

Field cancerisation in colorectal cancer: A new frontier or pastures past?

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of cancer biology, early diagnosis of colorectal cancer remains elusive. Based on the adenoma-carcinoma sequence, cancer develops through the progressive accumulation of mutations in key genes that regulate cell growth. However, recent mathematical modelling suggests that some of these genetic events occur prior to the development of any discernible histological abnormality. Cells acquire pro-tumourigenic mutations that are not able to produce morphological change but predispose to cancer formation. These cells can grow to form large patches of mucosa from which a cancer arises. This process has been termed "field cancerisation". It has received little attention in the scientific literature until recently. Several studies have now demonstrated cellular, genetic and epigenetic alterations in the macroscopically normal mucosa of colorectal cancer patients. In some reports, these changes were effectively utilised to identify patients with a neoplastic lesion suggesting potential application in the clinical setting. In this article, we present the scientific evidence to support field cancerisation in colorectal cancer and discuss important limitations that require further investigation. Characterisation of the field defect is necessary to enable early diagnosis of colorectal cancer and identify molecular targets for chemoprevention. Field cancerisation offers a promising prospect for experimental cancer research and has potential to improve patient outcomes in the clinical setting.

Key words: Colorectal cancer; Carcinogenesis; Biomarkers; Epigenetics; Synchronous

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Core tip: There is a great deal of interest in developing non-invasive tests that are able to detect colorectal cancer in the asymptomatic population. Most current research activity is focussed on investigating the biological changes found in tumour tissue itself. This

Abstract

Despite considerable advances in our understanding

review evaluates the biological alterations found in the normal mucosa around a neoplastic lesion and critically analyses the concept of field cancerisation. It highlights recent advances and identifies important molecular targets that could play a role in early colorectal carcinogenesis. In particular, the available evidence for field cancerisation is scrutinised and future avenues for further scientific enquiry are outlined.

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INTRODUCTION

Colorectal cancer is the third most common cancer worldwide, affecting 1.36 million people^[1] and is the largest killer amongst non-smokers in the United Kingdom^[2]. The greatest chance of cure is with disease confined to the bowel wall, hence, early diagnosis and prompt treatment are important^[3]. It is generally accepted that cancers develop through accumulation of mutations in key genes^[4,5]. Traditionally, a three step process comprising initiation, promotion and progression was proposed^[6,7]. Later, it became apparent that the colonic epithelium undergoes an ordered sequence of genetic events with corresponding histological abnormalities on its journey to cancer formation^[8]. However, several recent analyses have revealed that the mutations found in colorectal cancer occur long before the onset of a clinically visible lesion^[9,10]. In many cancers, cells have been shown to acquire pro tumorigenic mutations that are not able to produce morphological change but predispose to subsequent malignant transformation^[11-14]. These cells can expand creating patches of mucosa which have an increased risk of developing into cancer. This process has been described as "field cancerisation"^[15,16]. It is a concept that has previously received little attention in the scientific literature. Most studies investigating colorectal carcinogenesis have focused solely on the cancer tissue or assumed that the mucosa adjacent to the neoplastic lesion is normal^[17,18]. However, based upon the field cancerisation theory, characterization of the biological events that occur in the "mucosa at risk" could enable identification of the earliest steps in colorectal cancer formation. This could aid the scientist in discovery of how neoplasia develops and enable the clinician to develop more reliable tests to risk stratify patients. This article discusses the evidence for field cancerisation, its limitations and its potential clinical application to improve patient outcome.

FIELD CANCERISATION THEORY - DEFINITION AND MECHANISM

Field cancerisation was first described by Slaughter in 1953 for head and neck squamous cell carcinoma^[15]. It was based upon the observation that a statistically significant proportion of oral cancers developed in multifocal areas and often had histologically abnormal cells surrounding the cancer. Since its inception, field cancerisation has been applied to several other cancers including cancer of the oesophagus^[19], stomach^[20], lung^[21], bladder^[22], pancreas^[23] and skin^[24]. In the colon, it has been described as "the process whereby colonic epithelial cells acquire pro-tumorigenic mutations that are insufficient to cause morphological change but which predispose to tumour"^[25]. Multiple mechanisms have been proposed to explain how a patch of altered mucosa forms around a cancer (Figure 1). Genetic analysis has revealed that the field defect consists of a clonal proliferation of a mutated cell. Based on this observation, it was proposed that a mutation or epigenetic alteration in stem cells gives it reproductive advantage so that it generates clonal descendents that outcompete neighbouring stem cells^[26]. These stem cells replace other stem cells through the process of niche succession^[27] and eventually, the entire crypt is occupied by the mutant or epigenetically altered cells^[28,29]. Crypt fission of this mutated or epigenetically altered crypt results in a patch defect (Figure 1A). Other mechanisms include alteration in the adjacent mucosa by the presence of the tumour itself or as a result of chemicals released by the tumour^[30,31] (Figure 1B). Whilst others have proposed that, like oral cancer^[32,33] and bladder cancer, colonic epithelial cells shed in the lumen at one place could migrate to another site, seed and give rise to synchronous cancer^[16] (Figure 1C). Field changes could be more widespread which has led some authors to suggest that dietary exposure, for example, vitamin B and folate, could alter the methylation state of the entire colonic mucosa predisposing it to cancer^[25]. Further investigation is required to elucidate the precise biological mechanism and causal events that underly field cancerisation. Despite this, however, a large body of evidence exists that supports the presence of a field defect in colorectal cancer.

EARLY EVIDENCE BASED ON THE TRANSITIONAL MUCOSA

The term "transitional mucosa" was used to describe the patch of mucosa around a cancer that was abnormal compared to the rest of the mucosa. Although field cancerisation had not been formally proposed at the time, these were some of the early studies supporting the concept.

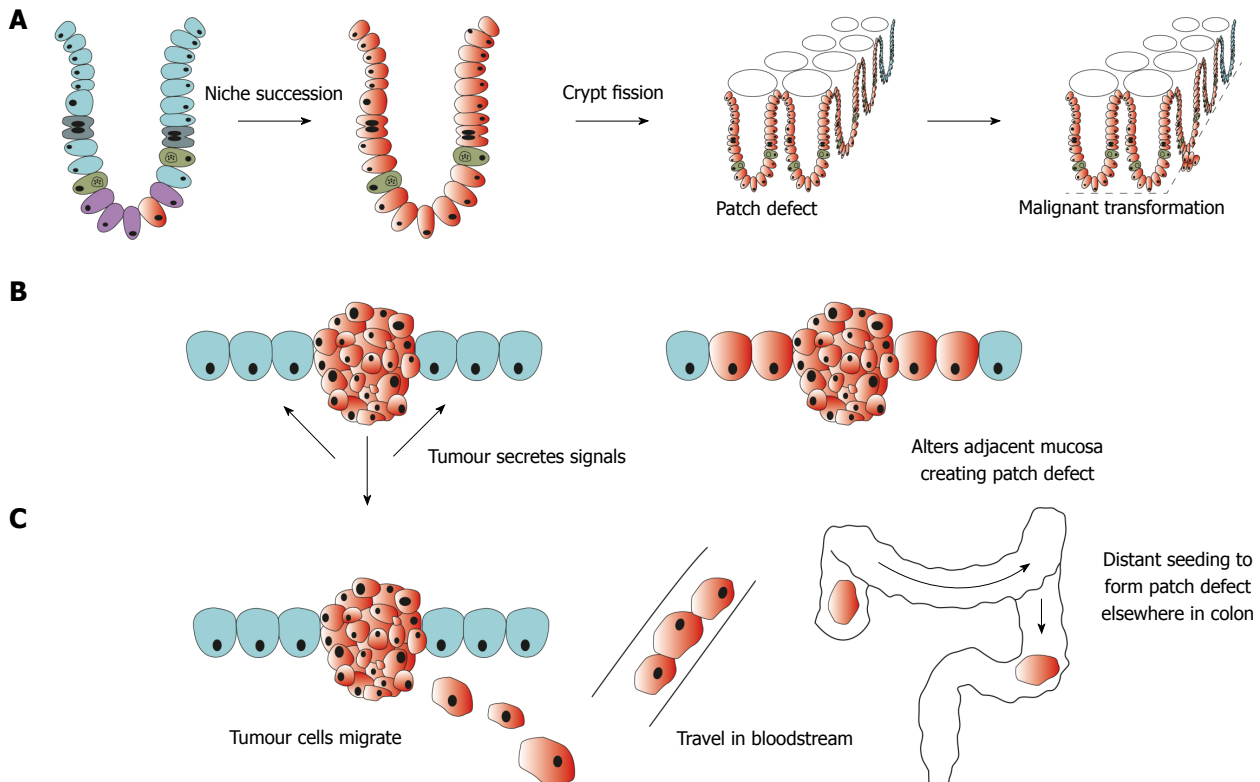


Figure 1 Schematic representation of proposed mechanisms for formation of field defect. A: A mutation or epigenetic alteration in a stem cell (depicted in red) is inherited by all cells within the crypt through niche succession. Crypt fission results in several crypts becoming biologically altered creating a patch defect. Further mutation within this field of altered mucosa leads to malignant transformation; B: Tumour secretes chemical signals that alter the adjacent mucosa resulting in a field defect; C: Malignant cells shed from a tumour travel in the bloodstream and seed in a distant site rendering the mucosa susceptible to malignant transformation.

In 1969, Filipe described abnormal histochemical properties in the transitional mucosa^[34] with a decrease in sulfomucins, usually found in normal colorectal tissue and concurrent increase in sialomucin content. Sulfomucins protect against luminal insults by increasing mucus viscosity which increases the resistance against bacterial degradation and microbe adhesion^[35]. Replacement by sialomucin has been previously observed in colorectal cancer tissue. Based on the finding of increased sialomucin content in the transitional mucosa, Filipe proposed that these changes could represent an early stage of carcinogenesis^[36]. Further exploration using light microscopy revealed that there were alterations in crypt morphology and cell type within this mucosa. Saffos and Rhatigan^[37] found an increase in the length of the crypts, increased distension and branching of the crypts and an increase in the number of goblet cells in the crypt. They were unable to demonstrate similar changes in tissue samples taken along the rest of the colon in these patients. They concluded that these changes were confined to the rim around the tumour.

In the late 1970's, several investigators characterised the ultrastructural properties of the transitional mucosa^[38,39]. They found that the crypts were larger in diameter and composed of larger cells with larger nuclei compared to those found in the normal colon. There was also a change in cell distribution with an increase

in mature goblet cells in the lower half of the crypt and an increase in immature goblet cells and "intermediate" cells in the upper half of the crypt.

Subsequent studies have highlighted alterations in the nuclear morphology of cells in the transitional mucosa, often as far as 50 mm from the tumour^[40,41]. Many nuclear features were found to differ in the transitional mucosa compared to that of healthy controls including total optical density, nuclear area, chromatin texture, chromatin coarseness, average optical density and increased tendency of peripherally placed chromatin^[42]. However, because of the considerable inter-patient and inter gland variation in these parameters, the authors cautioned against the use of any single feature to identify those at risk. Instead, it was suggested that a combination of parameters be used to develop a tool for risk stratifying patients. More recent studies, using computer based karyometric analysis^[43] or electron microscopy^[44] have confirmed these earlier findings. Although there are differences in nuclear appearance of cells in the transitional mucosa, the variability seen between patients and between samples taken from the same patient preclude the use of nuclear analysis as a discriminant factor to risk stratify patients. Investigators have therefore sought to identify other biological changes that could be indicative of a field defect.

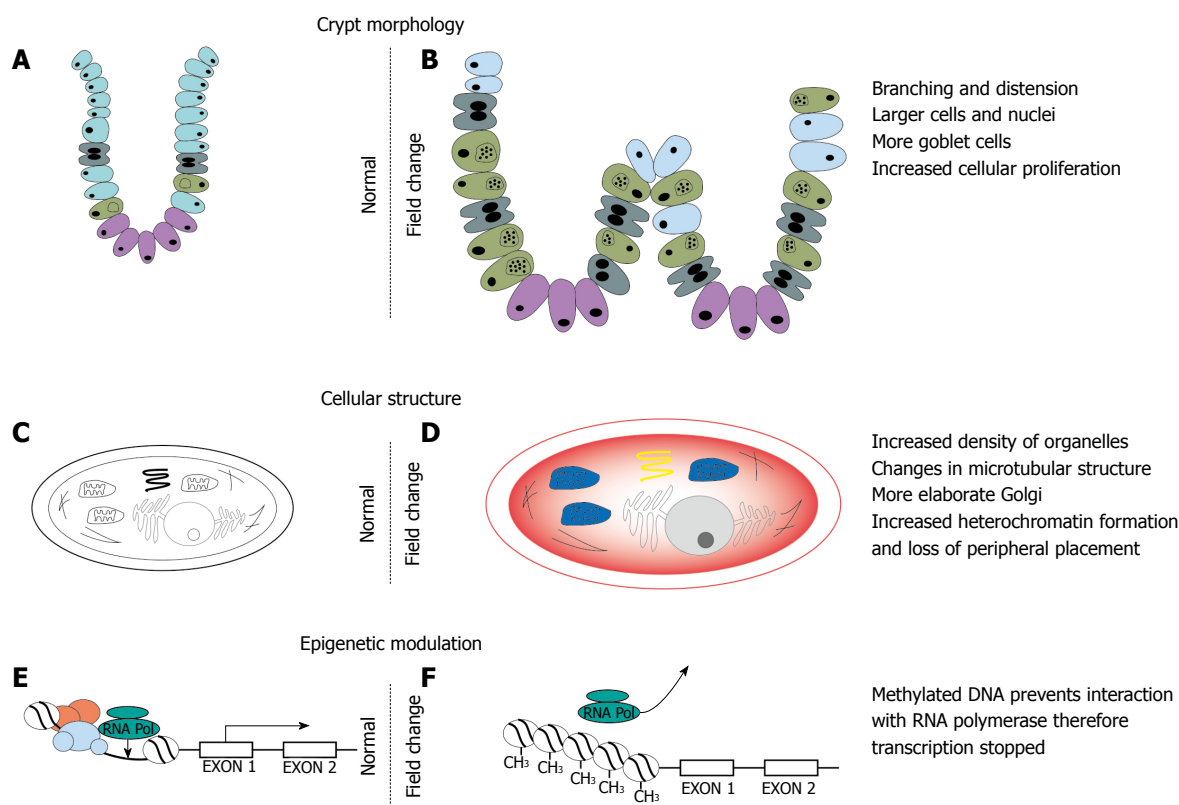


Figure 2 Changes in crypt morphology, cellular ultrastructure and epigenetic modulation in the field defect. A, B: Changes in crypt morphology characterised by increased branching and distension of crypts, increased cell division and a change in proportion of cells with increase in goblet cells; C, D: Changes in the cell cytoskeleton, organelles and nuclear composition; E, F: Epigenetic modulation of DNA leading to transcriptional silencing of certain genes involved in regulation of cell division, apoptosis and DNA repair.

SUPPORTING EVIDENCE BASED ON CANCER BIOLOGY

Colorectal cancer usually arises from multiple dysfunctional cellular processes which enable the cell to evade homeostatic signals and grow in an autonomous manner. Similarly, alterations at the genetic, epigenetic and protein level in a number of cellular processes and function have been described in the colonic field (Figure 2).

Cellular proliferation and apoptosis

Many studies have demonstrated that there is an increase in cellular proliferation and concurrent reduction in apoptosis in the macroscopically normal appearing mucosa around a malignant lesion^[45-48]. Using 3H thymidine autoradiography, the rate of proliferation in normal mucosa was found to be significantly higher in patients harbouring a colorectal cancer or large adenoma compared to those with small adenomas or healthy individuals^[45]. These changes were most prominent in the upper third of the mucosal crypt^[47,48] and could potentially be utilised as a predictive marker to identify patients with a neoplastic lesion^[49,50]. Not only have these changes in proliferation been linked to the presence of neoplastic lesions but they have also been shown to be predictive for the risk of polyp recurrence^[49]. Epithelial cell proliferation in

the macroscopically normal rectal mucosa of patients with and without polyps was assessed. The crypt was divided into five compartments along the longitudinal axis and the labelling index was calculated for the entire crypt and each of the five compartments. In the 22 patients whose polyps recurred, the upper compartments 3, 4 and 5 demonstrated a significantly higher labelling index compared to the 33 patients without recurrence. There was an upward shift in the proliferative zone of the crypt that was associated with polyp recurrence. Interestingly, there was no difference in the labelling index between the first and second biopsy suggesting that an underlying genetic defect or persisting environmental insult may have been responsible for the field defect detected in this study.

Genetic and epigenetic modulation

The genetic and epigenetic abnormalities found in colorectal cancer have also been shown to extend into the macroscopically normal mucosa supporting the field cancerisation theory. Early studies using flow cytometry confirmed that mucosa adjacent to diploid cancers is diploid in nature and in patients with aneuploid tumours, is often aneuploid^[51,52]. Similarly, epigenetic modulation of genes has been found to differ in patients with classical adenomas compared to

those with serrated polyps^[53]. These lesions represent the two different pathways to colorectal cancer. Classical adenomas are linked to carcinogenesis that occurs along the CIN (chromosomal instability) pathway compared to serrated polyps which are the precursor for the CpG island methylator phenotype (CIMP) pathway. The authors demonstrated that age-related methylation was inversely associated with the presence of classical adenomas compared to methylation of cancer specific genes that was more likely in patients with serrated polyps. This suggests that the background mucosa of these patients had detectable epigenetic differences that conferred a predisposition to a specific pathway of cancer prior to the development of any discernible histological abnormality.

Differences in expression of genes across a wide number of cellular processes have been implicated in the field defect and may potentially play a role in early tumourigenesis. Based on a 15-gene signature encompassing genes that play a role in the APC/Beta catenin, NFκB, cell cycle and inflammation pathways, significant alterations in gene expression were found in the normal mucosa of human cancer resection specimens, often extending into the margins^[54]. In a further study, based on analysis of a macroscopically normal appearing rectosigmoid biopsy, this 15-gene signature could discriminate between individuals with and without polyps^[55].

Epigenetic silencing of genes has been implicated in both mismatch repair deficient and CIMP cancer. Several investigators have reported epigenetic changes in the normal colonic mucosa in patients with cancer. Reduced protein expression of the DNA repair proteins, mismatch repair endonuclease (Pms2), DNA excision repair protein (ERCC1) and DNA excision repair protein XPF (ERCC4) was found up to 10 cm longitudinally from the tumour edge supporting the field theory^[56]. In a study by Shen *et al.*^[57], O-6-methylguanine-DNA methyltransferase (MGMT) methylation was found in the normal adjacent mucosa of 50% patients whose tumours also had methylated MGMT compared to only 6% when MGMT was not methylated in the tumour. In 10 out of 13 patients, methylation changes were seen as far as 10 cm away from the tumour and hypermethylation was more pronounced at 1 cm compared to 10 cm. These findings raise the possibility that MGMT methylation may play a role in the field defect representing an early step in the carcinogenesis pathway of tumours with hypermethylated MGMT. Similarly, others have also reported that MGMT hypermethylation is more likely to be found in the surrounding mucosa of microsatellite unstable tumours compared to microsatellite stable cancers^[58,59]. Other studies have investigated the methylation profile of combinations of multiple genes confirming that the apparently normal mucosal field has undergone significant epigenetic change that could represent the earliest stages of colorectal cancer development^[60,61].

Epigenetic modulation through methylation of micro-RNAs (miR) may also contribute to a field defect. Grady *et al.*^[62], found expression of hsa-miR-342, a microRNA encoded in an intron of the gene *EVL*, is commonly suppressed in human colorectal cancer. They found methylation at the *EVL*/hsa-miR-342 locus in 56% of histologically normal mucosa from patients with colorectal cancer compared to only 12% of patients without colorectal cancer. Similarly, methylation of miR-124a and miR-34b/c in the histologically normal mucosa, was observed in 59% and 26% of patients with cancer but was not found in patients without cancer or Ulcerative Colitis^[63]. In another study, the level of methylation of miR-137 was found to be higher in the macroscopically normal mucosa in cancer patients compared to healthy controls^[64] (10.3% vs 7.7%, $P = 0.035$). These findings suggest that changes at the micro-RNA level could also play an important role in field defect around a tumour.

Although, there are considerable genetic and epigenetic alterations in the "normal" mucosa surrounding a cancer, it is not yet clear which of these changes are most important. Epigenetic changes are particularly interesting as they can be modified by changes in diet or pharmacological agents unlike the germline mutations often linked with cancer. Elucidation of the specific epigenetic marker that underlies the field defect could enable specific chemopreventative agents to be designed to target these early changes prior to the development of any precancerous lesions such as adenomas.

FIELD CANCERISATION - POTENTIAL PITFALLS

Although there is sufficient evidence to support the field cancerisation theory in colorectal cancer, a number of pertinent questions remain.

Pre malignant change or a secondary phenomenon?

Similar changes in crypt and cellular morphology to those observed in the transitional mucosa have also been described in mucosa adjacent to squamous cell carcinoma of the anus^[65,66], sarcoma of the colon^[66,67] and in non-neoplastic lesions such as endometriosis^[67]. This has led to the conclusion that these alterations do not represent premalignant change but rather, a reactive phenomenon in response to tumour or non-neoplastic injury such as that induced by inflammation or necrosis. Early studies showed that the width of the transitional mucosa was related to the size of the tumour^[36] where it became larger with increasing stage of the tumour. However, if these changes precede cancer formation, it would be expected that as a tumour grows, it replaces the transitional mucosa from which it arose resulting in a smaller area containing the initial cellular changes. Therefore, one would expect that

the area of the transitional mucosa would be inversely related to the size of the tumour and the changes seen in the transitional mucosa would be demonstrated throughout the entire colonic mucosa. Investigators were unable to provide evidence to support this hypothesis which led to the proposal that the transitional mucosa represents the neoplastic phenotype, however, is likely to be a secondary phenomenon as a result of factors released by the tumour. This may also explain how it was found adjacent to non-neoplastic lesions which would be expected to release similar growth factors, in response to the inflammatory process, to those secreted by the tumour. However, subsequent studies have successfully correlated genetic mutations found in the tumour with those demonstrated in the surrounding mucosa confirming that these tumour cells share a common clonal origin. Also, the reports of these changes persisting despite removal of the offending lesion^[49,68] suggest that this is a primary phenomenon rather than reactive change in response to the presence of a neoplastic lesion. Hence, these field changes are most likely to be pre-malignant events that represent some of the very early steps along the path to colorectal cancer.

How far along the colon does a field defect extend?

If the macroscopically normal mucosa is biologically altered in response to the tumour, it would be limited to the area immediately in the vicinity of the tumour. In an early study investigating the field defect based on histochemical analysis, transitional mucosa was found in 90/100 cases, extending as far as 17 cm from the tumour^[69]. The change in sialomucin content that was identified in the transitional mucosa was found at the resection margins and in a subset of patients, it was a direct extension of the zone of altered mucosa surrounding the tumour. Several other studies have described biological changes in mucosa as far as 10 cm from the tumour^[43,70,71] whilst others have reported that the field defect extends as far as the rectum in these patients^[49,72]. Some authors have shown that the hypermethylation changes observed in the field defect are more pronounced 1 cm away from the tumour compared to 10 cm^[57] whereas others were unable to corroborate their findings with distance from the lesion^[54]. Some investigators have proposed that the field of altered mucosa does not occur in a contiguous manner but occurs in discrete patches. Bernstein *et al.*^[46] measured the bile salt induced apoptosis rate in 68 patients (17 colorectal cancer, 37 adenoma and 14 with neoplasia). Biopsies were taken 20 cm from the anal verge, caecum and descending colon. Site to site variability, both between regions of the colon and adjacent biopsies was greater than the inter-patient variability for individuals with a history of colorectal cancer suggesting that there was "patchiness" of the susceptibility of regions of the colon to bile acid

induced apoptosis. In other words, the field defect was not continuous along the entire colon; there were areas which showed greater changes in rate of apoptosis, however, these areas did not correspond to site of previous neoplasia. If these changes occur as a consequence of the interplay between an underlying genetic predisposition and environmental insult, patchy mucosal alterations could be explained by differences in luminal factors along the colon. Hence, there would be areas that are more susceptible to carcinogens found in the lumen or areas where cells are defective at protecting against the harmful effects of carcinogens. Further study is required to characterise the nature of the field defect and examine the causative agents responsible.

Are these alterations passengers or drivers in carcinogenesis?

Colon cancers have been found to contain a median of 76 non-silent sequence mutations of which, only 15 represent driver mutations^[73]. These are mutations in key oncogenes or tumour suppressor genes that confer a selective advantage to the cell enabling it to divide uncontrollably and survive in unfavourable conditions. In comparison, passenger mutations occur during normal cell division that takes place to replenish the colonic epithelium and have no role in driving carcinogenesis. They can be over-represented in cancer tissue due to aberrant DNA repair mechanisms and defective anti-apoptotic machinery. Similarly, it is difficult to discriminate which of the molecular changes found in the field defect are integral in driving cancer formation from those that are innocent bystanders. Roy *et al.*^[74], used 4-dimensional elastic light scattering fingerprinting (4D-ELF) to probe the nanoarchitecture of colonocytes in the Azoxymethane treated rat model vs the saline treated rat. They measured 4D-ELF at different time points and correlated the changes observed with the emergence of the aberrant crypt focus. Their finding that changes in 4D-ELF were apparent 2 wk prior to development of aberrant crypt foci (ACF) and that they correlated both spatially and temporally with subsequent development of ACF suggests that these changes were integral in early colorectal cancer formation.

Mathematical modelling suggests that it is not the rate of mutations which is important but rather the selection of clones of cells with specific advantages in autonomous growth that drives malignant transformation^[75]. It has also become apparent that this selective advantage is not conferred by mutations in one or few genes but is the accumulated benefit of several genes that have low individual selective advantage^[76]. Therefore, it is crucial that mechanistic studies are conducted based upon the gene targets found in the mucosal field to discern the driver mutations from those that are innocent bystanders.

CLINICAL APPLICATION

Despite some of these shortcomings, field cancerisation in colorectal cancer is a promising prospect upon which to develop potentially diagnostic and therapeutic modalities. Elucidation of the underlying molecular mechanism could enable more accurate screening tests to be designed that are able to identify individuals with a malignant lesion. Current research is focused on developing tools that are capable of identifying patients with colorectal cancer based on analysis of a "normal" biopsy from a distant site. Using light scattering technology, three manifestations of tissue alteration in the colonic field have been shown^[77]: changes in microcirculation [early increase in blood supply (EIBS)], changes in the extracellular matrix from abnormal cross linking and alignment of collagen fibres [as assessed by low coherence backscattering (LEBS)] and differences in the internal structure of colonocytes [as assessed using partial wave spectroscopy (PWS)]. EIBS can be detected within 30 cm of a polyp using a spectroscopic probe on 222 patients undergoing colonoscopy. The magnitude of EIBS correlated with the size and proximity of the adenoma. Based on a rectal biopsy, EIBS was found to be increased in 50% patients with an adenoma. A logistic regression model using EIBS, mucosal oxyhaemoglobin and patient age gave a sensitivity of 83% and specificity of 82% with an AUC of 0.88 for the detection of advanced adenomas^[72]. A progressive change from control patients to those with advanced adenomas was demonstrated using LEBS parameters^[78]. LEBS was able to discriminate between patients with and without advanced adenomas with 100% sensitivity, 80% specificity and an AUC of 0.90. An *in vivo* study was subsequently performed where a fibre optic probe was used to measure LEBS parameters in the rectum of 574 subjects^[79] and was shown to reliably identify patients with an advanced adenoma. Similarly, PWS has been shown to correlate with risk of developing colorectal cancer^[80]. The differences in EIBS, LEBS and PWS parameters detected in these studies were not confounded by demographics, presence of non-neoplastic lesions or site of adenoma suggesting true potential for development into a screening tool.

The presence of a field defect may indicate a higher risk of metachronous neoplastic lesions and could help to identify which patients require more radical surgery. Field cancerisation could also be utilized to ascertain risk of disease progression, hence, could enable risk stratification of patients with inflammatory bowel disease or a family history of colorectal cancer. However, the most exciting use of field cancerisation theory is its potential application in chemoprevention. Individuals at risk of malignancy could be identified based on field defects in their mucosa. Pharmacological therapy could be developed, targeted at the underlying signaling pathway, to modify the field change and reduce the risk of subsequent malignant transformation.

FUTURE DIRECTIONS

There is considerable evidence in the literature to support the field cancerisation theory in colorectal cancer. However, important questions about the underlying mechanism and extent of the field defect require further investigation before it can be applied in a clinical setting.

In other conditions that results in an increased risk of colorectal cancer such as Ulcerative Colitis, mutations in KRAS, CDKN2A (p16) and TP53 have been detected in non-tumour, non-dysplastic and dysplastic epithelium. In two patients, these changes were detected 4 years before the development of tumour suggesting that they represent some of the very early genetic events that led to colorectal carcinogenesis^[81].

Furthermore, a recent study using a mouse colitis model showed persisting epigenetic alteration in the mucosa despite removal of the toxic insult that initiated it^[68]. Lessons learnt from these studies could shed light upon the interactions that take place between the environment and the mucosa in the journey along the cancer pathway.

Cancer research has traditionally focused on characterization of the genetic/epigenetic events that occur in a malignant cell to understand the processes that contribute to malignancy. This approach seems somewhat backwards, especially in a disease where early intervention is important. Future research needs to identify early events that occur along the cancer pathway. Hence, a paradigm shift in scientific enquiry is required which focusses on the temporal sequence of mutational events to elucidate early molecular targets in colorectal cancer. The field cancerisation theory offers such an approach whereby, based on the changes occurring in the surrounding mucosa, the initial events leading to colorectal carcinogenesis can be discerned.

REFERENCES

- 1 Estimated Incidence, Mortality and Prevalence Worldwide in 2012. Available from: URL: http://globocan.iarc.fr/Pages/fact_sheets_cancer.aspx
- 2 Cancer Research UK Statistics on Colorectal Cancer. Available from: URL: <http://www.cancerresearchuk.org/cancer-info/cancerstats/types/bowel/?script=true>
- 3 Colorectal Cancer Survival by Stage - NCIN Data Briefing. Available from: URL: http://www.ncin.org.uk/publications/data_briefings/colorectal_cancer_survival_by_stage
- 4 **McCombs RS**, McCombs RP. A hypothesis on the causation of cancer. *Science* 1930; **72**: 423-424 [PMID: 17814892 DOI: 10.1126/science.72.1869.423]
- 5 **Nowell PC**. The clonal evolution of tumor cell populations. *Science* 1976; **194**: 23-28 [PMID: 959840 DOI: 10.1126/science.959840]
- 6 **Nordling CO**. A new theory on cancer-inducing mechanism. *Br J Cancer* 1953; **7**: 68-72 [PMID: 13051507 DOI: 10.1038/bjc.1953.8]
- 7 **Knudson AG**. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci USA* 1971; **68**: 820-823 [PMID: 5279523 DOI: 10.1073/pnas.68.4.820]
- 8 **Fearon ER**, Vogelstein B. A genetic model for colorectal

- tumorigenesis. *Cell* 1990; **61**: 759-767 [PMID: 2188735 DOI: 10.1016/0092-8674(90)90186-1]
- 9 **Stoler DL**, Chen N, Basik M, Kahlenberg MS, Rodriguez-Bigas MA, Petrelli NJ, Anderson GR. The onset and extent of genomic instability in sporadic colorectal tumor progression. *Proc Natl Acad Sci USA* 1999; **96**: 15121-15126 [PMID: 10611348 DOI: 10.1073/pnas.96.26.15121]
 - 10 **Tsao JL**, Yatabe Y, Salovaara R, Järvinen HJ, Mecklin JP, Aaltonen LA, Tavaré S, Shibata D. Genetic reconstruction of individual colorectal tumor histories. *Proc Natl Acad Sci USA* 2000; **97**: 1236-1241 [PMID: 10655514 DOI: 10.1073/pnas.97.3.1236]
 - 11 **Franklin WA**, Gazdar AF, Haney J, Wistuba II, La Rosa FG, Kennedy T, Ritchey DM, Miller YE. Widely dispersed p53 mutation in respiratory epithelium. A novel mechanism for field carcinogenesis. *J Clin Invest* 1997; **100**: 2133-2137 [PMID: 9329980 DOI: 10.1172/JCI119748]
 - 12 **Hafner C**, Toll A, Fernández-Casado A, Earl J, Marqués M, Acquadro F, Méndez-Pertuz M, Urioste M, Malats N, Burns JE, Knowles MA, Cigudosa JC, Hartmann A, Vogt T, Landthaler M, Pujol RM, Real FX. Multiple oncogenic mutations and clonal relationship in spatially distinct benign human epidermal tumors. *Proc Natl Acad Sci USA* 2010; **107**: 20780-20785 [PMID: 21078999 DOI: 10.1073/pnas.1008365107]
 - 13 **Leedham SJ**, Graham TA, Oukrif D, McDonald SA, Rodriguez-Justo M, Harrison RF, Shepherd NA, Novelli MR, Jankowski JA, Wright NA. Clonality, founder mutations, and field cancerization in human ulcerative colitis-associated neoplasia. *Gastroenterology* 2009; **136**: 542-550.e6 [PMID: 19103203 DOI: 10.1053/j.gastro.2008.10.086]
 - 14 **Maley CC**, Galipeau PC, Li X, Sanchez CA, Paulson TG, Reid BJ. Selectively advantageous mutations and hitchhikers in neoplasms: p16 lesions are selected in Barrett's esophagus. *Cancer Res* 2004; **64**: 3414-3427 [PMID: 15150093 DOI: 10.1158/0008-5472.CAN-03-3249]
 - 15 **Slaughter DP**, Southwick HW, Smejkal W. Field cancerization in oral stratified squamous epithelium; clinical implications of multicentric origin. *Cancer* 1953; **6**: 963-968 [PMID: 13094644 DOI: 10.1002/1097-0142(195309)6:5<963>
 - 16 **Braakhuis BJ**, Tabor MP, Kummer JA, Leemans CR, Brakenhoff RH. A genetic explanation of Slaughter's concept of field cancerization: evidence and clinical implications. *Cancer Res* 2003; **63**: 1727-1730 [PMID: 12702551]
 - 17 **Dakubo GD**, Jakupciak JP, Birch-Machin MA, Parr RL. Clinical implications and utility of field cancerization. *Cancer Cell Int* 2007; **7**: 2 [PMID: 17362521 DOI: 10.1186/1475-2867-7-2]
 - 18 **Rubin H**. Fields and field cancerization: the preneoplastic origins of cancer: asymptomatic hyperplastic fields are precursors of neoplasia, and their progression to tumors can be tracked by saturation density in culture. *Bioessays* 2011; **33**: 224-231 [PMID: 21254148 DOI: 10.1002/bies.201000067]
 - 19 **Wong DJ**, Paulson TG, Prevo LJ, Galipeau PC, Longton G, Blount PL, Reid BJ. p16(INK4a) lesions are common, early abnormalities that undergo clonal expansion in Barrett's metaplastic epithelium. *Cancer Res* 2001; **61**: 8284-8289 [PMID: 11719461]
 - 20 **Kim SK**, Jang HR, Kim JH, Noh SM, Song KS, Kim MR, Kim SY, Yeom YI, Kim NS, Yoo HS, Kim YS. The epigenetic silencing of LIMS2 in gastric cancer and its inhibitory effect on cell migration. *Biochem Biophys Res Commun* 2006; **349**: 1032-1040 [PMID: 16959213 DOI: 10.1016/j.bbrc.2006.08.128]
 - 21 **Grepmeier U**, Dietmaier W, Merk J, Wild PJ, Obermann EC, Pfeifer M, Hofstaedter F, Hartmann A, Woenckhaus M. Deletions at chromosome 2q and 12p are early and frequent molecular alterations in bronchial epithelium and NSCLC of long-term smokers. *Int J Oncol* 2005; **27**: 481-488 [PMID: 16010431]
 - 22 **Kakizoe T**. Development and progression of urothelial carcinoma. *Cancer Sci* 2006; **97**: 821-828 [PMID: 16822297 DOI: 10.1111/j.1349-7006.2006.00264.x]
 - 23 **Kitago M**, Ueda M, Aiura K, Suzuki K, Hoshimoto S, Takahashi S, Mukai M, Kitajima M. Comparison of K-ras point mutation distributions in intraductal papillary-mucinous tumors and ductal adenocarcinoma of the pancreas. *Int J Cancer* 2004; **110**: 177-182 [PMID: 15069678 DOI: 10.1002/ijc.20084]
 - 24 **Jonason AS**, Kunala S, Price GJ, Restifo RJ, Spinelli HM, Persing JA, Leffell DJ, Tarone RE, Brash DE. Frequent clones of p53-mutated keratinocytes in normal human skin. *Proc Natl Acad Sci USA* 1996; **93**: 14025-14029 [PMID: 8943054 DOI: 10.1073/pnas.93.24.14025]
 - 25 **Luo Y**, Yu M, Grady WM. Field cancerization in the colon: a role for aberrant DNA methylation? *Gastroenterol Rep (Oxf)* 2014; **2**: 16-20 [PMID: 24760232 DOI: 10.1093/gastro/got039]
 - 26 **Bernstein C**, Nfonsam V, Prasad AR, Bernstein H. Epigenetic field defects in progression to cancer. *World J Gastrointest Oncol* 2013; **5**: 43-49 [PMID: 23671730 DOI: 10.4253/wjgo.v5.i3.43]
 - 27 **Calabrese P**, Tavaré S, Shibata D. Pretumor progression: clonal evolution of human stem cell populations. *Am J Pathol* 2004; **164**: 1337-1346 [PMID: 15039221 DOI: 10.1016/S0002-9440(10)63220-8]
 - 28 **Kim KM**, Shibata D. Methylation reveals a niche: stem cell succession in human colon crypts. *Oncogene* 2002; **21**: 5441-5449 [PMID: 12154406 DOI: 10.1038/sj.onc.1205604]
 - 29 **Greaves LC**, Preston SL, Tadrous PJ, Taylor RW, Barron MJ, Oukrif D, Leedham SJ, Deheragoda M, Sasieni P, Novelli MR, Jankowski JA, Turnbull DM, Wright NA, McDonald SA. Mitochondrial DNA mutations are established in human colonic stem cells, and mutated clones expand by crypt fission. *Proc Natl Acad Sci USA* 2006; **103**: 714-719 [PMID: 16407113 DOI: 10.1073/pnas.0505903103]
 - 30 **Boland CR**, Kim YS. Transitional mucosa of the colon and tumor growth factors. *Med Hypotheses* 1987; **22**: 237-243 [PMID: 3473277 DOI: 10.1016/0306-9877(87)90189-7]
 - 31 **Kuniyasu H**, Yasui W, Shinohara H, Yano S, Ellis LM, Wilson MR, Bucana CD, Rikita T, Tahara E, Fidler IJ. Induction of angiogenesis by hyperplastic colonic mucosa adjacent to colon cancer. *Am J Pathol* 2000; **157**: 1523-1535 [PMID: 11073812 DOI: 10.1016/S0002-9440(10)64790-6]
 - 32 **Califano J**, Leong PL, Koch WM, Eisenberger CF, Sidransky D, Westra WH. Second esophageal tumors in patients with head and neck squamous cell carcinoma: an assessment of clonal relationships. *Clin Cancer Res* 1999; **5**: 1862-1867 [PMID: 10430093]
 - 33 **Bedi GC**, Westra WH, Gabrielson E, Koch W, Sidransky D. Multiple head and neck tumors: evidence for a common clonal origin. *Cancer Res* 1996; **56**: 2484-2487 [PMID: 8653681]
 - 34 **Filipe MI**. Value of histochemical reactions for mucosubstances in the diagnosis of certain pathological conditions of the colon and rectum. *Gut* 1969; **10**: 577-586 [PMID: 4241152 DOI: 10.1136/gut.10.7.577]
 - 35 **Nieuw Amerongen AV**, Bolscher JG, Bloemena E, Veerman EC. Sulfomucins in the human body. *Biol Chem* 1998; **379**: 1-18 [PMID: 9504711]
 - 36 **Filipe MI**, Branfoot AC. Abnormal patterns of mucus secretion in apparently normal mucosa of large intestine with carcinoma. *Cancer* 1974; **34**: 282-290 [PMID: 4850363 DOI: 10.1002/1097-0142(197408)34:2<282>
 - 37 **Saffos RO**, Rhatigan RM. Benign (nonpolyloid) mucosal changes adjacent to carcinomas of the colon. A light microscopic study of 20 cases. *Hum Pathol* 1977; **8**: 441-449 [PMID: 892796 DOI: 10.1016/S0046-8177(77)80008-7]
 - 38 **Dawson PA**, Filipe MI. An ultrastructural and histochemical study of the mucous membrane adjacent to and remote from carcinoma of the colon. *Cancer* 1976; **37**: 2388-2398 [PMID: 177188 DOI: 10.1002/1097-0142(197605)37:5<2388>
 - 39 **Riddell RH**, Levin B. Ultrastructure of the "transitional" mucosa adjacent to large bowel carcinoma. *Cancer* 1977; **40**: 2509-2522 [PMID: 922692 DOI: 10.1002/1097-0142(197711)40:5]
 - 40 **Bibbo M**, Michelassi F, Bartels PH, Dytch H, Bania C, Lerma E, Montag AG. Karyometric marker features in normal-appearing glands adjacent to human colonic adenocarcinoma. *Cancer Res* 1990; **50**: 147-151 [PMID: 1688372]
 - 41 **Verhest A**, Kiss R, d'Olne D, Larsimont D, Salmon I, de Launoit Y, Fourneau C, Pasteels JL, Pector JC. Characterization of human colorectal mucosa, polyps, and cancers by means of computerized

- morphonuclear image analyses. *Cancer* 1990; **65**: 2047-2054 [PMID: 1695545 DOI: 10.1002/1097-0142(19900501)65:9<2047]
- 42 **Montag AG**, Bartels PH, Dytch HE, Lerma-Puertas E, Michelassi F, Bibbo M. Karyometric features in nuclei near colonic adenocarcinoma. Statistical analysis. *Anal Quant Cytol Histol* 1991; **13**: 159-167 [PMID: 1716896]
- 43 **Alberts DS**, Einspahr JG, Krouse RS, Prasad A, Ranger-Moore J, Hamilton P, Ismail A, Lance P, Goldschmid S, Hess LM, Yozwiak M, Bartels HG, Bartels PH. Karyometry of the colonic mucosa. *Cancer Epidemiol Biomarkers Prev* 2007; **16**: 2704-2716 [PMID: 18086777 DOI: 10.1158/1055-9965.EPI-07-0595]
- 44 **Cherkezyan L**, Stypula-Cyrus Y, Subramanian H, White C, Dela Cruz M, Wali RK, Goldberg MJ, Bianchi LK, Roy HK, Backman V. Nanoscale changes in chromatin organization represent the initial steps of tumorigenesis: a transmission electron microscopy study. *BMC Cancer* 2014; **14**: 189 [PMID: 24629088 DOI: 10.1186/1471-2407-14-189]
- 45 **Terpstra OT**, van Blankenstein M, Dees J, Eilers GA. Abnormal pattern of cell proliferation in the entire colonic mucosa of patients with colon adenoma or cancer. *Gastroenterology* 1987; **92**: 704-708 [PMID: 3817391]
- 46 **Bernstein C**, Bernstein H, Garewal H, Dinning P, Jabi R, Sampliner RE, McCuskey MK, Panda M, Roe DJ, L'Heureux L, Payne C. A bile acid-induced apoptosis assay for colon cancer risk and associated quality control studies. *Cancer Res* 1999; **59**: 2353-2357 [PMID: 10344743]
- 47 **Anti M**, Armuzzi A, Morini S, Iacone E, Pignataro G, Coco C, Lorenzetti R, Paolucci M, Covino M, Gasbarrini A, Vecchio F, Gasbarrini G. Severe imbalance of cell proliferation and apoptosis in the left colon and in the rectosigmoid tract in subjects with a history of large adenomas. *Gut* 2001; **48**: 238-246 [PMID: 11156647]
- 48 **Badvie S**, Hanna-Morris A, Andreyev HJ, Cohen P, Saini S, Allen-Mersh TG. A "field change" of inhibited apoptosis occurs in colorectal mucosa adjacent to colorectal adenocarcinoma. *J Clin Pathol* 2006; **59**: 942-946 [PMID: 16679352 DOI: 10.1136/jcp.2005.033431]
- 49 **Anti M**, Marra G, Armelao F, Percesepe A, Ficarelli R, Ricciuto GM, Valenti A, Rapaccini GL, De Vitis I, D'Agostino G. Rectal epithelial cell proliferation patterns as predictors of adenomatous colorectal polyp recurrence. *Gut* 1993; **34**: 525-530 [PMID: 8491402]
- 50 **Hanna-Morris A**, Badvie S, Cohen P, McCullough T, Andreyev HJ, Allen-Mersh TG. Minichromosome maintenance protein 2 (MCM2) is a stronger discriminator of increased proliferation in mucosa adjacent to colorectal cancer than Ki-67. *J Clin Pathol* 2009; **62**: 325-330 [PMID: 18474544 DOI: 10.1136/jcp.2007.054643]
- 51 **Ngoi SS**, Staiano-Coico L, Godwin TA, Wong RJ, DeCosse JJ. Abnormal DNA ploidy and proliferative patterns in superficial colonic epithelium adjacent to colorectal cancer. *Cancer* 1990; **66**: 953-959 [PMID: 2386922 DOI: 10.1002/1097-0142(19900901)66:5<953>]
- 52 **Saccani Jotti G**, Fontanesi M, Orsi N, Sarli L, Pietra N, Peracchia A, Sansesebastiano G, Becchi G. DNA content in human colon cancer and non-neoplastic adjacent mucosa. *Int J Biol Markers* 1995; **10**: 11-16 [PMID: 7629421]
- 53 **Worthley DL**, Whitehall VL, Buttenshaw RL, Irahara N, Greco SA, Ramsnes I, Mallitt KA, Le Leu RK, Winter J, Hu Y, Ogino S, Young GP, Leggett BA. DNA methylation within the normal colorectal mucosa is associated with pathway-specific predisposition to cancer. *Oncogene* 2010; **29**: 1653-1662 [PMID: 19966864 DOI: 10.1038/onc.2009.449]
- 54 **Chen LC**, Hao CY, Chiu YS, Wong P, Melnick JS, Brotman M, Moretto J, Mendes F, Smith AP, Bennington JL, Moore D, Lee NM. Alteration of gene expression in normal-appearing colon mucosa of APC(min) mice and human cancer patients. *Cancer Res* 2004; **64**: 3694-3700 [PMID: 15150130 DOI: 10.1158/0008-5472.CAN-03-3264]
- 55 **Hao CY**, Moore DH, Chiu YS, Wong P, Bennington JL, Smith AP, Chen LC, Lee NM. Altered gene expression in normal colonic mucosa of individuals with polyps of the colon. *Dis Colon Rectum* 2005; **48**: 2329-2335 [PMID: 16400515 DOI: 10.1007/s10350-005-0153-2]
- 56 **Facista A**, Nguyen H, Lewis C, Prasad AR, Ramsey L, Zaitlin B, Nfonsam V, Krouse RS, Bernstein H, Payne CM, Stern S, Oatman N, Banerjee B, Bernstein C. Deficient expression of DNA repair enzymes in early progression to sporadic colon cancer. *Genome Integr* 2012; **3**: 3 [PMID: 22494821 DOI: 10.1186/2041-9414-3-3]
- 57 **Shen L**, Kondo Y, Rosner GL, Xiao L, Hernandez NS, Vilaythong J, Houlihan PS, Krouse RS, Prasad AR, Einspahr JG, Buckmeier J, Alberts DS, Hamilton SR, Issa JP. MGMT promoter methylation and field defect in sporadic colorectal cancer. *J Natl Cancer Inst* 2005; **97**: 1330-1338 [PMID: 16174854 DOI: 10.1093/jnci/dji275]
- 58 **Ramírez N**, Bandrés E, Navarro A, Pons A, Jansa S, Moreno I, Martínez-Rodenas F, Zárata R, Bitarte N, Monzó M, García-Foncillas J. Epigenetic events in normal colonic mucosa surrounding colorectal cancer lesions. *Eur J Cancer* 2008; **44**: 2689-2695 [PMID: 18938072 DOI: 10.1016/j.ejca.2008.09.004]
- 59 **Svrcek M**, Buhard O, Colas C, Coulet F, Dumont S, Massaoudi I, Lamri A, Hamelin R, Cosnes J, Oliveira C, Seruca R, Gaub MP, Legrain M, Collura A, Lascols O, Tiret E, Fléjou JF, Duval A. Methylation tolerance due to an O6-methylguanine DNA methyltransferase (MGMT) field defect in the colonic mucosa: an initiating step in the development of mismatch repair-deficient colorectal cancers. *Gut* 2010; **59**: 1516-1526 [PMID: 20947886 DOI: 10.1136/gut.2009.194787]
- 60 **Belshaw NJ**, Elliott GO, Foxall RJ, Dainty JR, Pal N, Coupe A, Garg D, Bradburn DM, Mathers JC, Johnson IT. Profiling CpG island field methylation in both morphologically normal and neoplastic human colonic mucosa. *Br J Cancer* 2008; **99**: 136-142 [PMID: 18542073 DOI: 10.1038/sj.bjc.6604432]
- 61 **Paun BC**, Kukuruga D, Jin Z, Mori Y, Cheng Y, Duncan M, Stass SA, Montgomery E, Hutcheon D, Meltzer SJ. Relation between normal rectal methylation, smoking status, and the presence or absence of colorectal adenomas. *Cancer* 2010; **116**: 4495-4501 [PMID: 20572039 DOI: 10.1002/cncr.25348]
- 62 **Grady WM**, Parkin RK, Mitchell PS, Lee JH, Kim YH, Tsuchiya KD, Washington MK, Paraskeva C, Willson JK, Kaz AM, Kroh EM, Allen A, Fritz BR, Markowitz SD, Tewari M. Epigenetic silencing of the intronic microRNA hsa-miR-342 and its host gene EVL in colorectal cancer. *Oncogene* 2008; **27**: 3880-3888 [PMID: 18264139 DOI: 10.1038/onc.2008.10]
- 63 **Deng G**, Kakar S, Kim YS. MicroRNA-124a and microRNA-34b/c are frequently methylated in all histological types of colorectal cancer and polyps, and in the adjacent normal mucosa. *Oncol Lett* 2011; **2**: 175-180 [PMID: 22870149 DOI: 10.3892/ol.2010.222]
- 64 **Balaguer F**, Link A, Lozano JJ, Cuatrecasas M, Nagasaka T, Boland CR, Goel A. Epigenetic silencing of miR-137 is an early event in colorectal carcinogenesis. *Cancer Res* 2010; **70**: 6609-6618 [PMID: 20682795 DOI: 10.1158/0008-5472.CAN-10-0622]
- 65 **Isaacson P**, Attwood PR. Failure to demonstrate specificity of the morphological and histochemical changes in mucosa adjacent to colonic carcinoma (transitional mucosa). *J Clin Pathol* 1979; **32**: 214-218 [PMID: 429587]
- 66 **Lev R**, Lance P, Camara P. Histochemical and morphologic studies of mucosa bordering rectosigmoid carcinomas: comparisons with normal, diseased, and malignant colonic epithelium. *Hum Pathol* 1985; **16**: 151-161 [PMID: 2579014]
- 67 **Listinsky CM**, Riddell RH. Patterns of mucin secretion in neoplastic and non-neoplastic diseases of the colon. *Hum Pathol* 1981; **12**: 923-929 [PMID: 6170566]
- 68 **Katsurano M**, Niwa T, Yasui Y, Shigematsu Y, Yamashita S, Takeshima H, Lee MS, Kim YJ, Tanaka T, Ushijima T. Early-stage formation of an epigenetic field defect in a mouse colitis model, and non-essential roles of T- and B-cells in DNA methylation induction. *Oncogene* 2012; **31**: 342-351 [PMID: 21685942 DOI: 10.1038/onc.2011.241]
- 69 **Dawson PM**, Habib NA, Rees HC, Wood CB. Mucosal field change in colorectal cancer. *Am J Surg* 1987; **153**: 281-284 [PMID: 2435183]
- 70 **Polley AC**, Mulholland F, Pin C, Williams EA, Bradburn DM,

- Mills SJ, Mathers JC, Johnson IT. Proteomic analysis reveals field-wide changes in protein expression in the morphologically normal mucosa of patients with colorectal neoplasia. *Cancer Res* 2006; **66**: 6553-6562 [PMID: 16818627 DOI: 10.1158/0008-5472.CAN-06-0534]
- 71 **Milicic A**, Harrison LA, Goodlad RA, Hardy RG, Nicholson AM, Presz M, Sieber O, Santander S, Pringle JH, Mandir N, East P, Obszynska J, Sanders S, Piazuolo E, Shaw J, Harrison R, Tomlinson IP, McDonald SA, Wright NA, Jankowski JA. Ectopic expression of P-cadherin correlates with promoter hypomethylation early in colorectal carcinogenesis and enhanced intestinal crypt fission in vivo. *Cancer Res* 2008; **68**: 7760-7768 [PMID: 18829530 DOI: 10.1158/0008-5472.CAN-08-0020]
- 72 **Gomes AJ**, Roy HK, Turzhitsky V, Kim Y, Rogers JD, Ruderman S, Stoyneva V, Goldberg MJ, Bianchi LK, Yen E, Kromine A, Jameel M, Backman V. Rectal mucosal microvascular blood supply increase is associated with colonic neoplasia. *Clin Cancer Res* 2009; **15**: 3110-3117 [PMID: 19383816 DOI: 10.1158/1078-0432.CCR-08-2880]
- 73 **Wood LD**, Parsons DW, Jones S, Lin J, Sjöblom T, Leary RJ, Shen D, Boca SM, Barber T, Ptak J, Silliman N, Szabo S, Dezso Z, Ustyanksky V, Nikolskaya T, Nikolsky Y, Karchin R, Wilson PA, Kaminker JS, Zhang Z, Croshaw R, Willis J, Dawson D, Shipitsin M, Willson JK, Sukumar S, Polyak K, Park BH, Pethiyagoda CL, Pant PV, Ballinger DG, Sparks AB, Hartigan J, Smith DR, Suh E, Papadopoulos N, Buckhaults P, Markowitz SD, Parmigiani G, Kinzler KW, Velculescu VE, Vogelstein B. The genomic landscapes of human breast and colorectal cancers. *Science* 2007; **318**: 1108-1113 [PMID: 17932254 DOI: 10.1126/science.1145720]
- 74 **Roy HK**, Liu Y, Wali RK, Kim YL, Kromine AK, Goldberg MJ, Backman V. Four-dimensional elastic light-scattering fingerprints as preneoplastic markers in the rat model of colon carcinogenesis. *Gastroenterology* 2004; **126**: 1071-1081; discussion 948 [PMID: 15057746]
- 75 **Schöllnberger H**, Beerenwinkel N, Hoogenveen R, Vineis P. Cell selection as driving force in lung and colon carcinogenesis. *Cancer Res* 2010; **70**: 6797-6803 [PMID: 20656803 DOI: 10.1158/0008-5472.CAN-09-4392]
- 76 **Bozic I**, Antal T, Ohtsuki H, Carter H, Kim D, Chen S, Karchin R, Kinzler KW, Vogelstein B, Nowak MA. Accumulation of driver and passenger mutations during tumor progression. *Proc Natl Acad Sci USA* 2010; **107**: 18545-18550 [PMID: 20876136 DOI: 10.1073/pnas.1010978107]
- 77 **Backman V**, Roy HK. Advances in biophotonics detection of field carcinogenesis for colon cancer risk stratification. *J Cancer* 2013; **4**: 251-261 [PMID: 23459690 DOI: 10.7150/jca.5838]
- 78 **Roy HK**, Turzhitsky V, Kim Y, Goldberg MJ, Watson P, Rogers JD, Gomes AJ, Kromine A, Brand RE, Jameel M, Bogovejic A, Pradhan P, Backman V. Association between rectal optical signatures and colonic neoplasia: potential applications for screening. *Cancer Res* 2009; **69**: 4476-4483 [PMID: 19417131 DOI: 10.1158/0008-5472.CAN-08-4780]
- 79 **Mutyal NN**, Radosevich A, Gould B, Rogers JD, Gomes A, Turzhitsky V, Backman V. A fiber optic probe design to measure depth-limited optical properties in-vivo with low-coherence enhanced backscattering (LEBS) spectroscopy. *Opt Express* 2012; **20**: 19643-19657 [PMID: 23037017]
- 80 **Damania D**, Roy HK, Subramanian H, Weinberg DS, Rex DK, Goldberg MJ, Muldoon J, Cherkezyan L, Zhu Y, Bianchi LK, Shah D, Pradhan P, Borkar M, Lynch H, Backman V. Nanocytology of rectal colonocytes to assess risk of colon cancer based on field cancerization. *Cancer Res* 2012; **72**: 2720-2727 [PMID: 22491589 DOI: 10.1158/0008-5472.CAN-11-3807]
- 81 **Galandiuk S**, Rodriguez-Justo M, Jeffery R, Nicholson AM, Cheng Y, Oukrif D, Elia G, Leedham SJ, McDonald SA, Wright NA, Graham TA. Field cancerization in the intestinal epithelium of patients with Crohn's ileocolitis. *Gastroenterology* 2012; **142**: 855-864.e8 [PMID: 22178590 DOI: 10.1053/j.gastro.2011.12.004]

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