

# Sono-photoacoustic imaging of gold nanoemulsions: Part I. Exposure thresholds



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## ABSTRACT

Integrating high contrast bubbles from ultrasound imaging with plasmonic absorbers from photoacoustic imaging is investigated. Nanoemulsion beads coated with gold nanospheres (NEB-GNS) are excited with simultaneous light (transient heat at the GNS's) and ultrasound (rarefactional pressure) resulting in a phase transition achievable under different scenarios, enhancing laser-induced acoustic signals and enabling specific detection of nanoprobe at lower concentration. An automated platform allowed dual parameter scans of both pressure and laser fluence while recording broadband acoustic signals. Two types of NEB-GNS and individual GNS were investigated and showed the great potential of this technique to enhance photoacoustic/acoustic signals. The NEB-GNS size distribution influences vaporization thresholds which can be reached at both permissible ultrasound and light exposures at deep penetration and at low concentrations of targets. This technique, called sono-photoacoustics, has great potential for targeted molecular imaging and therapy using compact nanoprobe with potentially high-penetrability into tissue.

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## 1. Introduction

Contrast-enhanced imaging with exogenous contrast agents is a rapidly developing technique for both photoacoustic (PA) and ultrasound (US) systems. Microbubbles have been used in US harmonic imaging [1] of vasculature, and for drug delivery [2] or cavitation-based treatments [3]. Their large size (typically 1–10  $\mu\text{m}$ ), however, inhibits high penetrability into tissue through leaky vasculature (300–500 nm endothelial gaps) and nanoscale pores, and through active mechanisms such as cellular uptake. Making stable nanobubbles at the dimensions required for enhanced transport is currently a challenge [4].

To solve this problem, phase-change contrast agents, which change state from liquid to gas if exposed to thermal and/or acoustic energy, were introduced. Liquid perfluorocarbon nanodroplets with a low-boiling point and a physical dimension appropriate for enhanced penetrability into tissue have been developed [5–8]. The

phase-transition of these nanodroplets into a microbubble (vaporization) produces high contrast for US imaging. Compared to other US contrast agents, nanoemulsions can have a long circulation in the body (up to 2 h [8]). Because of increased surface tension, however, US exposure parameters must be relatively high, usually involving high repetition rates [7] or long excitations ( $\sim 100$  ms) [6]. This means that heat can be involved in the vaporization, increasing the possibility to damage surrounding tissues or limiting the repetition rate of the imaging modality. This mechanism is also not efficient using low-frequency ultrasound because of low acoustic absorption.

To achieve molecular imaging at significant depths within tissue, nanoscale, laser-activated PA contrast agents have been introduced [9–13]. Plasmonic absorption enhances the efficiency of compact nanoprobe (10–200 nm) in PA generation. However, a significant concentration is usually required to get single shot measurements for real-time *in vivo* imaging (e.g.  $\sim 10$  nM) [9–13]. Recent studies have shown that combining nanodroplets with efficient optical absorbers in a single contrast agent can enhance the PA signal from the absorbers [12]. Here we build on this work to demonstrate a highly sensitive, and potentially specific, approach for molecular imaging and therapy using a nanoscale contrast agent integrating

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perfluorocarbon nanodroplets with plasmonic absorbers. In particular, we explore the non-linear acoustic response of these agents to simultaneous light and sound excitations at exposure levels well within safety limits for routine diagnostic imaging.

In our previous work, we introduced a nanoemulsion of perfluorohexane droplets (100–200 nm) surrounded by amphiphilic gold nanopsheres (12 nm in diameter). It was used as a non-linear PA contrast agent by inducing a phase transition of the oil through plasmonic absorption of light by the gold nanoparticles [14]. This approach can produce highly sensitive PA images with high background suppression (*i.e.*, also highly specific contrast), but it requires a relatively high optical fluence to observe non-linear contrast (at 750 nm, threshold at 4 mJ/cm<sup>2</sup>, 10 dB contrast around 6 mJ/cm<sup>2</sup>), limiting its use for deep imaging applications.

An approach to greatly reduce the optical fluence required for a non-linear response from these contrast agents is suggested by recent work showing that laser generation of water vapor bubbles around plasmonic absorbers can be enhanced by adding ultrasound pressure fields [15–17]. This technique, named “PA cavitation”, enables high contrast imaging at lower exposure thresholds and concentrations. Building on this work, we investigate here the PA cavitation exposure thresholds needed to trigger vaporization of a nanoemulsion bead coated with gold nanopsheres (NEB-GNS). This approach leverages short-lived (a few  $\mu$ s) bubbles made possible by the high boiling point (BP) of the perfluorocarbon used in this nanoemulsion. In contrast to another approach using long lifetime bubbles [12], a short-lifetime strategy using reversible phase transitions is preferred for molecular imaging applications requiring extended exposures. Using PA cavitation, the detection sensitivity for imaging can be enhanced by two physical effects: higher thermal expansion will enhance the PA response and the emitted ultrasound field will instantly scatter on the generated bubbles.

In the remainder of this paper, we investigate the benefit of clustering small gold nanopsheres around high-BP perfluorocarbon droplets for combined PA/US applications. After introducing the physical characteristics of the NEB-GNS agent, vaporization thresholds are quantified and compared to single gold nanopsheres. The potential use of the vaporization signals for imaging is then studied and discussed. To specifically retrieve the vaporization signal in tissues loaded with NEB-GNS, photoacoustic and backscattered ultrasound signals must be canceled by a procedure described in a future publication called “sono-photoacoustic” imaging.

## 2. Materials and methods

### 2.1. Nanoemulsion samples

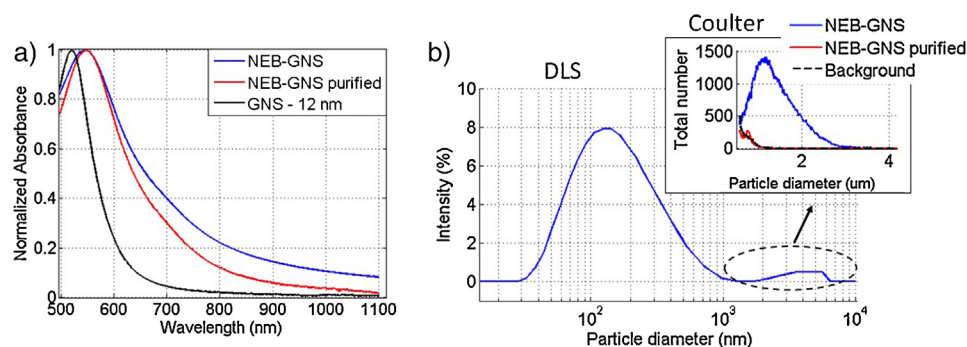
NEB-GNSs were synthesized using the procedure described in previous reports [18,19]. Colloidal GNSs (diameter 12 nm) were

synthesized using a citrate reduction method [20]. The particles were functionalized using PEG-thiol and butane-thiol (Sigma-Aldrich, St. Louis, MO, USA), with dosages of 0.8 chains/nm<sup>2</sup> Au and 700 molecules/nm<sup>2</sup> Au respectively. Attraction between the hydrophobic butane-thiol ligands resulted in clustered GNS dispersions. A solution of 1 vol% perfluorohexane (Sigma-Aldrich, St. Louis, MO, USA) and 0.012 vol% Au clusters in water was sonicated (102C, Branson, Danbury, CT, USA, pulsed regime – 1 s on, 4 s off) for 13 s in a cold water bath.

The absorbance was measured with a spectrophotometer (UV 1601, Shimadzu, Kyoto, Japan). The absorbance spectrum (*cf* Fig. 1a, blue curve) exhibits a red shift of the resonance from 520 nm to 547 nm compared to the original gold nanopsheres (black curve) and a broad tail enhancing absorption in the optical window ([600–1100] nm wavelengths). The size was measured by DLS as a broad dispersion between 30 nm and 1.2  $\mu$ m with a peak at 132.1 nm and a mean size of 303 nm (*cf* Fig. 1b). Additional complexes were identified between 2 and 6  $\mu$ m, but DLS may not be quantitative in this range as it is heavily affected by sedimentation or flotation [21]. In our case, the effective density of perfluorohexane beads (1.7 g/cm<sup>3</sup>) can be increased by the GNS (density 19.3) resulting in sedimentation speeds depending on size.

To confirm the presence and quantify the micron-size particle number, the size distribution and concentration of the emulsion were obtained prior to and after all experiments using a Coulter Multisizer III (Beckman Coulter, Miami, FL). A 20  $\mu$ m aperture was used, which can size particles with diameters from 0.56 to 12  $\mu$ m and considers any count below 0.56  $\mu$ m as the noise level. The sample was diluted  $\sim$ 1250 $\times$  on a 0.2  $\mu$ m filtered ISOTON II electrolyte (Beckman Coulter, Miami, FL). A 50  $\mu$ L sample was used each time, and all measurements were repeated 6 times using a volumetric count mode. Individual particles were sized and binned in 300 evenly spaced bins with 0.039  $\mu$ m width. All data are reported as a histogram with count vs. diameter, with the count (bin height) showing the number of particles in each bin interval. The reported concentration is computed for all ranges in question and accounts for the dilution factor and sampling volume used above.

A near Gaussian distribution of NEB-GNS was identified between 1 and 3  $\mu$ m (*cf* inset of Fig. 1b). The sample was then split in two volumes. Assuming an ideal covering of the bead by gold nanopsheres (80% surface density), the weight of each bead size was calculated. Then, a basic model of sedimentation (Stokes force = gravity) was used to calculate the sedimentation speeds of different particles: *e.g.*, in 24 h a 425 nm particle sinks 22 mm. In contrast, a 132 nm bead should sink only 6 mm in the same period. A sample of 10 ml was left to settle in a scintillation vial. As 22 mm was the height of our solution in the vial, we assume that all



**Fig. 1.** (a) Absorbance spectra of the nanoemulsion before (blue) and after purification (red), and of gold nanopsheres in suspension (black). (b) Sizing of the nanoemulsion by DLS (intensity). Inset: Sizing in the micron range using a Coulter counter: nanoemulsion before (blue) and after purification (red); background signal is shown in black.



marker spot when only the laser was fired. Then, US pulses were emitted and the echoes generated on the edge of the pipet were measured with the PVDF transducer. By changing the trigger delay controlling the duration between the acoustic emission and the lasing time, the arrival time of the PA signal was matched to the one of the 5th acoustic cycle, meaning the laser is fired on the sample when the 5th US cycle reaches it. Note, that the laser beam size of 6 mm is larger than the acoustic wavelength (1.2 mm) and covers multiple peak negative pressure sites. Thus, contrary to our preliminary experiment where the light was focused on a 1 mm area [28], the exact lasing time in the microsecond range will not affect the results. Broad illumination is also more representative of potential *in vivo* applications. The sample was then moved horizontally toward the transducer, allowing both the focal spot of the transducer and the laser spot to meet within the pipet bulb. As noted above, a 10-cycle ultrasound pulse emitted with the focused US transducer was used for all studies. The laser was fired at the arrival time of the 5th acoustic cycle at the transducer focus.

To cover all desired exposure ranges in terms of laser fluence and US pressure while recording enough data to perform statistics, an automated platform was designed. A TTL signal running at 20 Hz, corresponding to the flash lamp timing of the laser, is used as an input trigger to an ADC board (Razor 14X2 Express CompuScope, Gage, Lockport, IL, USA) controlled with Matlab. When the software is running, a trigger out is sent by the ADC board immediately after being triggered. This signal is then delayed by 250  $\mu$ s by a function generator (AFG-3252, Tektronix, OR, USA) to trigger the laser Q-switch. This approach limits exposure of the samples since lasing only occurs when a signal is actually being recorded. Moreover, the repetition rate was limited to 5 Hz by the Matlab software. The second channel of the generator was used to drive an RF amplifier (A150, ENI, USA) at a voltage also controlled by software to independently scan the pressure amplitude. Ultrasound was emitted earlier than the laser so that it could propagate to the region under study. The PVDF transducer finally received both the laser-induced PA signals and the scattered pressure waves originating from the focused transducer. This RF signal was captured by the ADC, displayed in real-time and saved on the computer hard-drive for further analysis. At each set of exposure parameters, 200 recordings were performed. The laser fluence was gradually increased. For each laser fluence, the ultrasound pressure was raised until significant cavitation activity was noted.

Fluence values were calibrated by placing an energy meter (J-power, Coherent, Santa Clara, CA) at a position matching the focus of the US transducer. A 2-mm pinhole was used, ensuring that it was at the maximum energy position of the laser beam. Then, light attenuation through 60-mm of water was taken into account to report all fluence values.

### 2.5. Data analysis

For each set of exposure parameters, a reference signal recorded without laser firing was subtracted from the 200 signals to remove the US scattered wave contributions from the plastic container, similar to methods used in our previous studies [28]. This subtraction was robust and produced residual signals outside the time window of interest much smaller than those obtained during laser excitation studies. The broadband noise signal was computed in a time window before the lasing time. If no bubble is present, the differential signal should correspond to pure noise. In contrast, the presence of vapor will automatically generate some scattered acoustic signals in all directions. To detect the vaporization events, a threshold on the magnitude of the signals equal to 2 times the noise value was chosen to count the number of cavitation events out of the 200 independent recordings. This

threshold was set arbitrarily and the results were not highly influenced by its value, indicating that our detection method is very sensitive. A sigmoidal function was fit to all broadband noise dependencies at each laser fluence to compute the cavitation threshold, defined as the peak negative acoustic pressure corresponding to a 50% probability of a cavitation event.

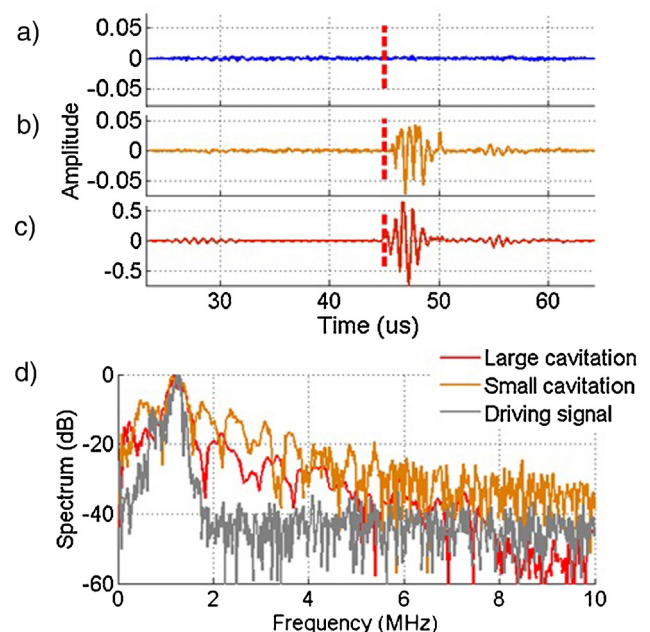
## 3. Results

### 3.1. Cavitation signals

After removing the contribution from the tube using a reference signal, the recorded signals clearly exhibited the presence of cavitation. With the same exposure parameters, a small cavitation event can be distinguished in Fig. 3b from no cavitation in Fig. 3a. Cavitation always occurred at the lasing time depicted by the red dotted line. A larger cavitation event is shown in Fig. 3c with 10-fold amplitude compared to the small cavitation (note scale change). Normalized spectra for both of these signals are shown in Fig. 3d. The broadband noise characteristics are present in both signals, although the main harmonic is relatively higher for the large cavitation event.

### 3.2. Vaporization probabilities

The resulting cavitation probabilities as a function of US peak negative pressure are shown for the NEB-GNS and purified NEB-GNS samples in Fig. 4a and b. For both samples, cavitation activity was not consistently observed using the 10-cycle ultrasound pulse alone, even at pressures exceeding 8 MPa (below 20% for the raw sample and 2% for the purified one; results are not shown). In contrast, a small fluence (0.64  $\text{mJ}/\text{cm}^2$ ) already creates consistent vaporization at 1.5 MPa for the raw sample and at 2.5 MPa for the purified one. Higher fluences decrease the vaporization threshold. Note that for the purified sample, and at such a low concentration,



**Fig. 3.** Wideband detection of cavitation events using a PVDF detector. Lasing time is shown with the red dotted line and the ultrasound 10-cycle pulse travels to the focus starting from the blue dotted line time. (a)–(c) Signals after differentiation. (a) and (b) No cavitation and cavitation (probability 5%) with exposures of  $F_0 = 0.641 \text{ mJ}/\text{cm}^2$ ,  $p = 1.74 \text{ MPa}$ . (c) Large cavitation ( $F_0 = 4.7 \text{ mJ}/\text{cm}^2$ ,  $p = 1.43 \text{ MPa}$ ). (d) Normalized spectra of differential signals for large (red), small (orange) and spectrum of the driving signal at 1.24 MHz.

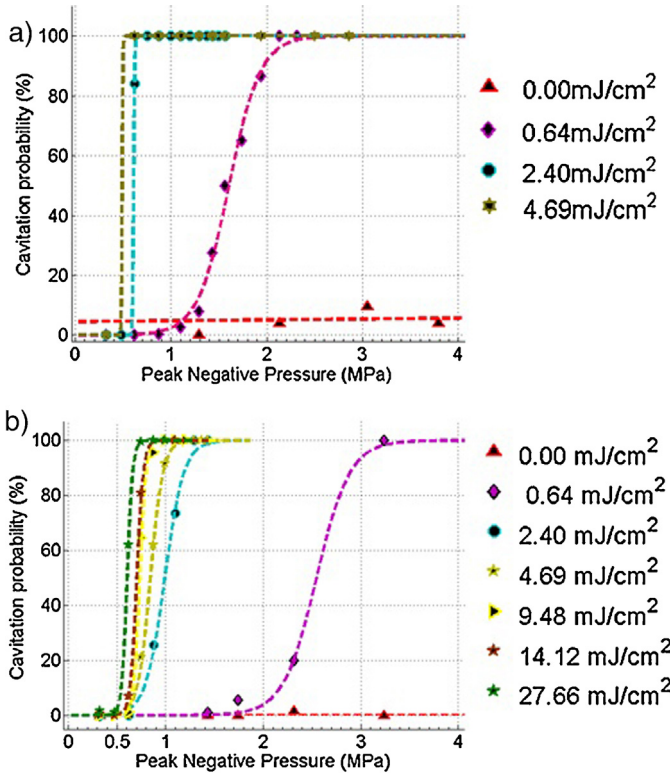


Fig. 4. Photoacoustic cavitation probabilities of emulsion sample #1 before (a) & after purification (b).

a very high laser fluence of 27.7 mJ/cm<sup>2</sup> is only efficient for vaporization at 0.6 MPa, and that raising the fluence from 9.48 to 27.7 mJ/cm<sup>2</sup> doesn't change the cavitation threshold dramatically.

These results show that combining light (transient heating) and short ultrasound pulse (pressure) exposures is much more efficient than using each of them separately.

### 3.3. Summary of the vaporization results and comparison with a GNS sample

A summary of the vaporization threshold (50% probability) of the GNS-NEB samples (raw and purified) and the GNS 12 nm sample is presented in Fig. 5. For a peak negative pressure of

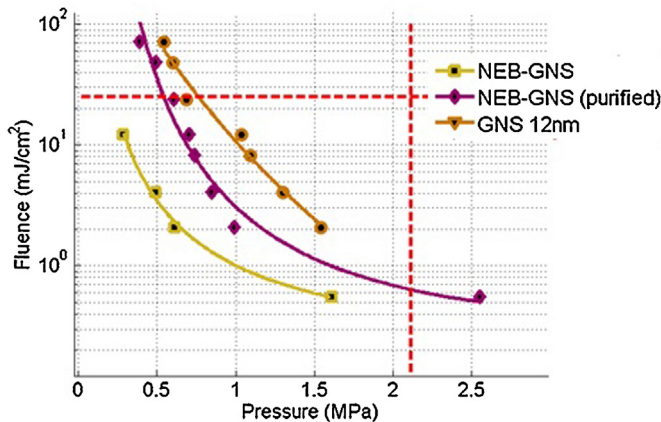


Fig. 5. Summary of vaporization thresholds. Each point in the Fluence/Peak negative pressure plane corresponds to a 50% vaporization probability. NEB-GNS, NEB-GNS purified and GNS. The red dotted lines indicate: the mechanical index (MI) FDA limitation of 1.9 for ultrasound imaging (vertical) and the permissible optical fluence exposure of 25 mJ/cm<sup>2</sup> (horizontal).

1.0 MPa, the fluence threshold of the GNS is reduced by a factor of 12 for the raw sample and by a factor of 6 for the filtered sample.

### 3.4. Sono-photoacoustic signal enhancement

For imaging purposes, the amplitude emitted by the bubbles will determine detection sensitivity. In addition, the non-linear nature of sono-photoacoustic signal enhancement may lead to a highly specific agent. To explore this hypothesis, the signals presenting cavitation within each data set were isolated. Then, the median value and the standard deviation of the maximum amplitude in time of these signals were calculated.

In Fig. 6, the sono-photoacoustic signal clearly evolves in a non-linear manner as a function of acoustic peak negative pressure for both the raw sample and the purified one. Note that amplitudes are higher in Fig. 6a than in Fig. 6b. The bubble signatures had in fact similar amplitudes at a fixed pressure. The error bars denote the standard deviation  $\sigma$  of the amplitudes within the ensemble of cavitation events with a length equal to  $2\sigma$ . While the cavitation threshold decreases with laser fluence, the amplitude of cavitation events does not change with pressure when the fluence is above a threshold (above 2.4 mJ/cm<sup>2</sup> for both samples). This result suggests the potential for deep sono-photoacoustic imaging.

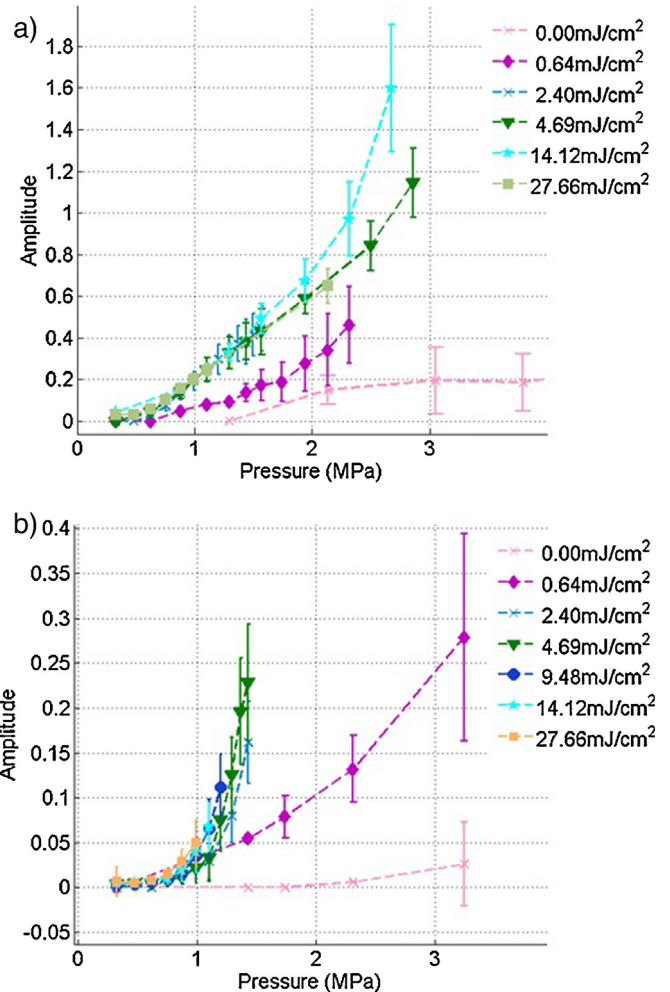


Fig. 6. Amplitude of sono-photoacoustic signals as a function of acoustic pressure at different fluences. (a) NEB-GNS sample. (b) NEB-GNS purified sample.

#### 4. Discussion

In this paper, we have demonstrated the potential of a novel contrast agent composed of nanoemulsion beads surrounded by gold nanospheres using simultaneous light and sound excitation. A robust emulsion characterization platform, which performs a 2-dimensional parameter scan (pressure and light) for cavitation threshold determination, was developed and introduced. This platform will be particularly beneficial for future testing and comparison of new nanoemulsion contrast agent formulations.

The exposure parameters required to achieve PA cavitation were extensively studied at a picomolar concentration of contrast agent ( $2\text{--}6 \times 10^{10}$  beads/ml). Ultrasound pressures were varied from 0.1 MPa to 9 MPa at focal peak negative pressures and laser fluences varied from 0.4 to 27.6 mJ/cm<sup>2</sup>. As laser fluence increased, the cavitation threshold (in terms of US peak negative pressure) decreased to values far below the FDA mechanical index ( $MI < 1.9$ ) limitation.

From Fig. 4, it is clear that the cavitation probability for ultrasound exposure alone remained low for pressures up to 4 MPa (<10%). The pressure for ultrasound exposure alone was scanned up to 9 MPa (not shown in Fig. 4) yet we did not reach a 50% probability of cavitation (dashed pink line). Previous work with similar nanoemulsions, in terms of boiling points and sizes, explored acoustic droplet vaporization thresholds of dodecafluoropentane (DDFP, boiling point in bulk = 29 °C) and of 2H,3H-perfluoropentane (H-PFP, boiling point in bulk = 55 °C) albumin-coated droplets with a mean diameter of approximately 260 nm, and 630 nm, respectively [24]. These nanoemulsions were included in albumin-acrylamide gel phantoms and sonicated with 10 cycle HIFU pulses ( $f = 2$  MHz, pulse repetition = 5 s), similar to our setup (1.24 MHz). For DDFP, an ultrasound peak negative pressure threshold was found at 8.50 MPa. In the case of H-PFP, the pressure was increased up to 8.7 MPa without evidence of vaporization. Our results are consistent with this study [24]. When exposed to 5 MHz, micron-scale perfluoropentane droplets vaporization thresholds decreases with droplet diameter [25] as surface tension decreases. As the purification process removes particles larger than 600 nm, the observed increase of vaporization threshold is consistent with previous studies. Additionally, many studies reported a decreasing vaporization pressure as a function of increasing US frequency [26,27]. As mentioned earlier, heat seems to be necessary to overcome surface tension stabilization. Our study shows that this heat can be delivered in a controlled and targeted way by plasmonic light absorption. Further work could study photoacoustic vaporization thresholds of NEB-GNS at higher frequencies to find the optimal configuration to implement real-time sono-photoacoustic imaging.

Preliminary imaging experiments (not shown here) suggest that particles accumulated at the bottom of the sample. Single nanoparticles are mostly subject to diffusion and gravity can usually be neglected. In contrast, gravity effects can be dominant for large oil droplets (1.7 density) and is amplified by the gold particles on the surface. We took advantage of these effects to purify our sample. A centrifugation technique could have been used, but it was not preferred because it could increase the probability of aggregation of beads at the bottom of the container. Similarly, extrusion through membrane filters could lead to more consistent size distributions, but it could also generate new distributions of beads since extrusion can actually help create droplets. It could also result in the loss of products trapped on the membrane surface.

Further work could investigate the best way to purify NEB-GNS samples. DLS measurements have to be complemented with other sizing techniques such as a Coulter counter to quantify the amount of micron-scale beads if high penetrability is needed (*i.e.* particles >500 nm will not penetrate the gaps in leaky vasculature). Even a

very small amount of larger beads can greatly affect the measured vaporization thresholds as their volume is far larger. A strategy using a broad distribution such as our raw sample (100 nm–6 μm) can image vasculature scales from large vessels to capillaries, with the ability to penetrate leaky vasculature only for the smaller particles.

Previous high speed photomicrography studies on the same emulsion type showed the short lifetimes  $\sim 1.2$  μs for the vapor bubbles under light exposure only [28], suggesting that the perfluorocarbon beads also go back to the liquid state when the ultrasound field vanishes. The fate of the gold nanoparticles during bubble expansion and after condensation has not been studied yet. Nevertheless, additional experiments have also shown a strong consistency in the cavitation activity after several thousand shots, indicating that the gold particles remain somehow coupled to the beads. Observing a single emulsion bead flowing into a small channel and exposed to light and sound could lead to better characterization [29].

Photoacoustic cavitation is surprisingly induced in the GNS sample composed of small particles (15 nm diameter) at a wavelength (750 nm) where optical absorption is very weak. Previous work has only studied photoacoustic cavitation of large diameter GNS (82 nm diameter) at the plasmonic resonance wavelength (532 nm) [17]. Here, the mean distance between GNS is 72 μm given the size and concentration, so coupling effects should not play a role in effective vapor bubble creation unless there is particle aggregation due to acoustic streaming, or other pressure-induced effects. Further modeling inspired by previous work [30,31] could focus on explaining this behavior. Compared to the emulsion beads, the liquid volume to vaporize is not the same and the total energies necessary to achieve vaporization might not be comparable.

As the probability of cavitation increases (see Fig. 6), we have shown that the amplitude of the resultant acoustic signal does not increase with laser fluence for fluences higher than 2.4 mJ/cm<sup>2</sup>. This means that when the phase transition occurs, a higher laser fluence does not affect the vaporization signals as much as the US pressure does. The laser pulse induces heat within a short 30–40 ns time window enabling the phase transition. Then, it appears that the bubble size is more influenced by the pressure field, which also controls the amplitude of bubble-generated acoustic waves. As extinction by soft tissue limits light penetration, a scenario of low laser fluence with a maximum permissible pressure would be ideal for contrast enhancement. Depending on the tissue types between the light source and the region of interest, such light penetration can be achieved from one to a few centimeters deep into tissue. A Monte Carlo simulation [31] of light transport was performed through skin (thickness  $\Delta Z = 1.12$  mm, absorption coefficient  $\mu_a = 0.49$  cm<sup>-1</sup>, scattering coefficient  $\mu_s = 82.6$  cm<sup>-1</sup>, anisotropy  $g = 0.75$ ), fat ( $\Delta Z = 2$  mm,  $\mu_a = 0.1$  cm<sup>-1</sup>,  $\mu_s = 115$  cm<sup>-1</sup>,  $g = 0.9$ ) and muscle ( $\Delta Z = 30$  mm,  $\mu_a = 0.09$  cm<sup>-1</sup>,  $\mu_s = 216$  cm<sup>-1</sup>,  $g = 0.9$ ). Using a gaussian beam (width: 2 cm,  $\lambda = 750$  nm, maximum fluence: 25 mJ/cm<sup>2</sup>), fluences of 2 mJ/cm<sup>2</sup> and 0.6 mJ/cm<sup>2</sup> were respectively reached at depths of 1.6 and 2.0 cm. Future studies could explore the penetration limits for different imaging applications.

Finally, thanks to the clustering of gold nanospheres on an emulsion bead, and to the resulting large signal from the bubble, these measurements succeeded even at low concentration in a regime where single shot linear PA imaging fails (0.25 cm<sup>-1</sup>). The nanoemulsion structure and synthesis can be further improved to reach higher efficiency, for example using another perfluorocarbon oil with a lower boiling point [25,32]. Future studies will explore the relationship between both threshold and acoustic signal strength as a function of concentration.

Based on this work, a real-time imaging system could achieve sono-photoacoustic imaging of NEB-GNS at depths exceeding several centimeters and at ultra-low concentration levels. This

imaging approach could rely on single cycle or multiple cycles US excitations, depending on the desired imaging range. However, to use this technique for *in vivo* imaging, scattered ultrasound and photoacoustic signals must both be canceled to retrieve the specificity of vaporization signals and perform accurate background suppression. A way to implement sono-photoacoustics in a real-time imaging sequence will be demonstrated in a companion publication. Moreover, inertial cavitation produced by this nanoemulsion already has been exploited for *in vitro* sonothrombolysis [28,33]. Further work can rely on this study to optimize the delivery of this treatment and its monitoring.

## 5. Conclusion

This work showed the potential of gold-coated nanoemulsion beads excited with simultaneous pulsed laser and ultrasound. A fully automated setup for dual exposure parameter scanning was designed for quantitative assessment of vaporization thresholds of the NEB-GNS. A phase transition of the nanoemulsion bead occurred at much lower peak negative pressure when laser fluence increased. We demonstrated that NEB-GNS vaporization could be achieved at a reduced exposure compared to the vaporization of water surrounding single GNS. The high amplitude of vaporization signals produced by simultaneous pulsed light and ultrasound exposure could be used to design an imaging modality allowing both high specificity and sensitivity of nano-contrast agent detection. The results could also be applied to drug delivery using nanodroplets with high encapsulation energy and to localized or targeted therapies using nanoagents.

## Conflict of interest statement

The authors declare that there are no conflicts of interest.

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## References

- [1] Wei K, Jayaweera AR, Firoozan S, Linka A, Skyba DM, Kaul S. Quantification of myocardial blood flow with ultrasound-induced destruction of microbubbles administered as a constant venous infusion. *Circulation* 1998;97:473–83.
- [2] Ferrara K, Pollard R, Borden M. Ultrasound microbubble contrast agents: fundamentals and application to gene and drug delivery. *Annu Rev Biomed Eng* 2007;9:415–47.
- [3] Unger EC, Porter T, Culp W, Labell R, Matsunaga T, Zutshi R. Therapeutic applications of lipid-coated microbubbles. *Adv Drug Deliv Rev* 2004;56:1291–314.
- [4] Cavalli R, Bisazza A, Trotta M, Argenziano M, Civra A, Donalizio M, et al. New chitosan nanobubbles for ultrasound-mediated gene delivery: preparation and *in vitro* characterization. *Int J Nanomed* 2012;7:3309–18.
- [5] Rapoport N, Gao Z, Kennedy A. Multifunctional nanoparticles for combining ultrasonic tumor imaging and targeted chemotherapy. *JNCI – J Natl Cancer Inst* 2007;99:1095–106.
- [6] Giesecke T, Hynynen K. Ultrasound-mediated cavitation thresholds of liquid perfluorocarbon droplets *in vitro*. *Ultrasound Med Biol* 2003;29:1359–65.
- [7] Kripfgans OD, Fowlkes JB, Miller DL, Eldevik OP, Carson PL. Acoustic droplet vaporization for therapeutic and diagnostic applications. *Ultrasound Med Biol* 2000;26:1177–89.
- [8] Rapoport N, Nam K-H, Gupta R, Gao Z, Mohan P, Payne A, et al. Ultrasound-mediated tumor imaging and nanotherapy using drug loaded, block copolymer stabilized perfluorocarbon nanoemulsions. *J Control Release* 2011;153:4–15.
- [9] Agarwal A, Huang SW, O'Donnell M, Day KC, Day M, Kotov N, et al. Targeted gold nanorod contrast agent for prostate cancer detection by photoacoustic imaging. *J Appl Phys* 2007;102:064701.
- [10] Chen Y-S, Frey W, Kim S, Kruizinga P, Homan K, Emelianov S. Silica-coated gold nanorods as photoacoustic signal nanoamplifiers. *Nano Lett* 2011;11:348–54.
- [11] Li P-C, Wei C-W, Liao C-K, Chen C-D, Pao K-C, Wang C-RC, et al. Photoacoustic imaging of multiple targets using gold nanorods. *IEEE Trans Ultrason Ferroelectr Freq Control* 2007;54:1642–7.

- [12] Wilson K, Homan K, Emelianov S. Biomedical photoacoustics beyond thermal expansion using triggered nanodroplet vaporization for contrast-enhanced imaging. *Nat Commun* 2012;3:618.
- [13] Kim K, Huang S-W, Ashkenazi S, O'Donnell M, Agarwal A, Kotov NA, et al. Photoacoustic imaging of early inflammatory response using gold nanorods. *Appl Phys Lett* 2007;90:223901.
- [14] Wei C, Lombardo M, Larson-Smith K, Pelivanov I, Perez C, Xia J, et al. Nonlinear contrast enhancement in photoacoustic molecular imaging with gold nanoparticle encapsulated nanoemulsions. *Appl Phys Lett* 2014;104:033701.
- [15] Farny CH, Wu T, Holt RG, Murray TW, Roy RA. Nucleating cavitation from laser-illuminated nano-particles. *Acoust Res Lett Online* 2005;6:138–43.
- [16] McLaughlan JR, Roy RA, Ju H, Murray TW. Ultrasonic enhancement of photoacoustic emissions by nanoparticle-targeted cavitation. *Opt Lett* 2010;35:2127–9.
- [17] Ju H, Roy RA, Murray TW. Gold nanoparticle targeted photoacoustic cavitation for potential deep tissue imaging and therapy. *Biomed Opt Express* 2013;4:66–76.
- [18] Larson-Smith K, Pozzo DC. Pickering emulsions stabilized by nanoparticle surfactants. *Langmuir* 2012;28:11725–32.
- [19] Larson-Smith K, Pozzo DC. Scalable synthesis of self-assembling nanoparticle clusters based on controlled steric interactions. *Soft Matter* 2011;7:5339.
- [20] Frens G. Controlled nucleation for the regulation of the particle size in monodisperse gold suspensions. *Nature* 1973;241:20–2.
- [21] Fattaccioli J, Baudry J, Émerard J-D, Bertrand E, Goubault C, Henry N, et al. Size and fluorescence measurements of individual droplets by flow cytometry. *Soft Matter* 2009;5:2232.
- [22] Draine BT, Flatau PJ. Discrete-dipole approximation for scattering calculations. *J Opt Soc Am A* 1994;11:1491.
- [23] Eisenmenger W, Staudenraus J. Fibre-optic probe hydrophone for ultrasonic and shock-wave measurements in water. *Ultrasonics* 1993;31:267–73.
- [24] Zhang P, Porter T. An *in vitro* study of a phase-shift nanoemulsion: a potential nucleation agent for bubble-enhanced HIFU tumor ablation. *Ultrasound Med Biol* 2010;36:1856–66.
- [25] Sheeran PS, Luo SH, Mullin LB, Matsunaga TO, Dayton PA. Design of ultrasonically-activatable nanoparticles using low boiling point perfluorocarbons. *Biomaterials* 2012;33:3262–9.
- [26] Kripfgans OD, Fowlkes JB, Woydt M, Eldevik OP, Carson PL. *In vivo* droplet vaporization for occlusion therapy and phase aberration correction. *IEEE Trans Ultrason Ferroelectr Freq Control* 2002;49:726–38.
- [27] Schad KC, Hynynen K. *In vitro* characterization of perfluorocarbon droplets for focused ultrasound therapy. *Phys Med Biol* 2010;55:4933.
- [28] Wei C, Xia J, Lombardo M, Perez C, Arnal B, Larson-Smith K, et al. Laser-induced cavitation in nanoemulsion with gold nanospheres for blood clot disruption: *in vitro* results. *Opt Lett* 2014;39:2599–602.
- [29] Perez C, Brayman A, Tu J, Swalwell J, Chen H, Matula T. Acoustic and optical characterization of ultrasound contrast agents via flow cytometry. *J Acoust Soc Am* 2012;132:1906.
- [30] Wu T, Farny CH, Roy RA, Holt RG. Modeling cavitation nucleation from laser-illuminated nanoparticles subjected to acoustic stress. *J Acoust Soc Am* 2011;130:3252–63.
- [31] (a) Wang L, Jacques SL, Zheng L. MCML – Monte Carlo modeling of light transport in multi-layered tissues. *Comput Methods Programs Biomed* 1995;47:131–46; (b) Krasovitski B, Kislev H, Kimmel E. Modeling photothermal and acoustical induced microbubble generation and growth. *Ultrasonics* 2007;47:90–101.
- [32] Sheeran PS, Wong VP, Luo S, McFarland RJ, Ross WD, Feingold S, et al. Decafluorobutane as a phase-change contrast agent for low-energy extravascular ultrasonic imaging. *Ultrasound Med Biol* 2011;37:1518–30.
- [33] Arnal B, Wei C-W, Xia J, Pelivanov IM, Lombardo M, Perez C, et al. Inertial cavitation in the nanostatic nanoemulsions with simultaneous pulsed laser and low frequency ultrasound excitation, vol. 8943. 2014. 89433E–89433E-8.



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