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## Increased *CYFIP1* dosage alters cellular and dendritic morphology and dysregulates mTOR

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### Abstract

Rare maternally inherited duplications at 15q11-13 are observed in about 1% of individuals with an Autism Spectrum Disorder (ASD), making it among the most common causes of ASD. 15q11-13 comprises a complex region, and because this CNV encompasses many genes, it is important to explore individual genotype-phenotype relationships. Cytoplasmic FMR1 interacting protein 1 (*CYFIP1*) is of particular interest because of its interaction with FMRP, its upregulation in transformed lymphoblastoid cell lines from patients with duplications at 15q11-13 and ASD, and the presence of smaller overlapping deletions of *CYFIP1* in patients with schizophrenia and intellectual disability. Here, we confirm that *CYFIP1* is upregulated in transformed lymphoblastoid cell lines, and demonstrate its upregulation in postmortem brain from 15q11-13 duplication patients for the first time. To investigate how increased *CYFIP1* dosage might predispose to neurodevelopmental disease, we studied the consequence of its overexpression in multiple systems. We show that overexpression of *CYFIP1* results in morphological abnormalities including cellular hypertrophy in SY5Y cells and differentiated mouse neuronal progenitors. We validate these results *in vivo* by generating a BAC transgenic mouse, which over-expresses

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#### Conflict of interest

The authors declare no conflict of interest. A provisional patent describing the potential utility of mTOR inhibitors in individuals with ASD harboring 15q11-13 duplications has been filed.

CYFIP1 under the endogenous promotor, observing an increase in the proportion of mature dendrite spines and dendritic spine density. Gene expression profiling at embryonic day 15 suggested dysregulation of mTOR signaling, which was confirmed at the protein level. Importantly, similar evidence of mTOR-related dysregulation was seen in brains from 15q11-13 duplication patients with ASD. Finally, treatment of differentiated mouse neuronal progenitors with an mTOR inhibitor (rapamycin) rescued the morphological abnormalities resulting from CYFIP1 overexpression. Together, these data show that CYFIP1 overexpression results in specific cellular phenotypes, and implicate modulation by mTOR signaling, further emphasizing its role as a potential convergent pathway in some forms of ASD.

## Keywords

CYFIP1; Autism; Fragile X syndrome; neural progenitor cells; signaling

## Introduction

Autism spectrum disorder (ASD) is a common and heterogeneous neuropsychiatric condition with deficits in social interactions and language development in addition to the presence of repetitive behavior and restricted interests [1]. ASD is considered a developmental brain disorder and recent studies have identified a number of rare and common variants that may contribute to etiology [2-11]. One of the most consistently reported genomic aberrations associated with ASD is duplication at chromosome 15q11-13 ((Dup)15q11-13) occurring in about 1% of cases [12-16]. Deletions at this same region give rise to Prader-Willi or Angelman syndrome based on parent of origin [17]. In all cases, instability of this region is mediated by three sets of low copy repeats (BP1, BP2, and BP3) that leads to alternative yet reproducible breakpoints [18]. Although previous work on 15q has focused largely on the imprinted BP2-BP3 region, there is evidence to suggest that one or more genes in the more proximal non-imprinted BP1-BP2 interval, and *CYFIP1* in particular, merit additional investigation. First, among individuals with deletions or gains at 15q11-13, those harboring larger events that include *CYFIP1* are typically most severely impacted [19, 20]. Second, consistent with a genomic imbalance model [21], smaller “BP1-BP2” deletions encompassing *CYFIP1* and three other genes are associated with developmental delay [22-25], epilepsies [26] and schizophrenia [27, 28]. Third, CYFIP1 was first identified as a protein that interacts with Fragile X mental retardation protein (FMRP) [29], which when mutated, causes Fragile X syndrome, another monogenic form of ASD [30]. Finally, *CYFIP1* is upregulated in transformed lymphoblastoid cell lines from patients with (dup)15q11-13 and ASD [31] and reduced in individuals with fragile X syndrome and ASD[32].

CYFIP1 has two known functions: regulating actin cytoskeletal dynamics [33], and repressing translational initiation [34]. CYFIP1 regulates the cytoskeleton through its competitive inhibition of interactions between FMRP and the small GTPase RAC1, which is implicated in the development and maintenance of neuronal structures [35, 36]. CYFIP1 regulates translation at the synapse through binding to Eukaryotic translation initiation factor 4E (eIF4E) in a complex with FMRP [34]. Work in *Drosophila melanogaster* [33] indicates

that null alleles are associated with abnormalities in neuronal morphology. No work, however, has been done in any mammalian system to look at the impact of ASD-relevant increases in *CYFIP1* gene dosage.

Here we show that CYFIP1 is increased in the brains of individuals with 15q11-13 duplications and ASD. We also generated a BAC transgenic mouse model and investigated the consequences of CYFIP1 over-expression *in vitro* and *in vivo*. We identified consistent abnormalities of cell morphology including increased cell size, changes in neurite outgrowth, and increased spine density on pyramidal neurons in the cerebral cortex of adult transgenic mice. This is consistent with previous work in mice with mutations in proteins involved in the same neuronal translation pathways, FMR1 and 4E-BP2 [35,36], which have either increased spine density or immature spines. Additionally, analysis of post mortem brain dendritic morphology also suggests that spine density is increased in some cases of ASD [34]. To begin to untangle the molecular pathways responsible for these pathological alterations, we performed gene expression analysis in embryonic mouse brain, identifying an enrichment for genes involved in mTOR-related signaling in transgenic mouse brain. We confirmed mTOR dysregulation in post mortem brain from individuals with 15q11 duplications and ASD. We next hypothesized that *Cyfp1* overexpression might impact cellular morphology through mTOR, which we tested via mTOR inhibition with rapamycin. Remarkably, we observed that rapamycin rescued the morphologic abnormalities in differentiated mouse neuronal progenitors (dMNPs). We also show that CYFIP1 and p-S6 are increased in the brains of individuals with 15q11-13 duplications and ASD, consistent with mTOR pathway activation in these individuals. Our demonstration of pathological consequences resulting from CYFIP1 overexpression, both *in vitro* and *in vivo*, identify for the first time a modulatory mechanism that may be of clinical utility.

## Materials and Methods

### Animals

Mice harboring a *Cyfp1* spanning BAC clone (RP24-333C15) were generated in the UCLA transgenic core (<http://tmc.ctrl.ucla.edu/tg-core/>) by injection into C57BL/6J pronuclei using standard protocols [37, 38]. DNA was prepared using a cesium chloride gradient, and checked for integrity by pulsed field gel electrophoresis. DNA was then microinjected into fertilized C57BL/6J pronuclei, which were implanted in the oviduct of pseudopregnant recipient female mice. Six potential founders were identified by screening tail DNA using transgene-specific primers (see Table S3). For the two founders found to give rise to transgene positive animals (Tg#8 and Tg#24), breeding colonies were established by backcrossing to C57BL/6J animals. Assessment of *Cyfp1* levels at each generation determined that transgenic mice failed to overexpress the protein at N4 and beyond, despite confirmed genotype and increased copy number. This was true for each of the two lines generated. To address this issue we not only restricted all experiments and analyses to N2 and N3 mice but also focused on abnormalities observed both *in vivo* and *in vitro*. Animals were kept on a 12 hr light/12 hr dark cycle with free access to food and water. All procedures involving animals were performed in accordance with a protocol approved by the UCLA Animal Research Committee.

## Histopathology

Mice were anesthetized with nembutal, and then transcardially perfused with PBS, followed by fixative (4% paraformaldehyde in PBS, pH 7.4). For Golgi staining, perfused brains were ultimately embedded in OCT as above and sectioned coronally at 60 $\mu$ m with a cryostat (Leica). For Hemotoxylin and Eosin (H/E) staining, fixed brains were embedded in paraffin and 7 $\mu$ m transverse sections obtained. Cell morphological tracing and quantitation of processes were carried out using Adobe Photoshop CS3 and NeuroMath [39]. Morphological and developmental spine characterization was carried out using standard methods [40-42]. Cell/tissue staining and morphological analysis details were described in the supplemental methods.

## Cell Culture

6 $\times$ 10<sup>6</sup> SY5Y cells were seeded in a T-75 flask in 20 ml of DMEM (Invitrogen) with 10% FBS (Gibco, NY, USA). Cell differentiation and transfection were carried out as described previously [31] and as described in the supplemental methods. MNPs were cultured for 2 days, infected with lentiviral constructs, and then differentiated for 14 days. Rapamycin and DMSO (vehicle) were purchased from Sigma-Aldrich.

## Patient Materials

Seven ASD individuals with maternally derived *CYFIP1* containing 15q11-13 duplications and six typically developing non-carrier controls were identified within the Autism Genetic Resource Exchange (AGRE) [31, 43, 44], Non-AU samples from Dr. N. Schanen (<http://www.bio.udel.edu/users/cschanen>) and frozen aliquots of EBV transformed lymphoblastoid cell lines were acquired (Supplemental Material, Table S1). Total RNA from lymphoblastoid cell lines, seeded at  $9 \times 10^6$  and harvested 24 hours later, was obtained using the RNeasy Mini system with DNase treatment (Qiagen). RNA quantity and quality were measured with a ND-100 (Nanodrop) and Bioanalyzer 2100 (Agilent), respectively. Three separate individual with ASD with maternally derived *CYFIP1* containing 15q11-13 duplications and five typically developing non-carrier controls were identified from the Autism Tissue Program cohort organized by Autism Speaks and the Harvard Brain Bank [45]; tissue samples from the superior temporal gyrus (STG, also known as Brodmann's area 41/42) were obtained (Supplemental Material, Table S2). RNA was obtained using the miRNeasy kit (Qiagen) and quantity and quality assessed as described above. Clinical information for all cases is available from AGRE and the ATP, respectively.

## qPCR

cDNA was generated from total RNA using random hexamers and Superscript III (Invitrogen, NY, USA). Real-time PCR was conducted on an HT-7900 (Applied Biosystems, Foster City, CA, USA) with Sybr-Rox SuperMix (Bio-Rad, Hercules, CA, USA) using published methods [31, 46, 47]. Intron-spanning primers are detailed in Supplemental Material (Table S3).

## Western Blotting

Western Blotting was performed by the modified of standard methods[48] and details were described in the supplemental methods.

## Microarray Gene Expression Profiling

Microarray profiling was performed as previously described [49, 50]. Briefly, total RNA was isolated from embryonic day 15 frontal cortex from Tg#8 transgenic animals (n=3) and littermate controls (n=3) using the Qiagen RNeasy Kit. RNA quantity and quality was assessed using a Nanodrop spectrophotometer (Nanodrop Technologies) and the Agilent Bioanalyzer (Agilent Technologies), respectively. Total RNA (200ng) was amplified, biotinylated and hybridized on Illumina Mouse ref-8 v2.0 BeadChip arrays, querying the expression of ~25,600 well-annotated Refseq transcripts, as per the manufacturer's protocol. Slides were scanned using Illumina BeadStation and signals extracted using Illumina BeadStudio software (Illumina, San Diego CA). Raw data were analyzed using Bioconductor packages ([www.bioconductor.org](http://www.bioconductor.org)). Quality-control analysis was performed using several indices including inter-array Pearson correlation, clustering based on variance, and the mean absolute deviation (MAD) using the top 1000 most variant probes [50]. Data were normalized using quantile normalization. Analysis of differential expression was performed using linear model fitting (LIMMA package). After linear model fitting, a Bayesian estimate of differential expression was calculated using a p-value threshold of 0.01. Pathway analyses were performed using DAVID Bioinformatics online tools (Database for Annotation, Visualization and Integrated Discovery; <http://david.abcc.ncifcrf.gov/>) and the Ingenuity Pathway Analysis System (IPA: Ingenuity® Systems, [www.ingenuity.com](http://www.ingenuity.com)). Raw and normalized expression data have been deposited in NCBI's Gene Expression Omnibus [51] and are accessible through GEO Series accession number GSE40852.

## Statistical Analysis

All results are expressed as mean  $\pm$ SEM. Western blots, qPCRs, and neuronal morphology comparisons between groups were analyzed with ANOVA.

## Results

### Increased gene dosage at 15q11-13 is associated with increased *CYFIP1* levels in transformed lymphoblastoid cells and postmortem brain

Prior to initiating mechanistic work in model systems, we sought to further validate the extent to which *CYFIP1* was upregulated in autistic probands with (dup)15q11-13. We conducted qPCR on transformed lymphoblastoid cell lines (4 carrier cases and 6 non-carrier controls) and human temporal cortex (3 carrier cases and 5 non-carrier controls). Confirming our previous results in lymphoblastoid cell lines [31], levels of *Cyfip1* mRNA were significantly upregulated in carriers cases versus non-carrier controls (2.5-fold increase; Figure 1A, Raw discrete expression data for individual patients; Figure S1A). To be relevant to neuropsychiatric symptoms one would expect that similar changes would be seen in brain tissue from carrier patients. Although very few samples from individuals with 15q11-13

duplications are available worldwide, we were able to that *Cyfp1* mRNA levels were also significantly increased in the superior temporal gyrus (STG, also known as Brodmann's area 41/42) in carriers versus controls (24-fold increase; Figure 1B, Raw discrete expression data for individual patients; Figure S1B). These data support the notion that dysregulation of *Cyfp1* may contribute to 15q11-13 related phenotypes by upregulation in patient cerebral cortex, and at the same time, confirm that transformed lymphoblastoid cell lines are a reliable biomarker for *Cyfp1* dysregulation in individuals harboring increased dosage at this locus.

### **Overexpression of *Cyfp1* *in vitro* gives rise to increased cell size and abnormal neurite outgrowth in neuronal cells**

SY5Y cells that overexpressed CYFIP1 (2-fold relative to mock transfected controls by western blot, Figure S2A) had significantly reduced neurite length (reduced ~ 50%) and longest neurite branch length (reduced ~ 60%) in addition to significantly increased neurite branch number (increased ~ 30%) and cell size (increased ~ 80%) -- Figure 2A and B. To determine whether increased levels of CYFIP1 also induced morphological changes in primary neurons, we performed similar experiments in dMNP's using lentiviral infection (2-fold relative to mock transfected controls by western blot, Figure S2B). Qualitatively indistinguishable results to our observations in the human SY5Y cells were seen in CYFIP1-overexpressing dMNP's (Figure 2C and D), which showed significantly reduced neurite length (reduced ~ 20%) and longest neurite branch length (reduced ~ 10%) in addition to significantly increased overall neurite branch number (increased ~ 20%) and cell size (increased ~ 20%). Together, these results show that CYFIP1 overexpression impacts neuronal morphology *in vitro* in a human cell line (SY5Y) and primary mouse neurons, also indicating parallel phenotypes in both mouse and human.

### **Development of transgenic mice to model *in vivo* consequences of increased *Cyfp1* dosage**

Based on the morphological changes observed *in vitro*, we attempted to confirm the *Cyfp1* dosage effects *in vivo*. Two lines of transgenic mice harboring a BAC spanning the mouse *Cyfp1* locus (Figure 2E) were produced as described in the Methods. Transgenic mice were born at normal Mendelian ratios in each strain (Tg #8 and #24), regardless of parent of origin. We screened adult hippocampus, where CYFIP1 is highly expressed, by western blot to demonstrate overexpression. Transgenic offspring from each line showed a significant increase in CYFIP1 protein levels relative to wild-type controls (~ 200% and 150% respectively, Figure 2F). Additional characterization of Tg #8 progeny by qPCR was also performed. *Cyfp1* transcript levels were significantly increased in transgenic relative to controls in embryonic day (e) 15 frontal cortex, postnatal day (P) 1 cerebellum, P11 hippocampus, and in adult frontal cortex and cerebellum (Supplemental Figure S3).

### **Overexpression of *Cyfp1* *in vivo* gives rise to increased cell size and abnormal neurite outgrowth in pyramidal neurons**

No significant differences in gross brain weight or length were observed between adult *Cyfp1* transgenic mice (Tg #8) and controls (Supplemental Figure S2). Similarly, no



significant histological differences were observed between adults of each genotype through examination of H&E stained sections of the cerebral cortex (Supplemental Figure S3) or hippocampus (data not shown).

Several lines of previous work raised the possibility that altered *CYFIP1* dosage might result in abnormal spine density and maturity. First, study of post-mortem brain materials from individuals with idiopathic ASD has found that spine density is increased in cases relative to controls [52]. Second, CYFIP1 is known to be a 4E-BP [34], and mice homozygous for a deletion in a distinct 4E-BP (eukaryotic initiation factor 4E-binding protein-2 or 4E-BP2) show an increased spine density [53]. Third, an increase in immature spines has been reported in mice harboring mutations in *Fmr1*, a demonstrated functional partner for CYFIP1 [40]. So, to more closely examine neuronal morphology in transgenic mice, we performed Golgi staining in two month old animals. Examination of layer II-III pyramidal neurons in frontal cortex revealed significant differences between transgenic and wild-type control mice (Figure 2G and H), similar to the effects we observed *in vitro*. Paralleling observations from our *in vitro* models, both transgenic strains showed significant alterations in dendritic morphology compared with controls, including reduced overall neurite length (~20% in both strains) and longest neurite branch length (~20% in both strains). Other phenotypic abnormalities paralleled expression levels as the Tg#8 strain displayed greater increases in neurite branch number (increased ~50% and 15% in Tg#8 and #24, respectively) and cell size than the Tg#24 strain, the latter having lower CYFIP1 expression (increased ~60% and 10% in Tg#8 and #24, respectively).

### **Overexpression of *Cyfp1* *in vivo* results in increased spine density and increased proportion of pyramidal neurons showing a mature spine morphology**

Further analyses of Golgi stained layer II-III pyramidal neurons from two month old mouse frontal cortex revealed a more than two-fold increase in the density of spines on basal dendrites in each transgenic strain relative to control mice (Figure 3A and B). Significant differences were not limited to the two-fold increase in spine density, but included spine structure as well; morphological and developmental spine characteristics [40-42, 54, 55] were seen to differ between transgenic mice and wild-type controls (Figures 3C through E). More specifically, these analyses revealed significant differences in the overall proportion of mushroom-type immature thin spines (Type B; reduced ~50% in both strains), mature stubby spines (Type D; increased 15% in both strains), and abnormal spines (Type E; increased ~500% in both strains). No differences between transgenic mice and wild-type controls were observed for either immature thin spines (Type A), or immature double mushroom-type spines (Type C). Abnormal spines in transgenic mice were seen to 'cluster' or 'clump' in a way not observed in controls (Figure 3E).

### **Transcriptome analyses in embryonic transgenic and control mice identify differentially expressed genes and highlight mTOR**

To investigate the mechanisms through which *Cyfp1* overexpression might give rise to the observed abnormalities, we carried out a comparative microarray analysis of Tg#8 transgenic mice versus controls at E15, a time when large differences in CYFIP1 were observed. We identified a robust group of 130 upregulated (red) and 149 downregulated

(green) genes (Bayesian t test,  $p < 0.01$ ; Figure 4A and B). qPCR verified five of the top differentiated genes including an approximate two-fold reduction in three ASD candidate genes (*Cntnap4*, *Nrxn1* and *Nlgn3*; Figure 4C).

As a first step towards understanding these data and underlying pathways that may be perturbed as a result of *Cyfp1* overexpression, we conducted gene ontology (GO) analysis using DAVID (Figure 4D). Consistent with the morphological changes observed in transgenic mice relative to controls, GO analyses highlighted an over-representation of differentially expressed genes within several functional groups including those relating to actin filament and microtubule binding. Separate pathway based analyses carried out using Ingenuity Pathway software (Ingenuity Systems, USA), highlighted an enrichment of mTOR pathway genes (Figure 4E,  $p < 0.01$ ). This was particularly interesting because CYFIP1 is involved in regulation of neuronal protein synthesis, and other published work has implicated mTOR signaling in the pathophysiology of other monogenic forms of ASD, including Fragile X syndrome [56], PTEN-related syndromes [57] and Tuberous Sclerosis [58].

### **Cyfp1-associated mTOR dysregulation seen in differentiated neuronal progenitors and human brain**

To build on our expression analyses and determine the extent to which the *Cyfp1*-associated mTOR dysregulation was generalizable to a variety of model systems, we performed additional protein-level characterization of dMNP and human post mortem samples. Mouse Neuronal Progenitors (MNPs) are multipotent precursors that have the ability to differentiate into neurons, astrocytes, and oligodendrocytes, and prior to differentiation have the capacity for self-renewal. The two-week differentiation protocol we employed resulted in most cells staining for MAP2 (neuronal marker, data not shown) and Tau (axonal marker, data not shown), respectively. Overexpression of CYFIP1 in dMNPs resulted in a 0.2-fold reduction in PTEN and a 15-fold increase in mTOR relative to control samples (Figure 5A). Importantly, knockdown of CYFIP1 resulted in the opposite effect, with a 1.2-fold increase in PTEN and a 0.5-fold decrease in mTOR (Figure 5B). These data indicate that CYFIP1 levels regulate elements of the mTOR cascade in mammalian neurons. To determine whether these results were relevant to human brain, we next investigated whether mRNA levels of mTOR were dysregulated in 15q11-13 duplication carriers with ASD relative to controls. These analyses, using the same samples from human superior temporal gyrus described above, showed a significant, approximately 6-fold upregulation of mTOR in patient carriers compared to non-carrier controls (Figure 5C, Raw discrete expression data for individual patients; Figure S6), suggesting that mTOR levels are sensitive to CYFIP1 dosage in human brain, as we observed in dMNPs.

### **Rapamycin, an mTOR inhibitor, rescues the morphological and molecular abnormalities resulting from CYFIP1 overexpression**

We next reasoned that if mTOR signaling was causing any of these abnormalities, that the mTOR inhibitor rapamycin might ameliorate any of the abnormalities resulting from CYFIP1 overexpression in dMNPs. We examined MNPs that were infected with either vector or *CYFIP1*, differentiated *in vitro* for 2 weeks, and subsequently exposed to 5 nM



rapamycin or vehicle (DMSO) for 24 hours. Qualitative and quantitative analyses of morphology showed that rapamycin treatment was associated with a normalization of neurite length, neurite branch length, overall neurite branch number, and cell size (Figure 5D and E). At the molecular level, although CYFIP1 was unchanged after 6 hours of rapamycin treatment, the ratio of phosphorylated mTOR to total mTOR was similar to that observed at baseline (Figure 5F and G). Importantly, we also observed a dramatic decrease in p-S6 (Ser 235/236) expression, a main downstream effector of mTOR and a readout of mTOR signaling activity, in the rapamycin-treated MNPs (Figure 5H and I).

Although no drop in CYFIP1 levels were observed after 6 hours of rapamycin, by 24 hours a reduction in CYFIP1 levels and a further decrease in total mTOR levels were observed (Figure S4). Thus, based on these time course data, a parsimonious mechanism for the reversal of morphological abnormalities associated with CYFIP1 over-expression is an effect on CYFIP1 levels through mTOR mediated p-S6 activation, which precedes changes in CYFIP1 protein levels.

To determine whether these results were relevant to human brain, we then examined whether levels of CYFIP1 and p-S6 proteins differed between 15q11-13 duplication carriers with ASD and non-carrier controls. Based on the above data, one would expect increase p-S6 in those with elevated CYFIP1 levels. Consistent with this prediction, we observed an approximate 2.5-fold upregulation of CYFIP1 and p-S6 in carriers (Figure 5J and K, Raw discrete expression data for individual patients; Figure S8), suggesting that mTOR activity is sensitive to *CYFIP1* dosage in human brain, as was observed in mouse. Our findings support a model (Figure 5L), where overexpression of CYFIP1 in neurons results in activation of mTOR signaling, which in turn gives rise to a series of morphological abnormalities.

## Discussion

Identifying individual contributory genes within multi-gene risk loci is essential for a mechanistic understanding of disease pathogenesis in ASD. The data presented here provide strong support for pathological consequences as a result of overexpression of *CYFIP1*, located at 15q11.2. Previous work has shown that deletions encompassing this four genes locus are associated with increased risk for ASD, intellectual disability and schizophrenia [27, 28]. Consistent with an important role in disease, we show that increased *CYFIP1* dosage gives rise to characteristic pathological cellular phenotypes in multiple model systems. Both *in vitro* and *in vivo*, overexpression of CYFIP1 resulted in a reduction in neurite length and longest neurite branch length, as well as an increase in neurite branch number and cell size. Golgi staining of transgenic mouse brain revealed alterations in dendritic spine morphology and density, outcomes observed with mutations in several other monogenic forms of ASD, including *Fmr1* [59], *Pten*[60], *Neurexins* [61] and *Neurologin1* [62]. We also investigated the potential mechanisms underlying these effects by characterizing expression differences in frontal cortex between transgenic mice overexpressing *Cyfp1* and wild-type controls. These genome-wide analyses, confirmed by functional experiments carried out *in vitro*, revealed a previous unknown link between CYFIP1 and mTOR signaling.

CYFIP1 was first identified through its functional interaction with the FMRP [29, 33], an RNA binding protein critical for the stability, intracellular transport, and translation of hundreds of target mRNAs [63]. Importantly, triplet repeat (CGG) expansions in the 5' UTR of *FMR1*, the gene that encodes FMRP, give rise to Fragile X syndrome, in which a higher than expected proportion of individuals have an ASD. FMRP target mRNAs [64], together with FMRP itself, form a complex with CYFIP1, which acts as an intermediary to elongation initiation factor 4E (eIF4E). CYFIP1 binding to eIF4E prevents the assembly of the multi-protein complex that mediates translational initiation [34]. Neuronal excitation mediated via BDNF activation of TrkB and/or signaling by metabotropic glutamate receptors, results in the dissociation of the FMRP-CYFIP1 complex from eIF4E, permitting translation. Interestingly, mice harboring mutations in *Fmr1* are known to have an increase in immature mushroom-shaped dendritic spines [65], the opposite to what we saw following CYFIP1 overexpression. Moreover, recent reports demonstrate that elimination of S6K1 in another model was able to rescue the abnormal excess of immature dendritic spines [66], also pointing to mTOR as an important mediator of the abnormal morphology. These results, together with a recent report found that immature spines were increased in number in *Cyfp1*-silenced neurons when compared to controls [36] suggests that FMRP and CYFIP1 operate cooperatively via mTOR, and support the notion that CYFIP1 action is sensitive to dosage.

We also observed that overexpression of CYFIP1 reduces levels of Phosphatase and tensin homolog on chromosome ten (PTEN), and conversely that knockdown of CYFIP1 increases PTEN levels. This is interesting, because it is well known that individuals harboring germline mutations in *Pten* often present with ASD and intellectual disability [57, 67, 68]. Moreover, *Pten* mutant mice, considered a model for ASD, show a subset of cellular abnormalities closely paralleling those we report here [69]. As a result of either CYFIP1 overexpression or *Pten* knockout in brain, both cell size and synapse density were increased relative to controls. Previous work has also demonstrated that inhibition of mTOR by CCI-779, a rapamycin derivative, was also able to rescue neuronal hypertrophy seen in *Pten* mutant mice [70]. This is consistent with our finding that rapamycin was able to rescue the morphological abnormalities we observed resulting from CYFIP1 overexpression. That abnormalities associated with both models can be rescued via inhibition of mTOR, suggests convergent, if not conserved mechanisms in these monogenic forms of ASD associated with protein translation dysregulation.

Another monogenic form of ASD, Tuberous Sclerosis, may also be mediated, at least partially, by abnormal mTOR signaling. Previous work showed that mosaic induction of *Tsc1* loss in neural progenitor cells resulted in the development of highly enlarged giant cells with enlarged vacuoles [71]. Consistent with this effect being mediated by mTOR, Carson and coworkers showed that postnatal rapamycin treatment completely reversed these phenotypes and rescued the mutants from epilepsy and premature death, despite prenatal onset of *Tsc1* loss and mTOR activation in the developing brain [72]. While our manuscript was under review a separate paper was published that looked at the consequence of *Cyfp1* overexpression on the morphology of neurons grown *in vitro* (hippocampal neurons from e18 rats grown for 21 days) [73]. Abnormalities they observed with regard to spine

morphology were different to those we saw in each of our transgenic mouse strains (layer II/III cortical neurons from adult Tg8 and Tg24 strains). Results from Pathania, M et al., are also unexpected in that they are inconsistent with the notion that reductions and increases in *CYFIP1* dosage have opposite effects on mTOR signaling (Figure 5A and B). More work will be required to determine whether apparent discrepancies are the result of differences relating to species, brain region, timing, culture vs. whole animal systems, or some other as yet unknown factor. Together, these results are consistent with the notion that regulation of local protein synthesis through PI3K and mTOR/S6K are likely to be of broad relevance to the mechanisms underlying the ASDs and related disorders. This is particularly interesting given the finding that previous work in post-mortem brain suggests that average spine density is increased in idiopathic ASDs [52]. Based on our results, we need to consider two potential mechanisms for rapamycin's reversal of *CYFIP1* induced morphologic abnormalities (Figure 5L). The first is a direct mTOR pathway effect on cytoskeletal proteins. The second is an indirect effect via mTOR activity and p-S6 diminishing *CYFIP1* levels, which, in turn, would normalize the morphological abnormalities we observed. Given that *CYFIP1* is known to regulate the actin cytoskeleton Rho GTPase via Rac1 [33] and recently published data [74] indicating that mice heterozygous for deletions showed rapamycin insensitive enhancements in long term depression it may also be that other abnormalities resulting from altered *Cyfp1* dosage may result from perturbation of distinct signaling pathways.

Although further work will be required to fully understand how loss and gain of *CYFIP1* impacts the mTOR cascade in the developing brain to increase risk for neuropsychiatric disorders, our work supports the notion that drugs acting on mTOR might be useful in individuals harboring either 15q11-13 duplications or smaller CNVs or sequence variants that result in changes in *CYFIP1* levels. Moreover, given the apparent relationship between *CYFIP1* dosage and expression in human transformed lymphoblastoid cell lines and brain, there is the possibility that mRNA expression levels in lymphoblastoid cell lines could be used as a biomarker to identify patients most suitable for treatment. It will be interesting to test the ability of rapamycin-like agents to ameliorate clinical outcomes in patients harboring genetic variants that encompass *CYFIP1*. At the same time, because both duplications and deletions at 15q11-13 appear to increase the risk for neuropsychiatric disease or lead to intellectual disability, careful attention to *CYFIP1* levels will likely be needed. Our data and published work from others shows that reductions and increases in *CYFIP1* levels give rise to opposite effects with regard to mTOR signaling, neuronal morphology, and structural aspects of disease related brain regions [27, 36]. Broadly speaking, these data are consistent with observations at other CNV loci including 5q35, 7q11.23, 16p11.2 where deletions and duplications have opposing effects on disease related endophenotypes, but at the same time, each increases risk for cognitive disability [75-79].

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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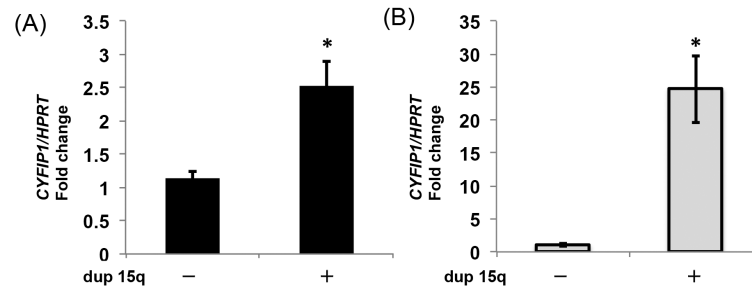
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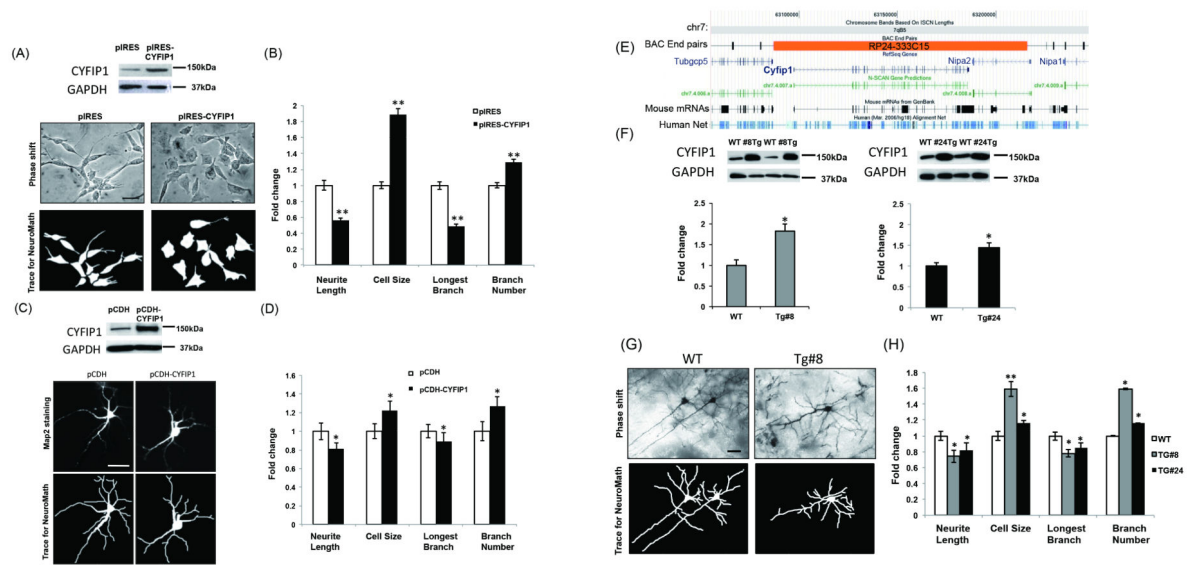
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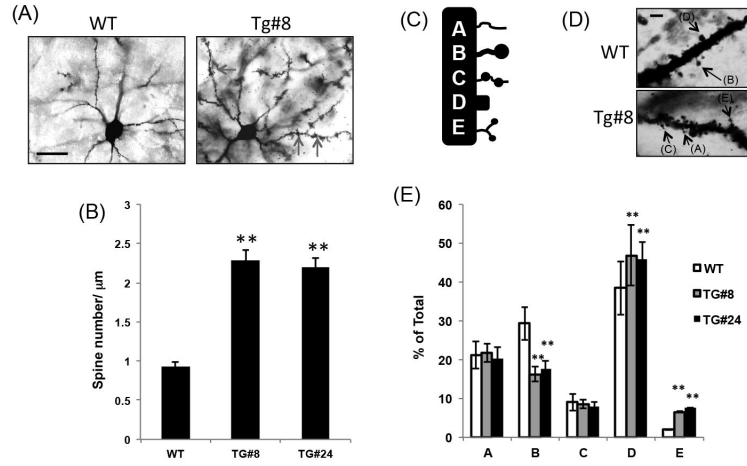


**Figure 1. Gains at 15q11-13 are associated with increased *CYFIP1* levels in transformed lymphoblastoid cell lines and postmortem brain**  
 Relative expression of *Cyfp1* mRNA was assessed by qPCR using *Hprt* as a reference. (A) In EBV transformed lymphoblastoid cell lines, levels were increased 2.5-fold in 15q11-13 duplication carriers (n=4) versus controls (n=6). (B) In postmortem superior temporal gyrus (STG, Brodmann's area 41/42), levels were increased 24-fold in carriers (n=3) versus controls (n=5). \* = p<0.05

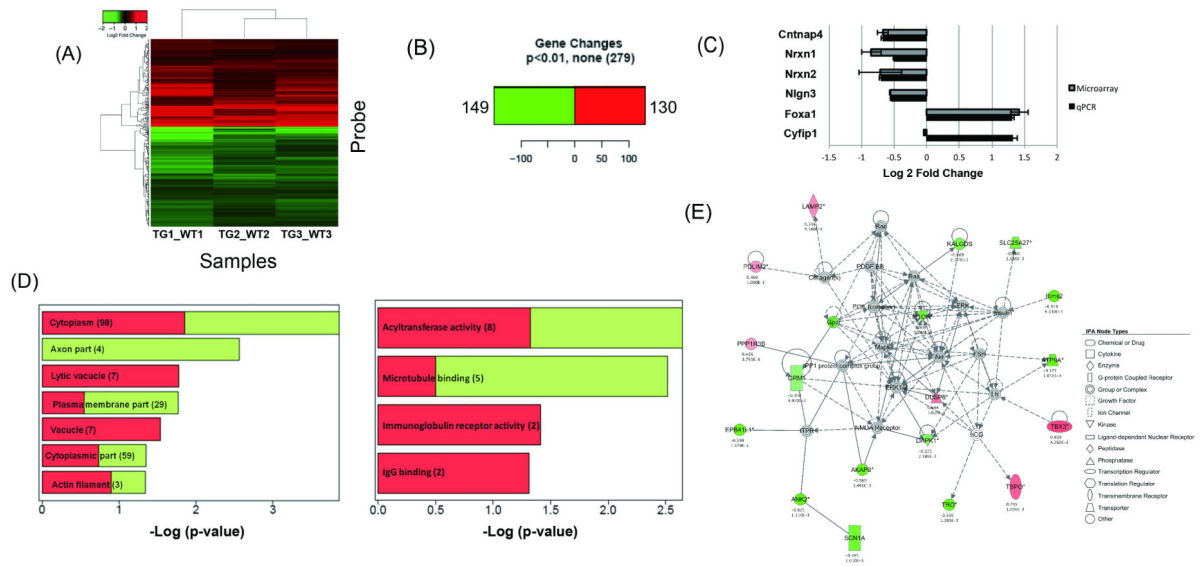


**Figure 2. Overexpression of *Cyfip1* gives rise to increased cell size and abnormal neurite outgrowth in neuronal cells both *in vitro* and *in vivo***

(A) SY5Y cells were transfected with either an empty pIRES vector or pIRES-CYFIP1 and protein levels assessed by Western blot (top panel). CYFIP1 protein levels were found to be increased in pIRES-CYFIP1 relative to pIRES vector. Characterization of cell morphology using NeuroMATH software identified morphological abnormalities in pIRES-CYFIP1 relative to pIRES vector (bottom panel, scale bar = 10 $\mu$ m). (B) Overexpression of CYFIP1 resulted in significant differences in neurite length (reduced ~ 50%), cell size (increased ~ 80%), longest neurite branch length (reduced ~ 60%), and overall neurite branch number (increased ~ 30%). (C) Differentiated mouse neuronal progenitors (dMNPs) were infected with either an empty pCDH vector or pCDH-CYFIP1 and protein levels were assessed by Western blot (top panel). CYFIP1 protein levels were found to be increased in pCDH-CYFIP1 relative to pCDH vector. Characterization of cell morphology of MAP2 stained dMNPs using NeuroMATH software identified morphological abnormalities in pCDH-CYFIP1 neurons relative to pCDH vector infected cells (bottom panel; scale bar = 10 $\mu$ m). (D) Results qualitatively indistinguishable from those for SY5Y cells were seen in CYFIP1-overexpressing dMNPs, with significant differences in neurite length (reduced ~ 20%), cell size (increased ~ 20%), longest neurite branch length (reduced ~ 10%), and overall neurite branch number (increased ~ 20%). (E) To determine the consequences of *Cyfip1* overexpression *in vivo*, we used a *Cyfip1* spanning bacterial artificial chromosome to generate transgenic mouse strains in which gene dosage was increased. (F) CYFIP1 protein levels were assessed by Western blot using samples obtained from adult hippocampus. Protein levels were found to be increased in transgenic mice from two independent transgenic strains (Tg#8 and #24) relative to control animals. (G and H) Paralleling the effects we observed *in vitro*, comparisons between transgenic mice and controls identified significant differences in neurite length (reduced ~ 20% in both strains), cell size (increased ~ 60% and 10% in Tg#8 and #24, respectively), longest neurite branch length (reduced ~ 20% in both strains), and overall neurite branch number (increased ~ 50% and 15% in Tg#8 and #24, respectively). Scale bar = 50 $\mu$ m. \* =  $p < 0.05$  and \*\* =  $p < 0.01$ .

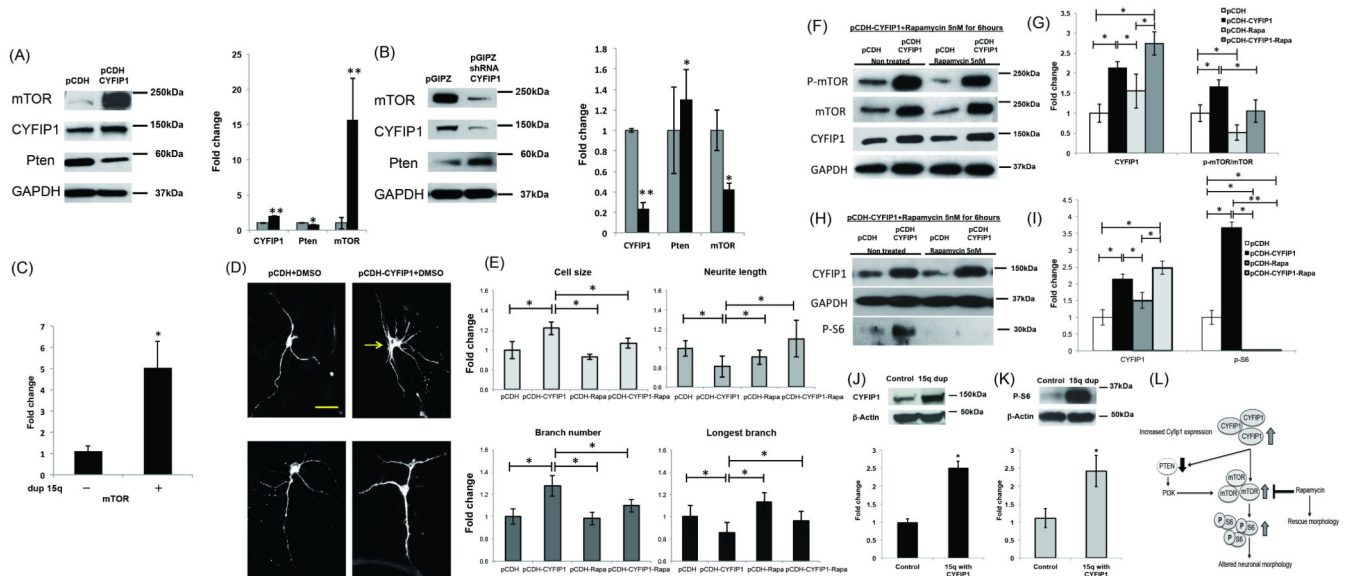


**Figure 3. Overexpression of *Cyfip1* in vivo results in increased spine density and increased proportion of pyramidal neurons showing a mature spine morphology**  
 Golgi staining of sections from two month old frontal cortex (layer II-III pyramidal neurons) revealed a (A) qualitative (red arrows; scale bar = 15μm), and (B) quantitative (two-fold increase) in spine density in transgenic mice (Tg#8 and #24 strains) relative to controls. (C) Categorical classification of spines highlight a (D) qualitative (black arrows; scale bar = 3μm), and (E) quantitative shift in the proportion of spines within each category in transgenic mice from either strain relative to controls. In both transgenic strains differences in the overall proportion of mushroom type immature thin spines (Type B; reduced ~50% in both strains), mature stubby spines (Type D; increased 15% in both strains), and abnormal spines (Type E; increased ~ 500% in both strains) were observed relative to control mice. No differences between transgenic mice and wild-type controls were observed for either immature thin spines (Type A) or immature double mushroom-type spines (Type C) \*\* = p<0.01.



**Figure 4. Global expression analyses on embryonic brain in transgenic and control mice identify differentially expressed genes and highlight mTOR**

(A) Heatmap depicting genes differentially expressed between embryonic day 15 Tg #8 *Cyfip1* BAC transgenic mice and controls at  $p < 0.01$ . Here and in all following panels part of this figure, red represents upregulation, whereas green represents downregulation (Log<sub>2</sub> scale). (B) 130 genes were found to be upregulated and 149 genes downregulated. (C) Between group differences for five of the top differentially expressed genes, including the autism-related genes *Nrxn1* and *Nlgn3*, were verified by qPCR. Fold changes are expressed as  $\log_2 \pm$  standard error. (D) Analysis of all differentially expressed genes ( $p < 0.01$ ) by gene ontology suggests significant enrichment in genes within actin filament and microtubule binding categories (Log<sub>2</sub> scale). (E) Ingenuity pathway analysis implicated the mTOR pathway (second “top” network,  $p = 0.05$ ). As above, red corresponds to upregulation, green represents downregulation, and grey genes not differentially expressed. A solid line indicates a direct interaction and a dashed line indicates an indirect interaction. A line without an arrowhead indicates binding. Where arrowheads are present, filled and dotted lines illustrate direct and indirect actions of one protein on another. IPA node type suggests the protein categories.



**Figure 5. Rapamycin, an mTOR inhibitor, rescues the morphological and molecular abnormalities resulting from CYFIP1 overexpression**

(A) Mouse neuronal progenitors (MNPs) were infected with either an empty pCDH vector or pCDH-CYFIP1, allowed to differentiate for two weeks, and then protein levels assessed by Western blot. 2-week differentiated MNPs (dMNP) overexpressing CYFIP1 showed increased levels of mTOR and decreased levels of PTEN. (B) MNPs were infected with either an empty pGIPZ vector or pGIPZ-CYFIP1 shRNA, allowed to differentiate for two weeks, and protein levels assessed by Western blot. CYFIP1 knockdown was found to result in decreased levels of mTOR and increased levels of PTEN. (C) mTOR mRNA levels in brain tissue samples obtained from patients with *CYFIP1* containing 15q11-13 duplications (n=3) and controls (n=5) were evaluated by qPCR, and found to be increased 6-fold in affected carriers. (D) Characterization of cell morphology of MAP2 stained dMNP using NeuroMath software identified morphological abnormalities in pCDH-CYFIP1 relative to pCDH vector infected cells (top panel; scale bar = 10 $\mu$ m). MNPs infected with pCDH-CYFIP1, allowed to differentiate for two weeks, and then treated for 24 hours with 5nM rapamycin were qualitatively (bottom panel), and (E) quantitatively similar to pCDH vector infected cells. pCDH: infected with vector and 24hours vehicle (DMSO) treatment, pCDH-CYFIP1: infected with *CYFIP1* vehicle (DMSO) treatment, pCDH-Rapa: infected with vector and 24 hours 5 nM rapamycin treatment, pCDH-CYFIP1-Rapa: infected with *CYFIP1* and 24 hours 5 nM rapamycin treatment. (F through I) Western blots were performed on the lysates of pCDH-CYFIP1 and pCDH vector infected dMNP treated for 6 hours with either DMSO or 5 nM rapamycin. We observed a normalization of mTOR activity at 6 hours after rapamycin treatment; the ratio of phosphorylated mTOR to total mTOR and levels of phosphorylated S6 Kinase were similar to that observed at baseline. Although rapamycin treated dMNP infected with pCDH-CYFIP1 showed a decrease of mTOR activity at 6 hours, CYFIP1 levels remained elevated. (J and K) Brain tissue samples obtained from *CYFIP1* containing 15q11-13 duplication carriers (n=3) and non-carrier controls (n=4) were evaluated by Western blot. CYFIP1 and p-S6 protein levels were found to be 2.5-fold increased in affected carriers. \* = p<0.05 and \*\* = p<0.01. (L) Scheme of



CYFIP1/mTOR signaling. Scheme showing the link between overexpression in CYFIP1 and neuronal morphological changes via overactivated mTOR. Results support a model where overexpression of CYFIP1 in neurons results in activation of mTOR signaling, which in turn gives rise to a series of morphological abnormalities. In response to CYFIP1 overexpression, levels of the negative regulator PTEN are reduced, mTOR levels are increased, and S6 activity is increased. Gross morphological abnormalities resulting from CYFIP1 overexpression are rescued by treatment with the mTOR inhibitor rapamycin, demonstrating causality.