ORIGINAL ARTICLE

The Analgesic Effects of Different Extracts of Aerial Parts of Coriandrum Sativum in Mice

Seyedeh Fatemeh Kazempor¹, Shabnam Vafadar langehbiz¹, Mahmoud Hosseini¹, Mohammad Naser Shafei², Ahmad Ghorbani³, Masoomeh Pourganji²

¹Neurocognitive Research Center & Department of Physiology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran; ²Neurogenic Inflammation Research Center & Department of Physiology, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran; ³Pharmacological Research Center of Medicinal Plants, School of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

ABSTRACT

Regarding the effects of Coriandrum sativum (C. sativum) on central nervous system, in the present study analgesic properties of different extracts of C. sativum aerial parts were investigated. The mice were treated by saline, morphine, three doses (20, 100 and 500 mg/kg) of aqueous, ethanolic, choloroformic extracts of C. sativum and one dose (100 mg/kg) of aqueous, two doses of ethanolic (100 and 500 mg/kg) and one dose of choloroformic (20 mg/kg) extracts of C. sativum pretreated by naloxone. Recording of the hot plate test was performed 10 min before injection of the drugs as a base and it was consequently repeated every 10 minutes after the extracts injection. The maximal percent effect (MPE) in the groups treated by three doses of aqueous, ethanolic and chloroformic extracts were significantly higher than saline group which were comparable to the effect of morphine. The effects of most effective doses of extracts were reversed by naloxone. The results of present study showed analgesic effect of aqueous, ethanolic and chloroformic extracts of C. sativum extract. These effects of the extracts may be mediated by opioid system. However, more investigations are needed to elucidate the exact responsible mechanism(s) and the effective compound(s) (Int J Biomed Sci 2015; 11 (1): 23-28).

Keywords: Analgesia; Coriandrum sativum; Hot plate; Mice; Morphine; Naloxone

Corresponding author: Mahmoud Hosseini, Department of Physiology, School of Medicine, Azadi Square, Mashhad, Iran. Postal Code: 9177948564; Tel: +98 511 8828565; Fax: +98 511 8828564. E-mail: Hosseinim@mums.ac.ir.

Received July 23, 2014; Accepted October 17, 2014

Copyright: © 2015 Seyedeh Fatemeh Kazempor et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.5/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

INTRODUCTION

Pain is an unpleasant sensation which is elicited by damaging or potentially damaging noxious stimuli. Several factors including sociocultural, psychological and biological conditions have important roles in pain perception (1-3). Pain relief can be achieved by nonpharmacologic approaches or by administration of a diversity of drugs. Analgesic drugs are used in single or in combination to affect peripheral or central nervous system (CNS) to decrease pain sensation (4). At present, the most widely used

medications for management of pain are acetaminophen, opioids, and nonsteroidal anti-inflammatory agents. However, the clinical uses of these drugs are accompanied with unpleasant side effects such as hepatic failure, gastrointestinal events, renal dysfunction, respiratory depression, and addiction (5). Therefore, the search for new analgesic agents with lesser side effects and more efficacies is of great interest.

Medicinal plants have been repeatedly considered as one of the main sources of medicines for treatment of several health problems of human. They are widely used to treat a wide range of CNS disorders including neurodegenerative diseases, stroke, dementia, seizures and insomnia (6-10).

Coriandrum sativum (C. sativum) is an annual herb belonging to the Apiaceae family. It is also known as coriander, cilantro, Arab parsley, Chinese parsley and Dhania (11). Although, all parts of the plant are edible, its fresh leaves and dried seeds are most frequently used in many cultures. C. sativum is widely used in traditional medicine to treat anxiety, dizziness, headache, edema, fever, digestive disorders, respiratory diseases, allergies, and burns (12).

Experimentally, *C. sativum* has been reported to have a wide range of biological activities including sedative, hypnotic, anti-inflammatory, antidiabetic, hypolipidemic, neuroprotective, and hepatoprotective effects (13-18). Also it has strong antioxidant activity which is superior to known antioxidants like ascorbic acid (16, 19-23).

In our previous work, it was found that aerial parts of this plant bearing compounds which show sedative/hypnotic effects in mice (13). Also, Emamghoreishi and colleagues reported that aqueous extract of *C. sativum* seed has anxiolytic and muscle relaxant effects (11). Considering these effects of *C. sativum* on nervous system, we aimed to test the possible analgesic effects of different extract of *C. sativum*.

MATERIALS AND METHODS

Drugs and chemicals

Morphine and naloxone was purchased from Temad Company (IRAN). Chloroform (99.8%) and ethanol (96%) were obtained from Merck Company.

Animal groups

In this study, 120 virgin male mice (27-32 g in weight) were used. The animals were maintained at the animal house under controlled conditions including 12 h light and

dark cycle, 22-24°C temperature and appropriate humidity with laboratory chow and water provided ad libitum. The study protocol using the laboratory rats complied with the general guidelines of the animal care of Mashhad University of Medical Sciences, Iran. The animals were divided into 15 groups (n=8 in each group)and treated as follows: (group1) saline as control group, (group 2) morphine (10 mg/kg), (groups 3-5) three doses of aqueous extract of C. sativum (20, 100 and 500 mg/kg), (groups 6-8) three doses of ethanolic extract of C. sativum (20, 100 and 500 mg/ kg), (groups 9-11) three doses of chloroformic extract of C. sativum (20, 100 and 500 mg/kg), (group 12) 100 mg/kg aqueous extract pretreated by naloxone (4 mg/kg), (groups 13-14) two doses of ethanolic extract (100 and 500 mg/kg) pretreated by naloxone (4 mg/kg) and (group 15) 20 mg/kg of choloroformic extract of C. sativum extract pretreated by naloxone (4 mg/kg).

Plant extracts

The aerial parts (leaves, stems, twigs) of *C. sativum* were collected from Neyshabur area, (Razavi Khorasan state, Iran). The identity of the plant was confirmed and for future reference a voucher specimen (10068) was deposited at the herbarium of school of Pharmacy (Mashhad University of Medical Sciences, Iran). The plant aerial parts were air-dried under shade at room temperature. Then, the dried aerial parts (50 g) were chopped, finely grounded, and extracted using a Soxhlet apparatus. Duration of extraction was 48 hours and 300 ml of distilled water, ethanol, or chloroform were used as solvent to prepare aqueous, ethanolic and chloroformic extracts, respectively (24-26). The extracts reduced to dryness with a rotary vacuum evaporator. The yields of aqueous, ethanolic and chloroformic extracts were 15%,17% and 5% respectively.

Nociceptive test

To assess nociceptive responses, hot plate method was used. The mice were placed on the hot plate with temperature setting controlled at 55 ± 0.2 °C. Cut-off time was 60 s. Nociceptive response was defined as licking forepaws or moving hind paws. Time duration between placing the animals on hot plate and licking fore paws or moving hind paws was considered as the reaction time. The hot plate test was performed as a base record 10 min before injection of the drugs and consequently it was repeated 5 times, every 10 min after injection. Analgesic effects of the extracts or vehicle were calculated as maximal possible effect (MPE) [MPE (%) = [(test response time-basal response time)/(cut-off time-basal response time) \times 100%] (1- 3, 27-29).

Statistical analysis

All data were expressed as Mean \pm SEM and analyzed by using ANOVA followed by Tukey's post hoc comparison test. P values less than 0.05 were considered to be statistically significant.

RESULTS

Analgesic activity of aqueous extract

The results showed that MPE in the animals treated by 20 mg/kg of aqueous extract was significantly higher than that of control group at 30 and 50 min after injection of the extract (P<0.05-P<0.01). Treatment of the animals by 100 mg/kg of the extract also increased MPE at10, 20, 30, 40 and 50 min compared to vehicle treated animals (P<0.05-P<0.001). MPE in animals treated by 500 mg/kg of the aqueous extract was higher than that of control only at 30 min after administration (P<0.01). Morphine administration increased MPE at all times after injection (P<0.001) (Fig. 1).

Analgesic activity of ethanolic extract

The results showed that MPE in the group treated by 20 mg/kg of ethanolic extract was significantly higher than that of control group at all times after injection of the extract (P<0.01-P<0.001). Treatment of the animals by

100 mg/kg of the extract also increased MPE at all times compared to vehicle treated animals (all P<0.001). MPE in animals treated by 500 mg/kg of the ethanolic extract was higher than that of control at 10-50 min after administration (P<0.01-P<0.001). Morphine administration increased MPE at all times after injection (P<0.001) (Fig. 2).

Analgesic activity of chloroformic extract

The results showed that MPE in the animal group treated by 20 mg/kg of chloroformic extract was significantly higher than that of control group all times after injection of the extract (P<0.01-P<0.001). Treatment of the animals by 100 mg/kg of the extract also increased MPE at 30, 40 and 50 min after injection compared to vehicle treated animals (P<0.01-P<0.001). MPE in animals treated by 500 mg/kg of the aqueous extract was higher than that of control at 10-50 min after administration (P<0.05-P<0.001). Morphine administration increased MPE at all times after injection (P<0.01-P<0.001) (Fig. 3).

Effect of naloxone on analgesic activity of the extracts

Pretreatment by naloxone reduced analgesic effect of 100 mg/kg of aqueous extract at 20 and 50 min (P<0.05-P<0.01, respectively) (Fig. 4). MPE of the animals pre-

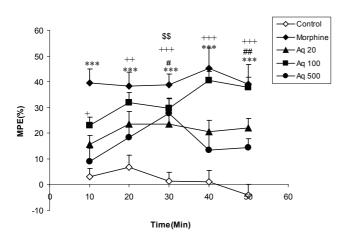


Figure 1. Comparison of MPE between three groups treated by 20, 100 and 500 mg/kg of aqueous extract of *C. sativum*(Aq 20, Aq 100, Aq 500 groups) morphine and control groups. $^{\#}P$ <0.05, $^{\#}P$ <0.01 comparison of 20 mg/kg of aqueous extract (Aq 20) and control. ^{+}P <0.05, ^{++}P <0.01, ^{++}P <0.001 comparison of 100 mg/kg of aqueous extract (Aq 100) and control. ^{58}P <0.01 comparison of 500 mg/kg of aqueous extract(Aq 500) and control. $^{**}P$ <0.001 comparison of morphine group (Mor) and control.

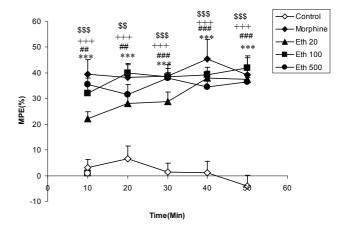


Figure 2. Comparison of MPE between three groups treated by 20, 100 and 500 mg/ kg of ethanolic extract of *C. sativum* (Eth 20, Eth 100, Eth 500 groups) morphine and control groups. ***P<0.01, ****P<0.001 comparison of 20 mg/ kg of ethanolic extract (Eth 20) and control. ****P<0.001 comparison of 100 mg/kg of ethanolic extract (Eth 100) and control. *\$\$P<0.01, \$\$\$\$P<0.001 comparison of 500 mg/kg of ethanolic extract (Eth 500) and control. ****P<0.001 comparison of morphine group (Mor) and control.

treated by naloxone before 100 mg/kg of the extract was significantly lower than that of animals treated only by 100 mg/kg of the extract at all times (P < 0.05-P < 0.01). The results also showed that naloxone reduced the analgesic activity of 500 mg/kg of the ethanolic extract at 10 and

50 min (both P<0.01) (Fig. 5). As the Figure 6 shows the MPE of the animals treated by naloxone before 20 mg/kg of chloroformic extract was significantly lower than that of the animals treated by only 20 mg/kg of the extract (P<0.05-P<0.001).

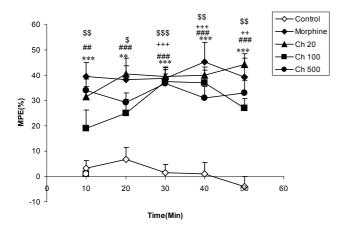


Figure 3. Comparison of MPE between three groups treated by 20, 100 and 500 mg/ kg of chloroformic extract of *C. sativum* (Eth 20, Eth 100, Eth 500 groups) morphine and control groups. ****P*<0.01, ******P*<0.001 comparison of 20 mg/kg of chloroformic extract (Ch 20) and control. ****P*<0.01, ******P*<0.001 comparison of 100 mg/kg of chloroformic extract (Ch 100) and control. ***P*<0.05, ****P*<0.01, *****P*<0.01 comparison of 500 mg/kg of chloroformic extract (Ch 500) and control. ****P*<0.01, *****P*<0.001 comparison of morphine group (Mor) and control.

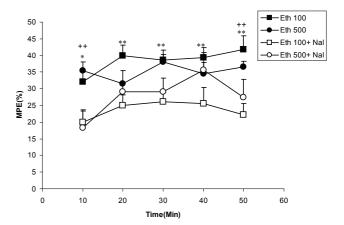


Figure 5. Comparison of MPE between groups treated by 100 mg/ kg of ethanolic extract of *C. sativum* (Eth 100 group) and a group pretreated by naloxone before 100 mg/kg of ethanolic extract of *C. sativum* (Eth 100 + Nal group). This figure also shows comparison of MPE between groups treated by 500 mg/kg of ethanolic extract of *C. sativum* (Eth 500 group) and a group pretreated by naloxone before 500 mg/kg of ethanolic extract of *C. sativum* (Eth 500 + Nal group). *P<0.01comparison between Eth 100 and Eth 100 + Nal groups. *P<0.01comparison between Eth 500 and Eth 500 + Nal groups.

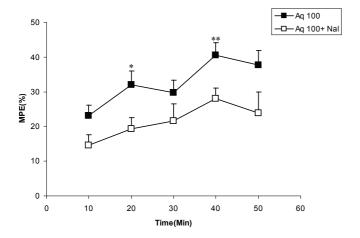


Figure 4. Comparison of MPE between groups treated by 100 mg/ kg of aqueous extract of C. sativum (Aq 100 group) and a group pretreated by naloxone before 100 mg/ kg of aqueous extract of C. sativum(Aq 100 + Nal group). *P <0.05, $^{**}P$ <0.01 comparison between Aq 100 and Aq 100 + Nal groups.

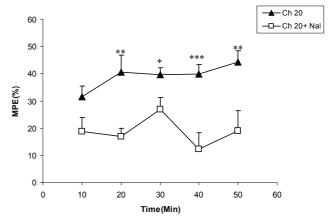


Figure 6. Comparison of MPE between groups treated by 20 mg/kg of chloroformic extract of C. sativum(Ch 20 group) and a group pretreated by naloxone before 20 mg/kg of aqueous extract of C. sativum(Ch 20 + Nal group). *P<0.05, **P<0.01, ***P<0.01 comparison between Ch 20 and Ch 20 + Nal groups.

DISCUSSION

The results of present study showed that three extracts (aqueous, ethanolic and chloroformic) of aerial parts of C. sativum had analgesic effects. The analgesic effect of the extracts was comparable to morphine and was attenuated by naloxone pretreatment. Consistent with this finding, it was previously shown that percolated methanolic extract of C. sativum had analgesic effect which was reversible by naloxone (30). In another study analgesic effects of the aqueous extract of the seeds of C. sativum had been reported using hot plate and tail flick tests (31). Because aerial parts of C. sativum are widely consumed as vegetable all over the world and no pharmacological studies have been yet evaluated the analgesic activity of different extracts of these parts of the plant, by evaluating we showed that C. sativum aerial parts had analgesic effects. The effects of ethanolic and chloroformic extracts were higher than aqueous extract; therefore it seems that the compounds that are soluble in ethanol or chloroform have potent analgesic effects.

The chemical compound(s) responsible for the analgesic effect of the extracts which didn't identify in the present study and need to be future studies. However, the presence of the flavonoids such as quercitin has been reported (32). It has been shown that the flavonoids have considerable anticonvulsant and analgesic effects (33-35). Sedative, CNS depressant and analgesic effects of falvonoids such as quercetin has been attributed to the affinity for the central benzodiazepine receptors (33, 34, 36-40). The beneficial effect of linalool in pentylenetetrazole seizure models as well its analgesic effects has been suggested (41, 42). It might be suggested that the beneficial effects of the extracts which were observed in the present study are at least in part due to linalool which is a main compound in coriander (43). Analgesic effects of polyphenols including rutin, caffeic acid, and gallic acid has also been reported which can be isolated from *C. sativum* (9, 44-49).

A huge number of other components have also been suggested for coriander, which are including ferrulic acid, chlorogenic acid, caroteinoids, anethole, borneol, camphene, camphor, carvone, cineole, citronelol, coriandrol, coriandrin, coumarins and hydroxy-coumarins (umbelliferone and scopoletin), p-cymene, euginol, geraniol, geranyl acetate, limonene, d(+)- linalool, myrcene, α - and β -phellandrene, α - and β -pinenes, α - and β -terpinene, 5- and 8-methoxypsoralens, tannins, and many others (9, 44, 49). Each of these compounds may also have a role in the analgesic effects of the extracts

which were seen in the present study however, it needs to be more investigated.

In conclusion, the present data, for the first time, demonstrated that different extracts from *C. sativum* aerial parts have analgesic activities. These activities were reversed be naloxone therefore; the interaction with opioid system might be suggested. Isolation of the active compound(s) from the extract may yield novel analgesic agent.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

ACKNOWLEDGMENTS

The authors would like to thank the Vice Presidency of Research, Mashhad University of Medical Sciences, for its financial supports.

REFERENCES

- Hosseini M, Taiarani Z, Hadjzadeh MA, Salehabadi S, et al. Different responses of nitric oxide synthase inhibition on morphine-induced antinociception in male and female rats. Pathophysiology. 2011; 18 (2): 143-149.
- Karami R, Hosseini M, Khodabandehloo F, Khatami L, et al. Different effects of L-arginine on morphine tolerance in sham and ovariectomized female mice. J. Zhejiang Univ Sci B. 2011; 12 (12): 1016-1023.
- Hosseini M, Taiarani Z, Karami R, Abad AA. The effect of chronic administration of L-arginine and L-NAME on morphine-induced antinociception in ovariectomized rats. *Indian J. Pharmacol.* 2011; 43 (5): 541-545.
- Raffa RB. Pharmacology of oral combination analgesics: rational therapy for pain. J. Clin. Pharm. Ther. 2001; 26 (4): 257-264.
- Berde CB, Sethna NF. Analgesics for the Treatment of Pain in Children. N. Engl. J. Med. 2002; 347 (14): 1094-1103.
- Kumar V. Potential medicinal plants for CNS disorders: an overview. *Phytother Res.* 2006; 20 (12): 1023-1035.
- Ghorbani A, Rakhshandeh H, Sadeghnia HR. Potentiating effects of Lactuca sativa on pentobarbital-induced sleep. Iran J Pharm Res. 2013;12(2):401-6.
- Asadpour E, Ghorbani A, Sadeghnia HR. Water-soluble compounds of lettuce inhibit DNA damage and lipid peroxidation induced by glucose/serum deprivation in N2a cells. *Acta. Pol. Pharm.* 2014; 71 (3): 409-413.
- Hosseini M, Harandizadeh F, Niazmand S, Soukhtanloo M, et al. The role for nitric oxide on the effects of hydroalcoholic extract of Achillea wilhelmsii on seizure. Avicenna J. Phytomed. 2014; 4 (4): 251-259.
- Hosseini M, Harandizadeh F, Niazamand S, Soukhtanloo M, et al. Antioxidant effect of Achillea wilhelmsii extract on pentylenetetrazole (seizure model)-induced oxidative brain damage in Wistar rats. Indian J. Physiol .Pharmacol. 2013; 57 (4): 418-424.
- Aissaoui A, Zizi S, Israili ZH, Lyoussi B. Hypoglycemic and hypolipidemic effects of Coriandrum sativum L. in Meriones shawi rats. *J. Ethnopharmacol.* 2011; 137 (1): 652-661.

- Emamghoreishi M, Khasaki M, Aazam MF. Coriandrum sativum: evaluation of its anxiolytic effect in the elevated plus-maze. *J. Ethno-pharmacol.* 2005; 96 (3): 365-370.
- Rakhshandeh H, Sadeghnia HR, Ghorbani A. Sleep-prolonging effect of Coriandrum sativum hydro-alcoholic extract in mice. *Nat. Prod. Res.* 2012 Oct 12; 26 (22): 2095-2098.
- Eidi M, Eidi A, Saeidi A, Molanaei S, et al. Effect of coriander seed (Coriandrum sativum L.) ethanol extract on insulin release from pancreatic beta cells in streptozotocin-induced diabetic rats. Phytother Res. 2009 Mar; 23 (3): 404-406.
- Dhanapakiam P, Joseph JM, Ramaswamy VK, Moorthi M, et al. The cholesterol lowering property of coriander seeds (Coriandrum sativum): mechanism of action. J. Environ. Biol. 2008; 29 (1): 53-56.
- Samojlik I, Lakic N, Mimica-Dukic N, Dakovic-Svajcer K, et al. Antioxidant and hepatoprotective potential of essential oils of coriander (Coriandrum sativum L.) and caraway (Carum carvi L.) (Apiaceae). J. Agric Food Chem. 2010; 58 (15): 8848-8853.
- Ghorbani A, Rakhshandeh H, Asadpour E, Sadeghnia HR. Effects of Coriandrum sativum extracts on glucose/serum deprivation-induced neuronal cell death. Avicenna J. Phytomed. 2012; 2 (1): 4-9.
- Zanusso-Junior G, Melo JO, Romero AL, Dantas JA, et al. Evaluation
 of the anti-inflammatory activity of coriander (Coriandrum sativum
 L.) in rodents. Revista Brasileira de Plantas Medicinais. 2011; 13 (1):
 17-23.
- de Almeida Melo E, Bion FM, Guerra NB. *In vivo* antioxidant effect of aqueous and etheric coriander (Coriandrum sativum L.) extracts. *Eur. J. Lipid Sci. Technol.* 2003; 105 (9): 483-487.
- Hashim MS, Lincy S, Remya V, Teena M, et al. Effect of polyphenolic compounds from Coriandrum sativum on H₂O₂-induced oxidative stress in human lymphocytes. Food Chemistry. 2005; 92 (4): 653-660.
- Misharina TA, Samusenko AL. Antioxidant properties of essential oils from lemon, grapefruit, coriander, clove, and their mixtures. *Applied Biochemistry and Microbiology*. 2008; 44 (4): 438-442.
- Sultana S, Ripa FA, Hamid K. Comparative antioxidant activity study of some commonly used spices in Bangladesh. *Pak. J. Biol. Sci.* 2010; 13: 340-343.
- Wangensteen H, Samuelsen AB, Malterud KE. Antioxidant activity in extracts from coriander. Food Chemistry. 2004; 88 (2): 293-297.
- Rakhshandah H, Hosseini M. Potentiation of pentobarbital hypnosis by Rosa damascena in mice. *Indian J. Exp. Biol.* 2006; 44 (11): 910-912.
- Mortazavian SM, Ghorbani A. Antiproliferative effect of viola tricolor on neuroblastoma cells in vitro. Aust. J. Med. Herbalism. 2012; 24 (3): 93-96.
- Mortazavian SM, Ghorbani A, Hesari TG. Effect of hydro-alcoholic extract of *Viola tricolor* and its fractions on proliferation of uterine cervix carcinoma cells. *Iran J. Obst. Gyncol Infertil.* 2012; 15 (22): 9-16.
- Kamkar Asl M, Nazariborun A, Hosseini M. Analgesic effect of the aqueous and ethanolic extracts of clove. *Avicenna J. Phytomed.* 2013; 3 (2): 186-192.
- 28. Hosseini M, Kamkar Asl M, Rakhshandeh H. Analgesic effect of clove essential oil in mice. *Avicenna J. Phytomed.* 2011; 1 (1): 1-6.
- 29. Hajzadeh MR, Rakhshandeh H, Esmaeilzadeh M, Ghorbani A. Analgesic and anti-inflammatory effects of *Portolaca oleracca* extract in mice and rat. *Koomesh.* 2004; 5 (3): 113-120.
- 30. Heidari MR, Aghili M, Soltaninezhad E. Evaluation of anti-inflammatory and analgesic effects of coriandrum sativum extract in mice. *J. Qazvin Univ. Med. Sci.* 2005; 8: 3-8.
- Taherian AA, Vafaei AA, Rashidy-Pour A, Emami-abarghoei M, Miladi-Gorgi H, Jarrahi M and Sadeghi H. Effects of aqueous extract of seed of Coriandrum sativum on acute pain in mice. J. Med. Plants

- 2005; 4: 30-35.
- 32. Kunzemann J, Herrmann K. Isolation and identification of flavon(ol)-O-glycosides in caraway (Carum carvi L.), fennel (Foeniculum vulgare Mill.), anise (Pimpinella anisum L.), and coriander (Coriandrum sativum L.), and of flavon-C-glycosides in anise. I. Phenolics of spices (author's transl). Z Lebensm Unters Forsch. 1977; 164 (3): 194-200.
- Rylski M, Duriasz-Rowinska H, Rewerski W. The analgesic action of some flavonoids in the hot plate test. *Acta. Physiol. Pol.* 1979; 30 (3): 385-388.
- Kaur R, Singh D, Chopra K. Participation of alpha2 receptors in the antinociceptive activity of quercetin. J. Med. Food. 2005; 8 (4): 529-532
- 35. Nassiri-Asl M, Mortazavi SR, Samiee-Rad F, Zangivand AA, *et al.* The effects of rutin on the development of pentylenetetrazole kindling and memory retrieval in rats. *Epilepsy Behav.* 2010; 18 (1-2): 50-53.
- Medina JH, Viola H, Wolfman C, Marder M, et al. Overview--flavonoids: a new family of benzodiazepine receptor ligands. Neurochem Res. 1997; 22 (4): 419-425.
- Griebel G, Perrault G, Tan S, Schoemaker H, et al. Pharmacological studies on synthetic flavonoids: comparison with diazepam. Neuropharmacology. 1999; 38 (7): 965-977.
- Paladini AC, Marder M, Viola H, Wolfman C, et al. Flavonoids and the central nervous system: from forgotten factors to potent anxiolytic compounds. J. Pharm. Pharmacol. 1999; 51 (5): 519-526.
- Youdim KA, Shukitt-Hale B, Joseph JA. Flavonoids and the brain: interactions at the blood-brain barrier and their physiological effects on the central nervous system. *Free Radic. Biol. Med.* 2004; 37 (11): 1683-1693.
- Kang TH, Jeong SJ, Kim NY, Higuchi R, et al. Sedative activity of two flavonol glycosides isolated from the flowers of Albizzia julibrissin Durazz. J. Ethnopharmacol. 2000; 71 (1-2): 321-323.
- Peana AT, D'Aquila PS, Chessa ML, Moretti MD, et al. (-)-Linalool produces antinociception in two experimental models of pain. Eur. J. Pharmacol. 2003; 460 (1): 37-41.
- Peana AT, Rubattu P, Piga GG, Fumagalli S, et al. Involvement of adenosine A1 and A2A receptors in (-)-linalool-induced antinociception. Life Sci. 2006; 78 (21): 2471-2474.
- 43. Usta J, Kreydiyyeh S, Knio K, Barnabe P, et al. Linalool decreases HepG2 viability by inhibiting mitochondrial complexes I and II, increasing reactive oxygen species and decreasing ATP and GSH levels. Chem. Biol. Interact. 2009; 180 (1): 39-46.
- 44. Kubo I, Fujita K, Kubo A, Nihei K, et al. Antibacterial activity of coriander volatile compounds against Salmonella choleraesuis. J. Agric Food Chem. 2004; 52 (11): 3329-3332.
- 45. Lapa Fda R, Gadotti VM, Missau FC, Pizzolatti MG, et al. Antinociceptive properties of the hydroalcoholic extract and the flavonoid rutin obtained from Polygala paniculata L. in mice. Basic Clin. Pharmacol. Toxicol. 2009; 104 (4): 306-315.
- de Campos Buzzi F, Franzoi CL, Antonini G, Fracasso M, et al. Antinociceptive properties of caffeic acid derivatives in mice. Eur. J. Med. Chem. 2009; 44 (11): 4596-4602.
- 47. Santos AR, De Campos RO, Miguel OG, Cechinel-Filho V, et al. The involvement of K+ channels and Gi/o protein in the antinociceptive action of the gallic acid ethyl ester. Eur. J. Pharmacol. 1999; 379 (1): 7.17
- 48. Mehrotra A, Shanbhag R, Chamallamudi MR, Singh VP, *et al.* Ameliorative effect of caffeic acid against inflammatory pain in rodents. *Eur. J. Pharmacol.* 2011; 666 (1-3): 80-86.
- Ishisaka A, Ichikawa S, Sakakibara H, Piskula MK, et al. Accumulation of orally administered quercetin in brain tissue and its antioxidative effects in rats. Free Radic Biol Med. 2011; 51 (7): 1329-1336.