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DNMT3A mutational status affects the results of dose-escalated induction therapy in acute myelogenous leukemia

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Abstract

Purpose—DNA methyltransferase 3A (*DNMT3A*) is one of the commonly mutated genes in acute myelogenous leukemia (AML). Reports on the prognostic significance of *DNMT3A* mutations have been inconsistent, and most of the data is available only for patients 60 years of age or younger. We hypothesized that this inconsistency is due to an interaction between the dose of anthracycline used in induction therapy and *DNMT3A* status. We studied whether patients with *DNMT3A*-mutated AML treated with standard dose anthracyclines had an inferior survival compared to patients with other mutation profiles or those who received high dose therapy.

Experimental design—152 patients in this retrospective cohort study (median age, 54 years) with *de-novo* AML underwent induction therapy and next-generation sequencing of 33 commonly mutated genes in hematologic malignancies, including *DNMT3A*, *FLT3-ITD*, *NPM1*, *and IDH1/2*. Cox regression was used to if those with *DNMT3A* mutations who were treated with standard dose anthracycline had inferior survival.

Results—*DNMT3A* mutations, found in 32% of patients, were not associated with an inferior survival. Dose escalation of anthracycline in the induction regimen was associated with improved survival in those with *DNMT3A* mutations but not those with wild-type *DNMT3A*. Patients with *DNMT3A* mutations who received standard dose induction had shorter survival time than other patient groups (10.1 months vs. 19.8 months, p=0.0129). This relationship remained significant (HR: 1.90, p=0.006) controlling for multiple variables.

Conclusions—Patients with *DNMT3A*-mutated AML have an inferior survival when treated with standard-dose anthracycline induction therapy. This group should be considered for high-dose induction therapy.

The authors declare no conflicts of interest.

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Keywords

Acute myeloid leukemia; DNMT3A; prognosis; induction chemotherapy

Introduction

The choice of induction and post-remission therapy in AML is guided by certain prognostic factors. Karyotype has historically been the largest determinant of prognosis ²³, but this inadequately predicts outcome in a large proportion of patients, particularly those with no karyotypic abnormalities. Recurrent gene mutations in *NPM1* and *CEBPA*, and internal tandem duplications (ITD) in *FLT3* have been recognized as important in AML pathogenesis and prognosis. ⁴ More recently, an additional class of genes recurrently mutated in AML genomes has been identified that normally function in the epigenetic regulation of transcription. These include *DNMT3A*, *TET2*, *IDH1/IDH2*, and *ASXL1*. ^{5,6789101} A growing body of evidence supports a pathogenic role for these mutations in AML. ¹¹

DNMT3A is one of the most commonly mutated genes in AML genomes and has been the topic of significant analysis since it was first noted by Ley et al. ¹² It encodes one of the DNA methyltransferases, and along with *DNMT3B*, is responsible for adding a methyl group to cytosine/guanine residues. The prevalence of mutations in *DNMT3A* ranges from 18–36% and is enriched in normal karyotype AML. ¹³¹²¹⁴¹⁵⁸¹⁶¹⁷¹⁸ The most frequently mutated residue of the *DNMT3A* gene occurs in the methyltransferase domain at Arginine 882, leading to decreased methylation activity *in vitro* ¹⁵ as well as decreased methylation levels in select genomic regions. ¹⁵¹² Additional mutations seen throughout the gene have also been described and are thought to also disrupt normal methylation activity. However, it has not been consistently associated with an altered gene expression pattern. ¹²

Despite an incomplete understanding of the functional changes induced by *DNMT3A* mutations, the initial studies of this gene mutation consistently showed that it conferred a poor prognosis. ¹²¹⁴⁸ However, more recent studies have contradicted this finding, and have shown no difference in overall survival based on *DNMT3A* mutational status in large, homogenously treated patient cohorts. ¹¹⁹¹⁸¹³ While the differences in prognostic significance in these studies may be due to a number of causes, including both patient factors and the location of the mutation, one interesting possibility that could account for these differences may be the intensity of therapy in these patient cohorts.

Patel et al recently noted that *DNMT3A* status affected the response to high-dose induction therapy in patients under age 60. ¹ In patients with wildtype *DNMT3A*, *NPM1* and *MLL*, there was no effect of dose escalation of daunorubicin from 45mg/m2 to 90mg/m2 on overall survival, whereas those with *DNMT3A* mutations did experience a survival benefit from a higher dose of daunorubicin. Since this observation may offer insight into the biologic characteristics of *DNMT3A* mutations and affect the choice of induction therapy, we further explored this relationship in a unique patient cohort. This cohort included many patients over age 60 in whom the value of high dose therapy is unclear.

Methods

Patient samples and treatment

Between January 2001 and August 2011, 172 patients with newly diagnosed AML consented to donation of their bone marrow or peripheral blood samples to the tissue bank at our institution. All patients consented to genetic analysis and clinical assessment on the basis of an institutional review board approved protocol with accompanying HIPAA authorization, and 167 underwent next-generation sequencing on the basis of available leukemia cell DNA.

Of these 167 patients, 152 underwent induction therapy and all analyses were restricted to this group (Supplemental Figure S1). The regimen selected for each patient was based on treating physician preference, but generally included 3 days of an anthracycline and 7 days of cytarabine. Patients without adequate cytoreduction at the day 14-marrow assessment were retreated at their nadir with the same drugs unless they were felt to have failed therapy. For the purposes of this study, we defined induction therapy as high-dose for those who received a cumulative dose of 270mg/m2 of daunorubicin as either a single induction of 90mg/m²/day or a double-induction with 45mg/m²/day-60mg/m²/day daunorubicin or 72mg/m2 of idarubicin, given as 12mg/m²/day on initiated on day 1 and again day 14. All other regimens were classified as standard dose therapy.

Cytogenetic analysis

All patients underwent cytogenetic analysis. Karyotype results were classified as good, intermediate, or poor risk according to the Medical Research Council criteria. ²⁰ Patients with missing cytogenetic data, including those with failed cytogenetic testing, were classified as unknown.

Next-generation sequencing

Mutational analysis was performed using a targeted next generation sequencing panel (*ASXL1, ATM, BRAF, CBL, CDKN2A, DDX3X, DNMT3A, ETV6, EZH2, FBXW7, FLT3* (ITD and TKD) *GNAS, IDH1, IDH2, JAK2, KIT, KLHL6, KRAS, MAPK1, MYD88, NOTCH1, NPM1, NRAS, PTEN, PTPN11, PHF6, RUNX1, SF3B1, TET2, TP53, WT1, XPO1, ZMYM3*). In short, DNA was quantified using a fluorescent based measurement (Qubit, Life Technologies, Ca) and 20–250 ng of DNA was used for custom target enrichment. Following library preparation with the TruSeq Amplicon assay (Ilumina, Ca) libraries were pooled and sequenced on the Miseq to an average depth of coverage greater than 1000x. This mean depth allowed for the most challenging amplicon to reach a minimum depth of coverage of 250 reads at all copy neutral loci. Data was then processed using a custom analysis pipeline composed of commercial, publically available and in house developed tools. ²¹

Statistical analysis

All hypothesis tests were 2-sided with statistical significance set as p<0.05. All analyses were performed in STATA Version 12.0 (StataCorp, College Station, TX). Baseline characteristics were compared between the mutated and wildtype *DNMT3A* status using the

chi-squared test for categorical variable and the Wilcoxon rank sum test for continuous variables.

Survival distributions for overall survival (OS) and relapse free survival (RFS) were computed using the Kaplan-Meier method and compared using the log-rank test to determine statistical differences in the distributions for the exposure groups. A Cox regression model was used to adjust for covariates including age over 60, cytogenetic risk group, sex, allogeneic transplant, and FLT3-ITD, NPM1, IDH1 and IDH2 mutations. A backwards elimination procedure was used to create the final multivariate model. Because an interaction between high-dose therapy and DNMT3A status was noted, an interaction term defined as DNMT3A mutated treated with standard-dose therapy compared to all other groups (DNMT3A wild type or DNMT3A mutated treated with high dose therapy) was retained in the multivariate model. We anticipated a sample size of 175 patients, with 22 (12.5%) in the DNMT3A-mutated/standard-dose group. A post-hoc calculation using bootstrap methods was used to estimate the power of the log-rank test used to test the hypothesis that there was a difference in survival among patients with a DNMT3A mutation who received standard dose anthracycline (n=33, 3-year survival rate=13.1%) compared those without a DNMT3A mutation and those with a DNMT3A mutation who received high dose anthracycline(n=119, 3-year survival=33.9%). The estimate of the power of the test was 73% (95% CI = 70% - 76%).

Results

Patient cohort

This patient cohort included all patients with a diagnosis of AML seen at the Hospital of the University of Pennsylvania between January 2001 and August 2011 who provided adequate tissue and gave informed consent for these studies (Supplementary Figure S1). Patient, disease, and treatment information is detailed in Table 1. Of note, the age range for this study was 19–86 years with a median age of 55, and 44% were 60 years. All cytogenetic risk groups are represented, with the intermediate risk group representing the largest fraction at 62%. High-dose induction therapy (as defined above) was given to 32% of all patients. The median follow-up time was 12.6 months.

Frequency and spectrum of DNMT3A mutations

Of the 152 patients assessed for mutation status, 52 (31.1%) harbored mutations in the *DNMT3A* gene (Supplementary Table S1). As expected, missense mutations in the R882 codon were the most common change, found in 57.7% (30/52) of those with *DNMT3A* mutations. Of those 30 patients, one also had a concurrent non-R882 mutation. An additional 21 patients had single non-R882-*DNMT3A* mutations and one additional patient had with 2 non-R882 mutations. For subsequent analyses, *DNMT3A* mutated included both missense mutations in the R882 codon as well as the non-R882 mutations.

Association of DNMT3A with patient, disease, and treatment characteristics

The association of *DNMT3A* mutations with patient, disease, and treatment characteristics is detailed in Table 1 and Supplemental Figure S1. At diagnosis, patients with *DNMT3A*

mutations were younger and less likely to be male compared to *DNMT3A* wild-type (33% vs. 50% were 60 years or older and 46% vs. 63% were male). More patients with *DNMT3A* mutations were in the intermediate cytogenetic risk group (79% vs 54%). The mean WBC count at diagnosis was also higher in those with *DNMT3A* mutations (74,700 vs 51,500).

DNMT3A mutations occurred concomitantly with *FLT3*-ITD, *NPM1*, and *IDH1* mutations more frequently than with wild-type *DNMT3A*, as seen in Table 1. When analysis was restricted to those with intermediate-risk cytogenetics, only *NPM1* remained associated with *DNMT3A* (70.9% vs 33.9%, p= <0.001). Concomitant mutations in *DNMT3A*, *NPM1* and *FLT3*-ITD occurred 18/167 patients as compared to the 6/167 expected by chance alone (p= 0.011). This triple-mutant genotype was initially noted by the Cancer Atlas Genome Study for AML and suggests biologic cooperation among these genes. ²²

Since we were interested in the interaction between the dose of anthracycline and *DNMT3A* status, we looked at the differences in induction chemotherapy dose in those with mutated or wild-type (wt) *DNMT3A*. The percentage of patients who received high dose induction therapy or double induction did not differ based on *DNMT3A* (Table 1).

Association of DNMT3A mutations with clinical outcomes

There was no difference in OS or RFS based on DNMT3A status alone, with median survival of 17.3 months and RFS of 13.8 months for DNMT3A-mutant compared to 16 and 13.1 for DNMT3A-wt (p=0.3297 and p=0.222, respectively) (Figure 1).

A mutational analysis of the ECOG 1900 trial patients demonstrated that the benefit of anthracycline-intensified induction was seen only in those with a particular mutation profile, including *DNMT3A* mutations. ¹ We found a similar pattern in our institution's cohort. Patients with mutated *DNMT3A* had an improved overall survival with high-dose therapy (p=0.017) as compared to those with DNMT3A-wt, who did not benefit from intensified therapy (Figure 2). Those with a mutated *DNMT3A* also had improved RFS with high-dose therapy, although this did not meet statistical significance (p=0.082).

Of the 152 patients who received induction therapy, 33 (21.7%) had both a *DNMT3A* mutation and received standard-dose induction. We found that patients with this profile had worse prognosis, with a median survival of 10.1 months compared to 19.8 months for all other patients (p=0.0129) (Figure 3a). Of note, there was no survival difference between the 3 patient subsets (DNMT3A-wildtype/standard dose; DNMT3A-wildtype/high dose; DNMT3A-mutant/high dose) that make up the comparator group (Figure 4) (p=0.845, 0.2637, 0.2767).

This relationship of poorer survival in the DNMT3A mutant/standard dose group persisted on multivariate analysis after adjustment for other known prognostic factors, including age >60 years, karyotype, *FLT3*-ITD, and *NPM1* mutations (HR: 1.89, p=0.006). With the exception of FLT3-ITD, all known prognostic factors were significantly associated with survival in the univariate analyses (Table 2). A similar effect was seen for RFS, with a median RFS of 10.1 months for those patients with a *DNMT3A* mutation treated with

standard dose therapy and 13.6 months for all others (p=0.020)(Figure 3b). This relationship was also significant in the multivariate analysis (Table 2).

Discussion

Mutational analysis in AML is being used to supplement traditional cytogenetic analysis in order to better understand prognosis and guide post-remission therapy. It has emerging implications for targeted therapy. The results of this study suggest it may also help to determine induction chemotherapy. Although *DNMT3A* mutations have a controversial impact on survival, this appears to be at least partially explained by an interaction with the dose of induction chemotherapy. Our study of a large single institutional cohort of AML patients confirms that patients with mutated, but not wildtype, *DNMT3A* have an inferior prognosis if treated with standard doses of anthracycline chemotherapy during induction therapy.

This finding was consistent throughout our analysis, including our multivariate analysis that adjusted for age, cytogenetics, *FLT3*-ITD and *NPM1*. As this was a retrospective study performed on patients who were not randomized to different doses of anthracyclines, it is possible that the inferior survival seen in *DNMT3A* mutant patients who received standard dose anthracycline was due to selection bias. Indeed, we were not able to collect and adjust for performance status. However, this finding was seen only in patients with *DNMT3A* mutations, the minority of the cohort, suggesting that the inferior survival of this group is due to a true biologic effect, not simply selection bias for those with a poor performance status who were unable to tolerate higher doses of anthracycline chemotherapy. Furthermore, even patients with mutated DNMT3A who achieved a complete response with standard dose anthracyclines had an inferior RFS, demonstrating that the improved survival seen in the patients who received high dose therapy was not the results of failure to provide a second induction to unfit patients with residual disease.

The biology driving this relationship is not certain. *DNMT3A* mutations have been linked to changes in methylation patterns in affected genomes ¹⁵¹² and extensive methylation loss when occurring with *FLT3*-ITD and *NPM1* mutations together. ²² It is possible that the pattern of changes in methylation through *DNMT3A* mutations could affect response to anthracyclines. Alternatively, *DNMT3A* mRNA and protein have been shown to be upregulated in response to increasing doses of doxorubicin in human colorectal cell lines, and silencing of *DNMT3A* increased the percentage of senescent cells in response to treatment with doxorubicin. ²³ *DNMT3A* mutations, particularly the single amino acid mutation, R882, has been shown to result in decreased function of the methyltransferase enzyme in *in vitro* studies. ¹⁵ It is plausible that the decreased function of *DNMT3A* allows for a better response to high dose anthracycline chemotherapy.

We defined high dose anthracycline for this study as either a cumulative dose of 270 mg/m^2 of daunorubicin (single induction of 90mg/m^2 /day or double induction with $45-60 \text{mg/m}^2$) or 72mg/m^2 of idarubicin (double induction of 12mg/m^2 of idarubicin). 49 patients received this high dose therapy; this included 24 patients who received "double-induction" with 2 rounds of standard induction. This was generally performed at the treating

physician's discretion in response to an inadequately ablated day 14 bone marrow biopsy. As such, we considered this a single high-dose regimen. We feel this is a reasonable and physiologic approach, and previous studies of anthracycline induction have been performed in a "response-adapted" method using double-induction as needed, then using the total dose of anthracycine received to guide subsequent trials. ²⁴²⁵²⁶

Our findings of inferior survival in patients with DNMT3A mutations who receive standard doses of anthracyclines support those of Patel et al in the ECOG 1900 cohort.¹ With 2 studies now revealing this interaction, it seems reasonable to use the findings to guide therapy. Anthracycline escalation led to an improved survival without an increase in toxicity in patients under age 60 in ECOG 1900.²⁷ Thus, we would not recommend a return to standard dose induction for those without DNMT3A mutations without further studies. However, in patients over age 60, the role of dose escalation is uncertain. A cooperative group study published by Lowenberg et al compared 45mg/m² to 90mg/m² of daunorubicin in patients over age 60 with newly diagnosed AML and found that while there was an improved CR rate in those who received high dose anthracycline, there was no difference in overall survival or in toxicity profile. ²⁸ Similarly, the Acute Leukemia French Association (ALFA)-9801 study found no difference in CR, OS, or EFS for dose-escalated therapy compared to standard dose idarubicin in patients age 50-70 with AML.²⁹ As such, high dose therapy has not been routinely adopted for this age group, although the lack of excess toxicity in these trials suggests that anthracycline induction may not need to be doseattenuated either. 30

Patients with *DNMT3A* mutations who are 60 years old may be a select group for whom higher dose anthracycline is reasonable. However, *DNMT3A* mutational analysis is often not feasible prior to the initiation of induction therapy. For example, current processing time for this test at our center is 7–10 days. Our cohort of patients receiving induction therapy ranged from age 19 to 79 including 59 patients age 60. Fourteen patients 60 years old received high dose therapy; 10 of whom received it as a double-induction. Therefore, one strategy in older patients may be to give standard dose induction with 45mg/m², and, if subsequently found to have a *DNMT3A* mutation, they could receive a second dose of daunorubicin at 45mg/m² on day 14 regardless of bone marrow biopsy results at that time to ensure that they receive full high dose anthracycline induction.

Mutational analysis of leukemic cells in patients with newly diagnosed AML is becoming more feasible and the number of clinical applications is growing. These results suggest that *DNMT3A* mutations alter the response to anthracycline chemotherapy, and define a group for whom high-dose therapy is particularly useful. Furthermore, it suggests that chemotherapy dose should be considered in the algorithm when incorporating comprehensive gene mutation signatures into risk-adapted post-remission therapy plans. Future studies are necessary to determine the biology that guides this relationship to allow further personalization of treatment plans in this AML subtype. Significantly larger patient cohorts are necessary to define the behavior of rare subtypes—such as the triple *DNMT3A*, *FLT3-ITD*, and *NPM1* mutant—and their response to chemotherapy.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

- Patel JP, Gonen M, Figueroa ME, Fernandez H, Sun Z, Racevskis J, et al. Prognostic relevance of integrated genetic profiling in acute myeloid leukemia. The New England journal of medicine. 2012; 366:1079–89. [PubMed: 22417203]
- Byrd JC, Mrozek K, Dodge RK, Carroll AJ, Edwards CG, Arthur DC, et al. Pretreatment cytogenetic abnormalities are predictive of induction success, cumulative incidence of relapse, and overall survival in adult patients with de novo acute myeloid leukemia: results from Cancer and Leukemia Group B (CALGB 8461). Blood. 2002; 100:4325–36. [PubMed: 12393746]
- Slovak ML, Kopecky KJ, Cassileth PA, Harrington DH, Theil KS, Mohamed A, et al. Karyotypic analysis predicts outcome of preremission and postremission therapy in adult acute myeloid leukemia: a Southwest Oncology Group/Eastern Cooperative Oncology Group Study. Blood. 2000; 96:4075–83. [PubMed: 11110676]
- Schlenk RF, Dohner K, Krauter J, Frohling S, Corbacioglu A, Bullinger L, et al. Mutations and treatment outcome in cytogenetically normal acute myeloid leukemia. The New England journal of medicine. 2008; 358:1909–18. [PubMed: 18450602]
- Tahiliani M, Koh KP, Shen Y, Pastor WA, Bandukwala H, Brudno Y, et al. Conversion of 5methylcytosine to 5-hydroxymethylcytosine in mammalian DNA by MLL partner TET1. Science. 2009; 324:930–5. [PubMed: 19372391]
- Delhommeau F, Dupont S, Della Valle V, James C, Trannoy S, Masse A, et al. Mutation in TET2 in myeloid cancers. The New England journal of medicine. 2009; 360:2289–301. [PubMed: 19474426]
- Mardis ER, Ding L, Dooling DJ, Larson DE, McLellan MD, Chen K, et al. Recurring mutations found by sequencing an acute myeloid leukemia genome. The New England journal of medicine. 2009; 361:1058–66. [PubMed: 19657110]
- Marcucci G, Haferlach T, Dohner H. Molecular genetics of adult acute myeloid leukemia: prognostic and therapeutic implications. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2011; 29:475–86. [PubMed: 21220609]
- Ward PS, Patel J, Wise DR, Abdel-Wahab O, Bennett BD, Coller HA, et al. The common feature of leukemia-associated IDH1 and IDH2 mutations is a neomorphic enzyme activity converting alphaketoglutarate to 2-hydroxyglutarate. Cancer cell. 2010; 17:225–34. [PubMed: 20171147]
- Metzeler KH, Becker H, Maharry K, Radmacher MD, Kohlschmidt J, Mrozek K, et al. ASXL1 mutations identify a high-risk subgroup of older patients with primary cytogenetically normal AML within the ELN Favorable genetic category. Blood. 2011; 118:6920–9. [PubMed: 22031865]
- 11. Shih AH, Abdel-Wahab O, Patel JP, Levine RL. The role of mutations in epigenetic regulators in myeloid malignancies. Nature reviews Cancer. 2012; 12:599–612.
- Ley TJ, Ding L, Walter MJ, McLellan MD, Lamprecht T, Larson DE, et al. DNMT3A mutations in acute myeloid leukemia. The New England journal of medicine. 2010; 363:2424–33. [PubMed: 21067377]
- Gaidzik VI, Schlenk RF, Paschka P, Stolzle A, Spath D, Kuendgen A, et al. Clinical impact of DNMT3A mutations in younger adult patients with acute myeloid leukemia: results of the AML Study Group (AMLSG). Blood. 2013; 121:4769–77. [PubMed: 23632886]
- Thol F, Damm F, Ludeking A, Winschel C, Wagner K, Morgan M, et al. Incidence and prognostic influence of DNMT3A mutations in acute myeloid leukemia. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2011; 29:2889–96. [PubMed: 21670448]

- Yan XJ, Xu J, Gu ZH, Pan CM, Lu G, Shen Y, et al. Exome sequencing identifies somatic mutations of DNA methyltransferase gene DNMT3A in acute monocytic leukemia. Nature genetics. 2011; 43:309–15. [PubMed: 21399634]
- Fried I, Bodner C, Pichler MM, Lind K, Beham-Schmid C, Quehenberger F, et al. Frequency, onset and clinical impact of somatic DNMT3A mutations in therapy-related and secondary acute myeloid leukemia. Haematologica. 2012; 97:246–50. [PubMed: 21993668]
- Renneville A, Boissel N, Nibourel O, Berthon C, Helevaut N, Gardin C, et al. Prognostic significance of DNA methyltransferase 3A mutations in cytogenetically normal acute myeloid leukemia: a study by the Acute Leukemia French Association. Leukemia. 2012; 26:1247–54. [PubMed: 22289988]
- Roller A, Grossmann V, Bacher U, Poetzinger F, Weissmann S, Nadarajah N, et al. Landmark analysis of DNMT3A mutations in hematological malignancies. Leukemia. 2013; 27:1573–8. [PubMed: 23519389]
- Markova J, Michkova P, Burckova K, Brezinova J, Michalova K, Dohnalova A, et al. Prognostic impact of DNMT3A mutations in patients with intermediate cytogenetic risk profile acute myeloid leukemia. European journal of haematology. 2012; 88:128–35. [PubMed: 21967546]
- 20. Grimwade D, Hills RK, Moorman AV, Walker H, Chatters S, Goldstone AH, et al. Refinement of cytogenetic classification in acute myeloid leukemia: determination of prognostic significance of rare recurring chromosomal abnormalities among 5876 younger adult patients treated in the United Kingdom Medical Research Council trials. Blood. 2010; 116:354–65. [PubMed: 20385793]
- 21. Daber RD, Sukhadia S, Morrissette JJD. Understanding the limitations of NGS informatics, an approach to clinical pipeline validation using artificial data sets. Cancer genetics.
- 22. Cancer Genome Atlas Research N. Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia. The New England journal of medicine. 2013; 368:2059–74. [PubMed: 23634996]
- 23. Zhang Y, Gao Y, Zhang G, Huang S, Dong Z, Kong C, et al. DNMT3a plays a role in switches between doxorubicin-induced senescence and apoptosis of colorectal cancer cells. International journal of cancer Journal international du cancer. 2011; 128:551–61. [PubMed: 20473858]
- 24. Ohno R, Kobayashi T, Tanimoto M, Hiraoka A, Imai K, Asou N, et al. Randomized study of individualized induction therapy with or without vincristine, and of maintenance-intensification therapy between 4 or 12 courses in adult acute myeloid leukemia. AML-87 Study of the Japan Adult Leukemia Study Group. Cancer. 1993; 71:3888–95. [PubMed: 8508355]
- 25. Kobayashi T, Miyawaki S, Tanimoto M, Kuriyama K, Murakami H, Yoshida M, et al. Randomized trials between behenoyl cytarabine and cytarabine in combination induction and consolidation therapy, and with or without ubenimex after maintenance/intensification therapy in adult acute myeloid leukemia. The Japan Leukemia Study Group. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 1996; 14:204–13. [PubMed: 8558199]
- 26. Miyawaki S, Tanimoto M, Kobayashi T, Minami S, Tamura J, Omoto E, et al. No beneficial effect from addition of etoposide to daunorubicin, cytarabine, and 6-mercaptopurine in individualized induction therapy of adult acute myeloid leukemia: the JALSG-AML92 study. Japan Adult Leukemia Study Group. International journal of hematology. 1999; 70:97–104. [PubMed: 10497848]
- Fernandez HF, Sun Z, Yao X, Litzow MR, Luger SM, Paietta EM, et al. Anthracycline dose intensification in acute myeloid leukemia. The New England journal of medicine. 2009; 361:1249–59. [PubMed: 19776406]
- Lowenberg B, Ossenkoppele GJ, van Putten W, Schouten HC, Graux C, Ferrant A, et al. Highdose daunorubicin in older patients with acute myeloid leukemia. The New England journal of medicine. 2009; 361:1235–48. [PubMed: 19776405]
- Pautas C, Merabet F, Thomas X, Raffoux E, Gardin C, Corm S, et al. Randomized study of intensified anthracycline doses for induction and recombinant interleukin-2 for maintenance in patients with acute myeloid leukemia age 50 to 70 years: results of the ALFA-9801 study. Journal of clinical oncology : official journal of the American Society of Clinical Oncology. 2010; 28:808–14. [PubMed: 20048183]

30. Luger SM. Treating the elderly patient with acute myelogenous leukemia. Hematology/the Education Program of the American Society of Hematology American Society of Hematology Education Program. 2010; 2010:62–9. [PubMed: 21239772]

Statement of Translational Relevance

The emergence of comprehensive mutational testing in AML has led to significant excitement in the leukemia community but also some concern for how to use the wide array of new genetic tests. One gene of interest is the *DNMT3A* gene. Previous work by Patel and colleagues¹ suggested that patients <60 years old with AML that express mutant *DNMT3A* require higher doses of anthracycline in their induction regimen in order to obtain equivalent results as *DNMT3A* wild type patients. However, this observation has not previously been reproduced. Here we describe our retrospective cohort study which confirms that mutated *DNMT3A* can predict for both overall and relapse-free survival with standard doses of anthracycline induction therapy, including patients 60 years. This decreased prognosis can be overcome by treating patients with higher doses of anthracycline. This confirms that patients with *DNMT3A* mutated AML should be treated with higher doses of anthracycline.



Figure 1. Survival by DNMT3A status

Figure 1A: Overall survival stratified by DNMT3A status, 1B: Relapse free survival stratified by DNMT3A status.



Figure 2. Effect of dose escalation depends on DNMT3A status

Figure 2 A: OS in DNMT3A-mutant, stratified by anthracycline dose B: OS in DNMT3Awildtype, stratified anthracycline dose C: RFS in DNMT3A-mutant, stratified by anthracycline dose D: RFS in DNMT3A-wildtype, stratified by anthracycline dose



Figure 3. Survival stratified by anthracycline dose and DNMT3A status

Figure 3 A: OS, comparing the DNMT3A mutant patients who received standard dose therapy to all other patients (DNMT3A-wildtype and DNMT3A mutant who received high dose therapy) B: RFS, comparing the DNMT3A mutant patients who received standard dose therapy to all other patients (DNMT3A-wildtype and DNMT3A-mutant who received high dose therapy)



Figure 4. Overall survival by DNMT3A status and anthracycline dose

Figure 4: Overall survival, as stratified by the presence or absence of a DNMT3A mutation and the anthracycline dose received.

Table 1

Patient, Disease, and Treatment Characteristics

	Full cohort (n=152)	DNMT3A mutant (n=49, 32%)	DNMT3A wild type (n=103, 68%)	Significance
Age at diagnosis, median, yrs (range)	54 (19–79)	54.4 (26–78)	54.1 (19–79)	0.6759
Age 60	39%	29%	44%	0.074
Male	57%	45%	63%	0.034
WBC at diagnosis (mean)	58,232	76,569	49,509	0.021
WBC 100,000	21%	31%	17%	0.057
Cytogenetics risk groups				0.003
Favorable	13%	0%	19%	
Intermediate	64%	82%	56%	
Poor	15%	10%	17%	
Unknown	7%	8%	7%	
FLT3-ITD mutant	32%	43%	26%	0.039
NPM1 mutant	33%	65%	25%	<0.001
IDH1 mutant	8%	16%	4%	0.008
IDH2 mutant	14%	14%	14%	0.908
High-dose therapy	32%	33%	32%	0.844
Double-induction	15%	12%	17%	0.493

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Abbreviations: n, number of patients; yrs, years

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Analysis	
Aultivariate	

Table 2

Covariate		Overall	Survival			Relapse F	ree Survival	
	Univariate HR	p-value	Multivariate HR	p-value	Univariate HR	p-value	Multivariate HR	p-value
DNMT3A mut and stud dose (vs. DNMT3A wt or DNMT3A mut and high dose)	1.71	0.014	1.90	0.006	1.47	0.070	1.69	0.017
DNMT3A mut (vs. DNMT3A wt)	1.21	0.331			1.02	0.911		
High dose (vs. stnd dose)	0.83	0.355			0.83	0.339		
Age 60 (vs. Age <60)	2.16	<0.001	2.62	< 0.001	2.25	<0.001	2.10	< 0.001
Intermediate Cytogenetics (vs. Favorable)	2.59	0.016	2.71	0.016	1.27	0.388	1.37	0.317
Poor Cytogenetics (vs. Favorable)	3.29	0.006	2.66	1.10	1.68	0.096	1.19	0.616
Unknown Cytogenetics (vs. Favorable)	2.55	0.0800	3.62	0.022	1.42	0.391	1.68	0.268
Female sex (vs. Male sex)	0.92	0.645			0.93	0.656		
FLT3-ITD (vs. no FLT3-ITD)	1.45	0.068	1.87	0.006	1.15	0.455	1.56	0.037
NPM1 mut (vs. NPM1 wt)	0.84	0.390	0.50	0.002	0.79	0.174	0.55	0.005
IDH1 mutated (vs. IDH1 wt)	0.91	0.792			0.75	0.375		
IDH2 mutated (vs. IDH2 wt)	0.93	0.782			0.87	0.585		
Allo tx (vs. no allo-fx)	0.46	<0.001			0.73	0.072		
Abbreviations: mut. mutant; vs. versus; wt. wildtype; stud. standard; Allo tx.	Allogeneic transpla	nt: HR. haza	rd ratio					

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