

## CROSSTALK

**CrossTalk proposal: Bradycardia in the trained athlete is attributable to high vagal tone**John H. Coote<sup>1</sup> and Michael J. White<sup>2</sup><sup>1</sup>*School of Clinical and Experimental Medicine, University of Birmingham, Birmingham B15 2TT, UK*<sup>2</sup>*School of Sport, Exercise and Rehabilitation Sciences, University of Birmingham, Birmingham B15 2TT, UK*

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Endurance trained athletes are well known to have low resting heart rates, with values below 30 beats min<sup>-1</sup> reported. Such low resting heart rates result from long periods of endurance training (Carter *et al.* 2003). We argue here that an increase in cardiac parasympathetic activity (vagal tone) is a major contributor.

The factors contributing to heart rhythm are complex but essentially there are three, the cardiac parasympathetic (vagal) nerves, the sympathetic nerves and the pacemaker cells, that set the intrinsic rate. At rest in healthy individuals the heart rate is determined by the balance between a high parasympathetic and low sympathetic activity changing around a fixed intrinsic pacemaker cell rhythm of around 105 beats min<sup>-1</sup> or lower depending on age (Jose & Collison, 1970). In humans and many large animals such as dogs, pigs and horses resting heart rate is lower than the intrinsic pacemaker rate. In healthy young humans heart rates of 60–70 beats min<sup>-1</sup> are common and atropine, given to block the effect of parasympathetic cholinergic nerves, results in increased heart rate to above the intrinsic rate (Maciel *et al.* 1985). This is not

the case in small animals such as guinea pigs, rats and mice, which have high sympathetic tone, and resting heart rates of 300–700 beats min<sup>-1</sup> are normal, with intrinsic rates from 170–500 beats min<sup>-1</sup> (Mohan *et al.* 2000; Danson & Paterson, 2003; D'Souza *et al.* 2014).

Studies to determine the underlying mechanisms causing the lowering of heart rate following endurance training have provided different explanations. This is partly due to interpretation of non-uniform methodology and partly to the difficulty of obtaining direct measurements of autonomic neural activity in humans. However, we consider there are clear indications that training bradycardia is largely caused by an augmentation of parasympathetic influence on the pacemaker cells. We are persuaded because it appears that neural pathways are necessary for post training resting bradycardia. For example, Ordway *et al.* (1982) showed that dogs with the cardiac autonomic nerves removed had no decrease in resting heart rate after endurance training compared to a control group, although other measures of the mild exercise training were similar in both groups. Furthermore, removal of baroreceptor influence by sinoaortic denervation in rats prevents an endurance training-induced increase in baroreflex–heart rate sensitivity and decrease in resting heart rate (Ceroni *et al.* 2009).

To appreciate the evidence it is essential to understand the organization of cardiac parasympathetic control.

**Cardiac vagal tone**

Cardiac parasympathetic nerves in dogs and cats have a low frequency (5 Hz) burst pattern of basal activity (Koizumi

*et al.* 1985) oscillating in phase with each cardiac beat and with a superimposed respiratory rhythm. Closer analysis of the cardiac preganglionic neurones in the brainstem by McAllen & Spyer (1978) has shown that the main generator of the activity is an excitatory input from the arterial baroreceptors via the nucleus of the tractus solitarius that fire during each heart beat. The effect of this excitatory input to cardiac preganglionic neurones is lessened during inspiration by an inhibitory input from nearby inspiratory neurones and increased during expiration (Gilbey *et al.* 1984). The magnitude of the respiratory-related rhythm is responsible for the fluctuations in heart rate known as respiratory sinus arrhythmia (RSA). Importantly there is a linear relationship between frequency of vagal activity and cardiac R–R interval (Parker *et al.* 1984) so that an increment of vagal firing would prolong the R–R interval by a fixed value independent of the initial R–R.

In humans indirect measurements of vagal tone based on RSA have been devised (e.g. Eckberg, 1983), developed and validated by animal studies (Katona & Jih, 1975). The most reliable indicator of vagal tone is that using frequency analysis of the cardiac beat known as power spectral analysis (PSA; Task Force of the European Society of Cardiology, North American Society of Pacing and Electrophysiology, 1996). PSA of heart rhythm reduces an R–R interval time series to its constituent sine wave frequency components. The spectral density of the signal describes the magnitude of the signal causing the variance at a particular frequency. It is the variation in the length of the inter-spike interval not the absolute length that is measured. In the spectrum the high frequency (HF) peak occurring at

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the individual's respiratory rate is abolished by atropine (Hollander & Bouman, 1975; Maciel *et al.* 1985) and by section of the vagus nerve in experimental animals (Ordway *et al.* 1982). A higher cardiac parasympathetic activity results in a more dominant respiratory rhythm so the HF peak is a robust surrogate for vagal tone (Fouad *et al.* 1984). However, it is only of significant quantitative value when changes in HF power are compared within a single individual and respiratory frequency and depth is controlled (Task Force, 1996).

With these caveats in mind a spectral analysis was done in a longitudinal study on a group of untrained young individuals who then underwent an aerobic endurance-training programme for 6 weeks. There was a marked and significant increase in HF power (Al-Ani *et al.* 1996) together with a significant decrease in resting heart rate. An enhanced vagal influence was further supported by measuring the immediate decrease in R–R interval in response to muscle contraction during a selected point in expiration and in inspiration. The immediate increase in heart rate caused by small muscle afferent fibres excited by muscle contraction occurs within the central nervous system. Therefore, their action reflects the excitability of neurones participating in the heart rate control circuits. Al-Ani *et al.* (1996) showed that the muscle-initiated decrease in R–R interval was significantly greater than the pre-training value. This is consistent with an increase in cardiac parasympathetic activity. The interpretation is supported by other studies showing that contraction-induced or exercise-induced immediate increase in heart rate is blocked by atropine (e.g. Maciel *et al.* 1985). The results accord with other longitudinal studies (Hottenrott *et al.* 2006). Thus these data strongly suggest an augmented contribution of cardiac parasympathetic nerve activity to training bradycardia.

Consistent with this Herrlich *et al.* (1960) showed that a training-induced decrease in resting heart rate in rats was associated with a considerable increase in atrial acetylcholine content. In accordance with this Paterson and his colleagues (Mohan *et al.* 2000; Danson & Paterson, 2003) have shown that presynaptic NO–cGMP-dependent acetylcholine release from cardiac parasympathetic terminals is enhanced in atria from exercise-trained mice. The cardiac parasympathetic NO–cGMP–cholinergic signalling pathway plays

an important role in the induction of bradycardia in guinea pigs, rats, mice, dogs and humans (Sears *et al.* 1998; Herring & Paterson, 2001, 2009; Chowdhary *et al.* 2002; Markos *et al.* 2002). Thus these studies clearly show that exercise-induced resting bradycardia is at least in part dependent on changes in parasympathetic activity.

A similar conclusion is reached by studies testing the efficacy or sensitivity of the baroreceptor–heart rate reflex. Activity in single fibres of the cardiac vagus nerve is increased by arterial baroreceptor stimulation (Kunze, 1972) and baroreflex sensitivity is increased in aerobic exercise-trained individuals (e.g. Lenard *et al.* 2005). Furthermore, removal of baroreceptor influence by sinoaortic denervation in rats prevents a training-induced increase in baroreflex–heart rate sensitivity (Ceroni *et al.* 2009).

#### Cardiac sympathetic activity

Using radiotracers to measure cardiac noradrenaline spillover from the coronary sinus in humans before and after endurance training, Meredith *et al.* (1991) found no significant change. However, skeletal muscle and kidney sympathetic activity is reduced following endurance training (Meredith *et al.* 1991; Grassi *et al.* 1994).

#### Concluding remarks

The foregoing evidence strongly suggests that resting bradycardia following endurance training is attributable to high cardiac vagal tone. However, we are aware that a study on rats by Bolter *et al.* (1973) showed that exercise training reduced the intrinsic heart rate in isolated atria. Boyett and colleagues (D'Souza *et al.* 2014) showed that this effect was due to down-regulation of the funny current  $I_f$ . Nonetheless, exercise training in rats also increases atrial acetylcholine, and vagal presynaptic NO–cGMP in trained mice. Therefore, it would seem reasonable to conclude that training-induced bradycardia in these small animals probably involves two mechanisms, an increase in cardiac parasympathetic activity and a decrease in intrinsic rate of pacemaker cells. In humans an increase in cardiac parasympathetic activity is a major contributor.

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#### References

- Al-Ani M, Munir SM, White M, Townend J & Coote JH (1996). Changes in R-R variability before and after endurance training measured by power spectral analysis and by the effect of isometric muscle contraction. *Eur J Appl Physiol* **74**, 397–403.
- Almeida MB & Araujo CGS (2003). Effects of aerobic training on heart rate. *Rev Bras Med Esporte* **9**, 1517–1530.
- Bolter CP, Hughson RL & Critz JB (1973). Intrinsic rate and cholinergic sensitivity of isolated atria from trained and sedentary rats. *Proc Soc Exp Biol Med* **144**, 364–367.
- Carter JB, Banister EW & Blaber AP (2003). Effect of endurance exercise on autonomic control of heart rate. *Sports Med* **33**, 33–46.
- Ceroni A, Chaar LJ, Bombein RL & Michelini LC (2009). Chronic absence of baroreceptor inputs prevents training-induced cardiovascular adjustments in normotensive and spontaneously hypertensive rats. *Exp Physiol* **94**, 630–640.
- Chowdhary S, Nuttall SL, Coote JH & Townend JN (2002). L-Arginine augments cardiac vagal control in healthy human subjects. *Hypertension* **39**, 51–56.
- Danson EJP & Paterson DJ (2003). Enhanced neuronal nitric oxide synthase expression is central to cardiac vagal phenotype in exercise trained mice. *J Physiol* **546**, 225–232.
- D'Souza A, Bucchi A, Johnsen AB, Logantha SJR, Monfredi O, Yanni J, Prehar S, Hardt G, Cartwright E, Wisloff U, Dobryznski H, DiFrancesco D, Morris GM & Boyett MR (2014). Exercise training reduces the resting heart rate via downregulation of the funny channel HCN4. *Nat Commun* **5**, 3775.
- Eckberg DL (1983). Human sinus arrhythmia as an index of vagal cardiac outflow. *J Appl Physiol* **54**, 961–966.
- Fouad FM, Tarazi RC, Ferrario CM, Fighaly S & Alicandri C (1984). Assessment of parasympathetic control of heart rate by a noninvasive method. *Am J Physiol Heart Circ Physiol* **246**, H838–H842.
- Gilbey MP, Jordan D, Richter DW & Spyer KM (1984). Synaptic mechanisms involved in the inspiratory modulation of vagal cardio-inhibitory neurones in the cat. *J Physiol* **356**, 65–78.

- Grassi G, Seravalle G, Calhoun DA & Mancia G (1994). Physical training and baroreceptor control of sympathetic activity. *Hypertension* **23**, 294–301.
- Herring N & Paterson DJ (2001). Nitric oxide-cGMP pathway facilitates acetylcholine release and bradycardia during vagal stimulation in the guinea-pig *in vitro*. *J Physiol* **535**, 507–518.
- Herring N & Paterson DJ (2009). Neuromodulators of peripheral cardiac sympatho-vagal balance. *Exp Physiol* **94**, 46–53.
- Herrlich HC, Raab W & Gige W (1960). Influence of muscular training and of catecholamines on cardiac acetylcholine and cholinesterase. *Arch Int Pharmacodyn Ther* **129**, 201–205.
- Hollander AP & Bouman LN (1975). Cardiac acceleration in man elicited by a muscle-heart rate reflex. *J Appl Physiol* **38**, 272–278.
- Hottenrott K, Hoos O & Esperer HD (2006). Heart rate variability and physical exercise. *Herz* **31**, 544–552.
- Jose AD & Collison D (1970). The normal range and determinants of the intrinsic heart rate in man. *Cardiovasc Res* **4**, 160–167.
- Katona PG & Jih F (1975). Respiratory sinus arrhythmia: noninvasive measure of parasympathetic cardiac control. *J Appl Physiol* **39**, 801–805.
- Koizumi K, Terui N & Kollai M (1985). Effect of cardiac vagal and sympathetic nerve activity on heart rate in rhythmic fluctuations. *J Auton Nerv Syst* **12**, 251–259.
- Kunze DL (1972). Reflex discharge patterns of cardiac vagal efferent fibres. *J Physiol* **222**, 1–15.
- Lenard Z, Studinger P, Mersich B, Pavlik G & Kollai M (2005). Cardiovascular autonomic function in sedentary and trained offspring of hypertensive parents. *J Physiol* **565**, 1031–1038.
- Maciel BC, Gallo JL, Marin NJA, Lima FEC, Terra FJ & Manco JC (1985). Parasympathetic contribution to bradycardia induced by endurance training in man. *Cardiovasc Res* **19**, 642–648.
- McAllen RM & Spyer KM (1978). The baroreceptor input to cardiac vagal motoneurons. *J Physiol* **282**, 365–374.
- Markos F, Snow HM, Kidd C & Conlon K (2002). Nitric oxide facilitates vagal control of heart rate via actions in the cardiac parasympathetic ganglia of the anaesthetised dog. *Exp Physiol* **87**, 49–52.
- Meredith IT, Friberg P, Jennings GL, Dewar EM, Fazio VA, Lambert GW & Esler MD (1991). Exercise training lowers resting renal but not cardiac sympathetic activity in humans. *Hypertension* **18**, 575–582.
- Mohan RM, Choate JK, Golding S, Herring N, Casadei B & Paterson DJ (2000). Peripheral pre-synaptic pathway reduces heart rate response to sympathetic activation following exercise training: role of NO. *Cardiovasc Res* **47**, 90–98.
- Parker P, Celler BG, Potter EK & McCloskey DI (1984). Vagal stimulation and cardiac slowing. *J Auton Nerv Syst* **11**, 226–231.
- Ordway GA, Charles JB, Randall DC, Billman GE & Wekstein DR (1982). Heart rate adaptation to exercise training in cardiac-denervated dogs. *J Appl Physiol Respir Environ Exerc Physiol* **52**, 1586–1590.
- Sears CE, Choate JK & Paterson DJ (1998). Effect of nitric oxide synthase inhibition on the sympatho-vagal control of heart rate. *J Auton Nerv Syst* **73**, 63–73.
- Task Force of the European Society of Cardiology, North American Society of Pacing and Electrophysiology (1996). Heart rate variability: standards of measurement, physiological interpretation and clinical use. *Circulation* **93**, 1043–1065.

#### Additional information

#### Competing interests

None declared.