MEK and PI3K inhibition in solid tumors: rationale and evidence to date

E. Jokinen and J.P. Koivunen

Abstract: PI3K-AKT-mTOR and Ras-Raf-MEK-ERK are the most commonly altered oncogenic pathways in solid malignancies. There has been a lot of enthusiasm to develop inhibitors to these pathways for cancer therapy. Unfortunately, the antitumor activities of single-agent therapies have generally been disappointing, excluding *B-Raf* mutant melanoma and renal cell cancer. Preclinical studies have suggested that concurrent targeting of the PI3K-AKT-mTOR and Ras-Raf-MEK-ERK pathways is an active combination in various solid malignancies. In the current work, we review the preclinical data of the PI3K and MEK dual targeting as a cancer therapy and the results of early-phase clinical trials, and propose future directions.

Keywords: inhibition, MEK, PI3K, solid tumors, targeted therapy

Constitutive activation of oncogenic pathways occurs in cancers with very high frequency. It is thought to be a central factor behind the hallmarks of cancer phenotype such as cell cycle progression, inhibition of apoptosis and reprogramming of metabolism. In solid malignancies, the PI3K-AKT-mTOR and Ras-Raf-MEK-ERK pathways are thought to play a central role in transmitting these oncogenic signals. Genetic alterations, such as receptor mutations or amplifications, or mutations in intermediate signal transducers, can lead to the constitutive activation of the pathways. The high frequency of genetic alterations causing constitutive activation of PI3K-AKT or Ras-Raf-MEK-ERK and the addiction of cancer cells on their signals have generated enthusiasm to develop pathway inhibitors.

Considering the central role of the pathways in transmitting upstream oncogenic signals, inhibition of these pathways could be an effective therapy in various cancer genotypes [Hanahan and Weinberg, 2011]. Despite the undisputed rationale of this approach, the clinical activity of the single pathway inhibition has been limited to B-Raf inhibition in *B-Raf* mutant melanoma and mammalian target of rapamycin (mTOR) inhibition in renal cell cancer [Solit *et al.* 2006; Hudes *et al.* 2007; Motzer *et al.* 2008]. The effectiveness of single pathway inhibition could be suppressed by *de novo* dependency on multiple signaling pathways or feedback activation of other signaling pathways [Faber *et al.* 2009; Chandarlapaty *et al.* 2010]. This has led to studies combining phosphoinositide-3 kinase (PI3K)/AKT/mTOR and Raf/mitogen-activated protein kinase kinase (MEK) inhibitors.

PI3K pathway

PI3Ks are divided by structure, regulation and lipid substrate specificity into three subclasses: class I, II and III. Of the three subclasses of PI3K, class I has been related to cancer. Class I PI3K is composed of a p110 catalytic subunit, coded by PIK3CA, PIK3CB, PIK3CD and PIK3CG genes, and a p85 regulatory subunit. The kinase activity of p110 is regulated downstream of receptor tyrosine kinases (RTKs) by the binding of tyrosine-phosphorylated proteins to its regulatory p85 subunit, resulting in reduction of its autoinhibitory activity. Furthermore, p110 has a Ras motif in which activated Ras proteins can bind leading to optimal lipid kinase activity. Activated PI3K produce phosphatidylinositol (3,4,5)-trisphosphate (PIP₃), which can activate downstream targets, such as AKT which further activates mTOR [Engelman et al. 2006].

Ther Adv Med Oncol

2015, Vol. 7(3) 170-180

DOI: 10.1177/ 1758834015571111

© The Author(s), 2015. Reprints and permissions: http://www.sagepub.co.uk/ journalsPermissions.nav

Correspondence to: Elina Jokinen, PhD Department of Oncology and Radiotherapy, Oulu University Hospital, Oulu, Finland, PB20, 90029 OYS elinamirjamijokinen@ gmail.com

Jussi P. Koivunen, MD, PhD Department of Oncology and Radiotherapy, Oulu University Hospital, Oulu, Finland

The PI3K-AKT-mTOR signaling pathway has an important role in cell survival and proliferation, and constitutive activation of the pathway is common in solid malignancies. The hyperactivation of the pathway is most frequently caused by the activation of RTKs, somatic mutations in specific signaling components of the pathway such as *PIK3CA*, or the loss of *PTEN* tumor suppressor. PIK3CA mutations are frequently detected in endometrial, breast, lung, urinary tract, colorectal, gliomas and gastric cancers [Cancer Genome Atlas Research Network, 2008, 2012a, 2012b, 2012c, 2013, 2014a, 2014b, 2014c] while amplification of the gene is commonly seen in head and neck, lung, gastric and ovarian cancers [Lin et al. 2005; Cancer Genome Atlas Research Network, 2011, 2012a, 2014a]. Genetic PTEN loss occurs recurrently in endometrial, prostate, gliomas, lung and breast cancers [Cancer Genome Atlas Research, 2008, 2012, 2012, 2013; Bismar et al. 2011] (Table 1, Figure 1).

PI3K-pathway inhibitors

Several small molecular inhibitors targeting the PI3K pathway have been developed, including dual PI3K–mTOR-, PI3K-, AKT- and mTOR inhibitors. Allosteric, rapalog analog mTOR inhibitors were the first to enter clinical trials but, unfortunately, showed activity only in a limited number of solid tumor types. Limited clinical activity of the rapalogs has been suggested to result from various drug-induced feedback loops leading to mTORC2-IRS-1 mediated hyperactivation of PI3K-AKT [O'Reilly *et al.* 2006].

Rapalog-induced PI3K-AKT hyperactivation has led to the development of upstream inhibitors, which could overcome the hyperactivation and have more clinical activity [Wander *et al.* 2011]. These upstream inhibitors include dual PI3KmTOR (BEZ235), pan PI3K (GDC-0941 and BKM120) and AKT inhibitors (MK-2206, GDC-0068) (Table 2). Of these drugs, most fall into adenosine triphosphate (ATP) competitive class, while both ATP and non-ATP competitive AKT inhibitors do exist. Recently, a newer generation of inhibitors has entered clinical trials including isoform specific PI3K inhibitors and ATP competitive inhibitors of mTORC1/2.

Currently, only the rapalog inhibitors of the PI3K-AKT-mTOR pathway have entered the clinic for the treatment of cancer. Rapalogs have previously been widely tested in various solid

malignancies but have shown clinical activity in a limited number of indications. Everolimus has been approved for renal cell cancer (RCC) and neuroendocrine cancers as a single-agent and for breast cancer in combination with hormonal therapy [Motzer et al. 2008; Yao et al. 2011; Baselga et al. 2012]. Another mTOR inhibitor, temsirolimus, has also been approved to the treatment of RCC [Hudes et al. 2007]. In contrast to most solid malignancies, activation of hypoxia-inducible factor 1-alpha (HIF-1 α) is a central oncogenic event in RCC, which might relate to the clinical activity of the rapalogs in this disease. mTOR is an indirect upstream regulator of HIF-1 α and therefore may represent an easier clinical target for mTOR inhibition compared with most solid malignancies in which the RTK pathways upstream of mTOR dominate in oncogenesis [Bellmunt et al. 2013].

Numerous clinical trials are testing the safety and efficiency of dual PI3K-mTOR, PI3K and AKT inhibitors in various solid malignancies. Treatment with these inhibitors seems to be well tolerated with dose-limiting toxicities (DLTs) from hyper-glycemia, rash, gastrointestinal side effects and stomatitis [Dienstmann *et al.* 2011; Bran and Siu, 2012]. Various methods have been used to analyze the dose–effect relationship of these drugs, such as the measurement of biomarkers pAKT and pS6 in cells and tissues. With the maximum tolerated dose (MTD), inhibitors induce 30–90% inhibition of pAKT and/or pS6 biomarkers [Bendell *et al.* 2011; Moreno Garcia *et al.* 2011].

Based on early-phase clinical trials, the singleagent clinical activity of PI3K-mTOR, PI3K and AKT inhibitors seems to be modest in unselected solid malignancies. The drugs appear to be more active in tumors bearing genetic alterations in PIK3CA or PTEN, but no clear correlation between genotype and response have been established [Brachmann et al. 2009; Weigelt et al. 2011]. General lack of clinical benefit could result from various factors, such as limited target inhibition or feedback activation of other pathways. As low as 30% target inhibition has been reported in clinical trials and it is unlikely that this magnitude of inhibition could result in clinical activity. Furthermore, feedback activation of other pathways has been reported in preclinical studies but general landscape of this phenomenon has not been characterized. Preclinical studies with PI3K and AKT inhibitors have shown that these drugs induce feedback activation of the extracellular-signal-regulated kinase (ERK) and human epidermal growth factor receptor (HER) family, which at least partly results in decreased anticancer activity [Chandarlapaty *et al.* 2010; Serra *et al.* 2011]. Similar feedback activation phenomenon of the ERK and HER family has also been recapitulated in clinical trials of the agents [Yan *et al.* 2013].

Ras-Raf-MEK-ERK pathway

The binding of cytokines, growth factors or mitogens to their receptors causes activation of the Shc/Grb2/SOS coupling complex, which in turn stimulates the inactive Ras (H-, K, N-isotypes). Stimulated Ras exchanges guanosine diphosphate (GDP) to guanosine triphosphate (GTP) and undergoes conformational change to active stage. Ras proteins have an intrinsic GTPase activity, required for inactivation of the GTP bound active state. The GTPase activity of Ras protein is strongly enhanced by binding of GTPase proteins such as nuclear factor 1 (NF1). Ras proteins have multiple downstream targets, with Raf kinase being the most well characterized. Raf kinase activates MEK1/2, which catalyzes activation of ERK1/2. Activation of ERK1/2 further phosphorylates a series of downstream targets involved in various cellular processes such as proliferation, differentiation, cell survival and angiogenesis.

Ras-Raf-MEK-ERK hyperactivation is very frequently evident in solid malignancies and is transforming in multiple cancer models. In cancers, activation of Ras-Raf-MEK-ERK occurs through various mechanisms. Hyperactivation of RTKs can induce constitutive activation of the pathway. Furthermore, genetic alterations in the pathway members are common mechanisms behind the hyperactivation. Ras mutations or loss of NF1 cause activation of the pathway at the level of Ras. Ras mutations are commonly found in pancreatic, colorectal, lung, and endometrial cancers [Almoguera et al. 1988; Cancer Genome Atlas Network, 2012b, 2013, 2014c] and NF1 mutations are seen in gliomas, sarcomas and lung cancers [Cancer Genome Atlas Research, 2008, 2014c; Barretina et al. 2010]. Mutations in the Raf genes are another level where genetic constitutive activation occurs repeatedly, while MEK or ERK mutations are rare. Raf mutations are frequent in melanoma and thyroid cancers [Davies et al. 2002; Kimura et al. 2003] (Table 1, Figure 1).



cell proliferation, growth and survival

Figure 1. Signaling of PI3K-AKT-mTOR and Ras-Raf-MEK-ERK pathways and inhibitors currently tested in cancer clinical trials. Signaling molecules are drawn with circles and inhibitors with boxes. Stars indicate signaling molecules that are frequently altered in solid malignancies.

AKTi, AKT inhibitor; DUALi, dual PI3K; ERK, extracellularsignal-regulated kinase; MEKi, mitogen-activated protein kinase kinase inhibitor; mTORi, mammalian target of rapamycin inhibitor; NF1, neurofibromin; PI3Ki, phosphoinositide-3 kinase inhibitor; RAFi, RAF inhibitor; RTK, receptor tyrosine kinase.

Inhibitors of Ras-Raf-MEK-ERK pathway

Considerable efforts have been made to develop Ras targeted drugs, such as farnesyl transferase inhibitors, but so far clinical trials have been unsuccessful [Sousa *et al.* 2008]. Various Raf inhibitors have been developed, of which sorafenib was the first to enter the clinic in the treatment of renal and hepatocellular cancers [Escudier *et al.* 2007; Llovet *et al.* 2008]. Even though sorafenib is an inhibitor of wildtype and mutant *B-Raf*, its clinical activity is limited in B-Raf driven cancers such as melanoma [Eisen *et al.* 2006; Gollob *et al.* 2006]. It is likely that the MTD of sorafenib does not lead to meaningful inhibition of mutant B-Raf [<10% by phosphorylated ERK (pERK) assessment] [Davies *et al.* 2012].

The clinical activity of sorafenib in renal and hepatocellular cancers is largely related to the other kinases inhibited by it such as VEGFR2-3, FGFR-1, PDGFR-b, Flt-3 and c-Kit [Wilhelm *et al.* 2008]. Later, inhibitors preferentially targeting mutant B-Raf, such as vemurafenib and dabrafenib, were developed. These inhibitors induce marked downregulation of B-Raf signaling and >80% inhibition of pERK in tumors correlates with responses [Bollag *et al.* 2010]. Dabrafenib

Gene	Cancer	Alteration
РІЗКСА		Mutation
	Endometrial	51%
	Breast	25%
	Lung (SQC)	16%
	Urinary tract	15%
	Glioma	15%
	Colon	12%
	Gastric	12%
РІЗКСА		Amplification
	Head and neck	43%
	Ovarian	18%
	Lung (SQC)	12%
	Gastric	12%
PTEN		Inactivation
	Endometrial	59%
	Prostate	42%
	Glioma	36%
	Lung (SQC)	15%
	Breast	10%
K-RAS		Mutation
	Pancreatic	95%
	Colorectal	43%
	Lung (AC)	32%
	Endometrial	18%
NF1		Inactivation
	Glioma	14%
	Sarcoma	11%
	Lung (AC)	11%
B-Raf		Mutation
	Melanoma	66%
	Thyroid (PAP)	36%
AC, adenocarcinoma; PAP, papillary; SCC, squamous cell carcinoma.		

Table 1. Frequencies of genetic alterations leading to activation of PI3K-AKT-mTOR and RAS-Raf-MEK-ERK pathways in specific solid cancers.

and vemurafenib have significant clinical activity in *B-Raf* mutant melanoma and have been approved for this indication [Chapman *et al.* 2011; Hauschild *et al.* 2012]. Conversely, mutant B-Raf targeted inhibitors paradoxically hyperactivate MEK/ERK signaling in cells with wildtype protein and therefore do not have antitumor activity in *B-Raf* wildtype tumors. Paradoxical hyperactivation of signaling by B-Raf inhibitors has been linked to increased C-Raf dimerizationmediated activation of downstream targets MEK/ ERK [Poulikakos *et al.* 2010]. Paradoxical MEK/ ERK hyperactivation in patients undergoing B-Raf inhibitor therapy can be manifested by presentation of skin squamous cell cancers, keratoachantomas or even leukemias. These clinical manifestations strongly associate with presence of *Ras* mutations in them [Callahan *et al.* 2012; Su *et al.* 2012].

MEK inhibitors are classed into ATP competitive and ATP non-competitive agents. Most of the known MEK inhibitors fall into the latter category, and hence are not directly competing for the ATP-binding site but bind to a unique allosteric binding site adjacent to the ATP binding site. Thus, non-ATP competitive MEK inhibitors have high specificity for the target [Wallace *et al.* 2005]. CI-1040 was the first allosteric MEK1/2 inhibitor to enter clinical trials. CI-1040 was shown to be well tolerated but possessed insufficient antitumor activity in patients [Rinehart et al. 2004]. Most of the newer MEK inhibitors following CI-1040 share a similar chemical structure but are more potent inhibitors of the target. These inhibitors include agents such as trametinib, selumetinib, GDC-0973 and BAY869766. Treatment with these agents seems to be well tolerated with DLTs from rash, diarrhea and periferial edema [Adjei et al. 2008; Bennouna et al. 2011; Infante et al. 2012; Lorusso et al. 2012]. Various methods have been used to analyze the dose-effect relationship of the drugs, such as measurement of pERK levels in normal tissues and tumors. With MTDs, these inhibitors induce >80% reduction of pERK levels in various tissues [Adjei et al. 2008; Weekes et al. 2013]. Even if true with B-Raf inhibitors, it is unknown whether MEK inhibitor induced target inhibition of >80% surpasses the required level for cytotoxicity.

MEK inhibitors are currently tested as single agents or in combination with other targeted agents or chemotherapy. Single-agent activity has been seen in *Ras* and *Raf* mutant patients [Zimmer *et al.* 2014]. The most promising results are in thyroid cancer in combination with I-131 and in *B-Raf* mutant melanoma in combination with B-Raf inhibitors [Flaherty *et al.* 2012; Ho *et al.* 2013] (Table 2).

Dual inhibition

Even though PI3K-AKT-mTOR and Ras-Raf-MEK-ERK are the most commonly altered signaling pathways in solid malignancies, the clinical efficiency of single pathway inhibitors has generally been disappointing with some exclusion such as B-Raf mutant melanoma or renal cell cancer. Cancers can be de novo dependent concurrently on these two parallel pathways and cross-signaling of the pathways is also evident, such as direct K-Ras mediated activation of PI3K [Faber et al. 2009; Chandarlapaty et al. 2010]. Many in vivo and in vitro studies have shown that PI3K-AKTmTOR and Ras-Raf-MEK-ERK pathways regulate each other's activity through feedback mechanisms, i.e. MEK inhibition causes PI3K/ Akt activation via ERBB receptors [Turke et al. 2012]. Furthermore, PI3K-AKT-mTOR and Ras-Raf-MEK-ERK have also been shown to coregulate shared downstream targets including forkhead box transcription factor class O (FOXO), Bcl-2-associated death promoter (BAD) and glycogen synthase kinase 3 (GSK3) [Mendoza et al.

2011]. The interaction of the parallel PI3K-AKTmTOR and Ras-Raf-MEK-ERK signaling pathways is thought to explain the inefficiency of single-agent treatments in most malignancies and gives rationale for the concurrent targeting of both pathways.

To date, most efforts to dual target the PI3K-AKT-mTOR and Ras-Raf-MEK-ERK pathways have been carried out with a combination of PI3K and MEK inhibitors. However, many studies with mTOR and AKT inhibitors do also exist. PI3K targeting agents are theoretically the most potent inhibitors of the pathway since they lack the downstream feedback activation of mTOR inhibitors [O'Reilly et al. 2006]. Furthermore, PI3K is also known to have other important, cancer-associated downstream targets than AKT making PI3K inhibitors more potent than AKT inhibitors from that perspective [Vasudevan et al. 2009]. Allosteric MEK inhibitors are highly specific for their targets and are known to also induce high inhibition of the pathway in patients [Adjei et al. 2008; Weekes et al. 2013], while the current RAF inhibitors are significantly less effective in inhibiting downstream signaling in tumors with wildtype Raf.

Preclinical models have shown that dual targeting with PI3K and MEK inhibitors possesses antitumor activity in various cancer models and genotypes. Activity is also seen in difficult to treat genotypes, such as Ras mutant tumors and basal like breast cancer [Engelman et al. 2008; Hoeflich et al. 2009; Sos et al. 2009]. Even though Ras is generally thought to be a direct upstream activator of Raf, responses to MEK inhibitors are variable in Ras mutant cancer models. Variable responses could be explained by Ras mediated activation of other signaling pathways such as PI3K and, therefore, dual targeting with MEK and PI3K inhibitors is potentially more effective [Mendoza et al. 2011]. Some models, such as basal-like breast cancer, have shown that treatment with MEK inhibitors induces feedback activation of RTKs which signal downstream to PI3K and therefore, dual targeting is more active [Hoeflich et al. 2009]. Many preclinical studies have investigated predictive factors for dual PI3K and MEK inhibitor therapy [Sos et al. 2009; Jokinen et al. 2012]. Even though some studies have suggested that responses are commonly seen in some cancer genotypes, no clear predictive markers have been found. Existence of measurable predictive factors would make the design and execution of clinical trials more fruitful.

Table 2. Examples of PI3K and MEK inhibitors tested in clinic.

PI3K inhibitors	
GDC-0941	GDC-0941 is a potent and selective, orally bioavailable class I PI3K inhibitor which inhibits the growth of a wide of human tumor cell lines. The compound has shown strong inhibition of the growth of IGROV-1 ovarian cancer and U87M glioblastoma xenografts. GDC-0941 has already been tested in clinical trials.
BEZ235	The imidazo[4,5- <i>c</i>]quinoline derivative BEZ235 is a dual PI3K-mTOR inhibitor. The compound blocks abnormal PI3K activation causing G1 cell cycle arrest. <i>In vivo</i> studies showed BEZ235 to be well tolerated and suitable for combinations studies. The inhibitor has entered clinical trials.
ВКМ120	The 2-morpholino pyrimidine derivative BKM120 is a selective pan-PI3K inhibitor which inhibits all four class I isoforms of PI3K. The compound preferentially inhibits tumor cells with PIK3CA mutations over KRAS and PTEN mutated tumor cells. The biologic, pharmacological and preclinical safety profile of BKM120 h support its clinical use and the compound has been tested in phase II clinical trials with cancer patients.
MEK inhibitors	
CI-1040	CI-1040 (PD-184352) is a specific small-molecule drug that inhibits MEK1/MEK2. The compound has been suggested to act as an allosteric inhibitor of MEK since it is known not to compete with the binding of ATP or protein substrates. CI-1040 blocks ERK phosphorylation and inhibits the growth of multiple human tumor cell lines as well as tumor growth in xenograft models. It has been shown that the cell growth inhibitory effect of CI-1040 is rapidly reversed after it is removed from the growth media. CI-1040 was the first MEK inhibitor to enter clinical trials where it was shown to be well tolerated, but was found to have insufficient antitumor activity in patients. PD-0325901 is a CI-1040 derivate with higher solubility and improved pharmacological properties currently being tested in clinical trials.
GDC-0973	GDC-0973 (XL518) is methanone derived, potent, orally bioavailable MEK1/2 inhibitor which has a 100-fold MEK1 selectivity over MEK2. The compound has shown tumor growth inhibition in BRAF and KRAS mutated cancer cell lines <i>in vivo</i> and <i>in vitro</i> .
Trametinib	Trametinib (GSK1120212), a selective MEK1 and MEK2 inhibitor, dephosphorylates phosphorylated MEK and also stabilizes the unphosphorylated form, thereby blocking the downstream signaling. At the moment, it is the only MEK inhibitor, which has showed promising results in phase III clinical trials.
Pimasertib	Pimasertib (AS703026, MSC1936369B) is an orally available, selective second generation inhibitor of MEK1/2 which binds ATP non-competitively to the distinctive allosteric site of MEK1/2 causing G0–G1 cell cycle arrest and apoptosis. Pimasertib has been shown to cause significant tumor regression in human multiple myeloma and KRAS mutated colorectal cancer xenografts.

ATP, adenosine triphosphate; ERK, extracellular-signal-regulated kinase; MEK, mitogen-activated protein kinase kinase; PI3K, phosphoinositide-3 kinase.

Dual inhibition: clinical studies

Numerous early-phase clinical studies investigating dual PI3K and MEK targeting are ongoing and some results have been presented in meetings during the past three years. Because of the heterogeneity and early-phase nature of the studies, it is difficult to derive a strong conclusion of the activity in various cancer subgroups and general toxicities. Therefore, we review four different phase I clinical studies and present their results.

Generally, combined PI3K and MEK inhibitor therapy seems to be feasible with manageable safety and toxicity profile. The most common adverse events (AEs) of therapy include diarrhea, rash, fatigue, vomiting and hyperglycemia. Unfortunately, the rate of response seems to be quite low with an overall response rate of 4.7%and a disease control rate of 19.2% when all four presented studies are combined. As in the preclinical models, no clear predictive factors for response have been identified. However, responses have been seen in *Ras or Raf* mutated cancers, and later studies have enriched these patient populations.

The clinical activity of GDC-0973 (MEK1/2 inhibitor) and GDC-0941 (class I PI3K inhibitor)

was studied in 78 patients with advanced solid tumors. The patients received daily both GDC-0973 and GDC-0941 with a 21 day on, 7 days off schedule or with an intermittent schedule where GDC-0973 was dosed on days 1, 4, 8, 11, 15 and 18 with a 28 day cycle and GDC-0941 daily with a 21/7 schedule. DLTs included elevation of grade 3 lipase and grade 4 creatine phosphokinase (CPK). Reported AEs were diarrhea, fatigue, nausea, rash, vomiting, dysgeusia, decreased appetite and CPK elevation. The combination therapy was found to be well tolerated, with toxicities comparable with those reported with single agents in phase I trials. Higher drug doses were tolerated with the intermitted dosing schedule. Partial responses were seen in three patients: one patient with B-Raf mutated melanoma, one with B-Raf mutated pancreatic cancer, and one with K-Ras mutated endometrial cancer. Stable disease lasting over 5 months was observed in five patients [Lorusso et al. 2012].

Daily dosing of BKM120 (pan-PI3K inhibitor) and trametinib (MEK inhibitor) was evaluated with 49 patients with advanced *Ras-* or *B-Raf*mutant cancers. The combination was found to be safe on patients. Grade 3 DLTs included stomatitis, dyspahgia, ejection fraction decrease, CPK elevation, nausea and anorexia. The most commonly observed AEs were listed as dermatitis, diarrhea, nausea, vomiting, rash, asthenia, increase in CPK, loss of appetite, pyrexia, stomatitis and hyperglycemia. Partial responses were seen in three patients with *K-Ras* mutant ovarian cancer and stable disease was observed with two patients with *B-Raf* mutated melanoma [Bedard *et al.* 2012].

In another combination study, 49 patients were treated with the pan-PI3K inhibitor copanlisib and the MEK inhibitor refametinib. DLTs included grade 3 aspartate transaminase (AST) or alanine aminotransferase (ALT) elevation, hyperglycemia, hypertension, diarrhea, mucositis and rash. The most common AEs included diarrhea, nausea, hyperglycemia, fatigue, rash, anorexia and hypertension. One partial response was seen in a patient with endometrial cancer, and stable disease lasting >4 cycles in 9 patients [Ramanathan *et al.* 2014].

The combination of BYL719 (PI3K α inhibitor) and binimetinib (MEK inhibitor) was studied in patients with advanced solid tumors with *Ras* or *B-Raf* mutations. A total of 58 patients were

treated once a day with BYL719 (80–270 mg) and twice a day with 30 or 45 mg of binimetinib. Common AEs were diarrhea, nausea, vomiting, reduced appetite, rash, pyrexia, fatigue and hyper-glycemia. Confirmed partial responses were seen with three of the four patients with *K-Ras*-mutant ovarian cancer, with one patient with *N-Ras* mutated melanoma. Stable disease status lasting for over 6 weeks was seen with 18 patients [Juric *et al.* 2014].

Further directions

A strong rationale and preclinical results have established the groundwork for the clinical development of dual PI3K and MEK inhibitor therapy to solid malignancies. However, early-phase clinical trials presented to date have only shown modest activity of the combination. There is room for new ideas and approaches to improve the antitumor activity and tolerability of the dual PI3K and MEK inhibitor therapy.

Optimal dosing schedule for PI3K and MEK inhibitor combination is unknown. Current clinical dosing regimens of PI3K and MEK inhibitors mainly investigate longer drug exposures with subtotal suppression of their targets. It is likely that more robust target inhibition with higher drug doses would have more antitumor activity. It is possible that higher drug doses with short exposure times could be clinically as tolerable as longer exposure with lower doses, but poses more antitumor activity. Some preclinical studies have shown that short, alternative dosing schedules of PI3K and MEK inhibitor combination can be as effective as longer exposures [Hoeflich et al. 2012; Jokinen et al. 2012]. Furthermore, it is currently unknown if both PI3K and MEK inhibitors need to be administered concurrently for the same period of time, or if either of the drugs could be used intermittently. There is some preclinical evidence for the intermittent uses of the drugs and some clinical studies have also tried to answer this question [Hoeflich et al. 2012; Jokinen et al. 2012; Lorusso et al. 2012]. We believe that studying numerous, highly dual PI3K and MEK therapysensitive preclinical models could be a useful way to figure out the optimal dosing schedules from the perspective of cytotoxicity. The dosing regimens could be brought forward for early-phase clinical trials to test their safety and toxicity.

An open question remains if the current PI3K and MEK inhibitors are optimal drugs for cancer

therapy. Allosteric MEK inhibitors are well-tolerated agents and do induce robust target inhibition with daily MTD [Adjei et al. 2008; Weekes et al. 2013]. Based on the clinical trials, current PI3K inhibitors are only able to induce modest target inhibition (30-90% inhibition of pAKT and pS6 biomarkers) with daily MTD [Bendell et al. 2011; Moreno Garcia et al. 2011]. Isoform-specific PI3K inhibitors could have less off-tumor effects and, therefore, enable more robust target inhibition with MTD. Promising phase I results of this approach (PI3K α and MEK inhibitor combination) have recently been presented [Juric et al. 2014]. Furthermore, it is possible that blocking other members of the PI3K-AKT-mTOR pathway could be more effective and tolerable than inhibiting PI3K itself.

Enriching the therapy-sensitive patients in clinical trials is an efficient way to speed up the development of an agent or a combination. Despite the robust preclinical and clinical efforts, no clear predictive factors for dual PI3K and MEK inhibitor therapy have been identified [Sos *et al.* 2009; Bedard *et al.* 2012; Jokinen *et al.* 2012; Lorusso *et al.* 2012]. From this perspective, it is unlikely that an easily measurable predictive factor to the dual therapy will be identified.

Cytotoxic drug combinations are the backbone of the current cancer therapy. Much less is known about targeted therapy combinations. It is possible that combining either cytotoxics or targeted agents to PI3K and MEK therapy could increase the number of tumors sensitive to them. As an example, novel preclinical works presented recently that antitumor activity of a MEK or mTOR inhibitors can be markedly increased by blocking of BCL-XL anti-apoptotic protein by BH3 mimetic drugs [Corcoran et al. 2012; Faber et al. 2014]. We feel that large-scale preclinical modeling would be appropriate to try to identify the best combinations for 'dual or triple inhibition' therapy. However, it remains highly challenging to bring two or three investigational agents concurrently to clinical trials.

Summary

PI3K-AKT-mTOR and Ras-Raf-MEK-ERK are the most commonly altered signaling pathways in solid malignancies, but the clinical efficiency of single-pathway inhibitors have generally been disappointing. Failure of the single-pathway targeting could result from various factors, such as feedback activation of the other pathways or shared downstream targets. Therefore, a strong rationale for dual PI3K and MEK inhibitor therapy for cancer treatment exists. Various preclinical models have identified this therapy to be efficient in various cancers and genotypes. Currently, numerous earlyphase clinical trials investigating dual PI3K and MEK therapy are ongoing. Clinical results presented to date have shown that dual targeting is feasible with manageable toxicity and safety profile, but the rate of response is quite low. It is possible that efficiency of dual targeting could be increased by alternative dosing schedules, newer generations of agents, intelligent drug combinations, or enriching the patients with predictive factors.

Funding

The study was supported by Cancer Society of Northern Finland, Emil Aaltonen Foundation, Finnish Foundation for Tuberculosis Resistance, Finnish Oncological Society, Oulu University Hospital, Orion-Farmos Science Foundation, Sigrid Juselius Foundation and Thelma Mäkikyrö foundation.

Conflict of interest statement

The authors declare no conflicts of interest in preparing this article.

References

Adjei, A., Cohen, R., Franklin, W., Morris, C., Wilson, D., Molina, J. *et al.* (2008) Phase I pharmacokinetic and pharmacodynamic study of the oral, small-molecule mitogen-activated protein kinase kinase 1/2 inhibitor AZD6244 (ARRY-142886) in patients with advanced cancers. *J Clin Oncol* 26: 2139–2146.

Almoguera, C., Shibata, D., Forrester, K., Martin, J., Arnheim, N. and Perucho, M. (1988) Most human carcinomas of the exocrine pancreas contain mutant C-K-Ras genes. *Cell* 53: 549–554.

Barretina, J., Taylor, B., Banerji, S., Ramos, A., Lagos-Quintana, M., Decarolis, P. *et al.* (2010) Subtypespecific genomic alterations define new targets for soft-tissue sarcoma therapy. *Nat Genet* 42: 715–721.

Baselga, J., Campone, M., Piccart, M., Burris, H., 3rd, Rugo, H., Sahmoud, T. *et al.* (2012) Everolimus in postmenopausal hormone-receptor-positive advanced breast cancer. *N Engl J Med* 366: 520–529.

Bedard, P., Tabernero, J., Kurzrock, R., Britten, C., Stathis, A., Perez-Garcia, J. *et al.* (2012) A phase Ib, open-label, multicenter, dose-escalation study of the oral PAN-PI3K inhibitor BKM120 in combination with the oral MEK1/2 inhibitor GSK1120212 in patients (pts) with selected advanced solid tumors. \mathcal{J} *Clin Oncol* 30(Suppl.): abstract 3003.

Bellmunt, J., Teh, B., Tortora, G. and Rosenberg, J. (2013) Molecular targets on the horizon for kidney and urothelial cancer. *Nat Rev Clin Oncol* 10: 557–570.

Bendell, J., Rodon, J., Burris, H., De Jonge, M., Verweij, J., Birle, D. *et al.* (2011) Phase I, doseescalation study of BKM120, an oral pan-class I PI3K inhibitor, in patients with advanced solid tumors. *J Clin Oncol* 30: 282–290.

Bennouna, J., Lang, I., Valladares-Ayerbes, M., Boer, K., Adenis, A., Escudero, P. *et al.* (2011) A phase II, open-label, randomised study to assess the efficacy and safety of the MEK1/2 Inhibitor AZD6244 (ARRY-142886) *versus* capecitabine monotherapy in patients with colorectal cancer who have failed one or two prior chemotherapeutic regimens. *Invest New Drugs* 29: 1021–1028.

Bismar, T., Yoshimoto, M., Vollmer, R., Duan, Q., Firszt, M., Corcos, J. *et al.* (2011) PTEN genomic deletion is an early event associated with ERG gene rearrangements in prostate cancer. *BJU Int* 107: 477–485.

Bollag, G., Hirth, P., Tsai, J., Zhang, J., Ibrahim, P., Cho, H. *et al.* (2010) Clinical efficacy of a RAF inhibitor needs broad target blockade in *BRAF*-mutant melanoma. *Nature* 467: 596–599.

Brachmann, S., Hofmann, I., Schnell, C., Fritsch, C., Wee, S., Lane, H. *et al.* (2009) Specific apoptosis induction by the dual PI3K/mTOR inhibitor NVP-BEZ235 in HER2 amplified and PIK3CA mutant breast cancer cells. *Proc Natl Acad Sci U S A* 106: 22299–22304.

Brana, I. and Siu, L. (2012) Clinical development of phosphatidylinositol 3-kinase inhibitors for cancer treatment. *BMC Med* 10: 161.

Callahan, M., Rampal, R., Harding, J., Klimek, V., Chung, Y., Merghoub, T. *et al.* (2012) Progression of RAS-mutant leukemia during raf inhibitor treatment. *N Engl J Med* 367: 2316–2321.

Cancer Genome Atlas Research Network (2008) Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* 455: 1061–1068.

Cancer Genome Atlas Research Network (2011) Integrated genomic analyses of ovarian carcinoma. *Nature* 474: 609–615.

Cancer Genome Atlas Research Network (2012a) Comprehensive genomic characterization of squamous cell lung cancers. *Nature* 489: 519–525.

Cancer Genome Atlas Network (2012b) Comprehensive molecular characterization of human colon and rectal cancer. *Nature* 487: 330–337. Cancer Genome Atlas Network (2012c) Comprehensive molecular portraits of human breast tumours. *Nature* 490: 61–70.

Cancer Genome Atlas Research Network (2013) Integrated genomic characterization of endometrial carcinoma. *Nature* 497: 67–73.

Cancer Genome Atlas Research Network (2014a) Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* 513: 202–209.

Cancer Genome Atlas Research Network (2014b) Comprehensive molecular characterization of urothelial bladder carcinoma. *Nature* 507: 315–322.

Cancer Genome Atlas Research Network (2014c) Comprehensive molecular profiling of lung adenocarcinoma. *Nature* 511: 543–550.

Chandarlapaty, S., Sawai, A., Scaltriti, M., Rodrik-Outmezguine, V., Grbovic-Huezo, O., Serra, V. *et al.* (2010) AKT inhibition relieves feedback suppression of receptor tyrosine kinase expression and activity. *Cancer Cell* 19: 58–71.

Chapman, P., Hauschild, A., Robert, C., Haanen, J., Ascierto, P., Larkin, J. *et al.* (2011) Improved survival with vemurafenib in melanoma with BRAF V600E mutation. *N Engl J Med* 364: 2507–2516.

Corcoran, R., Cheng, K., Hata, A., Faber, A., Ebi, H., Coffee, E. *et al.* (2012) Synthetic lethal interaction of combined BCL-XL and MEK inhibition promotes tumor regressions in KRAS mutant cancer models. *Cancer Cell* 23: 121–128.

Davies, H., Bignell, G., Cox, C., Stephens, P., Edkins, S., Clegg, S. *et al.* (2002) Mutations of the BRAF gene in human cancer. *Nature* 417: 949–954.

Davies, M., Fox, P., Papadopoulos, N., Bedikian, A., Hwu, W., Lazar, A. *et al.* (2012) Phase I study of the combination of sorafenib and temsirolimus in patients with metastatic melanoma. *Clin Cancer Res* 18: 1120–1128.

Dienstmann, R., Rodon, J., Markman, B. and Tabernero, J. (2011) Recent developments in anticancer agents targeting PI3K, AKT and mTORC1/2. *Recent Pat Anticancer Drug Discov* 6: 210–236.

Eisen, T., Ahmad, T., Flaherty, K., Gore, M., Kaye, S., Marais, R. *et al.* (2006) Sorafenib in advanced melanoma: a phase II randomised discontinuation trial analysis. *Br J Cancer* 95: 58–586.

Engelman, J., Chen, L., Tan, X., Crosby, K., Guimaraes, A., Upadhyay, R. *et al.* (2008) Effective use of PI3K and MEK inhibitors to treat mutant KRAS G12D and PIK3CA H1047R murine lung cancers. *Nat Med* 14: 1351–1356.

Engelman, J., Luo, J. and Cantley, L. (2006) The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat Rev Genet* 7: 606–619.

Escudier, B., Eisen, T., Stadler, W., Szczylik, C., Oudard, S., Siebels, M. *et al.* (2007) Sorafenib in advanced clear-cell renal-cell carcinoma. *N Engl J Med* 356: 125–134.

Faber, A., Coffee, E., Costa, C., Dastur, A., Ebi, H., Hata, A. *et al.* (2014) mTOR inhibition specifically sensitizes colorectal cancers with KRAS or BRAF mutations to BCL-2/BCL-XI inhibition by suppressing MCL-1. *Cancer Discov* 4: 42–52.

Faber, A., Li, D., Song, Y., Liang, M., Yeap, B., Bronson, R. *et al.* (2009) Differential Induction of apoptosis in HER2 and EGFR addicted cancers following PI3K inhibition. *Proc Natl Acad Sci U S A* 106: 19503–19508.

Flaherty, K., Infante, J., Daud, A., Gonzalez, R., Kefford, R., Sosman, J. *et al.* (2012) Combined BRAF and MEK inhibition in melanoma with BRAF V600 mutations. *N Engl J Med* 367: 1694–1703.

Gollob, J., Wilhelm, S., Carter, C. and Kelley, S. (2006) Role of RAF kinase in cancer: therapeutic potential of targeting the RAF/MEK/ERK signal transduction pathway. *Semin Oncol* 33: 392–406.

Hanahan, D. and Weinberg, R. (2011) Hallmarks of cancer: the next generation. *Cell* 144: 646–674.

Hauschild, A., Grob, J., Demidov, L., Jouary, T., Gutzmer, R., Millward, M. *et al.* (2012) Dabrafenib in BRAF-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial. *Lancet* 380: 358–365.

Ho, A., Grewal, R., Leboeuf, R., Sherman, E., Pfister, D., Deandreis, D. *et al.* (2013) Selumetinib-enhanced radioiodine uptake in advanced thyroid cancer. *N Engl* \mathcal{J} *Med* 368: 623–632.

Hoeflich, K., Merchant, M., Orr, C., Chan, J., Den Otter, D., Berry, L. *et al.* (2012) Intermittent administration of MEK inhibitor GDC-0973 plus PI3K inhibitor GDC-0941 Triggers robust apoptosis and tumor growth inhibition. *Cancer Res* 72: 210–219.

Hoeflich, K., O'Brien, C., Boyd, Z., Cavet, G., Guerrero, S., Jung, K. *et al.* (2009) *In vivo* antitumor activity of MEK and phosphatidylinositol 3-kinase inhibitors in basal-like breast cancer models. *Clin Cancer Res* 15: 4649–4664.

Hudes, G., Carducci, M., Tomczak, P., Dutcher, J., Figlin, R., Kapoor, A. *et al.* (2007) Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. *N Engl J Med* 356: 2271–2281.

Infante, J., Fecher, L., Falchook, G., Nallapareddy, S., Gordon, M., Becerra, C. *et al.* (2012) Safety, pharmacokinetic, pharmacodynamic, and efficacy data for the oral MEK inhibitor trametinib: a phase 1 dose-escalation trial. *Lancet Oncol* 13: 773–781.

Jokinen, E., Laurila, N. and Koivunen, J. (2012) Alternative dosing of dual PI3K and MEK inhibition in cancer therapy. *BMC Cancer* 12: 612. Juric, D., Soria, J., Sharma, S., Banerjee, U., Azaro, A., Desai, J. *et al.* (2014) A phase 1b Dose-escalation study of BYL719 plus binimetinib (MEK162) in patients with selected advanced solid tumors. *J Clin Oncol* 32:5s (Suppl.): abstract 9051.

Kimura, E., Nikiforova, M., Zhu, Z., Knauf, J., Nikiforov, Y. and Fagin, J. (2003) High prevalence of BRAF mutations in thyroid cancer: genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. *Cancer Res* 63: 1454–1457.

Lin, S., Liu, C., Ko, S., Chang, H., Liu, T. and Chang, K. (2005) Copy number amplification of 3Q26-27 oncogenes in microdissected oral squamous cell carcinoma and oral brushed samples from areca chewers. *J Pathol* 206: 417–422.

Llovet, J., Ricci, S., Mazzaferro, V., Hilgard, P., Gane, E., Blanc, J. *et al.* (2008) Sorafenib in advanced hepatocellular carcinoma. *N Engl J Med* 359: 37–390.

Lorusso, P., Shapiro, G., Panday, S., Kwak, E., Jones, C., Belvin, M. *et al.* (2012) A first-in-human phase Ib study to evaluate the MEK inhibitor GDC-0973, combined with the pan-PI3K Inhibitor GDC-0941, in patients with advanced solid tumors. *J Clin Oncol* 30(Suppl.): abstract 2566.

Mendoza, M., Er, E. and Blenis, J. (2011) The RAS-Erk and PI3K-mTOR pathways: cross-talk and compensation. *Trends Biochem Sci* 36: 320–328.

Moreno Garcia, V., Baird, R., Shah, K., Basu, B., Tunariu, N., Blanco, M. *et al.* (2011) A phase I study evaluating GDC-0941, an oral phosphoinositide-3 kinase (PI3K) inhibitor, in patients with advanced solid tumors or multiple myeloma. *J Clin Oncol* 29(Suppl.): abstract 3021.

Motzer, R., Escudier, B., Oudard, S., Hutson, T., Porta, C., Bracarda, S. *et al.* (2008) Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled phase III trial. *Lancet* 372: 449–456.

O'Reilly, K., Rojo, F., She, Q., Solit, D., Mills, G., Smith, D. *et al.* (2006) mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates AKT. *Cancer Res* 66: 1500–1508.

Poulikakos, P., Zhang, C., Bollag, G., Shokat, K. and Rosen, N. (2010) RAF inhibitors transactivate RAF dimers and ERK signalling in cells with wild-type BRAF. *Nature* 464: 427–430.

Ramanathan, R., Von Hoff, D., Eskens, F., Blumenschein, G., Richards, D., Renshaw, F. *et al.* (2014) A Phase 1b trial of PI3K inhibitor copanlisib (BAY 80-6946) combined with the allosteric-MEK Inhibitor refametinib (BAY 86-9766) in patients with advanced cancer. \mathcal{J} *Clin Oncol* 32:5s(Suppl.): abstract 2588. Rinehart, J., Adjei, A., Lorusso, P., Waterhouse, D., Hecht, J., Natale, R. *et al.* (2004) Multicenter phase II study of the oral MEK inhibitor, CI-1040, in patients with advanced non-small-cell lung, breast, colon, and pancreatic cancer. *J Clin Oncol* 22: 4456–4462.

Serra, V., Scaltriti, M., Prudkin, L., Eichhorn, P., Ibrahim, Y., Chandarlapaty, S. *et al.* (2011) PI3K inhibition results in enhanced HER Signaling and acquired ERK dependency in HER2-overexpressing breast cancer. *Oncogene* 30: 2547–2557.

Solit, D., Garraway, L., Pratilas, C., Sawai, A., Getz, G., Basso, A. *et al.* (2006) BRAF mutation predicts sensitivity to MEK inhibition. *Nature* 439: 358–362.

Sos, M., Fischer, S., Ullrich, R., Peifer, M., Heuckmann, J., Koker, M. *et al.* (2009) Identifying genotype-dependent efficacy of single and combined PI3K- and MAPK-pathway inhibition in cancer. *Proc Natl Acad Sci U S A* 106: 18351–18356.

Sousa, S., Fernandes, P. and Ramos, M. (2008) Farnesyltransferase inhibitors: a detailed chemical view on an elusive biological problem. *Curr Med Chem* 15: 1478–1492.

Su, F., Viros, A., Milagre, C., Trunzer, K., Bollag, G., Spleiss, O. *et al.* (2012) RAS mutations in cutaneous squamous-cell carcinomas in patients treated with BRAF inhibitors. *N Engl J Med* 366: 207–215.

Turke, A., Song, Y., Costa, C., Cook, R., Arteaga, C., Asara, J. *et al.* (2012) MEK inhibition leads to PI3K/ AKT activation by relieving a negative feedback on ERBB receptors. *Cancer Res* 72: 3228–3237.

Vasudevan, K., Barbie, D., Davies, M., Rabinovsky, R., McNear, C., Kim, J. *et al.* (2009) AKTindependent signaling downstream of oncogenic PIK3CA mutations in human cancer. *Cancer Cell* 16: 21–32. Wallace, E., Lyssikatos, J., Yeh, T., Winkler, J. and Koch, K. (2005) Progress towards therapeutic small molecule MEK inhibitors for use in cancer therapy. *Curr Top Med Chem* 5: 215–229.

Wander, S., Hennessy, B. and Slingerland, J. (2011) Next-generation mTOR inhibitors in clinical oncology: how pathway complexity informs therapeutic strategy. *J Clin Invest* 121: 1231–1241.

Weekes, C., Von Hoff, D., Adjei, A., Leffingwell, D., Eckhardt, S., Gore, L. *et al.* (2013) Multicenter phase I trial of the mitogen-activated protein kinase 1/2 inhibitor BAY 86-9766 in patients with advanced cancer. *Clin Cancer Res* 19: 1232–1243.

Weigelt, B., Warne, P. and Downward, J. (2011) PIK3CA mutation, but not PTEN loss of function, determines the sensitivity of breast cancer cells to mTOR inhibitory drugs. *Oncogene* 30: 3222–3233.

Wilhelm, S., Adnane, L., Newell, P., Villanueva, A., Llovet, J. and Lynch, M. (2008) Preclinical overview of sorafenib, a multikinase inhibitor that targets both RAF and VEGF and PDGF Receptor tyrosine kinase signaling. *Mol Cancer Ther* 7: 3129–3140.

Yan, Y., Serra, V., Prudkin, L., Scaltriti, M., Murli, S., Rodriguez, O. *et al.* (2013) Evaluation and clinical analyses of downstream targets of the AKT inhibitor GDC-0068. *Clin Cancer Res* 19: 6976–6986.

Yao, J., Shah, M., Ito, T., Bohas, C., Wolin, E., Van Cutsem, E. *et al.* (2011) Everolimus for advanced pancreatic neuroendocrine tumors. *N Engl J Med* 364: 514–523.

Zimmer, L., Barlesi, F., Martinez-Garcia, M., Dieras, V., Schellens, J., Spano, J. *et al.* (2014) Phase I expansion and pharmacodynamic study of the oral MEK inhibitor RO4987655 (CH4987655) in selected patients with advanced cancer with RAS-RAF mutations. *Clin Cancer Res* 20: 4251–4261.

Visit SAGE journals online

http://tam.sagepub.com

SAGE journals