

occurs in proximal nerves resulting in increased nerve size and increased proton spin density. In the early symptomatic stage, endoneurial oedema occurs with prolongation of T₂ relaxation time detected. This proximal pathology results in secondary axonal degeneration in distal nerves (Fig. 1A). In late stage disease, the pathology spreads to involve more distal nerves, at which point sural nerve biopsy may demonstrate amyloid deposition (Fig. 1B).

This paper represents an exciting development in TTR-FAP as it identifies the earliest biomarker of neuropathy to date and also provides interesting insights into disease pathogenesis. To translate this work into clinical practice, further studies are needed. First, the methods used here are time- and resource-intensive. Over 30 000 regions of interest were drawn by hand prior to analysis, and a large number of healthy controls were studied to allow normalization of signal intensities. Second, the data are presented as group analyses rather than to classify individual patients as ‘normal’ or ‘abnormal’. This would be necessary if these MRI techniques are to be used to distinguish mutation carriers who are presymptomatic and therefore would benefit from intervention, from those who are non-penetrant and would presumably receive no such benefit. Finally, as the authors acknowledge, longitudinal natural history studies are now needed to determine the usefulness of these biomarkers in identifying the optimal timing of a therapy in an individual patient, and the responsiveness of the

biomarkers as outcome measures, a key determinant of study power. Note should also be made of the increasing volume of research using MRI to quantify muscle changes secondary to neuropathic processes (Sinclair *et al.*, 2012), which is also a potential source of responsive biomarkers in this disease group.

TTR-related FAP is a devastating disease for which there are now a number of new and emerging therapies. As these treatments slow progression rather than reverse established damage, biomarkers of early stage and subclinical disease, before nerve conduction studies are abnormal, are urgently needed. Quantitative magnetic resonance neurography as demonstrated in the paper by Kollmer and colleagues not only provides intriguing insights into the anatomical and pathological progression of this devastating disease, but also shows promise as an early marker of the neuropathy that is the first manifestation of the disease in most patients.

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Jasper M. Morrow and Mary M. Reilly
MRC Centre for Neuromuscular Diseases,
UCL Institute of Neurology, Queen Square,
London WC1N 3BG, UK

Correspondence to: Mary M. Reilly,
E-mail: m.reilly@ucl.ac.uk

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References

- Adams D, Théaudin M, Cauquil C, Algalarrondo V, Slama M. FAP neuropathy and emerging treatments. *Curr Neurol Neurosci Rep* 2014; 14: 435.
- Berk JL, Suhr OB, Obici L, et al. Repurposing diflunisal for familial amyloid polyneuropathy: a randomized clinical trial. *JAMA* 2013; 310: 2658–67.
- Coelho T, Maia LF, Martins da Silva A, et al. Tafamidis for transthyretin familial amyloid polyneuropathy: a randomized, controlled trial. *Neurology* 2012; 79: 785–92.
- Hughes RAC, Bouche P, Cornblath DR, et al. European federation of neurological societies/peripheral nerve society guideline on management of chronic inflammatory demyelinating polyradiculoneuropathy: report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society. *Eur J Neurol Off J Eur Fed Neurol Soc* 2006; 13: 326.
- Kollmer J, Pham M. *In-vivo* detection of nerve injury in familial amyloid polyneuropathy by magnetic resonance-neurography. *Brain* 2015; 138: 549–62.
- Planté-Bordeneuve V, Lalu T, Misrahi M, et al. Genotypic-phenotypic variations in a series of 65 patients with familial amyloid polyneuropathy. *Neurology* 1998; 51: 708–14.
- Sinclair CDJ, Morrow JM, Miranda MA, et al. Skeletal muscle MRI magnetisation transfer ratio reflects clinical severity in peripheral neuropathies. *J Neurol Neurosurg Psychiatry* 2012; 83: 29–32.
- Wilczek HE, Larsson M, Ericzon B-G. FAPWTR. Long-term data from the Familial Amyloidotic Polyneuropathy World Transplant Registry (FAPWTR). *Amyloid Int J Exp Clin Investig Off J Int Soc Amyloidosis* 2011; 18 (Suppl 1): 193–5.

Temporal lobe epilepsy: a unique window into living human brain epigenetic gene regulation

This scientific commentary refers to ‘Differential DNA methylation profiles of coding and non-coding genes define hippocampal sclerosis in human temporal lobe epilepsy’ by Miller-Delaney *et al.* (10.1093/brain/awu373).

Epigenetic modification of the genome is a powerful mechanism for regulation of RNA expression. For the large majority of non-neoplastic brain disorders, a lack of human brain tissue suitable for complementary

epigenetic and RNA expression analyses poses a major challenge for translating data from experimental models to human disease. Temporal lobe epilepsy is a unique exception. Tissue from pharmaco-resistant patients who

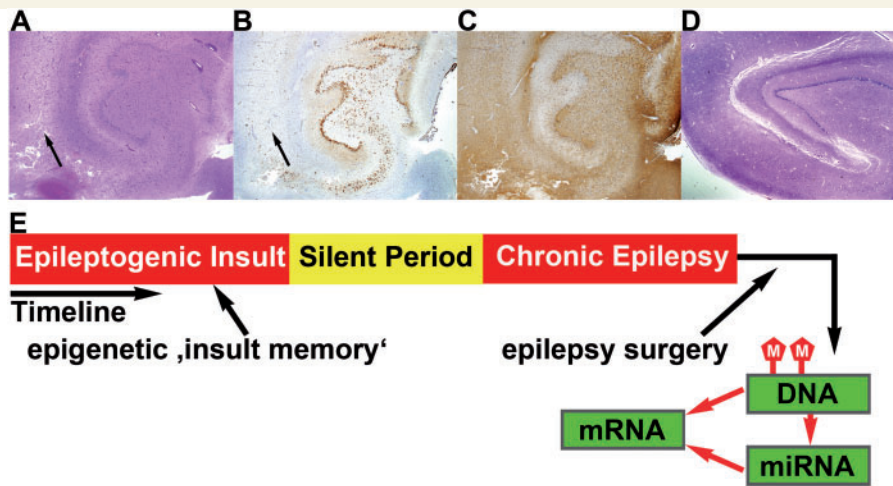


Figure 1 Epigenetic regulatory model in temporal lobe epilepsy. (A) Hippocampal biopsy specimen obtained during surgery in a patient with pharmacoresistant temporal lobe epilepsy (haematoxylin and eosin). The section shows hippocampal sclerosis, reflecting pathological alterations typical of those in the material used by Miller-Delaney *et al.* (2014). The main alterations comprise neuronal cell loss, particularly in the CA1 area (black arrows) visible in **A** and **B** (B: NeuN immunohistochemistry), and reactive astrogliosis (C: glial fibrillary acidic protein immunohistochemistry). (D) An autopsy section (haematoxylin and eosin) showing the hippocampal formation of a patient without neurological disorders, lacking segmental neurodegeneration and gliosis, and typical of an experimental control subject. (E) Differences in cellular composition between subjects with and without epilepsy are reflected in patterns of DNA methylation and mRNA and microRNA expression. The impact of other putative epileptogenic factors on hippocampal DNA methylation patterns constitutes a form of long-term ‘insult-induced memory’. (A–D) Magnification: $\times 1.25$.

undergo epilepsy surgery allow singular access for comprehensive molecular genetic studies. In this issue of *Brain*, Miller-Delaney and colleagues use this intriguing resource to report fundamental new insights into epigenetic regulation in human brain tissue (Miller-Delaney *et al.*, 2014).

Epigenetic dynamics under disease conditions are generally thought to represent wide-scale alterations in the expression of multiple classes of RNA. In the context of epilepsy, however, most studies of epigenetic modifications have focused on distinct structural changes and alterations in the expression of specific mRNAs (Kobow *et al.*, 2009; Ryley Parrish *et al.*, 2013). Much less is known about genome-wide epigenetic alterations and corresponding patterns of RNA expression in the disease. Miller-Delaney and colleagues addressed the challenge of molecular genetic analyses in human brain tissue by comparing genome-wide DNA methylation patterns in the hippocampi of patients with pharmaco-refractory temporal lobe epilepsy with those in autopsy samples of brain tissue from individuals with non-neurological

causes of death. The commonest lesion pattern in patients with temporal lobe epilepsy is hippocampal sclerosis, with segmental neurodegeneration and concomitant astrogliosis as pathological hallmarks (Fig. 1A–D). However, patterns of damage vary among individuals (Blümcke *et al.*, 2013) and Miller-Delaney *et al.* (2014) therefore subdivided their temporal lobe epilepsy group accordingly. Only 146 protein-coding genes were found to show altered DNA methylation in temporal lobe epilepsy, a surprisingly low number when compared to the complexity and functional consequences of this disorder. The vast majority of affected promoters showed hypermethylation, an observation that is in good agreement with the epigenetic data from key animal models of epilepsy (Kobow *et al.*, 2013). In a novel and intriguing finding, Miller-Delaney and colleagues also identified several methylation-sensitive microRNAs and long non-coding RNAs in the brain tissue.

By correlating these differential patterns of genome methylation with the corresponding RNA expression patterns, Miller-Delaney and co-workers

managed to fully exploit the potential of their experimental approach. Their analyses revealed a surprisingly weak overall correlation between promoter epigenetic dynamics and the expression of coding transcripts. However, for certain RNAs, promoter methylation patterns had a striking impact in human (epilepsy) brain tissue. This was true in particular for several microRNAs, as well as for certain non-coding RNAs. This difference in the degree of correspondence between epigenetic promoter programming and expression levels for coding mRNAs versus microRNAs might indicate that expression levels of the latter are more directly influenced by changes in promoter methylation—perhaps even under physiological conditions in the human brain. The abundance of coding mRNAs is regulated not only by promoter activity but also by factors such as the activity of microRNAs, which might complicate the link between epigenetics and mRNA levels. By contrast, the link between epigenetics and microRNA levels might be more direct. This differential impact of methylation on mRNA versus microRNA abundance could

have important consequences for the development of novel therapeutic strategies, with respect to predicting the molecular effects of targeting different RNA classes by epigenetic means in CNS disorders.

Epigenetic regulation has some particular characteristics that make it exceptionally attractive as a putative pathomechanism of temporal lobe epilepsy. One of these is the potential for long-lasting, stable effects on gene expression that outlive an initial transient signal. This may be especially relevant for post-mitotic neurons, which are subject to various insults with short- to long-lasting effects on their activity and connectivity (Guan *et al.*, 2002). In temporal lobe epilepsy, seizures do not generally start at birth in affected individuals but develop later in life (Pitkänen and Engel, 2014). Many patient histories reveal a transient insult in early childhood, which is followed by a silent interval, potentially lasting several years, before recurrent seizures emerge. What type of molecular modifications can be that long-lasting? Epigenetic remodelling could in fact constitute the brain's 'memory of transient epileptogenic insults' (Fig. 1E). However, many short- and long-term influences on DNA methylation will be reflected in an epigenetic pattern. In addition to transient potentially epileptogenic insults, the pattern of DNA methylation could also signal an increased disease risk owing to, for example, genomic imprinting or the gestational environment, which has been suggested to have pathogenetic relevance in other CNS disorders (Jiao *et al.*, 2013), or to differences in the cellular composition of the temporal lobes of patients with epilepsy versus control subjects (Fig. 1). More sophisticated experimental approaches that consider patients' family epigenetics and that use bioinformatics to control for cellular admixtures might be useful in the future (Guintivano *et al.*, 2013). Furthermore, epigenetic patterns may be rather dynamic (El-Osta *et al.*, 2008), and can also reflect relatively short-term changes, which can complicate data interpretation.

The work has some limitations, which Miller-Delaney and colleagues have obviously considered. One inherent difficulty with approaches that analyse human brain tissue relates to the choice of controls. Miller-Delaney *et al.* used autopsy brain tissue, but while DNA-methylation analyses can be carried out with this material, this approach clearly limits the comparability of genomic methylation dynamics and RNA expression patterns. Furthermore, the control and temporal lobe epilepsy cohorts will be subject to the methodological and statistical limitations inherent in large-scale studies based on such small sized groups. Miller-Delaney and colleagues have taken great care to control for group effects by minimizing differences in key parameters such as gender and age, wherever possible. But this clearly has limitations. As a result of the rather young age of the patients with temporal lobe epilepsy, the control group contains individuals who died from cardiovascular disease at unusually young ages. This raises suspicions of inherited impairments that increase cardiovascular disease risk and burden—might these have a potential epigenetic impact? Furthermore, the effects of antiepileptic pharmacotherapy in the group with temporal lobe epilepsy cannot easily be controlled for, and may be substantial. Stressing the fact that the present work is a pilot study, Miller-Delaney and colleagues fully acknowledge these shortcomings and suggest follow-up multicentre studies, which we eagerly anticipate will provide more detailed insights into this fast developing field of research with intriguing clinical implications.

Alexander Grote, Susanne Schoch and
Albert J. Becker
University of Bonn Medical Centre,
Germany

Correspondence to: Albert J. Becker
E-mail: albert_becker@uni-bonn.de

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References

- Blümcke I, Thom M, Aronica E, Armstrong DD, Bartolomei F, Bernasconi A, et al. International consensus classification of hippocampal sclerosis in temporal lobe epilepsy: a task force report from the ILAE Commission on Diagnostic Methods. *Epilepsia* 2013; 54: 1315–29.
- El-Osta A, Brasacchio D, Yao D, Pocaí A, Jones PL, Roeder RG, et al. Transient high glucose causes persistent epigenetic changes and altered gene expression during subsequent normoglycemia. *J Exp Med* 2008; 205: 2409–17.
- Guan Z, Giustetto M, Lomvardas S, Kim JH, Miniaci MC, Schwartz JH, et al. Integration of long-term-memory-related synaptic plasticity involves bidirectional regulation of gene expression and chromatin structure. *Cell* 2002; 111: 483–93.
- Guintivano J, Aryee MJ, Kaminsky ZA. A cell epigenotype specific model for the correction of brain cellular heterogeneity bias and its application to age, brain region and major depression. *Epigenetics* 2013; 8: 290–302.
- Jiao J, Opal MD, Dulawa SC. Gestational environment programs adult depression-like behavior through methylation of the calcitonin gene-related peptide gene. *Mol Psychiatry* 2013; 18: 1273–80.
- Kobow K, Jeske I, Hildebrandt M, Hauke J, Hahnen E, Buslei R, et al. Increased reelin promoter methylation is associated with granule cell dispersion in human temporal lobe epilepsy. *J Neuropathol Exp Neurol* 2009; 68: 356–64.
- Kobow K, Kaspi A, Harikrishnan KN, Kiese K, Ziemann M, Khurana I, et al. Deep sequencing reveals increased DNA methylation in chronic rat epilepsy. *Acta Neuropathol* 2013; 126: 741–56.
- Miller-Delaney SF, Bryan K, Das S, McKiernan RC, Bray IM, Reynolds JP, et al. Differential DNA methylation profiles of coding and noncoding genes define hippocampal sclerosis in human temporal lobe epilepsy. *Brain* 2015; 138: 601–16.
- Pitkänen A, Engel J Jr. Past and present definitions of epileptogenesis and its biomarkers. *Neurotherapeutics* 2014; 11: 231–41.
- Ryley Parrish R, Albertson AJ, Buckingham SC, Hablitz JJ, Mascia KL, Davis Haselden W, et al. Status epilepticus triggers early and late alterations in brain-derived neurotrophic factor and NMDA glutamate receptor Grin2b DNA methylation levels in the hippocampus. *Neuroscience* 2013; 248: 602–19.