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A transgenic mouse model to inducibly target prosurvival Bcl2 proteins with selective BH3 peptides *in vivo*

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Dear Editor,

BH3-mimetic drugs that antagonize Bcl-2 family prosurvival proteins are effective against some cancers, particularly those with abnormally high expression of the prosurvival protein target.¹ Despite the recent success of BH3-mimetics targeting Bcl-2, Bcl-x₁ and Bcl-w, mechanism-based cell killing in vivo by targeting other important prosurvival proteins such as Mcl-1 or Bfl-1 has vet to be demonstrated. In the absence of small molecules targeting these prosurvival proteins, BH3-domain peptides, the prototypes for this class of drug, are useful because their binding specificity profile can be manipulated.^{2,3} Such peptides have been applied in in vitro studies to validate the targeting of particular prosurvival proteins in certain tumor types.^{4,5} However, in vivo applications require technically challenging chemical modifications of peptides, and while mouse xenograft models of virally-infected BH3 domainexpressing tumor cells can be used, this does not allow evaluation of effects of the ligand on normal tissues.

Here we have generated transgenic mice in which peptidebased BH3 ligands can be inducibly expressed to evaluate their effects in vivo and provide proof-of-principle for similaracting drugs. To achieve this we adapted a previously described strategy allowing FLP-recombinase-mediated insertion of an expression cassette at an frt 'landing pad' at the type I collagen (Col1a1) locus in mouse embryonic stem cells.⁶ The cassette comprises sequences encoding a BH3domain protein under control of a tetracycline (tet)-regulated element (TRE) promoter. To develop a system that is broadly applicable to BH3 domains with different specificities, we employed the Bims BH3-only protein as a scaffold in which BH3 domains with different specificities could replace the native BH3 sequence (Figure 1a). Bim_S is an intrinsically unstructured protein that tolerates extensive mutation of its BH3 sequence, and Bim_sBH3 chimeras display the prosurvival protein specificity profile of the replacement BH3 domain.⁷ In this study we focussed on the BH3 domain of Bad because it targets the prosurvival proteins Bcl-2, Bcl-x, and Bcl-w. Hence Bim_sBad expression should mimic the BH3-mimetics ABT-737 and ABT-263 that have been extensively studied in vivo.1

We generated TRE-Bim_SBad transgenic mice and crossed them to mice expressing the rtTA (tet-on) transactivator under the control of the cytomegalovirus (CMV) promoter, which provides high-level expression in many tissues including blood.⁸ Bitransgenic mice were then treated with doxycycline (Dox) to induce Bim_sBad expression (Figure 1a). Western blot analysis verified Dox-inducible transgene expression in white blood cells of BimsBad: CMV-rtTA bitransgenic mice (Figure 1b). As ABT-737/263 induces thrombocytopenia in mice and humans due to antagonism of Bcl-x_L, we measured platelet levels as a biomarker for functional expression of the ligand.¹ Notably, blood cell analysis of these mice revealed a significant reduction (~65% decrease) in platelet counts (Figure 1c and d), a degree of thrombocytopenia comparable with that seen in patients administered with ABT-263.9 Importantly, platelet counts rebounded to normal levels following removal of Dox for 7 days (Figure 1d), illustrating the reversibility of the system. We also generated an additional transgenic mouse strain allowing inducible expression of a Bim_SBad construct possessing a BH3 sequence mutation (Bim_SBad^{mut}; Figure 1a) that decreases its affinity for Bcl-x_L by > 40-fold (K_D 8.5 nM *versus* < 0.2 nM as measured by surface plasmon resonance). Induction of Bim_SBad^{mut} expression (Figure 1b) did not alter blood cell or platelet counts (Figure 1d), indicating that the thrombocytopenia observed in Bim_SBad mice is due to Bcl-x_L inhibition. These data provide the first evidence that BH3-only proteins can be inducibly expressed in a mouse model, effectively mimicking at least one functional consequence (reduced platelet counts) seen in mice and humans treated with BH3-mimetic drugs of similar specificity. We envisage multiple applications for similarly engineered mice. For example, tet-regulated expression of different Bims variants in mice could reveal toxicities associated with neutralization of their prosurvival protein targets (individually or in combination), and tissue-specific effects could be addressed by crossing onto mice where rtTA expression is driven by different promoters. Moreover, mice crossed to different tumor-prone models could provide in vivo evidence for the tumor-killing efficacy associated with different BH3 specificities not yet available through small molecule

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BadBH3^{mut}: ¹⁰³NLWAAQRYGRELRRMSDEAVDSFKKG₁₂₈

b
TRE-BH3 ligan
CMV-rtTA



С

d

npg

2

	Doxycycline 7 days		
	+/+;+/+ (n=7)	TRE-Bim _s Bad (n=15)	TRE-Bim _s Bad; CMV-rtTA (n=17)
Erythrocytes (x10 ⁶ µL ⁻¹)	11.19 ± 0.17	11.32 ± 0.20	11.13 ± 0.13
Hematocrit (%)	54.91 ± 1.05	55.75 ± 0.56	55.48 ± 0.65
MCV (femtolitres)	49.07 ± 0.44	48.46 ± 0.30	49.85 ± 0.34
Leukocytes (x10 ³ µL ⁻¹)	9.81 ± 0.64	10.29 ± 0.53	9.12 ± 0.37
Neutrophils (x10 ³ μ L ⁻¹)	0.91 ± 0.20	0.80 ± 0.24	1.19 ± 0.20
Lymphocytes (x10 ³ μ L ⁻¹)	8.47 ± 0.55	8.93 ± 1.93	7.12 ± 1.12
Monocytes (x10 ³ µL ⁻¹)	0.17 ± 0.047	0.17 ± 0.058	0.25 ± 0.030
Platelets (x10 ³ µL ⁻¹)	1240 ± 61.82	1200 ± 36.12	439 ± 30.53****
MPV (femtolitres)	6.82 ± 0.23	6.93 ± 0.12	7.42 ± 0.098







Figure 1 Expression of Bim_sBad reduces platelet levels in mice. (a) Schematic of the CMV-rtTA and TRE-Bim_sBad transgenes. Upon Dox treatment of bitransgenic mice, the rtTA (tet-on) protein transactivates the TRE promoter to drive expression of N-terminally FLAG tagged Bim_sBad. Targeting vectors were generated by cloning Mlul-flanked FLAG-Bim_sBad/Bim_sBad^{mut} PCR amplicons into the Mlul site of a modified version of the pgkATGfrt vector⁶ in which the TRE promoter has been replaced with TREtight. BH3 domain numbering refers to the amino acid residue position within the respective Bim_s or Bad protein sequences. The BadBH3 residue in red (F121) was mutated to alanine in the Bim_sBad^{mut} mice. Targeted ES cell clones were injected into C57Bl/6 blastocysts and chimeras crossed to C57Bl/6 female mice. All mouse colonies were maintained by C57Bl/6 backcrossing. (b) Western blot of white blood cells isolated from TRE-Bim_sBad/Bim_sBad^{mut}, CMV-rtTA bitransgenic mice or wild-type or Bim_sBad/Bim_sBad^{mut} single transgenic control mice following 7 days of Dox food (600 mg/kg). Blood (100 µl) was cleared of red blood cells by 10-fold dilution in red cell lysis buffer for 5 min followed by centrifugation. The white blood cell-containing pellet was then resuspended in lysis buffer (20 mM Tris pH 7.4, 135 mM NaCl, 1.5 mM MgCl₂, 1 mM EGTA, 1% Triton X100 and 10% glycerol) for 1 h on ice, followed by centrifugation. The supernatant was then analyzed by Western blot probed with an anti- β -actin as loading control. * Indicates a non-specific band (c) Blood cell parameters after Dox treatment of bitransgenic and control animals for 7 days. *****P* < 0.0001 (*t*-test) compared with littermate controls (d) Platelet levels in TRE-Bim_sBad bitransgenic mice treated with Dox for 7 days rebound to normal levels after 7 days without Dox treatment. No changes in platelet levels were observed in Bim_sBad^{mut} mice following Dox treatment

drugs, and extended to studies on combination therapies with existing drugs.

Conflict of Interest

The authors declare no conflict of interest.

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- 1. Roy MJ et al. Br J Pharmacol 2014; 171: 1973–1987.
- 2. Lee EF et al. J Cell Biol 2008; 180: 341-355.

- 3. Foight GW et al. ACS Chem Biol 2014; 9: 1962-1968.
- 4. Glaser SP et al. Genes Dev 2012; 26: 120-125.
- 5. Del Gaizo Moore V, Letai A. Cancer Lett 2013; 332: 202-205.
- 6. Beard C et al. Genesis 2006; 44: 23-28.
- 7. Chen L et al. Mol Cell 2005; 17: 393-403.
- 8. Takiguchi M et al. PLoS One 2013; 8: e54009.
- 9. Roberts AW et al. J Clin Oncol 2012; 30: 488-496.

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