

● BRIEF REPORTS ●

Lewis blood genotypes of peptic ulcer and gastric cancer patients in Taiwan

Chi-Jung Yei, Jan-Gowth Chang, Mu-Chin Shih, Sheng-Fung Lin, Chao-Sung Chang, Fu-Tsong Ko, Kuang-Yang Lin, Ta-Chih Liu

Chi-Jung Yei, Blood Bank, Kaohsiung Medical University Hospital, Kaohsiung, Taiwan, China

Jan-Gowth Chang, Mu-Chin Shih, Department of Laboratory Medicine, China Medical University and Hospital, Taichung, Taiwan, China Sheng-Fung Lin, Chao-Sung Chang, Ta-Chih Liu, Division of Hematology/Oncology, Department of Internal Medicine, Kaohsiung Medical University Hospital, Kaohsiung, Taiwan, China

Fu-Tsong Ko, Kuang-Yang Lin, Department of Internal Medicine, Taipei Municipal Jen-Ai Hospital, Taipei, Taiwan, China

Correspondence to: Ta-Chih Liu, MD, Division of Hematology/ Oncology, Department of Internal Medicine, Kaohsiung Medical University Hospital, 100 Shih-Chuan 1st Road, Kaohsiung, Taiwan, China. d730093@cc.kmu.edu.tw

Telephone: +886-7-3121101-6113 Fax: +886-7-3162429 Received: 2005-01-04 Accepted: 2005-01-26

Abstract

AIM: The Lewis b (Le^b) antigen has been implicated as a possible binding site for attachment of *Helicobacter pylori* (*H pylori*) to gastric mucosa. However, studies both supporting and denying this association have been reported in the literature. Differences in secretor (Se) genotype have been suggested as a possible reason for previous discrepancies. Therefore, we investigated the relationship between Le and Se genotypes and *H pylori* infection rates in people with peptic ulcer or gastric cancer.

METHODS: Peripheral blood samples were obtained from 347 patients with endoscopic evidence of peptic ulcer disease (235 cases of duodenal ulcer, 62 of gastric ulcer, and 50 of combined duodenal ulcer/gastric ulcer) and 51 patients with gastric cancer on endoscopy. Peripheral blood specimens from 101 unrelated normal volunteers were used as controls. Lewis phenotype was determined using an antibody method, whereas Le and Se genotypes were determined by DNA amplification and restriction enzyme analysis. Gastric or duodenal biopsies taken from patients with endoscopic evidence of peptic ulcer or gastric cancer were cultured for *H pylori*. Isolates were identified as *H pylori* by morphology and production of urease and catalase. The *H pylori* infection status was also evaluated by rapid urease test (CLO test), and urea breath test (¹³C-UBT). Results of studies were analyzed by chi-square test (taken as significant).

RESULTS: *H pylori* was isolated from 83.7% (303/347) of patients with peptic ulcer disease. Statistical analysis did not show any significant difference in Lewis phenotype or genotype between patients with and without *H pylori* infection. No significant association was found between Lewis genotype and peptic ulcer or gastric cancer.

CONCLUSION: Lewis blood genotype or phenotype may not play a role in the pathogenesis of *H pylori* infection. However, bacterial strain differences and the presence of more than one attachment mechanism may limit the value of epidemiological studies in elucidating this matter.

 $\ensuremath{\mathbb{C}}$ 2005 The WJG Press and Elsevier Inc. All rights reserved.

Key words: Lewis histoblood group; *Helicobacter pylori*, Peptic ulcer; Gastric cancer

Yei CJ, Chang JG, Shih MC, Lin SF, Chang CS, Ko FT, Lin KY, Liu TC. Lewis blood genotypes of peptic ulcer and gastric cancer patients in Taiwan. *World J Gastroenterol* 2005; 11 (31): 4891-4894

http://www.wjgnet.com/1007-9327/11/4891.asp

INTRODUCTION

The Lewis blood group determinants are structurally related to the antigens of the ABO and H/h blood group systems. They are made by sequential addition of specific monosaccharides onto terminal saccharide precursor chains on glycolipids or glycoproteins. The glycolipids on which they reside on the erythrocyte surface are not synthesized in erythroid tissues, but are acquired by erythrocyte membranes from other tissues through circulating soluble forms that are bound to lipoproteins^[1].

The synthesis of the epitopes is dependent on the interaction of two different fucosyltransferases: alpha(1,2) fucosyltransferase encoded by the *FUT2* or secretor (Se) locus of the H/h blood group system, and alpha(1,3;1,4) fucosyltransferase (FUT3) encoded by the *FUT3* locus. Fucosylation by FUT3 gives rise to the Le^a epitope, whereas the action of both enzymes results in Le^b.

If FUT3 is not expressed, the phenotype is Le^{a-b-} regardless of whether FUT2 (Se) is expressed or not. If FUT3 and FUT2 are both expressed, the phenotype is Le^{a-b+} . If FUT3, but not FUT2 is expressed, the phenotype is Le^{a+b-} . A Le^{a+b+} phenotype may occur, if there is reduced expression of FUT2.

The Le^b glycan has been reported to mediate the attachment of *Helicobacter pylori* (*H pylori*) to human gastric mucosa^[2,3]. However, the clinical significance of this reported association remains a topic of debate. It has also been reported that people who do not secrete soluble Lewis b antigen are more susceptible to *H pylori* infection than people with secretor phenotypes^[4].

This study aimed to explore the association among Lewis antigen phenotypes and genotypes, infection with *H pylori*,

and consequent development of peptic ulcer or gastric cancer.

MATERIALS AND METHODS

Peripheral blood samples were collected as previously described^[4] from 347 patients with endoscopic evidence of peptic ulcer disease (235 cases of duodenal ulcer, 62 cases of gastric ulcer, 50 cases of duodenal ulcer and gastric ulcer) and 51 patients diagnosed with gastric cancer on endoscopy. Peripheral blood specimens from 101 unrelated healthy volunteers were used as normal controls. Subjects were enrolled from the Division of Gastroenterology, Department of Internal Medicine, Taipei Municipal Jen-Ai Hospital from August 1998 to December 2002.

The erythrocyte Lewis phenotype was determined by a tube method using monoclonal antibody (Gamma-CloneR, anti-Le^a, anti-Le^b Gamma Biologicals, Inc. Houston, TX, USA) at Taipei Municipal Jen-Ai Hospital.

Total genomic DNA was isolated from peripheral blood leukocytes as described previously^[5], and Le and Se genotypes were determined by DNA amplification (polymerase chain reaction, PCR) and restriction enzyme analysis^[6,7]. Oligonucleotide primer design and restriction enzyme analysis were carried out as previously described^[5]. The primer sequences and restriction enzymes used in this study are shown in Table 1. The amplified products were digested with appropriate restriction enzymes, followed by electrophoresis on 1.5-4% agarose gels. Direct sequencing of the PCR products in selected cases provided a check on the validity of the procedure.

Gastric or duodenal biopsies were taken from all patients with endoscopic evidence of peptic ulcer or gastric cancer and cultured for H pylori. Bacterial isolates were identified as H pylori by morphology and production of urease and catalase^[8]. The *H pylori* infection status was also evaluated by rapid urease test (CLO test), and urea breath test (¹³C-UBT). *H pylori* infection was defined as positive results by culture or two positive test results on histology, rapid urease test, and ¹³C-urea breath test^[9].

Results were analyzed by the chi-square test, P < 0.05 was considered statistically significant.

RESULTS

H pylori was isolated from 83.7% (303/347) of patients with peptic ulcer disease. Statistical analysis did not show a significant difference in Lewis phenotype (Table 2) or Lewis genotype (Table 3) between peptic ulcer patients with *H pylori* infection and those without infection. Among the 51 gastric cancer patients, *H pylori* infection prevalence in the different Lewis phenotypes was $Le^{a\cdotb+}$, 30/33 (90.9%); $Le^{a\cdotb-}$, 7/8 (87.5%); Le^{a+b-} , 3/3 (100%); and Le^{a+b+} , 6/7 (85.7%).

H pylori was detected in 109/124 (87.9%) of peptic ulcer patients with Se/Se genotype, 148/174 (85%) of those with Se/se genotype, and 53/58 (91.3%) of those with se/se genotype. The difference in proportion of infected patients was not significant (P = 0.436).

Similarly, no significant difference (P = 0.440) was found for presence of *H pylori* infection in peptic ulcer patients with Le/Le (86/97, 88.6%), Le/le (182/215, 84.6%) or le/le (31/34, 91.3%) genotypes.

No significant association was found between Lewis genotype and presence of peptic ulcer disease or gastric cancer (Table 4). Again, an analysis by Se genotype showed no significant difference between patients with peptic ulcer disease and normal controls (P = 0.915) or between patients with gastric cancer and normal controls (P = 0.741), whereas analysis by Le genotype gave similar, non-significant results

Table 1 Primer sequences and restriction enzymes of mutation in Se and Le genes

Mutation	Primer sequence	Enzyme	
Segene			
A385T	UP1: GATGGAGGAGGAATACCGCCTC	Ear I	
	DP2: GATCTCCTGGCGGAGGTGGTGGTAGAAGATC		
G428A	Identical to A358T primer pairs	Bgl II	
C571T	UP:AGGAGATCCTCCAGGAGTTCA	Dde I	
	DP:AGAAGGAGAAAAGGTCTCAAAGG		
G849A	Identical to C571T primer pairs	Dde I	
C628T	UP: AGTGTGGAAGGGGGTGGTGCC	Bgl I	
	DP:CCACTCTGGCAGGAAGGC		
Fusion gene	UP:CTGCCTCCTGACCATGTCC	Pst I	
	DP: identical to reverse primer of C628T		
Legenes			
T202C and C314T	UP:CCACCCTCCTGATCCTGCT <u>C</u>	Msp I (T202C	
	DP:GATATCCCAGTGGTGCACGATGATGATC	Bcl I (C314T)	
C445A	UP: identical to UP of T202 C and C314 T	BstN I	
	DP:GAGATTGAAGTATCTGTCCAAGGC		
G508A	UP:TCAACTTGGAGCCACACCCT	Alu I	
	DP:AGTTGGACACCGCCCAGGCCACCAG		
A1007C	UP:GCTCCTTCCGCTGGGCACTAG	Alu I	
	DP:TGGCCACAAAGGACTCCAGC		
T1067A	UP:GTACCAGACGGTGCG <u>AT</u> GCA	NsiI	
	DP: identical to DP of A1007 C		

¹UP: upstream primer; ²DP: downstream primer; The mutated base is underlined.

Table 2 Correlation between I	Lewis phenotype and	<i>H pylori</i> infection in peptic ulcer diseas	se
---------------------------------------	---------------------	--	----

Phenotypes	$Le^{a-b+}(n = 230)$	Le^{a-b-} (<i>n</i> = 59)	Le^{a+b-} (<i>n</i> = 14)	$\mathrm{Le}^{\mathrm{a+b+}} (n = 44)$	Р
H pylori status					0.5662
Positive	195	54	12	39	
Negative	35	5	2	5	
Prevalence %	84.7	91.5	85.7	88.5	

Table 3 Correlation between Lewis genotype and H pylori infection in peptic ulcer disease

Genotypes	SeLe/SeLe (n = 33)	SeLe/seLe $(n = 65)$	SeLe/Sele (n = 42)	$\frac{\text{SeLe/Sele}}{(n=90)}$	$\frac{\text{SeLe/Sele}}{(n=3)}$	SeLe/Sele $(n = 19)$	SeLe/Sele (<i>n</i> = 37)	SeLe/Sele (<i>n</i> = 46)	SeLe/Sele (n = 12)	Р
H pylori status										0.446
Positive	28	59	36	72	2	17	33	41	12	
Negative	5	6	6	18	1	2	4	5	0	
Prevalence %	84.8	90.7	85.7	80	66.6	89.4	89.1	89.1	100	

Table 4 Lewis genotypes in peptic ulcer and gastric cancer

Genotypes	Peptic ulcer $n = 347$	Gastric cancer $n = 51$	Normal <i>n</i> = 101	
SeLe/SeLe	33	2	7	
SeLe/seLe	65	11	25	
SeLe/Sele	42	4	10	
SeLe/sele	90	15	21	
Sele/Sele	3	3	5	
Sele/sele	19	4	7	
seLe/seLe	37	3	12	
seLe/sele	46	5	9	
sele/sele	12	4	5	
P	0.129^{1}	0.881^{2}		

 ${}^{1}P$ = 0.129 peptic ulcer *vs* normal controls; ${}^{2}P$ = 0.881 gastric cancer *vs* normal controls.

(P = 0.067 and P = 0.344 respectively).

Finally, no significant correlation was obtained between ABO blood group type and *H pylori* infection (Table 5).

DISCUSSION

H pylori is the main causative agent of gastric and duodenal ulcers^[10] and gastric adenocarcinoma^[11]. Attachment is a prerequisite for microbial colonization of epithelial surfaces and is mediated through interaction of adhesins on the bacterial surface and proteins or glycoconjugates on the surface of the epithelial cells^[12,13]. Borén *et al.*^[2], reported that the attachment of *H pylori* to gastric mucosa is mediated by the Lewis^b (Le^b) antigen and that the availability of receptors might therefore be reduced in individuals of blood groups A and B compared to people with blood group O.

Carneiro *et al.*^[14], found that there is a significant relationship between ABO blood group in combination with Lewis phenotype on the one hand and *H pylori* infection on the other. *H pylori* is present in 100% of those with Le^{b+}/O phenotype but in only 57% of Le^{b-}/A or B phenotype. However, infection is also present in 92% of Le^{b-}/O individuals and 86% of Le^{b+}/A or B individuals. Nonetheless, the Carneiro group challenged the finding of Niv *et al.*^[15], that positivity for *H pylori* is not associated with blood group O and their conclusion that their observations do not support the contention that the receptor for *H pylori* in the gastric mucosa is the Le^b antigen.

However, Clyne and Drumm^[16] found that adherence of *H pylori* to isolated human gastric cells is not dependent on Lewis antigen expression on the cells, and Umlauft *et al.*^[17], could not demonstrate any *in vivo* correlation between *H pylori* infection or disease and Le^b antigen. Taylor *et al.*^[18], also found that there is no correlation between Lewis antigen expression by *H pylori* and gastric epithelial cells in infected patients.

More recently, Keller *et al.*^[19], found that there is no significant association between secretor status or specific ABO blood group and *H pylori* infection or occurrence of gastro-duodenal ulcer. Aguiar^[20] also found that there is no significant association between the presence of *H pylori* and ABO, Lewis or secretor phenotype, while Nogueira *et al.*^[21], actually found that Le^b expression is nearly twice as common among children without *H pylori* (15/23, 65%) as in those with *H pylori* (16/47, 34%).

However, Yang et al.[22], found that there are significant

 Table 5
 ABO blood types and susceptibility to H pylori infection (n = 230)

Blood types	A (n = 44)	B (<i>n</i> = 54)	O (<i>n</i> = 122)	AB (n = 10)	Р
H pylori status					0.255
Positive	34	49	106	8	
Negative	10	5	16	2	
Prevalence %	77.2	90.7	86.8	80	

relationships between Lewis phenotype and H pylori infection. Expression of Le^a antigen (whether Le^{a+b-} or Le^{a+b+}) is associated with a higher infection rate but a lower bacterial density, a lower severity of chronic inflammation, and a lower frequency of lymphoid follicles in the gastric cardia. To complicate things even further, H pylori-infected patients expressing the Le^b antigen have a lower rate of gastro-duodenal ulcers but a higher bacterial density and inflammation severity in the gastric cardia. It is difficult to know how to interpret these results.

In vitro studies by Lindén *et al.*^[23], and Van de Bovenkamp *et al.*^[24], showed that binding of *H pylori* to human gastric MUC5AC mucin is blood group antigen-binding adhesin dependent and mediated by the Le^b structure in the mucin. However, it has also been shown that *H pylori* bacteria that do not express Lewis antigens but do express other complex carbohydrates that may still have the ability to form long-term colonies in the stomach^[25,26].

It may be that differences in reported results are due to strain differences in *H pylori*. Hennig *et al.*^[27], found that there is considerable heterogeneity among *H pylori* isolates in expression of the BabA adhesion, which is thought to bind to Le^b antigen present on the surface of gastric epithelial cells. Sheu *et al.*^[28], in a study of 188 dyspeptic patients with *H pylori* infection in Taiwan, found that all isolates have a positive babA2 genotype and that, among 139 patients with Le^b expression, *H pylori* density increases with Le^b intensity. In the 49 patients without gastric Le^b expression, *H pylori* density is positively correlated with Le^x and Le^a expression.

In conclusion, Lewis blood genotype or phenotype may not play a role in the pathogenesis of *H pylori* infection. However, bacterial strain differences and the presence of more than one attachment mechanism may limit the contribution of epidemiological studies toward elucidating this matter.

REFERENCES

- Marcus DM, Cass LE. Glycosphingolipids with Lewis blood group activity: uptake by human erythrocytes. *Science* 1969; 164: 553–555
- 2 Borén T, Falk P, Roth KA, Larson G, Normark S. Attachment of *Helicobacter pylori* to human gastric epithelium mediated by blood group antigens. *Science* 1993; 262: 1892-1895
- 3 Ilver D, Arnqvist A, Ogren J, Frick IM, Kersulyte D, Incecik ET, Berg DE, Covacci A, Engstrand L, Borén T. Helicobacter pylori adhesin binding fucosylated histo-blood group antigens revealed by retagging. Science 1998; 279: 373-377
- 4 **Borén T,** Normark S, Falk P. *Helicobacter pylori*: molecular basis for host recognition and bacterial adherence. *Trends Microbiol* 1994; **2**: 221-228
- 5 Chang JG, Chiou SS, Perng LI, Chen TC, Liu TC, Lee LS, Chen PH, Tang TK. Molecular characterization of glucose-6-phosphate dehydrogenase (G6PD) deficiency by naturally and amplification created restriction sites: five mutations account for most G6PD deficiency cases in Taiwan. *Blood* 1992; 80: 1079-1082
- 6 Chang JG, Yang TY, Liu TC, Lin TP, Hu CJ, Kao MC, Wang NM, Tsai FJ, Peng CT, Tsai CH. Molecular analysis of secretor type a (1,2)-fucosyltransferase gene mutations in the Chinese and Thai populations. *Transfusion* 1999; **39**: 1013-1017
- 7 Liu TC, Chang JG, Lin SF, Chang WC, Yang TY, Lin CL, Wang NM. Lewis (FUT3) genotypes in Taiwanese, Thai, and Filipino populations. *Ann Hematol* 2000; **79**: 599-603
- 8 Hazell SL, Evans DJ, Graham DY. *Helicobacter pylori* catalase. J Gen Microbiol 1991; **137**: 57-61

- 9 Wang WM, Lee SC, Wu DC, Chen LT, Liu CS, Peng CF, Ding HJ, Chen CY, Jan CM. Simplified ¹³C-urea breath test for the diagnosis of *Helicobacter pylori* infection-the availability of without fasting and without test meal. *Kaohsiung J Med Sci* 2000; **16**: 607-613
- Graham DY. *Helicobacter pylori*: its epidemiology and its role in gastroduodenal ulcer disease. J Gastroenterol Hepatol 1991; 6: 105-113
- 11 Eurogast Study Group. An international association between Helicobacter pylori infection and gastric cancer. The EUROGAST Study Group. Lancet 1993; 341: 1359-1362
- 12 **Ofek I**, Sharon N. Adhesins as lectins: specificity and role in infection. *Curr Top Microbiol Immunol* 1990; **151**: 91-113
- 13 Karlsson KA. Animal glycosphingolipids as membrane attachment sites for bacteria. Annu Rev Biochem 1989; 58: 309-350
- 14 Carneiro F, Amado M, Lago P, Taveira-Gomes A, Amil M, Barreira R, Soares J, Pinho C. *Helicobacter pylori* infection and blood groups. *Am J Gastroenterol* 1996; **91**: 2646-2647
- 15 Niv Y, Fraser G, Delpre G, Neeman A, Leiser A, Samra Z, Scapa E, Gilon E, Bar-Shany S. *Helicobacter pylori* infection and blood groups. *Am J Gastroenterol* 1996; 91: 101-104
- 16 Clyne M, Drumm B. Absence of effect of Lewis A and Lewis B expression on adherence of *helicobacter pylori* to human gastric cells. *Gastroenterology* 1997; 113: 72-80
- 17 Umlauft F, Keeffe EB, Offner F, Weiss G, Feichtinger H, Lehmann E, Kilga-Nogler S, Schwab G, Propst A, Grünewald K, Judmaier G. *Helicobacter pylori* infection and blood group antigens: lack of clinical association. *Am J Gastroenterol* 1996; **91**: 2135-2138
- 18 Taylor DE, Rasko DA, Sherburne R, Ho C, Jewell LD. Lack of correlation between Lewis antigen expression by *Helicobacter pylori* and gastric epithelial cells in infected patients. *Gastroenterology* 1998; **115**: 1113-1122
- 19 Keller R, Dinkel KC, Christl SU, Fischbach W. Interrelation between ABH blood group O, Lewis (B) blood group antigen, *Helicobacter pylori* infection, and occurrence of peptic ulcer. Z *Gastroenterol* 2002; 40: 273-276
- 20 Aguiar DC, Corvelo TC, Arajo M, Cruz EM, Daibes S, Assumpcao MB. Expression of ABH and Lewis antigens in chronic gastritis and pre-neoplastic alterations in gastric mucosa. Arq Gastroenterol 2002; 39: 222-232
- 21 Nogueira AM, Marques T, Soares PC, David L, Reis CA, Serpa J, Queiroz DM, Rocha GA, Rocha AC. Lewis antigen expression in gastric mucosa in children: relationship with *Helicobacter pylori* infection. J Pediatr Gastroenterol Nutr 2004; **38**: 85-91
- 22 Yang HB, Sheu BS, Chen RC, Wu JJ, Lin XZ. Erythrocyte Lewis antigen phenotypes of dyspeptic patients in Taiwancorrelation of host factor with *Helicobacter pylori* infection. J Formos Med Assoc 2001; 100: 227-232
- 23 Lindén S, Nordman H, Hedenbro J, Hurtig M, Borén T, Carlstedt I. Strain- and blood group-dependent binding of *Helicobacter pylori* to human gastric MUC5AC glycoforms. *Gastroenterology* 2002; 123: 1923-1930
- 24 **Van de Bovenkamp JH**, Mahdavi J, Korteland-Van Male AM, Buller HA, Einerhand AW, Boren T, Dekker J. The MUC5AC glycoprotein is the primary receptor for *Helicobacter pylori* in the human stomach. *Helicobacter* 2003; **8**: 521-532
- 25 **Rasko DA**, Wilson TJ, Zopf D, Taylor DE. Lewis antigen expression and stability in *Helicobacter pylori* isolated from serial gastric biopsies. *J Infect Dis* 2000; **181**: 1089-1095
- 26 Altman E, Smironova N, Li J, Aubry A, Logan SM. Occurrence of a nontypable *Helicobacter pylori* strain lacking Lewis blood group O antigens and DD-heptoglycan: evidence for the role of the core alpha-1,6-glucan chain in colonization. *Glycobiology* 2003; **13**: 777-783
- 27 Hennig EE, Mernaugh R, Edl J, Cao P, Cover TL. Heterogeneity among *Helicobacter pylori* strains in expression of the outer membrane protein BabA. *Infect Immun* 2004; 72: 3429-3435
- 28 Sheu BS, Sheu SM, Yang HB, Huang AH, Wu JJ. Host gastric Lewis expression determines the bacterial density of *Helicobacter pylori* in babA2 genopositive infection. *Gut* 2003; **52**: 927-932

Science Editor Wang XL and Guo SY Language Editor Elsevier HK