

HHS Public Access

Tuberculosis (Edinb). Author manuscript; available in PMC 2015 April 12.

Published in final edited form as:

Author manuscript

Tuberculosis (Edinb). 2014 March; 94(2): 162–169. doi:10.1016/j.tube.2013.12.001.

Bayesian Models for Screening and TB Mobile for Target Inference with *Mycobacterium tuberculosis*

Sean Ekins^{1,2,*}, Allen C. Casey³, David Roberts³, Tanya Parish³, and Barry A. Bunin¹ ¹Collaborative Drug Discovery, 1633 Bayshore Highway, Suite 342, Burlingame, CA 94010, USA ²Collaborations in Chemistry, 5616 Hilltop Needmore Road, Fuquay-Varina, NC 27526, USA ³Infectious Disease Research Institute, Seattle, WA USA

Abstract

The search for compounds active against *Mycobacterium tuberculosis* is reliant upon high throughput screening (HTS) in whole cells. We have used Bayesian machine learning models which can predict anti-tubercular activity to filter an internal library of over 150,000 compounds prior to *in vitro* testing. We used this to select and test 48 compounds *in vitro*; 11 were active with MIC values ranging from $0.4 \,\mu$ M to $10.2 \,\mu$ M, giving a high hit rate of 22.9%. Among the hits, we identified several compounds belonging to the same series including five quinolones (including ciprofloxacin), three molecules with long aliphatic linkers and three singletons. This approach represents a rapid method to prioritize compounds for testing that can be used alongside medicinal chemistry insight and other filters to identify active molecules. Such models can significantly increase the hit rate of HTS, above the usual 1% or lower rates seen. In addition, the potential targets for the 11 molecules were predicted using TB Mobile and clustering alongside a set of over 740 molecules with known *M. tuberculosis* target annotations. These predictions may serve as a mechanism for prioritizing compounds for further optimization.

Keywords

Bayesian models; Collaborative Drug Discovery Tuberculosis database; function class fingerprints; Virtual Screening; *Mycobacterium tuberculosis*

Supporting Information Available

Conflict of interest statement

^{© 2013} Elsevier Ltd. All rights reserved

^{*}To whom correspondence should be addressed. (Sean Ekins, Collaborations in Chemistry, 5616 Hilltop Needmore Road, Fuquay-Varina, NC 27526, USA, ekinssean@yahoo.com Phone 215-687-1320).

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Supplemental material is available online. The Bayesian models created in Discovery Studio using the previously published data in CDD are available from the authors upon written request.

Sean Ekins is a consultant for Collaborative Drug Discovery Inc. Barry A. Bunin is the Founder and CEO of Collaborative Drug Discovery Inc.

Introduction

The search for drugs to prevent or treat infectious diseases is an urgent research focus both in academia and across the pharmaceutical industry. In recent years there has been an increase in the efforts around high throughput screening (HTS) for *Mycobacterium tuberculosis*, in order to find compounds as therapeutics against tuberculosis (TB) ^{1–6}, A recent review of the state of TB research has summarized the limited pipeline of molecules in various drug discovery/development stages ⁷. Collaborative efforts that coordinate fragmented TB research efforts by individual groups will be critical to improve the chances of success in both identifying new targets and finding new molecules that could target them. Such efforts include the initiatives funded by NIAID, the Bill and Melinda Gates Foundation (BMGF) and the FP7 funded More Medicines For Tuberculosis (MM4TB) project.

The pipeline for TB therapeutics had not produced a new approved drug in over 40 years until the recently FDA-approved Bedaquiline ⁸⁻¹⁰, although there are several candidates in the clinic ^{9, 11}. Only a tiny fraction of TB targets have been addressed with approved drugs or early leads ¹² and recent testing has targeted additional proteins (e.g. MmpL3¹³). The relative lack of success with target-based screening compared with whole cell phenotypic screening is a pattern observed for other antibacterial targets, reflecting the difficulty of target-based high-throughput screening for novel antibiotics ¹⁴. In pharmaceutical companies, computational approaches are widely used to aid in drug discovery, but these have not been as exhaustively applied or validated for TB research. For example, virtual screening of compound libraries is used as a complement to high-throughput screening in *vitro* for many diseases ^{15–21}. Computational approaches applied to TB have been generally used by specialists focused on a single target or series of compounds and rarely in combination with other computational tools $^{22, 23}$. We recently exhaustively reviewed this topic ^{22, 24}, as computational methods are used in workflows by many pharmaceutical company project teams ¹⁸. We found several gaps when we look at how computational methods could be used in TB drug discovery including limited use of filtering for druglikeness or lead-likeness ²⁵, target deconvolution ²⁶, lack of sequential virtual and biochemical screening and lack of in silico ADME/Tox model use²². A clear disconnect was noted between the generation, utilization, dissemination, sharing and reuse of computational models and the entire drug discovery process 22 .

We have proposed using recently retrospectively validated Bayesian machine learning models for *M. tuberculosis* ^{25, 27, 28} for prospective compound evaluation. Three recent studies have also explored the optimization of these models by combining bioactivity and cytotoxicity data ^{29–31} and delivered hit rates in excess of 20%. In the current study we have validated the use of three Bayesian machine learning models by prospectively selecting a small percentage of an in house library for testing. We have identified 11 compounds with *in vitro* activity and predicted their potential targets using ligand-based computational approaches.

Experimental Methods

Chemicals

Compounds were purchased from ChemBridge (San Diego, CA), ChemDiv (San Diego, CA), Maybridge/Thermo Fisher Scientific Inc. (Waltham, MA) and Sigma - Aldrich (St. Louis, Mo).

CDD Database and SRI datasets

The development of the CDD TB database (Collaborative Drug Discovery Inc. Burlingame, CA) has been previously described ²⁵. The Tuberculosis Antimicrobial Acquisition and Coordinating Facility (TAACF) and Molecular Libraries Small Molecule Repository (MLSMR) screening datasets ^{2–4} were collected and uploaded in CDD TB from sdf files and mapped to custom protocols ³². All of the public *M. tuberculosis* datasets are available for free public read-only access and mining upon registration, making them a valuable molecule resource for researchers along with available contextual data on these samples from other non *M. tuberculosis* assays. These datasets are also publically available in PubChem ³³. The IDRI database and screening data used in modeling is proprietary.

Machine learning models for *M. tuberculosis*

We have previously described the generation and validation of Laplacian-corrected Bayesian classifier models ^{25, 27, 28} developed with single point screening and dose response data. In this study we have generated Laplacian-corrected Bayesian classifier models using Discovery Studio 2.5.5 34-38 Molecular function class fingerprints of maximum diameter 6 (FCFP 6) ³⁹, AlogP, molecular weight, number of rotatable bonds, number of rings, number of aromatic rings, number of hydrogen bond acceptors, number of hydrogen bond donors, and molecular fractional polar surface area were calculated from input sdf files using the "calculate molecular properties" protocol to distinguish between compounds that are active against M. tuberculosis and those that are inactive in this study. A Bayesian classifier model with the molecular descriptors described above was built using the "create Bayesian model" protocol and IDRI % inhibition at 20 µM for 1106 samples (308 active with >90% inhibition)⁴⁰. Each model was validated using leave-one-out cross-validation. Each sample was left out one at a time, and a model built using the results of the samples, and that model used to predict the left-out sample. Once all the samples had predictions, a receiver operator curve (ROC) plot was generated, and the cross validated (XV) ROC area under the curve (AUC) calculated (Table 1). All models generated were additionally evaluated by leaving out 50% of the data and rebuilding the model 100 times using a custom protocol for validation, in order to generate the XV ROC and AUC (Table 1). These models were also used for screening the "Infectious Disease Research Institute (IDRI) library" of 156,719 compounds with *M. tuberculosis* activity.

M. tuberculosis assays for biological activity

Molecules were screened at a single concentration of 20 μ M in Middlebrook 7H9 medium plus 10% v/v OADC (oleic acid, albumen, dextrose, catalase) and 0.05 % w/v Tween 80; actives were classified as having 90% inhibition of growth of *M. tuberculosis* H37Rv after

5 d ⁴⁰. MICs were determined in liquid medium ⁴¹; briefly a 10 point serial dilution of compounds was run and % growth of *M. tuberculosis* determined after 5 days incubation ⁴¹. Curves were generated using the Gompertz fit and MICs determined as minimal concentration required to inhibit growth completely.

Target prediction for IDRI compounds

Over 700 compounds with known *M. tuberculosis* targets were collated from the literature ⁴² and made available in the mobile application TB Mobile (Collaborative Drug Discovery Inc. Burlingame, CA) which is freely available for iOS and Android platforms ^{12, 43}. This dataset was recently updated to 745 compounds and covers over 70 targets. Molecules representing hits from screening in this study were input as queries in TB Mobile and the similarity of all molecules calculated in the application. The top most structurally similar compounds were used to infer *M. tuberculosis* targets. In most cases multiple targets are shown were the top 2–3 molecules had different targets. The 745 compounds with known *M. tuberculosis* targets and the hit compounds from this study were used to generate a Principal Component Analysis (PCA) using the interpretable descriptors used for machine learning model building previously in Discovery Studio (AlogP, molecular weight, number of rotatable bonds, number of rings, number of aromatic rings, number of hydrogen bond acceptors, number of hydrogen bond donors, and molecular fractional polar surface area). 1200 *M. tuberculosis* screening hits (actives and non-toxic only from the SRI screens ^{29–31}) were used to show how they covered the target-chemistry PCA space alongside the 745 compounds.

The 745 compounds with known *M. tuberculosis* targets and the hit compounds from this study were also clustered (100 clusters) using MDL fingerprints in Discovery Studio, and the position of the screening hits in specific clusters identified along with the targets of the other molecules in these clusters. In cases when a hit was a singleton, the identity of targets for clusters around a hit was noted. This clustering approach can also be used to infer targets alongside TB Mobile.

Results

IDRI – Bayesian model

A Bayesian model was generated with whole cell *M. tuberculosis* data for 1106 previously described TAACF and MLSMR actives and inactives [34]. The leave one out ROC was 0.82 and this decreased slightly (0.77) with internal validation with leave out $50\% \times 100$ (Table 2). The concordance (73.4%), specificity (77.3%) and selectivity (63.4%) were in line with the other models described previously (Table 1) ^{25, 27, 29, 30}. Using the FCFP-6 descriptors we can identify those substructure descriptors that contribute to the *M. tuberculosis* activity in the training set including imidazole, benzothiazole and quinolone, (Figure 1) and those that are not present in active compounds including acetamide, thioether, pyrrole, phenylether and piperazine (Figure 2).

IDRI - Prospective testing of the Bayesian Models

The previously published MLSMR dose response model ²⁵, MLSMR dose response and cytotoxicity model ²⁹ and the IDRI Bayesian model were used to rank the "IDRI library" of

156,719 compounds for *M. tuberculosis* activity. This library can be considered leadlike based on the mean Molecular weight (344.5), log P (3.3), hydrogen bond donors (1.0), hydrogen bond acceptors (3.6) and other properties (Supplemental Figure 1). After ranking the library with the Bayesian score derived from each Bayesian model, the top 1000 compounds were selected and analyzed. There was minimal overlap between all three models and compounds in the top scoring 1000 (Figure 3). The MLSMR models overlapped to the greatest degree (over 20% of the top 1000). Forty eight compounds were selected from these ranked lists and tested *in vitro*; 11 of these were classed as hits (22.9% hit rate) as they possessed anti-tubercular activity with MIC <10 μ M (Table 2). To illustrate the diversity of hits (Table 2), this included five quinolones including ciprofloxacin, three azole containing molecules with long aliphatic linkers and three singletons. Six compounds were found with the MLSMR dose response model, four were found with the IDRI model and one with the MLSMR dose response and cytotoxicity model. The Tanimoto similarity of the 11 compounds were compared to all the publically accessible TB related datasets and these ranged from 64–100%.

Target prediction for IDRI compounds

The PCA model of compounds with annotated *M. tuberculosis* targets represents the targetchemistry space and 88.7% of the variance is explained by the 3 principal components. The 11 hit compounds from this study were also added to this set and show they are clustered in a relatively narrow region (Figure 4A). Similarly, the hits from previous SRI screens only partially cover the target space (Figure 4B). Clustering the 11 hits with the 745 compounds with annotated target information enabled complementary target predictions with those based on molecular similarity performed with TB mobile (Table 2, Supplemental Figure 2). The known gyrase inhibitor class, fluoroquinolones were well predicted by both target inference methods. The remaining compounds had divergent predictions apart from the azoles, which were predicted to be InhA inhibitors.

Discussion

Drug discovery is time consuming and very costly ^{44, 45} such that any tools that can point out liabilities earlier will have considerable value ^{21, 46, 47}. The need for new anti-tubercular therapies is unquestioned in the face of drug resistance that has progressed to the point of the identification of totally drug resistant strains in India ⁴⁸ and a call for re-opening the TB sanatoria that were closed more than 60 years ago ⁴⁹. To address the challenge of drug resistance in TB infection, many groups have turned to HTS campaigns with chemically diverse libraries of small molecules to identify novel starting points for drug discovery ^{1, 50}. The TB community must now ask how to *mine efficiently and leverage* this growing database to provide new drug candidates, in the face of well known complications such as latency and persistence ⁵¹ and the numerous issues associated with typical HTS data ⁵². To help answer this question, we have identified a significant opportunity for the tuberculosis drug discovery community to harness pharmaceutical industry-tested computational methodologies ²². Subsequently, we turned in part to the cheminformatics methods which occupy an important place in the industrial drug discovery workflow. Ligand- and proteinbased methods, for example, have been used as a complement to high-throughput screening

in vitro ¹⁵. In order to validate the predictions from such methods we are required to test molecules for their whole cell TB activity.

We have developed and utilized machine learning models for *M. tuberculosis* ^{25, 27, 28} using large publically accessible HTS data sets ^{3, 4}. During retrospective validation of these models we observed at least 4-10 fold enrichment in identifying TB actives in the top scoring molecules ²⁸. These results indicated that using whole cell screening data from one laboratory for computational models can be used to benefit other laboratories via predictions of their compounds of interest and narrowing down the number of compounds to be tested in vitro²⁸. We have recently updated our approach to incorporate cytotoxicity data into the models $^{29-31}$. These previously published Bayesian models had considerably higher hit rates than random HTS screening ^{29, 30}. One study virtually screened over 82,000 molecules, 550 were tested *in vitro* and 124 actives were identified in total (22.5% hit rate)³⁰. A second study virtually screened over 38,000 molecules, tested 106 in vitro and identified 17 actives (22.5% hit rate)²⁹. In the current study we utilized several previously published models as well as a newly constructed model generated with new data from >1000 previously published active compounds. Three Bayesian models were ultimately used to screen the in house library of 156,719 molecules and 48 (0.03%) of the compounds were tested in vitro resulting in 11 hits. Again this confirmed the hit rates previously observed with a value of 22.9%. In our experience and as a point of contrast, the whole cell HTS hit rate for the IDRI group has varied from 0.6 - 2% depending on the assay (unpublished).

Using several Bayesian models for *M. tuberculosis* activity to prioritize compounds from a screening library of this size is by far the largest such analysis we have performed to date to our knowledge ^{29–31}. The results obtained further validate the hypothesis that Bayesian models ^{25, 27–31, 42} identify subsets of compound libraries enriched with active compounds, therefore requiring the testing of far fewer compounds. Future research will involve investigating open source descriptors and algorithms that can enable deploying such models more widely ^{53, 54}. This Bayesian modeling and virtual screening approach is also applicable to other neglected diseases.

This study also further utilized a recently developed mobile application for inferring potential *M. tuberculosis* targets for the 11 hits (Table 2, Supplemental Figure 2). This application draws together known small molecules and their annotated targets as well as other information relevant to the pathway targeted, essentiality, human ortholog etc. ^{12, 42}. Generating predictions with this application was also complemented by using clustering of the known compounds with targets and assessing which clusters the 11 compounds were in. The fluoroquinolone compounds are not surprisingly predicted as gyrase inhibitors using both target inference methods, apart from IDR-0173634 which is also predicted as a potential inhibitor of InhA. Although azoles are well known cytochrome P450 inhibitors ^{55, 56} they are predominantly predicted targets with no concordance with the target inference methods would be worthy of testing *in vitro*. Our analysis of the 11 hits suggest, as one would expect, that they are covering a very narrow section of chemistry and target space. In particular we have multiple fluoroquinolones and azoles (Table 2) so these may essentially count as a single data point in each case. Our approach (using known compounds

with *Mtb* targets) to infer potential targets for similar compounds is more conservative than methods which would use similarity to compounds known to be active against targets in other organisms. Such target prediction efforts would help us to prioritize targets to test.

In conclusion we have presented an approach using multiple Bayesian models to prioritize compounds for testing which identified active compounds. These in turn were used with TB Mobile 12 and clustering as mechanisms for predicting potential targets for compounds in *M. tuberculosis*, thereby serving as an approach for further identifying the best compounds for optimization. Such computational workflows leveraging prior knowledge further our aim of optimally using and integrating the data and resources available to us in order to accelerate drug discovery for *M. tuberculosis*²².

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgement

S.E. acknowledges colleagues at CDD for developing the software. Dr. Alex Clark and Dr. Malabika Sarker are acknowledged for assistance with TB Mobile. Accelrys are kindly acknowledged for providing Discovery Studio. S.E. acknowledges Dr. Joel Freundlich and Dr. Robert Reynolds for numerous discussions on TB and Bayesian models. We thank Torey Alling, Mai Ann Bailey and Juliane Ollinger for running the MICs at IDRI, Susantha Chandrasekera, Edward Kesicki and Joshua Odingo for assistance with compound structural information and Alfredo Blakeley for technical assistance with compound handling.

Funding

The CDD TB has been developed thanks to funding from the Bill and Melinda Gates Foundation (Grant#49852 "Collaborative drug discovery for TB through a novel database of SAR data optimized to promote data archiving and sharing"). The project described was supported by Award Number R43 LM011152-01 "Biocomputation across distributed private datasets to enhance drug discovery" from the National Library of Medicine. TB Mobile was developed with funding from Award Number 2R42AI088893-02 "Identification of novel therapeutics for tuberculosis combining cheminformatics, diverse databases and logic based pathway analysis" from the National Institutes of Allergy and Infectious Diseases.

R.C.R. and S. G. F. acknowledge the American Reinvestment and Recovery Act Grant 1RC1AI086677-01 (National Institutes of Health (NIH), National Institute of Allergy and Infectious Diseases (NIAID)) – "Targeting MDR-TB." The work at IDRI was funded in part by Eli Lilly and Company in support of the mission of the Lilly TB Drug Discovery Initiative and with Grant #42844 from the Bill and Melinda Gates Foundation.

References

- 1. Ballel L, Field RA, Duncan K, Young RJ. New small-molecule synthetic antimycobacterials. Antimicrobial agents and chemotherapy. 2005; 49:2153–2163. [PubMed: 15917508]
- Reynolds, RC.; Ananthan, S.; Faaleolea, E.; Hobrath, JV.; Kwong, CD.; Maddox, C.; Rasmussen, L.; Sosa, MI.; Thammasuvimol, E.; White, EL.; Zhang, W.; Secrist, JA, 3rd. Tuberculosis. Scotland: Edinburgh; 2011. High throughput screening of a library based on kinase inhibitor scaffolds against Mycobacterium tuberculosis H37Rv.
- Maddry, JA.; Ananthan, S.; Goldman, RC.; Hobrath, JV.; Kwong, CD.; Maddox, C.; Rasmussen, L.; Reynolds, RC.; Secrist, JA., 3rd; Sosa, MI.; White, EL.; Zhang, W. Tuberculosis. Vol. 89. Scotland: Edinburgh; 2009. Antituberculosis activity of the molecular libraries screening center network library; p. 354-363.
- Ananthan, S.; Faaleolea, ER.; Goldman, RC.; Hobrath, JV.; Kwong, CD.; Laughon, BE.; Maddry, JA.; Mehta, A.; Rasmussen, L.; Reynolds, RC.; Secrist, JA., 3rd; Shindo, N.; Showe, DN.; Sosa, MI.; Suling, WJ.; White, EL. Tuberculosis. Vol. 89. Scotland: Edinburgh; 2009. High-throughput screening for inhibitors of Mycobacterium tuberculosis H37Rv; p. 334-353.

- 5. Mak PA, Rao SP, Ping Tan M, Lin X, Chyba J, Tay J, Ng SH, Tan BH, Cherian J, Duraiswamy J, Bifani P, Lim V, Lee BH, Ling Ma N, Beer D, Thayalan P, Kuhen K, Chatterjee A, Supek F, Glynne R, Zheng J, Boshoff HI, Barry CE 3rd, Dick T, Pethe K, Camacho LR. A High-Throughput Screen To Identify Inhibitors of ATP Homeostasis in Non-replicating Mycobacterium tuberculosis. ACS Chem Biol. 2012; 7:1190–1197. [PubMed: 22500615]
- Stanley SA, Grant SS, Kawate T, Iwase N, Shimizu M, Wivagg C, Silvis M, Kazyanskaya E, Aquadro J, Golas A, Fitzgerald M, Dai H, Zhang L, Hung DT. Identification of Novel Inhibitors of M. tuberculosis Growth Using Whole Cell Based High-Throughput Screening. ACS Chem Biol. 2012; 7:1377–1384. [PubMed: 22577943]
- 7. Thayer A, Taking down TB. Chem Eng News. 2007 Sep 24.:21–32.
- Voelker R. MDR-TB has new drug foe after fast-track approval. Jama. 2013; 309:430. [PubMed: 23385248]
- 9. Koul A, Arnoult E, Lounis N, Guillemont J, Andries K. The challenge of new drug discovery for tuberculosis. Nature. 2011; 469:483–490. [PubMed: 21270886]
- Andries, K.; Verhasselt, P.; Guillemont, J.; Gohlmann, HW.; Neefs, JM.; Winkler, H.; Van Gestel, J.; Timmerman, P.; Zhu, M.; Lee, E.; Williams, P.; de Chaffoy, D.; Huitric, E.; Hoffner, S.; Cambau, E.; Truffot-Pernot, C.; Lounis, N.; Jarlier, V. Science. Vol. 307. New York, NY: 2005. A diarylquinoline drug active on the ATP synthase of Mycobacterium tuberculosis; p. 223-227.
- Kaneko T, Cooper C, Mdluli K. Challenges and opportunities in developing novel drugs for TB. Future Med Chem. 2011; 3:1373–1400. [PubMed: 21879843]
- Ekins S, Clark AM, Sarker M. TB Mobile: A Mobile App for Anti-tuberculosis Molecules with Known Targets. J Cheminform. 2013; 5:13. [PubMed: 23497706]
- Tahlan K, Wilson R, Kastrinsky DB, Arora K, Nair V, Fischer E, Barnes SW, Walker JR, Alland D, Barry CE 3rd, Boshoff HI. SQ109 targets MmpL3, a membrane transporter of trehalose monomycolate involved in mycolic acid donation to the cell wall core of Mycobacterium tuberculosis. Antimicrobial agents and chemotherapy. 2012; 56:1797–1809. [PubMed: 22252828]
- 14. Payne DA, Gwynn MN, Holmes DJ, Pompliano DL. Drugs for bad bugs: confronting the challenges of antibacterial discovery. Nat Rev Drug Disc. 2007; 6:29–40.
- 15. Schneider G. Virtual screening: an endless staircase? Nature reviews. 2010; 9:273–276.
- Zhang L, Fourches D, Sedykh A, Zhu H, Golbraikh A, Ekins S, Clark J, Connelly MC, Sigal M, Hodges D, Guiguemde A, Guy RK, Tropsha A. Discovery of Novel Antimalarial Compounds Enabled by QSAR-Based Virtual Screening. Journal of chemical information and modeling. 2013; 53:475–492. [PubMed: 23252936]
- Scior T, Bender A, Tresadern G, Medina-Franco JL, Martinez-Mayorga K, Langer T, Cuanalo-Contreras K, Agrafiotis DK. Recognizing Pitfalls in Virtual Screening: A Critical Review. Journal of chemical information and modeling. 2012
- Duffy BC, Zhu L, Decornez H, Kitchen DB. Early phase drug discovery: cheminformatics and computational techniques in identifying lead series. Bioorganic & medicinal chemistry. 2012; 20:5324–5342. [PubMed: 22938785]
- Krueger BA, Weil T, Schneider G. Comparative virtual screening and novelty detection for NMDA-GlycineB antagonists. Journal of computer-aided molecular design. 2009; 23:869–881. [PubMed: 19890609]
- Noeske T, Jirgensons A, Starchenkovs I, Renner S, Jaunzeme I, Trifanova D, Hechenberger M, Bauer T, Kauss V, Parsons CG, Schneider G, Weil T. Virtual Screening for Selective Allosteric mGluR1 Antagonists and Structure-Activity Relationship Investigations for Coumarine Derivatives. ChemMedChem. 2007; 2:1763–1773. [PubMed: 17868161]
- 21. Ekins S, Mestres J, Testa B. In silico pharmacology for drug discovery: methods for virtual ligand screening and profiling. Br J Pharmacol. 2007; 152:9–20. [PubMed: 17549047]
- Ekins S, Freundlich JS, Choi I, Sarker M, Talcott C. Computational Databases, Pathway and Cheminformatics Tools for Tuberculosis Drug Discovery. Trends in microbiology. 2011; 19:65– 74. [PubMed: 21129975]
- 23. Ballester PJ, Mangold M, Howard NI, Robinson RL, Abell C, Blumberger J, Mitchell JB. Hierarchical virtual screening for the discovery of new molecular scaffolds in antibacterial hit identification. J R Soc Interface. 2012; 9:3196–3207. [PubMed: 22933186]

- 24. Ekins, S.; Freundlich, JS. Methods in molecular biology. Vol. 993. Clifton, NJ: 2013. Computational models for tuberculosis drug discovery; p. 245-262.
- 25. Ekins S, Bradford J, Dole K, Spektor A, Gregory K, Blondeau D, Hohman M, Bunin B. A Collaborative Database And Computational Models For Tuberculosis Drug Discovery. Mol BioSystems. 2010; 6:840–851.
- Prathipati P, Ma NL, Manjunatha UH, Bender A. Fishing the target of antitubercular compounds: in silico target deconvolution model development and validation. Journal of proteome research. 2009; 8:2788–2798. [PubMed: 19301903]
- 27. Ekins S, Kaneko T, Lipinksi CA, Bradford J, Dole K, Spektor A, Gregory K, Blondeau D, Ernst S, Yang J, Goncharoff N, Hohman M, Bunin B. Analysis and hit filtering of a very large library of compounds screened against Mycobacterium tuberculosis. Molecular bioSystems. 2010; 6:2316– 2324. [PubMed: 20835433]
- 28. Ekins S, Freundlich JS. Validating new tuberculosis computational models with public whole cell screening aerobic activity datasets. Pharm Res. 2011; 28:1859–1869. [PubMed: 21547522]
- Ekins S, Reynolds R, Kim H, Koo M-S, Ekonomidis M, Talaue M, Paget SD, Woolhiser LK, Lenaerts AJ, Bunin BA, Connell N, Freundlich JS. Bayesian Models Leveraging Bioactivity and Cytotoxicity Information for Drug Discovery. Chem Biol. 2013; 20:370–378. [PubMed: 23521795]
- 30. Ekins S, Reynolds RC, Franzblau SG, Wan B, Freundlich JS, Bunin BA. Enhancing Hit Identification in Mycobacterium tuberculosis Drug Discovery Using Validated Dual-Event Bayesian Models. PLOSONE. 2013; 8:e63240.
- Ekins S, Freundlich JS, Hobrath JV, White EL, Reynolds RC. Combining Computational Methods for Hit to Lead Optimization in Mycobacterium tuberculosis Drug Discovery. Pharm Res. 2013 In Press.
- 32. Anon. Collaborative Drug Discovery, Inc. http://www.collaborativedrug.com/register
- 33. Anon. The PubChem Database. http://pubchem.ncbi.nlm.nih.gov/
- Prathipati P, Ma NL, Keller TH. Global Bayesian models for the prioritization of antitubercular agents. Journal of chemical information and modeling. 2008; 48:2362–2370. [PubMed: 19053518]
- 35. Bender A, Scheiber J, Glick M, Davies JW, Azzaoui K, Hamon J, Urban L, Whitebread S, Jenkins JL. Analysis of Pharmacology Data and the Prediction of Adverse Drug Reactions and Off-Target Effects from Chemical Structure. ChemMedChem. 2007; 2:861–873. [PubMed: 17477341]
- Klon AE, Lowrie JF, Diller DJ. Improved naive Bayesian modeling of numerical data for absorption, distribution, metabolism and excretion (ADME) property prediction. Journal of chemical information and modeling. 2006; 46:1945–1956. [PubMed: 16995725]
- Hassan M, Brown RD, Varma-O'brien S, Rogers D. Cheminformatics analysis and learning in a data pipelining environment. Mol Divers. 2006; 10:283–299. [PubMed: 17031533]
- Rogers D, Brown RD, Hahn M. Using extended-connectivity fingerprints with Laplacian-modified Bayesian analysis in high-throughput screening follow-up. J Biomol Screen. 2005; 10:682–686. [PubMed: 16170046]
- Jones DR, Ekins S, Li L, Hall SD. Computational approaches that predict metabolic intermediate complex formation with CYP3A4 (+b5). Drug Metab Dispos. 2007; 35:1466–1475. [PubMed: 17537872]
- 40. Roberts D, Ollinger J, Bailey M, Casey A, Parish T. Development of a whole cell high-throughput screen to identify inhibitors of Mycobacterium tuberculosis. Unpublished. 2012
- 41. Ollinger J, Bailey MA, Moraski GC, Casey A, Florio S, Alling T, Miller MJ, Parish T. A dual read-out assay to evaluate the potency of compounds active against Mycobacterium tuberculosis. PloS one. 2013; 8:e60531. [PubMed: 23593234]
- 42. Sarker M, Talcott C, Madrid P, Chopra S, Bunin BA, Lamichhane G, Freundlich JS, Ekins S. Combining cheminformatics methods and pathway analysis to identify molecules with whole-cell activity against Mycobacterium tuberculosis. Pharm Res. 2012; 29:2115–2127. [PubMed: 22477069]
- 43. TB Mobile. https://itunes.apple.com/us/app/tb-mobile/id567461644?mt=8

- 44. Paul SM, Mytelka DS, Dunwiddie CT, Persinger CC, Munos BH, Lindborg SR, Schacht AL. How to improve R&D productivity: the pharmaceutical industry's grand challenge. Nature reviews. 2010; 9:203–214.
- 45. Munos B. Lessons from 60 years of pharmaceutical innovation. Nature reviews. 2009; 8:959-968.
- 46. Ekins S, Waller CL, Swaan PW, Cruciani G, Wrighton SA, Wikel JH. Progress in predicting human ADME parameters in silico. J Pharmacol Toxicol Methods. 2000; 44:251–272. [PubMed: 11274894]
- 47. Ekins S, Mestres J, Testa B. In silico pharmacology for drug discovery: applications to targets and beyond. Br J Pharmacol. 2007; 152:21–37. [PubMed: 17549046]
- Udwadia ZF, Amale RA, Ajbani KK, Rodrigues C. Totally drug-resistant tuberculosis in India. Clin Infect Dis. 2012; 54:579–581. [PubMed: 22190562]
- 49. Dheda K, Migliori GB. The global rise of extensively drug-resistant tuberculosis: is the time to bring back sanatoria now overdue? Lancet. 2012; 379:773–775. [PubMed: 22033020]
- Sacchettini JC, Rubin EJ, Freundlich JS. Drugs versus bugs: in pursuit of the persistent predator Mycobacterium tuberculosis. Nat Rev Microbiol. 2008; 6:41–52. [PubMed: 18079742]
- Zhang Y. The magic bullets and tuberculosis drug targets. Annu Rev Pharmacol Toxicol. 2005; 45:529–564. [PubMed: 15822188]
- 52. Malo N, Hanley JA, Cerquozzi S, Pelletier J, Nadon R. Statistical practice in high-throughput screening data analysis. Nature biotechnology. 2006; 24:167–175.
- 53. Gupta RR, Gifford EM, Liston T, Waller CL, Bunin B, Ekins S. Using open source computational tools for predicting human metabolic stability and additional ADME/TOX properties. Drug metabolism and disposition: the biological fate of chemicals. 2010; 38:2083–2090. [PubMed: 20693417]
- 54. Ekins S, Gupta RR, Gifford E, Bunin BA, Waller CL. Chemical space: missing pieces in cheminformatics. Pharm Res. 2010; 27:2035–2039. [PubMed: 20683645]
- 55. Rupp B, Raub S, Marian C, Holtje HD. Molecular design of two sterol 14alpha-demethylase homology models and their interactions with the azole antifungals ketoconazole and bifonazole. Journal of computer-aided molecular design. 2005; 19:149–163. [PubMed: 16059669]
- Hitchcock CA, Dickinson K, Brown SB, Evans EG, Adams DJ. Interaction of azole antifungal antibiotics with cytochrome P-450-dependent 14 alpha-sterol demethylase purified from Candida albicans. Biochem J. 1990; 266

Substructure	Good	Bad	Bayesian Score	0	10	20	30	40	50
G1: 1303769626	13	0	1.139						
G2: 679321016	17	1	1.129			-			
G3: 636230097	12	0	1.127						
G4: 5470945	12	0	1.127						
G5: 1461049343	32	6	1.083						
G6: 800166649	15	2	1.059			r			
G7: 878342080	15	2	1.059						
G8: 1119859782	35	8	1.057						
G9: 862165829	34	8	1.05						
G10: 427038716	34	8	1.05						
G11: 182940121	11	1	1.047						
G12: 1427303459	14	2	1.043						
G13: 246195966	7	0	1.024						
G14: 1748509072	15	3	1.011						
G15: 852486437	9	1	1						
G16: 1456819197	14	3	0.994			• · ·			
G17: 1096308665	6	0	0.988						
G18: 1570979257	16	4	0.984						
G19: 2011863436	16	4	0.984						
G20: 1715199747	16	4	0.984						
				1	1		1	1	
Sum	343	57							

Figure 1.

Good features identified in the IDRI Bayesian Model

Substructure	Good	Bad	Bayesian Score	0	10	20	30	40	50	60	i
B1: 61357702	0	32	-2.258								
B2: 146973761	0	26	-2.075	_							
B3: 2056203474	0	23	-1.968	_							
B4: 1972798083	0	22	-1.93	_							
B5: 950918369	0	21	-1.89	_							
B6: 585722152	0	16	-1.665								
B7: 641200682	0	16	-1.665								
B8: 356782498	0	15	-1.613								
B9: 1922787335	0	14	-1.558								
B10: 511917765	0	13	-1.5								
B11: 262463794	0	13	-1.5								
B12: 194716698	3	59	-1.482								
B13: 140430755	2	42	-1.449								
B14: 625525848	0	12	-1.438	_							
B15: 120714312	0	12	-1.438								
B16: 447966750	0	12	-1.438	_							
B17: 178882354	0	12	-1.438								
B18: 205448520	0	12	-1.438	_							
B19: 452555611	0	11	-1.372	_							
B20: 303882603	0	11	-1.372								
Sum	5	394									

Figure 2.

Bad features identified in the IDRI Bayesian Model

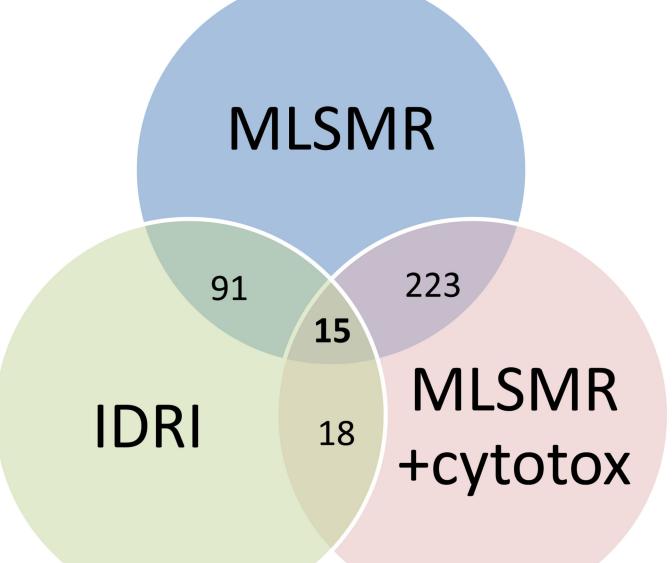
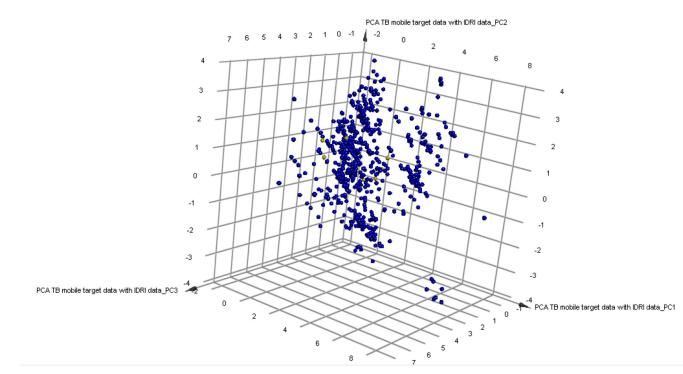


Figure 3.

Venn diagram showing the overlap of IDRI library compounds selected with the MLSMR dose response model, the MLSMR dose response and cytotoxicity model and the IDRI model for *M. tuberculosis* whole cell activity.

Ekins et al.





Β.

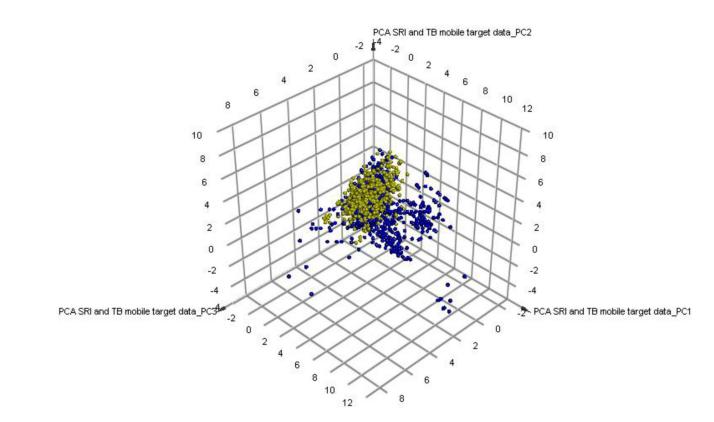


Figure 4.

Principal Component Analysis of 745 compounds with A. known *M. tuberculosis* targets (Blue) from TB Mobile and 11 screening hits (yellow) and B. 1200 active and non toxic compounds from SRI screens (yellow)

Table 1

Mean (SD) leave one out and leave out $50\% \times 100$ cross validation of *M. tuberculosis* Bayesian models (ROC = receiver operator characteristic)

Ekins et al.

Datasets (number of molecules)	Leave one out ROC	Leave out 50% × 100 External ROC Score	Leave out 50% × 100Internal ROC Score	Leave out 50% × 100 Concordance	Leave out 50% × 100 Specificity	Leave out 50% $\times 100$ Sensitivity
IDRI % inhibition at 20uM (1106)	0.82	0.77 ± 0.02	0.78 ± 0.02	73.4 ± 2.98	77.3 ± 6.28	63.4 ± 7.62

Author Manuscript

.

Author Manuscript

Author Manuscript

Table 2

Hits from IDRI library selected with Bayesian models. Promiscuity is calculated from activity of compound in PubChem as this represents a very large database of compounds and bioassays. Tanimoto similarity is reported when compared to the CDD database which contains over 300,000 compounds screened in vitro against Mtb. Target prediction was performed with TB Mobile and clustering as described in the Materials and Methods.

Prediction with Clustering	GyrA (Rv0006) cluster 53	GyrA (Rv0006) GyrB (Rv0005) murD (Rv2155c) cluster 84	Singleton in cluster 89 Dxs1 (Rv2682c) cluster 88 MbyA (Rv2384) cluster 90	Molecule a singleton in cluster 72 RpJJ (Rv0651) TJyA (Rv1694) cluster 71 Alr (Rv3423c) Cluster 73 Cluster 73	RpoB (R v0667) cluster 95
Prediction with TB Mobile	InhA (Rv 1484)	GyrA (Rv0006)	FolP1 (Rv3608c) FolP2 (Rv1207) Rv1885c	PtpA (Rv2234)	ThiL (Rv2977c)
Highest Tanimoto similarity in CDD public TB datasets	100%*	80%	%68	100%*	64%
NIH PubChem Notes	No Data	Ciprofloxacin - active in 6698 out of 8773 bioaasays including against M tuberculosis	Tested in 1 bioassay - inactive	No Data	No Data
Promiscuity	0	0.76	0	0	0
МІС ₉₀ (µМ)	2.2	0.4	2.4	6.3	6.6
Bayesian Model & Score	MLSMR 17.91	MLSMR 27.39	IDRI 20.79	MLSMR+ cytotox 38.11	MLSMR 21.71
Structure	······································	HIN A HO		° ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	
Compound	IDR- 0173634	IDR- 0229683*	IDR- 0198303	IDR- 0204586	IDR- 0218592

a 50	
Prediction with Clustering	GyrA (Rv0006) GyrB (Rv0005) MurD (Rv2155c) cluster 84
Prediction with TB Mobile	GyrA (Rv0006) GyrB (Rv0005)
Highest Tanimoto similarity in CDD public TB datasets	100%
NIH PubChem Notes	Perfloxacin - active in 185 out of 233 bioassays including <i>Mycobacteriuim</i> <i>leprae</i>
Promiscuity	1993991
MIC ₉₀ (µM)	8.7
Bayesian Model & Score	MLSMR 22.48
Structure	CH ₃
	웃

* not cytotoxic based on Vero cell toxicity data in CDD.