

HHS Public Access

Author manuscript *Clin Liver Dis*. Author manuscript; available in PMC 2015 April 02.

Published in final edited form as:

Clin Liver Dis. 2012 February ; 16(1): 95–131. doi:10.1016/j.cld.2011.12.009.

Malnutrition in Cirrhosis: Contribution and Consequences of Sarcopenia on Metabolic and Clinical Responses

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Keywords

Malnutrition; Cirrhosis; Sarcopenia; Metabolism

Cirrhosis with portosystemic shunting is associated with malnutrition, which is the most frequent, yet potentially reversible complication that worsens with disease progression and adversely affects outcome in these patients.¹⁻⁵ Malnutrition in cirrhosis is associated with major complications that include sepsis, uncontrolled ascites, hepatic encephalopathy (HE), spontaneous bacterial peritonitis, and hepatorenal syndrome that develop in 65% of malnourished patients versus 12% of well-nourished patients.^{4,6-10} Several recent reviews have discussed the current clinical problems and therapy for malnutrition in cirrhosis.¹¹⁻¹³ However, the major limitation of these is the lack of focus on the recent advances and potentially exciting data from diverse fields besides hepatology. This review focuses on the current understanding of malnutrition and the newer molecular pathways and targets that are likely to result in novel and specific therapies to reverse its components. Malnutrition in cirrhosis consists of a loss of skeletal muscle and adipose tissue mass. Even though it is being recognized that this combination should be defined as cachexia, $14,15$ the predominant loss of muscle mass in cirrhosis suggests that sarcopenia or loss of skeletal muscle mass is the primary nutritional consequence.^{16,17} In patients with cirrhosis, the prevalence of malnutrition characterized by loss of lean body mass and diminished skeletal muscle weight is estimated to be between 20% to 60% in different studies.^{5,18-21} Most studies have focused on quantifying lean body mass using different instruments, but the skeletal muscle constitutes between 40% and 50% of the lean body mass.22 More precise measures of skeletal muscle mass that are being recognized are the direct measures using imaging techniques.16,17 Skeletal muscle loss in cirrhosis worsens with advancing severity of liver disease as measured by Child's score and the development of portosystemic shunting.^{19,23-25}

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There has been limited success using several nutritional and other interventions in reversing malnutrition and low skeletal muscle mass in cirrhosis.^{1,2,23,26,27} Only partial improvement in anthropometric measures and body weight occur when enteral nutrition or parenteral amino acid mixtures are given.^{28,29} Neither recombinant growth hormone nor insulinlike growth factor 1 (IGF1) in human and animal models of cirrhosis were able to result in complete recovery of skeletal muscle mass.30-32 These poor results are likely related to the limited understanding of the pathophysiologic mechanisms responsible for diminished muscle mass in cirrhosis and portosystemic shunting.^{1,33} Several factors may contribute to this and include the predominantly descriptive nature of the human studies, heterogeneity in the definitions for malnutrition, limited mechanistic studies on skeletal muscle loss in cirrhosis, and the preponderance of publications on skeletal muscle biology in nonliver journals.34,35 Despite the number of publications in this area, there are few studies that have reconciled the recent and exciting data obtained from studies on skeletal muscle biology into our current understanding of malnutrition in cirrhosis.22,36 The present review aims toward integrating our current understanding of the clinical consequences, mechanisms, and therapeutic targets and approaches toward reversing the major complication of cirrhosis, sarcopenia, or loss of skeletal muscle mass.

Recent studies in animal models and cell culture systems have contributed significantly to our understanding of potential mechanisms responsible for sarcopenia in portosystemic shunting in cirrhosis.37-42

Our understanding of the metabolic processes in cirrhosis, skeletal muscle, and whole body protein, fat, and carbohydrate metabolism has increased over the past 2 decades, during which time liver transplantation has become a viable and definitive treatment option for endstage liver disease. Several questions, however, remain unanswered. These include the precise definition of malnutrition in cirrhosis; prevalence of malnutrition in cirrhosis that is affected by the method used to define malnutrition; the impact of malnutrition on outcome before, during, and after liver transplantation; the available therapeutic options; and the outcome in response to these interventions. Additionally, recent exciting and novel data from the authors' laboratory, and that of others, to identify the role of molecular signaling pathways are expected to provide novel insights into the management of patients with cirrhosis.36,43

MALNUTRITION IN LIVER DISEASE: DEFINITIONS

There is wide heterogeneity in the definition of malnutrition in cirrhosis, primarily because adult malnutrition is not well defined. In children, malnutrition is clearly defined as predominantly protein malnutrition or kwashiorkor and combined protein and calorie malnutrition or marasmus. In humans, most proteins are located in the skeletal muscle, $44-47$ and we have, therefore, defined clinical adult protein malnutrition as primarily skeletal muscle loss. Energy malnutrition is more difficult to define clearly, but because adipose tissue is the largest repository of calories, adult fat malnutrition can be defined as a reduction in whole body fat mass. Loss of skeletal muscle mass is also known as sarcopenia, even though this term has traditionally been used to define loss of muscle mass with aging.14,15 More recently, other terms have been used that include cachexia, which is

defined as loss of both muscle and fat mass that is not responsive to providing adequate dietary intake, and precachexia, which is based on the percentile values of the measured muscle and fat mass compared with controls.^{14,15} However, it must be reiterated that these consensus definitions are being developed, but their relevance to the complex metabolic and nutritional derangements in cirrhosis have not been evaluated. It may, however, be summarized that based on our current understanding, malnutrition in cirrhosis comprises reduced muscle mass and strength, called sarcopenia, as well as loss of subcutaneous and visceral fat mass that may be called *adipopenia*. The term hepatic cachexia can be used to define the proportionate loss of both muscle and adipose tissue mass. Finally, the rapid increase in prevalence of fatty liver–related cirrhosis is increasing the number of patients who have sarcopenic obesity characterized by a disproportionate loss of skeletal muscle mass with preserved or increased visceral or subcutaneous adipose tissue mass. Given these reasons, it may be best to avoid the term, malnutrition in cirrhosis, because it can be used to refer to sarcopenia, adipopenia, cachexia, precachexia, obesity, sarcopenic obesity, and micronutrient deficiencies. Precision in definition will permit a clear definition of the patient population being studied and the outcome measures being quantified. Given the lack of such a consensus definition in patients with cirrhosis, the authors have defined these terms in the specific population of patients with cirrhosis (Table 1).

METHODS TO ASSESS MALNUTRITION IN CIRRHOSIS

As previously stated, most publications on malnutrition in cirrhosis use heterogeneous definitions. Standard nutritional assessment instruments use laboratory tests, such as prothrombin time; albumin; prealbumin; transferrin; creatinine height index; and on tests of immune function, such as the delayed-type hypersensitivity reactions.⁴⁸⁻⁵⁰ Because endstage liver disease or cirrhosis confound the common measures of nutritional status, their utility in these patients is reduced. Patients with cirrhosis have significant impairment in their hepatic synthetic function that results in low serum albumin, prealbumin, transferrin levels, and prolonged prothrombin time. These levels will result in an overestimation of the prevalence of malnutrition in these patients.^{51,52} Renal impairment is common in cirrhosis, making the creatinine height index an imprecise measure of malnutrition.53 Anthropometric measures are affected by altered fluid status caused by ascites, peripheral edema, diuretic and salt intake, and concomitant rental failure that makes weight changes difficult to interpret.54,55 Skinfold thickness that measures subcutaneous fat mass, upper-arm measure of muscle area (midarm muscle area), and subjective global assessment (SGA) has additional limitations, including interobserver variability.^{56,57} Furthermore, with the change in demographics and socioeconomic patterns, there are changes in the normal values, and concurrent norms should be used for defining criteria for sarcopenia and cachexia.58 Finally, the anergy in cirrhosis makes delayed-type hypersensitivity an inaccurate gauge of malnutrition.51,59

Several indirect, in vivo methods have been used to quantify body composition in cirrhosis. These methods include total-body electrical conductivity, bioelectrical impedance, dual energy x-ray absorptiometry, deuterium dilution, air displacement plethysmography, and magnetic resonance spectroscopy. $60-62$ These methods are based on the principle that at least 2 components exist in the body fat mass and fat-free mass that is essentially water, protein,

and mineral.63 By determining the whole body weight and fat mass, it is assumed that the remaining weight is nonfat or lean mass. Because 40% to 60% of lean body mass in humans and rodents is contributed by skeletal muscle mass, quantification of lean body or fat-free mass is considered to be a measure of whole body skeletal muscle mass.⁶⁴ There are also concerns expressed about the 2-compartment model obtained from these studies, and alternative 3-component and 4-component models have been proposed.63,65-67 These multicomponent models suffer from limitations in cirrhosis because of the alteration in hydration, bone mineralization, and fluid shifts. Hence, there seems to be no true gold standard or reference technique to quantify malnutrition in cirrhosis. The choice of application is based on cost, logistics, availability, and the need for accuracy and segmental body composition. Based on published studies on malnutrition, the authors' definitions of protein malnutrition to be reflected by skeletal muscle mass and fat malnutrition quantified by the loss of subcutaneous and visceral fat mass as well as altered thermogenesis seem most clinically relevant and can be applied at the bedside.

Recently, psoas muscle area quantified on a single section of computed tomography (CT) of the abdomen at the L3/4 level has been validated as a reliable, noninvasive measure of reduced whole body skeletal muscle mass in cirrhosis.^{16,17} The authors have observed this to be equally reliable for quantifying visceral fat mass on the same section. Because CT of the abdomen is routinely used to screen for lesions in patients with cirrhosis, this can also be used to quantify skeletal muscle and fat mass in these patients. Despite its simplicity, cost and irradiation are 2 considerations that need to be taken into account when using this method.

PREVALENCE OF MALNUTRITION IN CIRRHOSIS

A high prevalence of malnutrition has been reported in patients with cirrhosis in studies in which visceral protein status and immunologic measures are included in the nutritional assessment.48,68,69 The prevalence of nutritional disorders is lower when malnutrition is diagnosed by anthropometric measures only.54,55 Differences in the cause and severity of disease also affect the estimated prevalence of malnutrition in cirrhosis.^{48,70-73} A review of studies published that examined the prevalence of malnutrition using defined criteria is shown in Table 1.4,18,19,49,59,65,68,71-82 It can be summarized from these data that the prevalence of malnutrition depends primarily on the definition chosen, cause of the liver disease, the stage of the disease, and the methods used to quantify malnutrition. Anthropometric measures have been considered to be most dependable; using only these criteria, the prevalence is significantly lower than previously estimated. The lowest estimate from the largest study in 114,703 hospitalized patients with cirrhosis compared with hospitalized patients without cirrhosis showed a prevalence of 6.1% in cirrhosis compared with 1.9% in controls.⁸¹ The major limitation of this study is that malnutrition was diagnosed imprecisely based on a clinical discharge diagnosis. The investigators acknowledge the limitations but suggest that their data support previous published literature on the high (more than 4 fold) prevalence of malnutrition in cirrhosis compared with patients without cirrhosis. Other studies have confirmed that using a combination of biochemical and immunologic studies overestimates the prevalence of muscle and fat loss as estimated by clinical and anthropometric methods.^{19,72} Given these observations, it would

be appropriate to have a standardized method of assessment of protein and fat malnutrition in cirrhosis. An extensive review of the data suggests that the modified SGA that is appropriate in cirrhosis and precise upper-extremity anthropometric measures may be the best available option.4,49 A recent study by the authors' group in 97 hospitalized patients has shown that grip strength and SGA remain the most feasible instruments in assessing the nutritional status and outcome.

These data suggest that based on the definition of the specific component of malnutrition, an appropriate measurement instrument should be chosen. Increasing interest in imaging methods is because of the ability to distinguish the reduction of skeletal muscle and visceral and adipose tissue mass. However, functional measures of muscle strength remain one of the most relevant measures of sarcopenia.⁸³

SEVERITY OF LIVER DISEASE WORSENS SARCOPENIA

Malnutrition has also been related to the severity of liver disease as estimated by Child's score.⁷³ Several modifications of the original Child's score have been used, including the Child-Turcotte, Campbell Child, and Pugh-Child scoring systems.80 In the Pugh modification, the nutritional status was replaced by prothrombin time; the rationale for this was that the nutritional assessment used in the other versions had a significant subjective evaluation, whereas the prothrombin time in combination with serum albumin provides a more objective measure of long-term nutritional evaluation.⁸⁴ However, as has been discussed earlier, these are truly measures of hepatic function and are likely to show greater abnormality with worsening severity of liver disease. The authors have specifically excluded those investigators who used the Child-Turcotte and Campbell Child scoring system because these have nutritional evaluation incorporated into them and are, therefore, biased in favor of a higher prevalence of malnutrition in advanced disease. In 3 published studies that evaluated the impact of severity of underlying liver disease as measured by the Child-Pugh scoring system showed that there is evidence of malnutrition as assessed by grip strength, body cell mass, body fat mass, and ideal body weight early in the course of the disease.73,74,85 These measures of nutritional deficiency become worse with progressive severity of liver disease.^{73,74,85} Other measures of severity of liver disease, including the model for end-stage liver disease (MELD) score, have not been systematically assessed for their relation to the severity of sarcopenia, cachexia, or malnutrition.

These observations suggest that clinical and anthropometric measures of loss of muscle mass and fat mass are common in cirrhosis and worsen with the progression of liver disease.

CLINICAL IMPLICATIONS OF MALNUTRITION IN CIRRHOSIS

For practical clinical purposes, the impact of malnutrition in cirrhosis on outcome can be examined by the effect of skeletal muscle loss on survival and complications of cirrhosis. With the availability of liver transplantation, aggressive intensive care, antibiotics, renal support, and endoscopic interventions to prevent and treat the complications of cirrhosis, there is a resurgence of interest in the nutritional management of these patients. Several studies have consistently shown that malnutrition in cirrhosis affects the survival and the development of the complications of cirrhosis.

Malnutrition and Survival in Cirrhosis

Several investigators have examined the impact of malnutrition, primarily using instruments that measure sarcopenia, and observed that worsening severity of muscle loss is accompanied by higher mortality (Table 2).^{3-5,10,16,17,76,86-91} It is interesting that despite a large number of studies across the world demonstrating that sarcopenia and malnutrition worsen survival in cirrhosis, no studies have documented improved survival with reversal of sarcopenia. In this context, it is interesting that the authors' studies on reversal of sarcopenia after transjugular intrahepatic portosystemic shunt (TIPS) have demonstrated better survival after TIPS in patients in whom skeletal muscle mass increased compared with those in whom skeletal muscle mass did not change or became less.⁹²

Malnutrition and Quality of Life

Quality of life in cirrhosis is significantly lower than that in controls (Table 3).^{6,93-98} This finding has been related to the severity of underlying liver disease as assessed by the Child's scoring system. Because the Child's score relates to the severity and prevalence of malnutrition, it is expected that malnutrition will be related to the quality of life. Recently, in a prospective study of 61 patients with cirrhosis, those with malnutrition as defined by SGA had impairment in 6 of the 8 quality-of-life scales on the SF-36.⁹⁴ However, in patients with hepatocellular carcinoma, quality of life was not related to tumor mass or hepatocellular failure.⁹⁵ Similarly in patients with primary biliary cirrhosis, a condition with the most severe reduction in fat and muscle mass, the Nottingham health profile, a measure of quality of life, was not related to severity or duration of the disease.⁹⁶ However, in neither of these studies was the relation between malnutrition and quality of life evaluated. In summary, based on existing data, patients with cirrhosis and malnutrition as assessed by SGA had a worse quality of life than those with preserved muscle and fat mass. These findings were independent of the complications of cirrhosis. More recently, previous episodes of HE, even after complete resolution, impact the quality of life in patients with cirrhosis.99 However, in a prospective study, minimal HE did not seem to have a significant impact on quality of life in patients with cirrhosis.98 This finding was in contrast to clinical expectations and recent interest on the impact of minimal HE on driving skills and motor vehicle–related accidents.100,101

Malnutrition and Clinical Complications of Cirrhosis

The known major life-threatening complications of cirrhosis that include ascites, spontaneous bacterial peritonitis, portal hypertension and gastrointestinal bleeding, HE and hepatorenal syndrome, and all of these are adversely affected by malnutrition and sarcopenia (Table 4).4,8,9,81,102-104 Other complications include hepatocellular carcinoma and pulmonary and cardiac complications of cirrhosis.105 Each of these complications aggravates the catabolic state by their impact on circulating cytokines and hormones and results in the reduction of muscle mass.35 However, few studies have systematically evaluated the impact of malnutrition on the development and progression of these complications. In a classical study by Moller, 106 the development of portal hypertension, portosystemic collaterals, and varices were more severe and common in malnourished patients. In this study, nutritional status was scored by a subjective assessment scale of 1 to

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4, with 4 being cachexia. Even though the investigators do not describe the validation process of this scoring system, this study demonstrates that malnourished patients had more severe portal hypertension and risk of variceal bleeding.

In states of hepatocellular dysfunction and portal hypertension, plasma concentration of ammonia is elevated, and the skeletal muscle has been suggested to play a significant role in ammonia detoxification.^{42,107} Because hyperammonemia is considered to be the major pathogenic factor in the development of HE, it has been speculated that low muscle mass will predispose to and aggravate the severity of HE. There are 2 studies that have specifically examined the impact of malnutrition on the development and outcome of HE with conflicting results.^{7,108,109}

Both of these are single time-point cross-sectional assessments for malnutrition as defined by anthropometric and other criteria. In the study by Kalaitzakis and colleagues⁷ in 128 patients with cirrhosis of varied causes, HE was diagnosed as overt by West Haven criteria and the number connection test. Malnutrition was defined by anthropometric measurement less than the fifth percentile of established norms for the general population, body mass index less than 20 mg/m², or weight loss of greater than or equal to 5% to 10% in the previous 3 to 6 months. Among these patients, 40% had malnutrition and 34% had HE. Patients with malnutrition had HE more frequently, and malnutrition was an independent risk factor for HE. In contrast, in another prospective study by Soros and colleagues,¹⁰⁸ nutritional assessment was performed using body mass index, anthropometrics using the triceps skinfold thickness and arm muscle area, and bioelectrical impedance. It is interesting that the 2 studies yielded conflicting results in terms of the impact of malnutrition on the development of HE. Unfortunately, both were cross-sectional studies and did not specifically examine the impact of sarcopenia on the development of HE. One of the major confounding factors in these assessments is that previous episodes of HE are also likely to worsen sarcopenia by a combination of hyperammonemia, poor oral intake, and hospitalizations. In a prospective study, the authors demonstrated that the frequency of HE was higher in patients who had evidence of sarcopenia.¹¹⁰ The authors' studies in an animal model of hyperammonemia (portacaval anastomosis [PCA] rat) and in murine myoblasts have suggested that ammonia induces the expression of myostatin, a transforming growth factor (TGF) β superfamily member that is known to worsen sarcopenia.^{111,112} The authors' data suggest that hyperammonemia of cirrhosis induces HE and worsens sarcopenia. The development and progression of sarcopenia then begins a self-destructive cycle of recurrent HE and further loss of muscle mass caused by impaired nonhepatic ammonia disposal.

PATHOGENESIS AND MECHANISMS OF SARCOPENIA IN CIRRHOSIS

Because sarcopenia is the major contributor to malnutrition, functional status, and outcomes in cirrhosis, an understanding of the biochemical and cellular mechanisms that result in loss of muscle mass is critical to identify therapeutic targets. Initial works in understanding the metabolic alterations in cirrhosis were based on isotopic tracer methodology.113 Despite initial enthusiasm, these were predominantly descriptive studies; and only recently, the advances in myology, gerontology, and molecular biology are being translated into

identifying precise molecular abnormalities in the skeletal muscle and their dysregulation in cirrhosis.15,114

Maintenance of Skeletal Muscle Mass

To understand the mechanisms of sarcopenia, an understanding of the mechanisms of maintenance of muscle mass is necessary (Fig. 1). Skeletal muscle mass is maintained by a balance between muscle protein synthesis, protein breakdown, and satellite cell proliferation and differentiation.¹¹⁵ Satellite cells are myogenically committed precursor cells that contribute nuclei to the myocytes for maintenance and growth of mature skeletal muscle.116,117 Increase in skeletal muscle protein synthesis and satellite cell proliferation and differentiation are necessary for skeletal muscle growth.115 Satellite cells constitute 2% to 4% of adult skeletal muscle, whereas skeletal muscle structural protein is the major contributor to skeletal muscle mass. Therefore, alterations in skeletal muscle mass are primarily caused by changes in the structural protein content. Another critical concept that needs to be reiterated is that even though both impaired protein synthesis and increased protein breakdown contribute to the reduced muscle mass in cirrhosis, their contributions are distinct. A reduction in protein synthesis alone results only in the failure to accrete protein mass, whereas an increase in proteolysis is necessary for the loss of muscle mass. However, continued enhanced proteolysis precludes cell survival and needs to be regulated. Because protein synthesis and proteolysis do not occur independently, rather are highly integrated, it is the relative contribution that determines the muscle mass. Current methods to quantify skeletal muscle protein synthesis and proteolysis lack sufficient sensitivity to identify the small changes that occur with the disease.¹¹⁸ These methods are being supplemented by quantifying whole muscle protein synthesis instead of the traditional fractional synthesis rate in animal studies^{36,119} but have not been developed in humans yet. Furthermore, there are no studies that have directly quantified skeletal muscle protein synthesis or breakdown in human cirrhosis. The use of isotopic tracers using stable isotope-labeled amino acids have examined whole body protein metabolism in cirrhosis.¹

Protein Metabolism in Cirrhosis with Portosystemic Shunting

Protein turnover studies in cirrhosis using tracer isotopes have yielded conflicting results.120-123 These differences may be related to confounding variables, such as differences in disease severity, nutritional status, and the methodology used to quantify protein turnover. The estimation of rates of whole body protein breakdown using $[1¹³C]$ leucine, in humans with stable cirrhosis (defined as Child's class A or B) in the fasted state, were not different from those in healthy controls.120 Studies using phenylalanine tracer showed a decreased whole body protein breakdown in patients with cirrhosis of Child's class B and C (decompensated) and no difference between compensated patients with cirrhosis (Child's class A and B) and healthy controls.^{124,125} Contradictory results have been reported using different isotopic tracers, such as $[15N]$ glycine and $[14C]$ tyrosine.¹²⁶

Several methods have been used to examine protein synthesis in vivo.¹²⁷ However, muscle biopsies are required for precise quantification of skeletal muscle protein synthesis. Even though these have not been reported in human patients with cirrhosis, whole body amino

acid kinetic studies showed lower rates of protein synthesis in patients with cirrhosis than in controls.121,123 Arteriovenous differences in amino acid concentration in the lower extremity showed that proteolysis and protein synthesis were lower.¹²³ These abnormalities may persist or worsen in the postprandial state in patients with cirrhosis.¹²¹

Data from studies in animal models are equally conflicting. In the rat model of carbon tetrachloride–induced cirrhosis, lower rate of protein breakdown and lower protein synthesis was observed.128 In the PCA rat, a lower rate of liver and brain protein synthesis was reported 3 weeks after anastamosis.129 In contrast, another study in PCA rats showed no difference in the rate of protein synthesis in different organs.¹³⁰ An increased skeletal muscle proteolysis mediated by the ubiquitin proteasome pathway has been described in the bile duct ligated rat.¹³¹ However, the bile duct ligated rat as a model of cirrhosis differs from human cirrhosis because secondary biliary cirrhosis in humans is extremely rare and steatorrhea and malabsorption that accompany this procedure affect the muscle mass independent of cirrhosis.

Another pathway of protein breakdown that is being increasingly examined is the lysosomal cathepsin–mediated autophagy.132,133 Autophagy serves to remove long-lived and abnormal proteins and dysfunctional organelles and helps recycle the substrates generated to permit protein synthesis. Autophagy is enhanced during states of nutrient deprivation and cellular stress. Preliminary studies from the authors' laboratory have shown an increased skeletal muscle autophagy. However, with the impaired muscle protein synthesis in cirrhosis, autophagy may be futile and contributes to sarcopenia, especially in the presence of reduced ubiquitin-proteasome–mediated proteolysis.

Despite the heterogeneity in the disease and methodologies used, the preponderance of evidence based on studies in humans and animals suggest an unchanged rate of protein breakdown and a decrease in the rate of protein synthesis in cirrhosis.113,121,123 Several confounding variables may have contributed to the differences in observations and include the stage of the disease at the time of study, underlying cause of cirrhosis, duration of illness, muscle mass before disease development, and comorbid conditions that also contribute to whole body and skeletal muscle protein metabolism.

Satellite Cells and Skeletal Muscle Mass

Skeletal muscle fibers in adults are composed of terminally differentiated myocytes that do not replicate.^{116,117} Their growth and adaptation to injury depend on a small population of stem cells called satellite cells that are committed to a myogenic lineage and are closely associated with the periphery of the muscle fibers.117 Proliferation and differentiation of satellite cells contribute to the accretion of myonuclei in mature muscle cells and growth of skeletal muscle.^{117,134} Impaired satellite cell proliferation and differentiation occur in sarcopenia of aging, calorie restriction, hind limb unloading, and immobilization.¹³⁵⁻¹³⁷ However, the contribution of impaired satellite cell function to the diminished muscle mass in cirrhosis is unknown. The authors have shown in the PCA rat an impaired satellite cell proliferation and differentiation as evidenced by the low expression of proliferating cell nuclear antigen (PCNA) and myogenic regulatory factors (myoD, myf5, and myogenin).38,39 The authors' in vivo immunohistochemical studies using 5 bromo 2'

deoxyuridine incorporation have also shown that following PCA, there is a significantly lower mitotic index of satellitel cells compared with the control animals. These data suggest that satellite cell function is impaired in portosystemic shunting and may play a role in sarcopenia of cirrhosis. The enhancement of satellite cell function is a potential therapeutic target in these patients.

Molecular Regulation of Skeletal Muscle Protein Metabolism and Satellite Cell Function

There are 2 regulatory pathways that contribute to skeletal muscle growth: (1) enhanced protein synthesis in existing muscle fibers and (2) proliferation and differentiation of myogenic satellite cells that fuse with the exiting muscle fibers (Fig. 2). Myostatin and IGF1 are the 2 major upstream regulators of these functions in the skeletal muscle. An increase in skeletal muscle protein synthesis results from the activation of components of the highly regulated components of the canonical IGF1/PI3K/Akt/mTOR signaling pathway (see Fig. 1).115 Activation of Akt and mTOR by phosphorylation results in the stimulation of ribosomal protein translation by the effector proteins, p70s6k and 4E-BP.138 The impairment of component proteins in this pathway results in reduced protein synthesis.^{115,139} Proteolysis that is responsible for the reduction in muscle mass is also regulated by myostatin and IGF1. Impaired phosphorylation and activation of Akt results in increased ubiquitin proteasome–mediated proteolysis, and reduced mTOR activation results in enhanced autophagy.34,140 These results demonstrate the complex crosstalk at different components between the critical regulators of muscle mass and their ultimate targets and functional consequences.

Myostatin, a member of the TGF β superfamily expressed in the skeletal muscle, is a potent inhibitor of muscle protein synthesis and satellite cell function.¹⁴¹ The authors have shown that the PCA rat is an appropriate model to examine the mechanisms responsible for failure to increase lean body weight and gain skeletal muscle mass with portosystemic shunting in cirrhosis.40,111 The authors have previously reported that an increased expression of myostatin occurred 2 weeks after PCA and accompanied the failure to gain skeletal muscle mass and impaired satellite cell function.^{38,39} This finding was accompanied by an impaired skeletal muscle protein synthetic response and decreased phosphorylation of mTOR and its downstream targets, p70s6 kinase and 4 E binding protein 1.139 Others have shown that myostatin blocks an upstream regulator or mTOR (ie, protein kinase B or Akt.)^{142,143} The administration of follistatin reversed the myostatin-induced loss of lean body mass, skeletal muscle weight, and impaired phosphorylation of mTOR and p70s6k.³⁶ Myostatin has also been shown to activate the ubiquitin proteasome–mediated proteolysis and the lysosomal autophagy.140,144

IGF1

In addition to myostatin, IGF1 is the other major factor that regulates skeletal muscle protein metabolism.34 There is some evidence that locally produced IGF1 in the skeletal muscle (mechano-growth factor) mediates these effects rather than the circulating.145 IGF1 increases muscle mass by promoting protein synthesis, inhibiting protein breakdown, and increasing satellite cell proliferation and differentiation. The intracellular signaling pathways downstream of IGF1 binding to its receptor, IGF1 receptor α (IGF1R α) have been well

characterized.115 Increased skeletal muscle protein synthesis and satellite cell proliferation in response to IGF1 are mediated by the activating Akt and sequential phosphorylation and activation of its downstream targets.146,147 These data suggest that Akt is the central mediator of critical components of the pathway of protein synthesis in the skeletal muscle.

There is intense interest in the regulation of skeletal muscle IGF1 and myostatin in liver disease. Identification of the binding sites of both androgen receptor and nuclear factor kB on the promoter region of myostatin also holds promise as effective therapeutic targets. Translation of these data from animal and in vitro cell culture studies to humans is essential.

AGING PATIENTS WITH CIRRHOSIS

As the global population ages, this contributes to the progressive worsening of sarcopenia. It is estimated that after the age of 50 years, approximately 1% of skeletal muscle loss occurs per year.148,149 The impact and interaction of sarcopenia of aging and cirrhosis are not known. The adverse effects of both of these processes on muscle mass and function may be exponential, contributing to an urgent need to increase our understanding of the mechanisms and identification of therapies.

POSTTRANSPLANTED PATIENTS WITH CIRRHOSIS

Pretransplant malnutrition, specifically sarcopenia, adversely impacts the perioperative and immediate posttransplant outcomes (Table 5). Additionally, pretransplant sarcopenia is associated with worse outcomes after liver transplantation.^{5,17} It is thought that liver transplantation is curative for cirrhosis. However, it must be reiterated that this option is available only to a minority of patients. Furthermore, posttransplant metabolic syndrome and the attendant insulin resistance adds to the worsening of muscle loss.^{150,151} Finally, mTOR inhibitors, calcineurin inhibitors, and corticosteroids, commonly used immunosuppressants used after transplantation, alter the expression and activity of critical regulators of muscle protein metabolism.152-154 These observations support the urgent need to develop therapies to reverse and treat sarcopenia in cirrhosis because this casts a long shadow from before transplantation to after the procedure.

THERAPEUTIC OPTIONS

Given the human data that suggest that cirrhosis is a state of accelerated starvation,¹⁵⁵ several nutrient interventions have been tried with limited success in long-term improvement in protein or energy metabolism.156,157 Several hormonal alterations have resulted in interventions that use anabolic androgens, IGF, and growth hormone with no benefit and several adverse effects.¹⁵⁸⁻¹⁶¹ Current studies underway on understanding the mechanisms of alteration in protein and fat malnutrition in cirrhosis are likely to provide the basis of novel treatment options (Table 6). Methods to determine the nutritional needs have also been devised and include quantification of the resting energy expenditure (REE), respiratory quotient (RQ), and daily protein needs.

The standard method to measure REE and RQ is the use of a metabolic cart. However, cost, logistics, and complexity of the test have led to this being used only in research settings.

Interest has increased recently in the use of a handheld calorimeter that has been found to be more precise than a variety of predictive equations. In hospitalized patients with cirrhosis, the authors have found the handheld respiratory calorimeter (MedGem, Microlife Medical Home Solutions, Inc., CO, USA) to be as precise as a metabolic cart in the clinical research unit in quantifying REE. The European Society for Clinical Nutrition and Metabolism guidelines suggest no protein restriction in patients with cirrhosis based on published studies.¹⁵⁷

NUTRITIONAL SUPPLEMENTATION

Because skeletal muscle is the major whole body protein store, with a reduction in muscle mass, whole body protein content is lower. Increased protein intake has been demonstrated to be safe, well tolerated, and beneficial in patients with cirrhosis, but the long-term anabolic effects on muscle mass and function have not yet been established.33,156,162-166 Several nutritional interventions have been examined that have focused on 2 specific areas: decrease the intermeal frequency and increase caloric and protein intake (Table 7).^{93,103,167-171} However, as stated earlier, recent advances in our understanding of skeletal muscle biology, regulatory pathways, and targeted interventions have not been evaluated. Separation of adipocyte and skeletal muscle responses to specific interventions are likely to result in the reversal of sarcopenia without the accompanying increase in fat mass and to avoid the development of sarcopenic obesity.

Dietary modification and supplements have been examined with conflicting results. Frequent snacks, late-evening snacks, branched chain amino acid supplementation, breakfast, and protein supplementation have been examined with beneficial results but have not been incorporated into routine clinical practice.^{22,171-173} There is increasing evidence that shortening the interval between meals will reduce the severity and prevalence of malnutrition in cirrhosis.²² Hence, the emphasis has been on late-evening snacks and, recently, on breakfast on waking up.^{22,171} Both these measures have the benefit of increasing the availability of amino acids and suppressing gluconeogenesis from amino acids derived from endogenous proteolysis. Furthermore, both splanchnic and whole body protein breakdown are suppressed by dietary intake. Late-evening snacks have been shown to improve whole body protein kinetics with lower protein breakdown and increased protein synthesis. However, these are short-term effects. Animal data in the portacaval shunted rat model suggest that early in the course of the illness, there is increased proteolysis; and later, once loss of muscle and fat mass is established, there is an impaired skeletal muscle protein synthesis.³⁹ This interpretation is supported by studies in stable patients with cirrhosis in whom there is increased whole body protein breakdown.³³ The stage of the disease and underlying cause affect the severity of malnutrition in humans. However, the authors' animal data suggest that the duration of illness plays a significant role, with increased proteolysis early and impaired protein synthesis late in the disease.³⁶ Therefore, the therapeutic strategy will be to focus on reducing muscle proteolysis early in the disease and promote muscle protein synthesis later in the disease once muscle loss is established. Furthermore, the authors' observation of increased skeletal muscle autophagy is novel and needs further studies for its implications in the pathogenesis and reversal of sarcopenia in cirrhosis.

Of the essential amino acids, leucine holds the most promise as an intervention to reverse sarcopenia in aging.^{174,175} Leucine is not only an essential amino acid substrate for protein synthesis but also functions as a direct activator of the critical protein synthesis and autophagy regulator, $mTOR$ ¹⁷⁶ Additionally, leucine stimulates insulin release from the pancreatic β cells that functions as an anabolic hormone in the skeletal muscle.¹⁷⁷ Finally, leucine is an energy substrate in the skeletal muscle. However, because the administration of leucine will stimulate muscle protein synthesis, other essential amino acids may become limiting and need to be replaced.¹⁷⁸ Leucine-enriched essential amino acids can, therefore, be considered in the long-term management of sarcopenia of cirrhosis. It has been identified that reversing sarcopenia and cachexia can improve outcome in other disorders, like cancer.179 A similar therapeutic approach in patients with cirrhosis is likely to improve survival, quality of life, and the development of other complications. Such outcome measures have not yet been reported. The authors recently showed that in response to TIPS, a subgroup of patients had an improvement in muscle size measured on CT. These patients had significantly better survival compared with those who either did not increase or had a reduction in muscle size.⁹²

Micronutrient Replacement

Even though the authors have not focused on micronutrient replacement in cirrhosis, deficiency of vitamin D and zinc are well recognized and need to be identified and treated.180-184

Exercise

The role of aerobic and resistance exercise on skeletal muscle insulin signaling, protein synthesis response, AMP kinase activity, and satellite cell function has been studied extensively in aging.^{185,186} However, fatigue; reduced maximum exercise capacity in patients with cirrhosis; and the presence of limiting complications, including ascites, encephalopathy, and portal hypertension, have limited the translation of the data or the elegant designs of the studies performed in patients without cirrhosis.187 Resistance exercise increases portal hypertension, and even transient increases in portal hypertension can result in catastrophic variceal bleeding and death.188 It is, therefore, critical that the data on the impact of exercise on muscle mass and function be translated very judiciously in patients with cirrhosis.

Novel strategies to reverse cachexia, including myostatin antagonists, are also of clinical interest, especially given recent data that myostatin may play a critical role in cirrhotic sarcopenia.35,189 The authors' data in an animal model that the adverse consequences of increased myostatin expression can be reversed without impacting the underlying liver disease are especially exciting³⁶ because liver transplantation is not a universally available treatment option and reversing hepatic cachexia-sarcopenia should be a major therapeutic option for cirrhosis. Given the paucity of data, the understudied nature of the problem, sarcopenia in cirrhosis deserves to be recognized as an area of unmet need with the potential to improve the outcome of the large number of patients with cirrhosis. One potential strategy for the development of novel and successful therapies is the need for consilience between

the diverse and seemingly unrelated fields of aging, molecular signaling, nutraceuticals, hepatology, transplant immunology, clinical nutrition, and transplant surgeons.

References

- 1. Tessari P. Protein metabolism in liver cirrhosis: from albumin to muscle myofibrils. Curr Opin Clin Nutr Metab Care. 2003; 6(1):79–85. [PubMed: 12496684]
- 2. Bianchi G, Marzocchi R, Agostini F, et al. Update on nutritional supplementation with branchedchain amino acids. Curr Opin Clin Nutr Metab Care. 2005; 8(1):83–7. [PubMed: 15586005]
- 3. Bilbao I, Armadans L, Lazaro JL, et al. Predictive factors for early mortality following liver transplantation. Clin Transplant. 2003; 17(5):401–11. [PubMed: 14703921]
- 4. Alvares-da-Silva MR, Reverbel da ST. Comparison between handgrip strength, subjective global assessment, and prognostic nutritional index in assessing malnutrition and predicting clinical outcome in cirrhotic outpatients. Nutrition. 2005; 21(2):113–7. [PubMed: 15723736]
- 5. Selberg O, Bottcher J, Tusch G, et al. Identification of high- and low-risk patients before liver transplantation: a prospective cohort study of nutritional and metabolic parameters in 150 patients. Hepatology. 1997; 25(3):652–7. [PubMed: 9049214]
- 6. Kalaitzakis E, Simren M, Olsson R, et al. Gastrointestinal symptoms in patients with liver cirrhosis: associations with nutritional status and health-related quality of life. Scand J Gastroenterol. 2006; 41(12):1464–72. [PubMed: 17101578]
- 7. Kalaitzakis E, Olsson R, Henfridsson P, et al. Malnutrition and diabetes mellitus are related to hepatic encephalopathy in patients with liver cirrhosis. Liver Int. 2007; 27(9):1194–201. [PubMed: 17919230]
- 8. Huisman EJ, Trip EJ, Siersema PD, et al. Protein energy malnutrition predicts complications in liver cirrhosis. Eur J Gastroenterol Hepatol. 2011; 23(11):982–9. [PubMed: 21971339]
- 9. Merli M, Lucidi C, Giannelli V, et al. Cirrhotic patients are at risk for health care-associated bacterial infections. Clin Gastroenterol Hepatol. 2010; 8(11):979–85. [PubMed: 20621200]
- 10. Merli M, Giusto M, Gentili F, et al. Nutritional status: its influence on the outcome of patients undergoing liver transplantation. Liver Int. 2010; 30(2):208–14. [PubMed: 19840246]
- 11. O'Brien A, Williams R. Nutrition in end-stage liver disease: principles and practice. Gastroenterology. 2008; 134(6):1729–40. [PubMed: 18471550]
- 12. Kerwin AJ, Nussbaum MS. Adjuvant nutrition management of patients with liver failure, including transplant. Surg Clin North Am. 2011; 91(3):565–78. [PubMed: 21621696]
- 13. Ferreira LG, Anastacio LR, Correia MI. The impact of nutrition on cirrhotic patients awaiting liver transplantation. Curr Opin Clin Nutr Metab Care. 2010; 13(5):554–61. [PubMed: 20531175]
- 14. Argiles JM, Anker SD, Evans WJ, et al. Consensus on cachexia definitions. J Am Med Dir Assoc. 2010; 11(4):229–30. [PubMed: 20439040]
- 15. Evans WJ, Morley JE, Argiles J, et al. Cachexia: a new definition. Clin Nutr. 2008; 27(6):793–9. [PubMed: 18718696]
- 16. Montano-Loza AJ, Meza-Junco J, Prado CM, et al. Sarcopenia is associated with mortality in patients with cirrhosis. Clin Gastroenterol Hepatol. 2011 Epub ahead of print.
- 17. Englesbe MJ, Patel SP, He K, et al. Sarcopenia and mortality after liver transplantation. J Am Coll Surg. 2010; 211(2):271–8. [PubMed: 20670867]
- 18. Peng S, Plank LD, McCall JL, et al. Body composition, muscle function, and energy expenditure in patients with liver cirrhosis: a comprehensive study. Am J Clin Nutr. 2007; 85(5):1257–66. [PubMed: 17490961]
- 19. Alberino F, Gatta A, Amodio P, et al. Nutrition and survival in patients with liver cirrhosis. Nutrition. 2001; 17(6):445–50. [PubMed: 11399401]
- 20. Campillo B, Richardet JP, Bories PN. Enteral nutrition in severely malnourished and anorectic cirrhotic patients in clinical practice. Gastroenterol Clin Biol. 2005; 29(6–7):645–51. [PubMed: 16141996]
- 21. Plauth M, Schutz ET. Cachexia in liver cirrhosis. Int J Cardiol. 2002; 85(1):83–7. [PubMed: 12163212]

- 22. Tsien CD, McCullough AJ, Dasarathy S. Late evening snack exploiting a period of anabolic opportunity in cirrhosis. J Gastroenterol Hepatol. 2011 Epub ahead of print.
- 23. Nutritional status in cirrhosis. Italian multicentre cooperative project on nutrition in liver cirrhosis. J Hepatol. 1994; 21(3):317–25. [PubMed: 7836699]
- 24. Lata J, Husova L, Jurankova J, et al. Factors participating in the development and mortality of variceal bleeding in portal hypertension–possible effects of the kidney damage and malnutrition. Hepatogastroenterology. 2006; 53(69):420–5. [PubMed: 16795985]
- 25. Gunsar F, Raimondo ML, Jones S, et al. Nutritional status and prognosis in cirrhotic patients. Aliment Pharmacol Ther. 2006; 24(4):563–72. [PubMed: 16827812]
- 26. Marchesini G, Marzocchi R, Noia M, et al. Branched-chain amino acid supplementation in patients with liver diseases. J Nutr. 2005; 135(Suppl 6):1596S–601S. [PubMed: 15930476]
- 27. Kato M, Miwa Y, Tajika M, et al. Preferential use of branched-chain amino acids as an energy substrate in patients with liver cirrhosis. Intern Med. 1998; 37(5):429–34. [PubMed: 9652895]
- 28. Tsiaousi ET, Hatzitolios AI, Trygonis SK, et al. Malnutrition in end stage liver disease: recommendations and nutritional support. J Gastroenterol Hepatol. 2008; 23(4):527–33. [PubMed: 18397483]
- 29. Charlton M. Branched-chain amino acid enriched supplements as therapy for liver disease. J Nutr. 2006; 136(Suppl 1):295S–8S. [PubMed: 16365102]
- 30. Conchillo M, de Knegt RJ, Payeras M, et al. Insulin-like growth factor I (IGF-I) replacement therapy increases albumin concentration in liver cirrhosis: results of a pilot randomized controlled clinical trial. J Hepatol. 2005; 43(4):630–6. [PubMed: 16024131]
- 31. Zaratiegui M, Castilla-Cortazar I, Garcia M, et al. IGF1 gene transfer into skeletal muscle using recombinant adeno-associated virus in a rat model of liver cirrhosis. J Physiol Biochem. 2002; 58(3):169–76. [PubMed: 12603011]
- 32. Bucuvalas JC, Horn JA, Chernausek SD. Resistance to growth hormone in children with chronic liver disease. Pediatr Transplant. 1997; 1(1):73–9. [PubMed: 10084790]
- 33. Biolo G, Antonione R, Barazzoni R, et al. Mechanisms of altered protein turnover in chronic diseases: a review of human kinetic studies. Curr Opin Clin Nutr Metab Care. 2003; 6(1):55–63. [PubMed: 12496681]
- 34. Glass D, Roubenoff R. Recent advances in the biology and therapy of muscle wasting. Ann N Y Acad Sci. 2010; 1211:25–36. [PubMed: 21062293]
- 35. Ruegg MA, Glass DJ. Molecular mechanisms and treatment options for muscle wasting diseases. Annu Rev Pharmacol Toxicol. 2011; 51:373–95. [PubMed: 20936944]
- 36. Dasarathy S, McCullough AJ, Muc S, et al. Sarcopenia associated with portosystemic shunting is reversed by follistatin. J Hepatol. 2011; 54(5):915–21. [PubMed: 21145817]
- 37. Gayan-Ramirez G, van de Casteele M, Rollier H, et al. Biliary cirrhosis induces type IIx/b fiber atrophy in rat diaphragm and skeletal muscle, and decreases IGF-I mRNA in the liver but not in muscle. J Hepatol. 1998; 29(2):241–9. [PubMed: 9722205]
- 38. Dasarathy S, Dodig M, Muc SM, et al. Skeletal muscle atrophy is associated with an increased expression of myostatin and impaired satellite cell function in the portacaval anastomosis rat. Am J Physiol Gastrointest Liver Physiol. 2004; 287(6):G1124–30. [PubMed: 15256363]
- 39. Dasarathy S, Muc S, Hisamuddin K, et al. Altered expression of genes regulating skeletal muscle mass in the portacaval anastomosis rat. Am J Physiol Gastrointest Liver Physiol. 2007; 292(4):G1105–13. [PubMed: 17185634]
- 40. Dasarathy S, Mullen KD, Dodig M, et al. Inhibition of aromatase improves nutritional status following portacaval anastomosis in male rats. J Hepatol. 2006; 45(2):214–20. [PubMed: 16684577]
- 41. Canturk NZ, Canturk Z, Ozden M, et al. Protective effect of IGF-1 on experimental liver cirrhosisinduced common bile duct ligation. Hepatogastroenterology. 2003; 50(54):2061–6. [PubMed: 14696465]
- 42. Holecek M, Kandar R, Sispera L, et al. Acute hyperammonemia activates branched-chain amino acid catabolism and decreases their extracellular concentrations: different sensitivity of red and white muscle. Amino Acids. 2011; 40(2):575–84. [PubMed: 20614225]

- 43. Sriram S, Subramanian S, Sathiakumar D, et al. Modulation of reactive oxygen species in skeletal muscle by myostatin is mediated through NF-kappaB. Aging Cell. 2011; 10(6):931–48. [PubMed: 21771249]
- 44. Chinn KS. Prediction of muscle and remaining tissue protein in man. J Appl Physiol. 1967; 23(5): 713–5. [PubMed: 6061385]
- 45. Krzywicki HJ, Chinn KS. Body composition of a military population, Fort Carson, 1963. I. Body density, fat, and potassium 40. Am J Clin Nutr. 1967; 20(7):708–15. [PubMed: 6036259]
- 46. Krzywicki HJ, Chinn KS. Human body density and fat of an adult male population as measured by water displacement. Am J Clin Nutr. 1967; 20(4):305–10. [PubMed: 6022006]
- 47. Krzywicki HJ, Chinn KS. Body composition of a military population, Fort Carson, 1963. I. Body density, fat and potassium 40. Rep US Army Med Res Nutr Lab Denver. 1966; 22:1–17. Lab Rep 296.
- 48. DiCecco SR, Wieners EJ, Wiesner RH, et al. Assessment of nutritional status of patients with endstage liver disease undergoing liver transplantation. Mayo Clin Proc. 1989; 64(1):95–102. [PubMed: 2492064]
- 49. Hasse J, Strong S, Gorman MA, et al. Subjective global assessment: alternative nutritionassessment technique for liver-transplant candidates. Nutrition. 1993; 9(4):339–43. [PubMed: 8400590]
- 50. Hasse JM. Nutritional implications of liver transplantation. Henry Ford Hosp Med J. 1990; 38(4): 235–40. [PubMed: 2086551]
- 51. O'Keefe SJ, El-Zayadi AR, Carraher TE, et al. Malnutrition and immunoincompetence in patients with liver disease. Lancet. 1980; 2(8195 pt 1):615–7. [PubMed: 6107408]
- 52. Sobhonslidsuk A, Roongpisuthipong C, Nantiruj K, et al. Impact of liver cirrhosis on nutritional and immunological status. J Med Assoc Thai. 2001; 84(7):982–8. [PubMed: 11759979]
- 53. Francoz C, Prie D, Abdelrazek W, et al. Inaccuracies of creatinine and creatinine-based equations in candidates for liver transplantation with low creat-inine: impact on the model for end-stage liver disease score. Liver Transpl. 2010; 16(10):1169–77. [PubMed: 20879015]
- 54. Thuluvath PJ, Triger DR. How valid are our reference standards of nutrition? Nutrition. 1995; 11(6):731–3. [PubMed: 8719130]
- 55. Thuluvath PJ, Triger DR. Evaluation of nutritional status by using anthropometry in adults with alcoholic and nonalcoholic liver disease. Am J Clin Nutr. 1994; 60(2):269–73. [PubMed: 8030606]
- 56. Pikul J, Sharpe MD, Lowndes R, et al. Degree of preoperative malnutrition is predictive of postoperative morbidity and mortality in liver transplant recipients. Transplantation. 1994; 57(3): 469–72. [PubMed: 8108888]
- 57. Fuller NJ, Jebb SA, Goldberg GR, et al. Inter-observer variability in the measurement of body composition. Eur J Clin Nutr. 1991; 45(1):43–9. [PubMed: 1855499]
- 58. Godoy R, Goodman E, Levins R, et al. Anthropometric variability in the USA: 1971-2002. Ann Hum Biol. 2005; 32(4):469–86. [PubMed: 16147396]
- 59. Akerman PA, Jenkins RL, Bistrian BR. Preoperative nutrition assessment in liver transplantation. Nutrition. 1993; 9(4):350–6. [PubMed: 8400592]
- 60. Horber FF, Thomi F, Casez JP, et al. Impact of hydration status on body composition as measured by dual energy x-ray absorptiometry in normal volunteers and patients on haemodialysis. Br J Radiol. 1992; 65(778):895–900. [PubMed: 1422663]
- 61. Pirlich M, Schutz T, Spachos T, et al. Bioelectrical impedance analysis is a useful bedside technique to assess malnutrition in cirrhotic patients with and without ascites. Hepatology. 2000; 32(6):1208–15. [PubMed: 11093726]
- 62. Madden AM, Morgan MY. The potential role of dual-energy x-ray absorptiometry in the assessment of body composition in cirrhotic patients. Nutrition. 1997; 13(1):40–5. [PubMed: 9058447]
- 63. Morgan MY, Madden AM, Jennings G, et al. Two-component models are of limited value for the assessment of body composition in patients with cirrhosis. Am J Clin Nutr. 2006; 84(5):1151–62. [PubMed: 17093169]

- 64. Chinn KS, Hannon JP. Relationship of muscle protein to other components of the fat-free mass. Am J Physiol. 1966; 211(4):993–7. [PubMed: 5926591]
- 65. Morgan MY, Madden AM. The assessment of body composition in patients with cirrhosis. Eur J Nucl Med. 1996; 23(2):213–25. [PubMed: 8925859]
- 66. Madden AM, Morgan MY. A comparison of skinfold anthropometry and bioelectrical impedance analysis for measuring percentage body fat in patients with cirrhosis. J Hepatol. 1994; 21(5):878– 83. [PubMed: 7890906]
- 67. Fuller NJ, Jebb SA, Laskey MA, et al. Four-component model for the assessment of body composition in humans: comparison with alternative methods, and evaluation of the density and hydration of fat-free mass. Clin Sci (Lond). 1992; 82(6):687–93. [PubMed: 1320550]
- 68. Mendenhall CL, Anderson S, Weesner RE, et al. Protein-calorie malnutrition associated with alcoholic hepatitis. Veterans Administration Cooperative Study Group on Alcoholic Hepatitis. Am J Med. 1984; 76(2):211–22. [PubMed: 6421159]
- 69. Lautz HU, Selberg O, Korber J, et al. Protein-calorie malnutrition in liver cirrhosis. Clin Investig. 1992; 70(6):478–86.
- 70. Addolorato G, Capristo E, Greco AV, et al. Influence of chronic alcohol abuse on body weight and energy metabolism: is excess ethanol consumption a risk factor for obesity or malnutrition? J Intern Med. 1998; 244(5):387–95. [PubMed: 9845854]
- 71. Caly WR, Strauss E, Carrilho FJ, et al. Different degrees of malnutrition and immunological alterations according to the aetiology of cirrhosis: a prospective and sequential study. Nutr J. 2003; 2:10. [PubMed: 14613508]
- 72. Caregaro L, Alberino F, Amodio P, et al. Malnutrition in alcoholic and virus-related cirrhosis. Am J Clin Nutr. 1996; 63(4):602–9. [PubMed: 8599326]
- 73. Roongpisuthipong C, Sobhonslidsuk A, Nantiruj K, et al. Nutritional assessment in various stages of liver cirrhosis. Nutrition. 2001; 17(9):761–5. [PubMed: 11527674]
- 74. Figueiredo FA, De Mello PR, Kondo M. Effect of liver cirrhosis on body composition: evidence of significant depletion even in mild disease. J Gastroenterol Hepatol. 2005; 20(2):209–16. [PubMed: 15683423]
- 75. Lehnert ME, Clarke DD, Gibbons JG, et al. Estimation of body water compartments in cirrhosis by multiple-frequency bioelectrical-impedance analysis. Nutrition. 2001; 17(1):31–4. [PubMed: 11165885]
- 76. de Carvalho L, Parise ER, Samuel D. Factors associated with nutritional status in liver transplant patients who survived the first year after transplantation. J Gastroenterol Hepatol. 2010; 25(2): 391–6. [PubMed: 19929929]
- 77. Hehir DJ, Jenkins RL, Bistrian BR, et al. Nutrition in patients undergoing orthotopic liver transplant. JPEN J Parenter Enteral Nutr. 1985; 9(6):695–700. [PubMed: 3934404]
- 78. Loguercio C, Sava E, Sicolo P, et al. Nutritional status and survival of patients with liver cirrhosis: anthropometric evaluation. Minerva Gastroenterol Dietol. 1996; 42(2):57–60. [PubMed: 8783865]
- 79. Mills PR, Shenkin A, Anthony RS, et al. Assessment of nutritional status and in vivo immune responses in alcoholic liver disease. Am J Clin Nutr. 1983; 38(6):849–59. [PubMed: 6650446]
- 80. Reisman Y, Gips CH, Lavelle SM. Assessment of liver cirrhosis severity in 1015 patients of the Euricterus database with Campbell-Child, Pugh-Child and with ascites and ascites-nutritional state (ANS) related classifications. Euricterus Project Management Group. Hepatogastroenterology. 1997; 44(17):1376–84. [PubMed: 9356858]
- 81. Sam J, Nguyen GC. Protein-calorie malnutrition as a prognostic indicator of mortality among patients hospitalized with cirrhosis and portal hypertension. Liver Int. 2009; 29(9):1396–402. [PubMed: 19602136]
- 82. Tai ML, Goh KL, Mohd-Taib SH, et al. Anthropometric, biochemical and clinical assessment of malnutrition in Malaysian patients with advanced cirrhosis. Nutr J. 2010; 9:27. [PubMed: 20576106]
- 83. Rantanen T, Harris T, Leveille SG, et al. Muscle strength and body mass index as long-term predictors of mortality in initially healthy men. J Gerontol A Biol Sci Med Sci. 2000; 55(3):M168–73. [PubMed: 10795731]

- 84. Durand F, Valla D. Assessment of the prognosis of cirrhosis: Child-Pugh versus MELD. J Hepatol. 2005; 42(Suppl 1):S100–7. [PubMed: 15777564]
- 85. Figueiredo FA, Perez RM, Freitas MM, et al. Comparison of three methods of nutritional assessment in liver cirrhosis: subjective global assessment, traditional nutritional parameters, and body composition analysis. J Gastroenterol. 2006; 41(5):476–82. [PubMed: 16799890]
- 86. Bathgate AJ, Hynd P, Sommerville D, et al. The prediction of acute cellular rejection in orthotopic liver transplantation. Liver Transpl Surg. 1999; 5(6):475–9. [PubMed: 10545533]
- 87. Deschenes M, Villeneuve JP, Dagenais M, et al. Lack of relationship between preoperative measures of the severity of cirrhosis and short-term survival after liver transplantation. Liver Transpl Surg. 1997; 3(5):532–7. [PubMed: 9346797]
- 88. Figueiredo F, Dickson ER, Pasha T, et al. Impact of nutritional status on outcomes after liver transplantation. Transplantation. 2000; 70(9):1347–52. [PubMed: 11087151]
- 89. Le Cornu KA, McKiernan FJ, Kapadia SA, et al. A prospective randomized study of preoperative nutritional supplementation in patients awaiting elective orthotopic liver transplantation. Transplantation. 2000; 69(7):1364–9. [PubMed: 10798755]
- 90. Muller MJ, Lautz HU, Plogmann B, et al. Energy expenditure and substrate oxidation in patients with cirrhosis: the impact of cause, clinical staging and nutritional state. Hepatology. 1992; 15(5): 782–94. [PubMed: 1568718]
- 91. Shahid M, Johnson J, Nightingale P, et al. Nutritional markers in liver allograft recipients. Transplantation. 2005; 79(3):359–62. [PubMed: 15699770]
- 92. Tsien CD, Shah S, Runkana A, et al. Reversal of sarcopenia predicts survival after transjugular intrahepatic portosystemic stent. Hepatology. 2011 Epub ahead of print.
- 93. Les I, Doval E, Flavia M, et al. Quality of life in cirrhosis is related to potentially treatable factors. Eur J Gastroenterol Hepatol. 2010; 22(2):221–7. [PubMed: 19794311]
- 94. Norman K, Kirchner H, Lochs H, et al. Malnutrition affects quality of life in gastroenterology patients. World J Gastroenterol. 2006; 12(21):3380–5. [PubMed: 16733855]
- 95. Poon RT, Yu WC, Fan ST, et al. Long-term oral branched chain amino acids in patients undergoing chemoembolization for hepatocellular carcinoma: a randomized trial. Aliment Pharmacol Ther. 2004; 19(7):779–88. [PubMed: 15043519]
- 96. Poupon RE, Chretien Y, Chazouilleres O, et al. Quality of life in patients with primary biliary cirrhosis. Hepatology. 2004; 40(2):489–94. [PubMed: 15368455]
- 97. Arguedas MR, DeLawrence TG, McGuire BM. Influence of hepatic encephalopathy on healthrelated quality of life in patients with cirrhosis. Dig Dis Sci. 2003; 48(8):1622–6. [PubMed: 12924658]
- 98. Wunsch E, Szymanik B, Post M, et al. Minimal hepatic encephalopathy does not impair healthrelated quality of life in patients with cirrhosis: a prospective study. Liver Int. 2011; 31(7):980–4. [PubMed: 21733087]
- 99. Moscucci F, Nardelli S, Pentassuglio I, et al. Previous overt hepatic encephalopathy rather than minimal hepatic encephalopathy impairs health-related quality of life in cirrhotic patients. Liver Int. 2011; 31(10):1505–10. [PubMed: 22093325]
- 100. Bajaj JS, Saeian K, Schubert CM, et al. Minimal hepatic encephalopathy is associated with motor vehicle crashes: the reality beyond the driving test. Hepatology. 2009; 50(4):1175–83. [PubMed: 19670416]
- 101. Bajaj JS. Minimal hepatic encephalopathy matters in daily life. World J Gastroenterol. 2008; 14(23):3609–15. [PubMed: 18595126]
- 102. Montomoli J, Holland-Fischer P, Bianchi G, et al. Body composition changes after transjugular intrahepatic portosystemic shunt in patients with cirrhosis. World J Gastroenterol. 2010; 16(3): 348–53. [PubMed: 20082481]
- 103. Campillo B, Bories PN, Pornin B, et al. Influence of liver failure, ascites, and energy expenditure on the response to oral nutrition in alcoholic liver cirrhosis. Nutrition. 1997; 13(7–8):613–21. [PubMed: 9263252]
- 104. Ndraha S, Hasan I, Simadibrata M. The effect of L-ornithine L-aspartate and branch chain amino acids on encephalopathy and nutritional status in liver cirrhosis with malnutrition. Acta Med Indones. 2011; 43(1):18–22. [PubMed: 21339541]

- 105. Dong MH, Saab S. Complications of cirrhosis. Dis Mon. 2008; 54(7):445–56. [PubMed: 18570914]
- 106. Moller S, Bendtsen F, Christensen E, et al. Prognostic variables in patients with cirrhosis and oesophageal varices without prior bleeding. J Hepatol. 1994; 21(6):940–6. [PubMed: 7699257]
- 107. Olde Damink SW, Jalan R, Dejong CH. Interorgan ammonia trafficking in liver disease. Metab Brain Dis. 2009; 24(1):169–81. [PubMed: 19067143]
- 108. Soros P, Bottcher J, Weissenborn K, et al. Malnutrition and hypermetabolism are not risk factors for the presence of hepatic encephalopathy: a cross-sectional study. J Gastroenterol Hepatol. 2008; 23(4):606–10. [PubMed: 18005013]
- 109. Kalaitzakis E, Bjornsson E. Hepatic encephalopathy in patients with liver cirrhosis: is there a role of malnutrition? World J Gastroenterol. 2008; 14(21):3438–9. [PubMed: 18528945]
- 110. Periyalwar P, Dasarathy J, Tsien C, et al. Sarcopenia of cirrhosis results in more severe and frequent episodes of hepatic encephalopathy. Hepatology. 2011 Epub ahead of print.
- 111. Dasarathy S, Mullen KD, Conjeevaram HS, et al. Preservation of portal pressure improves growth and metabolic profile in the male portacaval-shunted rat. Dig Dis Sci. 2002; 47(9):1936–42. [PubMed: 12353833]
- 112. Dasarathy S, Yang Y, Muc S, et al. Ammonia causes elevated skeletal muscle myostatin and reduced myoblast proliferation. Hepatology. 2009 Epub ahead of print.
- 113. McCullough AJ, Mullen KD, Kalhan SC. Defective nonoxidative leucine degradation and endogenous leucine flux in cirrhosis during an amino acid infusion. Hepatology. 1998; 28(5): 1357–64. [PubMed: 9794922]
- 114. Kung T, Springer J, Doehner W, et al. Novel treatment approaches to cachexia and sarcopenia: highlights from the 5th Cachexia Conference. Expert Opin Investig Drugs. 2010; 19(4):579–85.
- 115. Glass DJ. Skeletal muscle hypertrophy and atrophy signaling pathways. Int J Biochem Cell Biol. 2005; 37(10):1974–84. [PubMed: 16087388]
- 116. Wagers AJ, Conboy IM. Cellular and molecular signatures of muscle regeneration: current concepts and controversies in adult myogenesis. Cell. 2005; 122(5):659–67. [PubMed: 16143100]
- 117. Le GF, Rudnicki MA. Skeletal muscle satellite cells and adult myogenesis. Curr Opin Cell Biol. 2007; 19(6):628–33. [PubMed: 17996437]
- 118. Garlick PJ, McNurlan MA, Essen P, et al. Measurement of tissue protein synthesis rates in vivo: a critical analysis of contrasting methods. Am J Physiol. 1994; 266(3 Pt 1):E287–97. [PubMed: 8166249]
- 119. Welle S, Bhatt K, Pinkert CA. Myofibrillar protein synthesis in myostatin-deficient mice. Am J Physiol Endocrinol Metab. 2006; 290(3):E409–15. [PubMed: 16219664]
- 120. Mullen KD, Denne SC, McCullough AJ, et al. Leucine metabolism in stable cirrhosis. Hepatology. 1986; 6(4):622–30. [PubMed: 3089896]
- 121. Tessari P, Barazzoni R, Kiwanuka E, et al. Impairment of albumin and whole body postprandial protein synthesis in compensated liver cirrhosis. Am J Physiol Endocrinol Metab. 2002; 282(2):E304–11. [PubMed: 11788361]
- 122. Tessari P, Inchiostro S, Barazzoni R, et al. Fasting and postprandial phenylalanine and leucine kinetics in liver cirrhosis. Am J Physiol. 1994; 267(1 Pt 1):E140–9. [PubMed: 8048503]
- 123. Morrison WL, Bouchier IA, Gibson JN, et al. Skeletal muscle and whole-body protein turnover in cirrhosis. Clin Sci (Lond). 1990; 78(6):613–9. [PubMed: 2165895]
- 124. Tessari P, Biolo G, Inchiostro S, et al. Leucine and phenylalanine kinetics in compensated liver cirrhosis: effects of insulin. Gastroenterology. 1993; 104(6):1712–21. [PubMed: 8500731]
- 125. Tugtekin I, Wachter U, Barth E, et al. Phenylalanine kinetics in healthy volunteers and liver cirrhotics: implications for the phenylalanine breath test. Am J Physiol Endocrinol Metab. 2002; 283(6):E1223–31. [PubMed: 12424105]
- 126. McCullough AJ, Tavill AS. Disordered energy and protein metabolism in liver disease. Semin Liver Dis. 1991; 11(4):265–77. [PubMed: 1763333]

Periyalwar and Dasarathy Page 20

- 127. Guillet C, Boirie Y, Walrand S. An integrative approach to in-vivo protein synthesis measurement: from whole tissue to specific proteins. Curr Opin Clin Nutr Metab Care. 2004; 7(5):531–8. [PubMed: 15295273]
- 128. Holecek M, Skopec F, Sprongl L. Protein metabolism in cirrhotic rats: effect of dietary restriction. Ann Nutr Metab. 1995; 39(6):346–54. [PubMed: 8678470]
- 129. Lundborg H, Hamberger A. Effects of portacaval anastomosis on liver and brain protein synthesis in rats. Surgery. 1977; 82(5):643–7. [PubMed: 918852]
- 130. Dunlop DS, Kaufman H, Zanchin G, et al. Protein synthesis rates in rats with portacaval shunts. J Neurochem. 1984; 43(5):1487–9. [PubMed: 6491663]
- 131. Wang YY, Lin SY, Chuang YH, et al. Protein nitration is associated with increased proteolysis in skeletal muscle of bile duct ligation-induced cirrhotic rats. Metabolism. 2010; 59(4):468–72. [PubMed: 19846167]
- 132. Mizushima N, Komatsu M. Autophagy: renovation of cells and tissues. Cell. 2011; 147(4):728– 41. [PubMed: 22078875]
- 133. Masiero E, Agatea L, Mammucari C, et al. Autophagy is required to maintain muscle mass. Cell Metab. 2009; 10(6):507–15. [PubMed: 19945408]
- 134. Kandarian SC, Jackman RW. Intracellular signaling during skeletal muscle atrophy. Muscle Nerve. 2006; 33(2):155–65. [PubMed: 16228971]
- 135. Machida S, Booth FW. Regrowth of skeletal muscle atrophied from inactivity. Med Sci Sports Exerc. 2004; 36(1):52–9. [PubMed: 14707768]
- 136. Bornemann A, Maier F, Kuschel R. Satellite cells as players and targets in normal and diseased muscle. Neuropediatrics. 1999; 30(4):167–75. [PubMed: 10569207]
- 137. Hepple RT. Dividing to keep muscle together: the role of satellite cells in aging skeletal muscle. Sci Aging Knowledge Environ. 2006; 2006(3):e3.
- 138. Anthony JC, Yoshizawa F, Anthony TG, et al. Leucine stimulates translation initiation in skeletal muscle of postabsorptive rats via a rapamycin-sensitive pathway. J Nutr. 2000; 130(10):2413–9. [PubMed: 11015466]
- 139. Bodine SC, Stitt TN, Gonzalez M, et al. Akt/mTOR pathway is a crucial regulator of skeletal muscle hypertrophy and can prevent muscle atrophy in vivo. Nat Cell Biol. 2001; 3(11):1014–9. [PubMed: 11715023]
- 140. Lokireddy S, Mouly V, Butler-Browne G, et al. Myostatin promotes the wasting of human myoblast cultures through promoting ubiquitin-proteasome pathway-mediated loss of sarcomeric proteins. Am J Physiol Cell Physiol. 2011; 301(6):C1316–24. [PubMed: 21900687]
- 141. Zimmers TA, Davies MV, Koniaris LG, et al. Induction of cachexia in mice by systemically administered myostatin. Science. 2002; 296(5572):1486–8. [PubMed: 12029139]
- 142. Morissette MR, Cook SA, Buranasombati C, et al. Myostatin inhibits IGF-I-induced myotube hypertrophy through Akt. Am J Physiol Cell Physiol. 2009; 297(5):C1124–32. [PubMed: 19759331]
- 143. Trendelenburg AU, Meyer A, Rohner D, et al. Myostatin reduces Akt/TORC1/p70S6K signaling, inhibiting myoblast differentiation and myotube size. Am J Physiol Cell Physiol. 2009; 296(6):C1258–70. [PubMed: 19357233]
- 144. Lee JY, Hopkinson NS, Kemp PR. Myostatin induces autophagy in skeletal muscle in vitro. Biochem Biophys Res Commun. 2011; 415(4):632–6. [PubMed: 22079631]
- 145. Goldspink G. Age-related muscle loss and progressive dysfunction in mechano-sensitive growth factor signaling. Ann N Y Acad Sci. 2004; 1019:294–8. [PubMed: 15247032]
- 146. Bush JA, Kimball SR, O'Connor PM, et al. Translational control of protein synthesis in muscle and liver of growth hormone-treated pigs. Endocrinology. 2003; 144(4):1273–83. [PubMed: 12639910]
- 147. Latres E, Amini AR, Amini AA, et al. Insulin-like growth factor-1 (IGF-1) inversely regulates atrophy-induced genes via the phosphatidylinositol 3-kinase/Akt/mammalian target of rapamycin (PI3K/Akt/mTOR) pathway. J Biol Chem. 2005; 280(4):2737–44. [PubMed: 15550386]
- 148. Berger MJ, Doherty TJ. Sarcopenia: prevalence, mechanisms, and functional consequences. Interdiscip Top Gerontol. 2010; 37:94–114. [PubMed: 20703058]

- 149. Doherty TJ. Invited review: aging and sarcopenia. J Appl Physiol. 2003; 95(4):1717–27. [PubMed: 12970377]
- 150. Pagadala M, Dasarathy S, Eghtesad B, et al. Posttransplant metabolic syndrome: an epidemic waiting to happen. Liver Transpl. 2009; 15(12):1662–70. [PubMed: 19938136]
- 151. Guillet C, Boirie Y. Insulin resistance: a contributing factor to age-related muscle mass loss? Diabetes Metab. 2005; 31(Spec No 2):5S20–6. [PubMed: 16415762]
- 152. Michel RN, Chin ER, Chakkalakal JV, et al. Ca2+/calmodulin-based signalling in the regulation of the muscle fibre phenotype and its therapeutic potential via modulation of utrophin A and myostatin expression. Appl Physiol Nutr Metab. 2007; 32(5):921–9. [PubMed: 18059617]
- 153. Sakuma K, Nakao R, Aoi W, et al. Cyclosporin A treatment upregulates Id1 and Smad3 expression and delays skeletal muscle regeneration. Acta Neuropathol. 2005; 110(3):269–80. [PubMed: 15986223]
- 154. Lipina C, Kendall H, McPherron AC, et al. Mechanisms involved in the enhancement of mammalian target of rapamycin signalling and hypertrophy in skeletal muscle of myostatindeficient mice. FEBS Lett. 2010; 584(11):2403–8. [PubMed: 20412806]
- 155. Scolapio JS, Bowen J, Stoner G, et al. Substrate oxidation in patients with cirrhosis: comparison with other nutritional markers. JPEN J Parenter Enteral Nutr. 2000; 24(3):150–3. [PubMed: 10850939]
- 156. Cabre E, Gonzalez-Huix F, bad-Lacruz A, et al. Effect of total enteral nutrition on the short-term outcome of severely malnourished cirrhotics. A randomized controlled trial. Gastroenterology. 1990; 98(3):715–20. [PubMed: 2105256]
- 157. Plauth M, Cabre E, Riggio O, et al. ESPEN guidelines on enteral nutrition: liver disease. Clin Nutr. 2006; 25(2):285–94. [PubMed: 16707194]
- 158. Bucuvalas JC, Cutfield W, Horn J, et al. Resistance to the growth-promoting and metabolic effects of growth hormone in children with chronic liver disease. J Pediatr. 1990; 117(3):397– 402. [PubMed: 2391594]
- 159. Moller S, Becker U, Gronbaek M, et al. Short-term effect of recombinant human growth hormone in patients with alcoholic cirrhosis. J Hepatol. 1994; 21(5):710–7. [PubMed: 7534320]
- 160. Donaghy A, Ross R, Wicks C, et al. Growth hormone therapy in patients with cirrhosis: a pilot study of efficacy and safety. Gastroenterology. 1997; 113(5):1617–22. [PubMed: 9352864]
- 161. Wallace JD, bbott-Johnson WJ, Crawford DH, et al. GH treatment in adults with chronic liver disease: a randomized, double-blind, placebo-controlled, crossover study. J Clin Endocrinol Metab. 2002; 87(6):2751–9. [PubMed: 12050245]
- 162. Cabral CM, Burns DL. Low-protein diets for hepatic encephalopathy debunked: let them eat steak. Nutr Clin Pract. 2011; 26(2):155–9. [PubMed: 21447768]
- 163. Gundling F, Seidl H, Pehl C, et al. How close do gastroenterologists follow specific guidelines for nutrition recommendations in liver cirrhosis? A survey of current practice. Eur J Gastroenterol Hepatol. 2009; 21(7):756–61. [PubMed: 19322099]
- 164. Heyman JK, Whitfield CJ, Brock KE, et al. Dietary protein intakes in patients with hepatic encephalopathy and cirrhosis: current practice in NSW and ACT. Med J Aust. 2006; 185(10): 542–3. [PubMed: 17115965]
- 165. Kondrup J, Nielsen K, Juul A. Effect of long-term refeeding on protein metabolism in patients with cirrhosis of the liver. Br J Nutr. 1997; 77(2):197–212. [PubMed: 9135367]
- 166. Porter C, Cohen NH. Indirect calorimetry in critically ill patients: role of the clinical dietitian in interpreting results. J Am Diet Assoc. 1996; 96(1):49–57. [PubMed: 8537570]
- 167. Kondrup J, Muller MJ. Energy and protein requirements of patients with chronic liver disease. J Hepatol. 1997; 27(1):239–47. [PubMed: 9252101]
- 168. Nielsen K, Kondrup J, Martinsen L, et al. Long-term oral refeeding of patients with cirrhosis of the liver. Br J Nutr. 1995; 74(4):557–67. [PubMed: 7577893]
- 169. Plauth M, Roske AE, Romaniuk P, et al. Post-feeding hyperammonaemia in patients with transjugular intrahepatic portosystemic shunt and liver cirrhosis: role of small intestinal ammonia release and route of nutrient administration. Gut. 2000; 46(6):849–55. [PubMed: 10807899]

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- 170. Swart GR, van den Berg JW, van Vuure JK, et al. Minimum protein requirements in liver cirrhosis determined by nitrogen balance measurements at three levels of protein intake. Clin Nutr. 1989; 8(6):329–36. [PubMed: 16837309]
- 171. Vaisman N, Katzman H, Carmiel-Haggai M, et al. Breakfast improves cognitive function in cirrhotic patients with cognitive impairment. Am J Clin Nutr. 2010; 92(1):137–40. [PubMed: 20444959]
- 172. Khanna S, Gopalan S. Role of branched-chain amino acids in liver disease: the evidence for and against. Curr Opin Clin Nutr Metab Care. 2007; 10(3):297–303. [PubMed: 17414498]
- 173. Kachaamy T, Bajaj JS. Diet and cognition in chronic liver disease. Curr Opin Gastroenterol. 2011; 27(2):174–9. [PubMed: 20975555]
- 174. Anthony JC, Anthony TG, Kimball SR, et al. Signaling pathways involved in translational control of protein synthesis in skeletal muscle by leucine. J Nutr. 2001; 131(3):856S–60S. [PubMed: 11238774]
- 175. Dreyer HC, Drummond MJ, Pennings B, et al. Leucine-enriched essential amino acid and carbohydrate ingestion following resistance exercise enhances mTOR signaling and protein synthesis in human muscle. Am J Physiol Endocrinol Metab. 2008; 294(2):E392–400. [PubMed: 18056791]
- 176. Drummond MJ, Rasmussen BB. Leucine-enriched nutrients and the regulation of mammalian target of rapamycin signalling and human skeletal muscle protein synthesis. Curr Opin Clin Nutr Metab Care. 2008; 11(3):222–6. [PubMed: 18403916]
- 177. Koopman R, Wagenmakers AJ, Manders RJ, et al. Combined ingestion of protein and free leucine with carbohydrate increases postexercise muscle protein synthesis in vivo in male subjects. Am J Physiol Endocrinol Metab. 2005; 288(4):E645–53. [PubMed: 15562251]
- 178. Katsanos CS, Kobayashi H, Sheffield-Moore M, et al. A high proportion of leucine is required for optimal stimulation of the rate of muscle protein synthesis by essential amino acids in the elderly. Am J Physiol Endocrinol Metab. 2006; 291(2):E381–7. [PubMed: 16507602]
- 179. Lee SJ, Glass DJ. Treating cancer cachexia to treat cancer. Skelet Muscle. 2011; 1(1):2. [PubMed: 21798080]
- 180. Rode A, Fourlanos S, Nicoll A. Oral vitamin D replacement is effective in chronic liver disease. Gastroenterol Clin Biol. 2010; 34(11):618–20. [PubMed: 20801590]
- 181. Stamoulis I, Kouraklis G, Theocharis S. Zinc and the liver: an active interaction. Dig Dis Sci. 2007; 52(7):1595–612. [PubMed: 17415640]
- 182. Yoshida Y, Higashi T, Nouso K, et al. Effects of zinc deficiency/zinc supplementation on ammonia metabolism in patients with decompensated liver cirrhosis. Acta Med Okayama. 2001; 55(6):349–55. [PubMed: 11779097]
- 183. Bavdekar A, Bhave S, Pandit A. Nutrition management in chronic liver disease. Indian J Pediatr. 2002; 69(5):427–31. [PubMed: 12061678]
- 184. Crawford BA, Labio ED, Strasser SI, et al. Vitamin D replacement for cirrhosis-related bone disease. Nat Clin Pract Gastroenterol Hepatol. 2006; 3(12):689–99. [PubMed: 17130879]
- 185. Drummond MJ, Dreyer HC, Pennings B, et al. Skeletal muscle protein anabolic response to resistance exercise and essential amino acids is delayed with aging. J Appl Physiol. 2008; 104(5): 1452–61. [PubMed: 18323467]
- 186. Wilborn CD, Taylor LW, Greenwood M, et al. Effects of different intensities of resistance exercise on regulators of myogenesis. J Strength Cond Res. 2009; 23(8):2179–87. [PubMed: 19826309]
- 187. Koopman R, Saris WH, Wagenmakers AJ, et al. Nutritional interventions to promote postexercise muscle protein synthesis. Sports Med. 2007; 37(10):895–906. [PubMed: 17887813]
- 188. Garcia-Pagan JC, Santos C, Barbera JA, et al. Physical exercise increases portal pressure in patients with cirrhosis and portal hypertension. Gastroenterology. 1996; 111(5):1300–6. [PubMed: 8898644]
- 189. Garcia PS, Cabbabe A, Kambadur R, et al. Brief-reports: elevated myostatin levels in patients with liver disease: a potential contributor to skeletal muscle wasting. Anesth Analg. 2010; 111(3):707–9. [PubMed: 20686014]

- 190. Harrison J, McKiernan J, Neuberger JM. A prospective study on the effect of recipient nutritional status on outcome in liver transplantation. Transpl Int. 1997; 10(5):369–74. [PubMed: 9287402]
- 191. Stephenson GR, Moretti EW, El-Moalem H, et al. Malnutrition in liver transplant patients: preoperative subjective global assessment is predictive of outcome after liver transplantation. Transplantation. 2001; 72(4):666–70. [PubMed: 11544428]
- 192. Campillo B, Richardet JP, Scherman E, et al. Evaluation of nutritional practice in hospitalized cirrhotic patients: results of a prospective study. Nutrition. 2003; 19(6):515–21. [PubMed: 12781851]

Proliferation

Differentiation

Fig. 1.

Regulation of skeletal muscle mass. The protein synthesis and satellite cell (myogenically committed stem cells) contribute to muscle growth and reversal of atrophy. These are regulated primarily by myostatin and IGF1. The proteolysis is mediated primarily by the ubiquitin proteasome pathway with a variable contribution by the lysosomal cathepsin mediated autophagy pathway.

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Fig. 2.

Integration of the 3 major pathways that regulate skeletal muscle mass. Myostatin and IGF1 regulate muscle growth via transcriptional and posttranslational regulation of myogenic genes. The ubiquitin proteasome pathway is responsible for proteolysis. All 3 pathways crosstalk at multiple levels, including Akt, mTOR, AMP kinase, and FOXO. PIP2, phosphatidyl inositor bisphosphate; PIP3, phosphatidylinositol 3 phosphate; IGF1R, IGF1 receptor; Alk5, activinlike kinase 5, forms a heterodimeric complex with generic TGFβ receptor for myostatin; Act IIbr, activin II b receptor; GSK, 3β glycogen synthase kinase; eIF, eukaryotic initiation factor; 4E BP1, 4 E binding protein 1; IRS 1, insulin receptor substrate that is downstream of both insulin and IGF1 receptor; MURf, muscle ring finger protein, final component in the ubiquitin proteasome pathway with atrogin; PCNA, proliferating cell nuclear antigen, a marker of satellite cell proliferation.

Table 1

Prevalence of malnutrition in cirrhosis Prevalence of malnutrition in cirrhosis

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Clin Liver Dis. Author manuscript; available in PMC 2015 April 02.

Tai et al,⁸² 2010 36 MAMC 50 50 MAMC

 36

Tai et al, 82 2010

MAMC
SGA

 SGA 40 40

 $60 + 40$

Alcohol
Viral

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Abbreviations: BIABCM, bioelectrical impedance analyzer measured body cell mass; BIATBP, bioelectrical impedance analyzer measured total body protein; BMI, body mass index; SGA, subjective Abbreviations: BIABCM, bioelectrical impedance analyzer measured body cell mass; BIATBP, bioelectrical impedance analyzer measured total body protein; BMI, body mass index; SGA, subjective global assessment; TBF, total body fat; TBP, total body protein; TSF, triceps skinfold thickness. global assessment; TBF, total body fat; TBP, total body protein; TSF, triceps skinfold thickness.

Summary: Eighteen studies, with a total of 3041 of subjects enrolled, measure malnutrition by a variety of methods, including SGA, MAMC, TSF, BIA, and clinical assessment. The prevalence of Summary: Eighteen studies, with a total of 3041 of subjects enrolled, measure malnutrition by a variety of methods, including SGA, MAMC, TSF, BIA, and clinical assessment. The prevalence of malnutrition ranges from 6.1% to 100.0%. Cause of cirrhosis included alcohol, viral, primary biliary, autoimmune, sclerosing cholangitis, and Wilson disease. malnutrition ranges from 6.1% to 100.0%. Cause of cirrhosis included alcohol, viral, primary biliary, autoimmune, sclerosing cholangitis, and Wilson disease.

Table 2

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Impact of malnutrition on survival in cirrhosis Impact of malnutrition on survival in cirrhosis

Summary: The impact of malnutrition on survival in cirrhosis is described in these 13 studies (n = 1187). The prevalence of pretransplant malnutrition was between 9.4% and 69.0%. Malnutrition is defined Summary: The impact of malnutrition on survival in cirrhosis is described in these 13 studies (n = 1187). The prevalence of pretransplant malnutrition was between 9.4% and 69.0%. Malnutrition is defined by BIA, HG, TSF, SGA, and total psoas area. The presence of malnutrition increased morbidity and mortality, including acute rejection after transplant and prolonged ICU stay. There is a statistically
significant correlatio by BIA, HG, TSF, SGA, and total psoas area. The presence of malnutrition increased morbidity and mortality, including acute rejection after transplant and prolonged ICU stay. There is a statistically significant correlation between the severity of malnutrition and mortality.

Table 3

Clin Liver Dis. Author manuscript; available in PMC 2015 April 02.

Summary: The impact of malnutrition on quality of life in cirrhosis is described in the previous 7 studies (n = 1104). Malnutrition is defined by BIA, MAMC, TSF, SGA, and branched chain amino acids levels. The prevalence of malnutrition ranged from 19% to 59%. Malnourished patients had statistically significant increase in gastrointestinal symptoms and decreased quality of life on the CLQD, SF-36,

Summary: The impact of malnutrition on quality of life in cirrhoss is described in the previous 7 studies (n = 1104). Malnutrition is defined by BIA, MAMC, TSF, SGA, and branched chain amino acids
levels. The prevalence o

and NHP questionnaire.

Table 4

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P = .001); 33.3% of malnourished subjects

1.8% of well-nourished subjects developed uncontrolled ascites,

P = .007) complications at follow-

P = .00), hepatorenal (13% vs 0%, *P*

influenced only by malnutrition (SGA A-B, C) HR 0.18; $P =$.

0003

Portal HTN Clinical diagnosis 6.1 PCM was higher among patients with cirrhosis and PHTN compared with general medical inpatients

 $\overline{61}$

Clinical diagnosis

114,703

Portal HTN

Sam and Nguyen, 81 2009

(6.1% vs 1.9%,

Sam and Nguyen,⁸¹ 2009 In patients with cirrhosis and PHTN, significantly higher rates of malnutrition in patients with alcohol

abuse compared with those without (7.6% vs 4.7%,

abuse compared with those without (7.6% vs 4.7%, $P<0.001$), greater prevalence of ascites (64.6% vs 4.7.9%, $P<0.001$), and hepatorenal syndrome (5.1% vs 2.8% $P<0.001$) but not HE (60.8% vs 59.7%, $P = 1.1$) among patien

In patients with cirrhosis and PHTN, significantly higher rates of malnutrition in patients with alcohol

P<.0001), and hepatorenal syndrome (5.1% vs 2.8%

= .11) among patients with malnutrition and cirrhosis with PHTN

Montomoli et al,¹⁰² 2010 21 BIA 30 BIA 30 TIPS procedure lowered portal pressure before compared with after $(6.0 \pm 2.1$ mm Hg vs 15.8 ± 4.8 mm Hg,

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ВIА

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Montomoli et al, 102 2010

 kg to 12.7 ± 5.6 kg,

P<.001). After TIPS, normal-weight patients had an increase in dry lean mass (from 10.9 ± 5.9

P = .031) and TBW (from 34.5 ± 7.6 L to 40.2 ± 10.8 L,

40 Patients with vs without malnutrition had more frequent episodes of HE (46% vs 27%;

P = .202), severity of cirrhosis by CP score (2.5% vs 2.2%,

37%, = .275)

was no difference in episodes of HE comparing alcohol cause of liver disease vs other cause (49% vs

Patients with vs without malnutrition had more frequent episodes of HE (46% vs 27% ; $P = .031$); There

P = .007)

P = .031); There

P = .09) or by MELD (6% vs 5%, *P*

P<.0001), greater prevalence of ascites (64.6% vs

P<.0001) but not HE (60.8% vs 59.7%, *P*

P<.0001; OR: 1.55, 95% CI: 1.4–1.7)

PCM was higher among patients with cirrhosis and PHTN compared with general medical inpatients (6.1% vs 1.9%, P<.0001; OR: 1.55, 95% CI: 1.4-1.7)

HE 128 Anthropometry, MAC <fifth

128

 $H\!E$

percentile

Anthropometry, MAC <fifth

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MELD, model end-stage liver disease; NAP, nonascitic patients with cirrhosis; OR, odds ratio; PCM, protein calorie malnutrition; PHTN, portal hypertension; PNI, prognostic nutritional index; SBP, A Discretions. Deviations, Deviations, Deviations, O. Controllers, OR, odds ratio; PCM, protein calorie malmurition; PHTN, portal hypertension; PNT, prognostic nutritional index; SBP, portal index; SBP, and a basic of the spontaneous bacterial peritonitis; TAP, patients with cirrhosis with tense ascites; TBW, total body water; TSF, triceps skinfold. spontaneous bacterial peritonitis; TAP, patients with cirrhosis with tense ascites; TBW, total body water; TSF, triceps skinfold.

Summary: Seven studies (n = 751) describe the impact of malnutrition on the complications of liver disease. Malnutrition is defined by BMI, HG, MAMC, TSF, and SGA. Prevalence of malnutrition is
6.1% to 67.0%. Complications 6.1% to 67.0%. Complications of liver disease included are ascites, SBP, portal hypertension, hepatorenal syndrome, and HE. There is a statistically significant increase in complications in patients with Summary: Seven studies (n = 751) describe the impact of malnutrition on the complications of liver disease. Malnutrition is defined by BMI, HG, MAMC, TSF, and SGA. Prevalence of malnutrition is malnutrition. malnutrition.

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Table 5

Impact of malnutrition on liver transplant outcomes in cirrhosis Impact of malnutrition on liver transplant outcomes in cirrhosis

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Abbreviations: BCM, body cell mass; BIA, bioelectric impedance; BMI, body mass index; CI, confidence interval; HR, hazard ratio; ICU, intensive care unit; MAMC, midarm muscle circumference; OLT, Abbreviations: BCM, body cell mass; BIA, bioelectric impedance; BMI, body mass index; CI, confidence interval; HR, hazard ratio; ICU, intensive care unit; MAMC, midarm muscle circumference; OLT, orthotopic liver transplant; PCM, protein caloric malnutrition; REE, resting energy expenditure; TPA, total psoas area; TSF, triceps skinfold; UNOS, united network of organ sharing. orthotopic liver transplant; PCM, protein calorie malnutrition; REE, resting energy expenditure; TPA, total psoas area; TSF, triceps skinfold; UNOS, united network of organ sharing.

Summary: Eight studies (n = 754) assessed the impact of malnutrition on liver-transplant outcomes. Prevalence of malnutrition ranged form 9.4% to 79%. Malnutrition was defined by BIA, HG, MAMC, Summary. Eight studies (n = 754) assessed the impact of malnutrition on liver-transplant outcomes. Prevalence of malnutrition ranged form 9.4% to 79%. Malnutrition was defined by BIA, HG, MAMC, TSF, and SGA. Severe malnutrition independently predicted intraoperative complications, increased episodes of infection, increased length of stay in the ICU, acute cellular rejection, and increased TSF, and SGA. Severe malnutrition independently predicted intraoperative complications, increased episodes of infection, increased length of stay in the ICU, acute cellular rejection, and increased transfusion requirements. transfusion requirements.

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Table 6

Therapeutic options for malnutrition in cirrhosis Therapeutic options for malnutrition in cirrhosis

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Abbreviations: BCAA, branched chain amino acid; BW, body weight; EtOH cirrhosis, alcoholic cirrhosis; FM, fat mass; LBM, lean body mass; MAMA, midarm muscle area; MAMC, midarm muscle Abbreviations: BCAA, branched chain amino acid; BW, body weight; EtOH cirrhosis, alcoholic cirrhosis; FM, fat mass; LBM, lean body mass; MAMA, midarm muscle area; MAMC, midarm muscle circumference; MJ, megajoule; PCM, protein calorie malnutrition; TSF, triceps skinfold. circumference; MJ, megajoule; PCM, protein calorie malnutrition; TSF, triceps skinfold.

Summary: Six studies (n = 126) describe the therapeutic interventions for malnutrition in cirrhosis. Malnutrition is defined by BCAA, BW, LBM, MAMC, serum ammonia and glutamine level, and TSF. Summary. Six studies (n = 126) describe the therapeutic interventions for malnutrition in cirrhosis. Malnutrition is defined by BCAA, BW, LBM, MAMC, serum ammonia and glutamine level, and TSF. Malnourished patients are noted to have increase in other complications of cirrhosis, including ascites, spontaneous bacterial peritonitis, portal hypertension, hepatorenal syndrome, and HE. There is a Malnourished patients are noted to have increase in other complications of cirrhosis, including ascites, spontaneous bacterial peritonitis, portal hypertension, hepatorenal syndrome, and HE. There is a Prevalence of malnutrition is 50% to 73% and was not assessed in 2 studies. Therapeutic options include modified eating patterns with increased protein through either oral nutrition or parenteral. Prevalence of malnutrition is 50% to 73% and was not assessed in 2 studies. Therapeutic options include modified eating patterns with increased protein through either oral nutrition or parenteral. statistically significant increase in LBM and FM in patients with cirrhosis with improved oral or parenteral nutrition. statistically significant increase in LBM and FM in patients with cirrhosis with improved oral or parenteral nutrition.

Table 7

Pathophysiology-based therapeutic options

Abbreviation: BCAA, branched chain amino acid.