, Perrillo RP. Molecular basis for persistent hepatitis B virus infection in the liver after clearance of serum hepatitis B surface antigen. *Hepatology* 1998; 27: 1736-1742 [PMID: 9620351]
- 64 Zoulim F. New insight on hepatitis B virus persistence from the study of intrahepatic viral cccDNA. *J Hepatol* 2005; 42: 302-308 [PMID: 15710212]
- 65 Lucifora J, Durantel D, Testoni B, Hantz O, Levrero M, Zoulim F. Control of hepatitis B virus replication by innate response of HepaRG cells. *Hepatology* 2010; 51: 63-72 [PMID: 19877170 DOI: 10.1002/hep.23230]
- Kaur P, Paliwal A, Durantel D, Hainaut P, Scoazec JY, Zoulim F, Chemin I, Herceg Z. DNA methylation of hepatitis B virus (HBV) genome associated with the development of hepatocellular carcinoma and occult HBV infection. *J Infect Dis* 2010; 202: 700-704 [PMID: 20653444 DOI: 10.1086/655398]
- 67 Lucifora J, Arzberger S, Durantel D, Belloni L, Strubin M, Levrero M, Zoulim F, Hantz O, Protzer U. Hepatitis B virus X protein is essential to initiate and maintain virus replication after infection. J Hepatol 2011; 55: 996-1003 [PMID: 21376091 DOI: 10.1016/



- j.jhep.2011.02.015]
- 68 Pollicino T, Amaddeo G, Restuccia A, Raffa G, Alibrandi A, Cutroneo G, Favaloro A, Maimone S, Squadrito G, Raimondo G. Impact of hepatitis B virus (HBV) preS/S genomic variability on HBV surface antigen and HBV DNA serum levels. *Hepatology* 2012; 56: 434-443 [PMID: 22271491 DOI: 10.1002/hep.25592]
- 69 McCullough J. Progress toward a pathogen-free blood supply. Clin Infect Dis 2003; 37: 88-95 [PMID: 12830413 DOI: 10.1086/375232]
- 70 Schlenke P. Pathogen inactivation technologies for cellular blood components: an update. *Transfus Med Hemother* 2014; 41: 309-325 [PMID: 25254027 DOI: 10.1159/000365646]
 - P- Reviewer: Arsenijevic N, Gherlan GS, Romeo R, Silva LD S- Editor: Ji FF L- Editor: A E- Editor: Wu HL





Published by Baishideng Publishing Group Inc

8226 Regency Drive, Pleasanton, CA 94588, USA

Telephone: +1-925-223-8242

Fax: +1-925-223-8243

E-mail: bpgoffice@wjgnet.com
Help Desk: http://www.wjgnet.com/esps/helpdesk.aspx
http://www.wjgnet.com

