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The Gene Expression Database for Mouse Development (GXD): putting developmental expression information at your fingertips

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Abstract

Because molecular mechanisms of development are extraordinarily complex, the understanding of these processes requires the integration of pertinent research data. Using the Gene Expression Database for Mouse Development (GXD) as an example, we illustrate the progress made towards this goal, and discuss relevant issues that apply to developmental databases and developmental research in general. Since its first release in 1998, GXD has served the scientific community by integrating multiple types of expression data from publications and electronic submissions and by making these data freely and widely available. Focusing on endogenous gene expression in wild-type and mutant mice and covering data from RNA *in situ* hybridization, *in situ* reporter (knock-in), immunohistochemistry, RT-PCR, northern blot and western blot experiments, the database has grown tremendously over the years in terms of data content and search utilities. Currently, GXD includes over 1.4 million annotated expression results and over 260,000 images. All these data and images are readily accessible to many types of database searches. Here we describe the data and search tools of GXD; explain how to use the database most effectively; discuss how we acquire, curate, and integrate developmental expression information; and describe how the research community can help in this process.

Keywords

literature curation; data integration; online resource; *in situ* hybridization; immunohistochemistry; anatomy ontology

INTRODUCTION

Gene expression data provide crucial insights into the molecular mechanisms of development, differentiation, and disease. However, the data are voluminous, complex, and heterogeneous. They are generated by many different laboratories and scattered through thousands of publications. Without the help of centralized databases, it is impossible to keep abreast of all this information, let alone to access and search these data in a cohesive and integrated way.

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The Gene Expression Database for Mouse Development (GXD) was one of the first databases to address these critical issues (Ringwald et al., 1994; Ringwald et al., 1999). As a mammalian model system, the mouse is heavily used for developmental research. Tissues from all developmental stages and from different mouse strains and mutants are subject to detailed expression studies. For many years, the GXD project has been curating mouse developmental expression data from the published literature, as well as acquiring data through direct submissions and collaboration with efforts that generate pertinent expression data at a large-scale. For example, GXD has incorporated the *in situ* hybridization data from the EurExpress (Diez-Roux et al., 2011), GenePaint (Visel et al., 2004), GUDMAP (Genitourinary Molecular Anatomy Project; Harding et al., 2011), and BGEM (Brain Gene Expression Map; Magdaleno et al., 2006) projects. GXD integrates data from all these different sources and, as a major component of the Mouse Genome Informatics (MGI) resource (www.informatics.jax.org), combines the expression information with genetic, functional, and phenotypic data. Therefore, these expression data are readily accessible to many types of database searches (Smith et al., 2014; Finger et al., 2011; Blake et al., 2014).

Here we describe the current status of GXD with a particular emphasis on its search utilities. Further, we illustrate issues of data curation and integration that apply to developmental research in general.

CONCEPTS, SCOPE, AND EXPRESSION DATA CONTENT

GXD covers all developmental stages and all organ systems and comprises expression data from wild-type and mutant mice. The main focus is on endogenous gene expression data during development. As data accumulate, GXD aims to provide increasingly complete information about which RNA and protein products are made from a given gene, where and when these products are expressed, and how their expression varies in different mouse strains and mutants. Because there is no single assay type that can provide answers to all these questions, GXD is designed as a system that can integrate different types of expression data. At this point, GXD captures data from RNA *in situ* hybridization, *in situ* reporter (knock-in), immunohistochemistry, RT-PCR, northern blot and western blot experiments.

Expression patterns (i.e. the time and space of gene expression) are described in a standardized way by using an extensive anatomical ontology that has been developed in collaboration with the eMouseAtlas (EMAP) project (Hayamizu et al., 2013; Bard et al., 1998). The ontology is structured hierarchically, allowing the integrated description of expression patterns from experiments with differing spatial resolution, as well as enabling searches that include anatomical structures and their substructures (described in detail below).

As illustrated in Figure 1, each database record describes the results obtained for each specimen, including the level and pattern of expression for each anatomical structure examined, as well as the molecular probe and the experimental conditions used. Images of the original expression data accompany the annotations whenever possible. By capturing these elemental data, different types of expression data can be represented and integrated in a robust manner. Genes and mutant alleles are recorded using official nomenclature, and all

data are associated with a reference. Genes, probes, alleles, anatomical structures, and references are key points of data integration that tie genetic, genomic, expression, functional, and phenotypic data closely together, enabling search capabilities unique to GXD/MGI.

GXD currently includes detailed expression records for > 13,800 genes. The data come from > 67,000 expression assays and include > 1.4 million expression result annotations and > 260,000 images of primary expression data. 82 % of the data are from RNA *in situ* hybridization studies and 10 % from RT-PCR experiments, reflecting the detailed spatial resolution and sensitivity required in developmental expression studies. In addition to data from different strains of wild-type mice, GXD currently includes expression data from > 1,900 mouse mutants. In the following we illustrate different ways to effectively search these data.

SEARCHING FOR EXPRESSION DATA

The GXD Home Page

One way to access the expression data for a given gene is to use the Quick Search, accessible from all MGI pages, in order to find the corresponding gene detail page and then follow the links in the “Expression” section (see Fig. 2). However, GXD’s search tools allow you to do much more than look up expression data gene by gene. To take full advantage of GXD, we recommend the GXD Home Page as a starting point: <http://www.informatics.jax.org/expression.shtml>. This page gives access to GXD’s search forms and provides more information about GXD, including user help, news announcements, and instructions for submitting data to GXD. We now describe the use of three of GXD’s search forms in more detail.

Gene Expression Data Query

The Gene Expression Data Query form (<http://www.informatics.jax.org/gxd>) is the most versatile and powerful search form for detailed expression data in GXD. It offers two different query utilities via the Standard Search and Differential Expression Search tabs.

The Standard Search tab (shown in Fig. 3) allows investigators to ask both broad and very specific questions pertinent to their research interests and quickly obtain matching expression results. Researchers can find expression data for specific genes, or for sets of genes as defined by their biological function or by association with annotated mouse phenotypes or human diseases. They can search for expression data for specific anatomical structures and/or specific developmental stages. Searches can be limited to expression data from wild-type mice or to expression data from mice that have been mutated in specific genes. Assay type(s) can also be chosen. Further, using combinations of the query parameters described above makes it possible to formulate complex queries. For example, one could search for genes associated with DiGeorge Syndrome expressed in heart; for genes involved in left/right asymmetry that are expressed in the primitive streak; or for genes involved in signaling pathways that are expressed in the eye of *Pax6* mutant mice.

The Differential Expression Search tab allows querying for genes that are expressed in one anatomical structure but not in another and/or at some developmental stages but not others. For example, one can search for genes that are expressed in the epithalamus but not in the hypothalamus; or for genes that are expressed at the morula stage (Theiler stage 3) but not at the blastocyst stages (Theiler stages 4 and 5). These searches will return a list of genes whose expression has been shown to be absent (not detected), as well as genes whose expression has not been analyzed or recorded in the database for the specified structures and stages. The two cases can be distinguished by filtering the results summaries for instances where expression was not detected (see below).

The Standard Search and Differential Expression Search return a results page with tabbed summaries for assay results, assays, genes, and images (see Fig. 4). This allows users to see the desired level of detail and focus on the data they are most interested in. The four tabs indicate the number of records returned, respectively. One can narrow down the returned results further by modifying the search or by filtering the expression results by Anatomical System, Assay Type, Detected/Not Detected, Theiler Stage, or Wild type/Mutant. Filters can be selected and de-selected to interactively revise and refine the data summaries. Most of the data columns of the summaries are sortable. Further, data from the Assay Results and Genes summaries can be downloaded in text or spreadsheet formats allowing upload into other applications.

Each row in the assays, assay results, and image summaries includes links to detailed expression records. Figure 1 shows, as an example, an entry for an immunohistochemistry experiment, illustrating the detail in which data in GXD are annotated. These Assay Detail pages display image panes together with their annotations. Each image pane is linked to the corresponding image detail page that shows the publication figure, which is often multi-paned, thus presenting the image pane in context. Image detail pages might also include links to external resources. For example, detail pages for the EurExpress and GenePaint images link back to the corresponding records at these sites where utilities such as serial-section browsers and high-resolution images are available. Some image detail pages also have links to the Edinburgh Mouse Atlas and Gene Expression Database (EMAGE). GXD makes all its RNA *in situ* and immunohistochemistry images, together with their annotations, available to EMAGE (Richardson et al., 2014), so that the expression patterns can be mapped and queried spatially. If *in situ* images have been spatially mapped, GXD provides links from image detail pages to the corresponding mapped images in EMAGE. However, only a subset of the wild-type expression images can be spatially mapped (the 3D atlas is based on wild-type embryos). The standardized text-based description of expression patterns employed by GXD is essential as it allows the representation and integration of all types of expression data from wild-type and mutant mice, as well as the further integration with other data that relate to anatomy, such as mouse phenotype and human disease data.

In short, the Gene Expression Data Query form enables researchers to quickly find specific sets of expression data. Query summaries can be interactively refined and lead to the detailed expression records.

Mouse Developmental Anatomy Browser

As discussed above, GXD and EMAP have developed an extensive ontology for mouse developmental anatomy to describe the time and space of gene expression in a standardized way. This enables intuitive and comprehensive expression searches at variable anatomical resolution. The ontology is structured hierarchically. Each term can have multiple parents, allowing the anatomy to be represented and searched from different perspectives. For example, “brain” is represented as part of the “nervous system” and as part of the “head”. Searches for expression in the “nervous system” or in the “head” will both return expression data for the brain.

The newest version of Mouse Developmental Anatomy comprises all 28 Theiler stages, including the embryonic (TS 1-26), newborn (TS 27), and postnatal mouse (TS 28). There is one “abstract” (non-stage specific) representation of the mouse anatomy that lists the anatomical structures for all Theiler stages, as well as the stage range during which each structure is present. We refer to these terms as EMAPA terms (A for abstract). In addition, there are 28 stage-specific representations (derived from the abstract version). Currently, the ontology includes more than 6500 EMAPA terms and more than 25,000 stage-specific terms (referred to as EMAPS terms). Expression data in GXD are annotated to the stage-specific anatomical terms.

The Mouse Developmental Anatomy Browser (<http://www.informatics.jax.org/vocab/gxd/anatomy>) allows users to navigate through this anatomical ontology, view a specific anatomical structure, and to obtain the expression data associated with that structure and its substructures (see Fig. 5). The browser consists of three interactive sections: a search section for finding and selecting anatomical terms; a detail section that provides additional information for a selected term and lets users toggle between the abstract and stage-specific versions of the anatomical ontology; and a tree view section that allows users to view the terms in their hierarchical context, to expand and collapse branches of the hierarchy, and to retrieve the expression data for specific anatomical structures. The expression result summaries accessed from the Mouse Developmental Anatomy Browser have the same features and utilities as those obtained upon using the Gene Expression Data Query Form. They can be sorted and filtered and link to the detailed expression records, as described above.

Terms from the mouse developmental ontology are also being used to label the EMAP 3D atlas and to describe expression patterns in EMAGE (Armit et al., 2012). Efforts to establish cross-references to anatomical ontologies from other model organism and human databases are underway (Mungall et al., 2012; Dahdul et al., 2012; Hayamizu et al., 2012; Van Slyke et al., 2014; Segerdell et al., 2013, Costa et al., 2013). This will foster the comparative analysis of expression patterns between model organisms used in developmental research.

Gene Expression Literature Query

GXD provides researchers with an effective way to search the mouse embryonic expression literature. Our curators survey journals to find all publications that contain the types of data that GXD collects. As a first annotation step, they index all these publications with regard to

the genes that have been studied, the expression assay types used, and the ages analyzed. These annotations are then combined with bibliographic information from PubMed to generate the Gene Expression Literature Index. The index is complete and up-to-date from 1990 onwards for all major journals (~150). Currently, the index covers > 21,800 references reporting expression data for > 15,100 genes. An average of 1,200 papers are added to the index per year. All this information is available via the Gene Expression Literature Query form (<http://www.informatics.jax.org/gxdlit>). The query form allows researchers to quickly find publications that report specific sets of expression data (see Fig. 6). These searches are more effective and complete than PubMed searches because the index uses standard nomenclature for genes, assay types, and ages and because the annotations are based on the entire article, including supplemental data.

ACQUIRING AND CURATING DATA

Data are acquired from the literature, via electronic data submissions, and through collaborations with projects that generate pertinent expression data at a large-scale. All data are reviewed by curators and annotated in standardized ways by making extensive use of controlled vocabularies and ontologies as illustrated in Fig. 1. This is a prerequisite for the data integration and search capabilities that GXD provides.

GXD is the only effort that curates mouse developmental expression data from the literature in a systematic way. The first step in this process is populating the Gene Expression Literature Index as described above. The next step is to annotate the details of the expression data from these articles, including supplemental data, as illustrated in Fig. 1. Papers are prioritized for detailed annotation based on the information in the Gene Expression Literature Index. Prioritization criteria include: genes for which there is no detailed data in the database; genes associated with human diseases; genes which, based on the number of publications, are underrepresented in the detailed portion of the database; publications that include a large amount of expression data; and publications that characterize the developmental expression pattern of a given gene in detail (often the first publications for that gene).

Large-scale electronic data sets are reviewed by a combination of computational and manual checks to make sure that the probe-to-gene assignments are up-to-date and that data entries are complete. Nomenclature issues, data ambiguities, and questions that might arise during the mapping of submitted data to ontologies are resolved together with the data provider before the data are loaded into GXD.

All data, including those from large-scale data sets, gain significant value when integrated with the other data in GXD and the larger MGI resource. All data can be explored together and searches can be done that are unavailable elsewhere. Further, GXD ensures that data and data connections are maintained and kept up-to-date after large-scale projects have ended and gene models or the gene names used in the literature have changed.

Authors base their conclusions on much more primary data than will appear in a publication, they are specialists in their fields, and they have detailed knowledge about specific experimental parameters and potential pitfalls that is not available to curators. For these

reasons, GXD curators rely on the text of the manuscript, or of data submissions, to derive annotations of expression patterns. These annotations are standardized by using terms from the anatomical ontology (which is expanded as required). However, curators do not interpret images themselves, thereby trying to derive additional expression results that have not been asserted in the paper. For the reasons discussed above, interpretation of images by curators to infer expression or absence of expression would be problematic and error-prone. Instead, GXD displays standardized text annotations together with the corresponding images so users can see the original expression data (see Fig. 1). We have obtained permissions to include images from all major developmental journals, as well as from many others. Currently, over 70% of the result annotations are accompanied by images. Also in cases where GXD does not have the permission to include images, we annotate the data based on the information in the manuscript and provide a reference to the corresponding figure panes.

It is thus important to note that the detail of expression annotations in GXD is determined by the details provided in the text of published articles and electronic data submissions. Detailed descriptions of expression results will lead to more fine-grained and complete annotations. Further, a clear correspondence in the publication between stated expression results, probes, specimens, and images facilitates data annotation. Concise, unambiguous descriptions of expression patterns result in robust annotations. Issues of completeness, consistency, data identity, and clarity apply not only to GXD but to all databases that curate data from the published scientific literature. This includes other developmental organism databases such as GEISHA (Antin et al., 2014), Xenbase (James-Zorn et al., 2013), ZFIN (Howe et al., 2013), FlyBase (St. Pierre et al., 2014), and Wormbase (Harris et al., 2014). Publications that present data, results, and interpretations clearly and identify genes, strains, reagents, and methods adequately can significantly facilitate data curation and dissemination, thus greatly benefiting the researchers who generated the data initially as well as improving data access to the research community at large.

THE CASE FOR ELECTRONIC DATA SUBMISSION

Both journal publications and databases are essential for the research enterprise, and they fulfill complementary roles. Journal publications excel in the narrative descriptions of novel discoveries and discussions of data implications and future applications. However, they usually include only part of the data pertinent to a study due to space constraints or because authors report only the results most relevant to the papers' narrative. Expression (or non-expression) in anatomical systems peripheral to the narrative is often not reported. Further, journal publications cannot and do not provide a framework for data integration and regular updates, with the result that the data cannot be searched adequately. Databases, on the other hand, do not have space constraints or an incentive to focus on specific scientific narratives; and they excel at the handling, maintenance, and integration of data, and in making them accessible to searches. Currently, data generated from conventional (non large-scale) laboratories are primarily reported in free-text descriptions in journal publications. Unless database curators extract data from these publications and bring them into formats proper for integration and searching, these data will remain poorly integrated into our scientific knowledge base. This is clearly not a good solution for making research data accessible.

An obvious and effective solution to address this problem is to combine journal publications with electronic data submissions. Following the example of sequence and array expression data, the types of developmental expression data discussed here could be submitted to pertinent public databases in conjunction with publications. Submitters would receive accession numbers that can be cited in the publications. Without causing an undue burden on submitters, they could, for example, provide an expanded legend for each image that includes more complete descriptions of the expression patterns seen. Thus, electronic submission can include more primary data and results can be described in greater detail and in more standardized ways. While such submissions would still require review and annotation by database curators, it would significantly facilitate the acquisition, integration, and dissemination of data. Having the data widely accessible to many types of searches would benefit everyone: the journal (because all the data would be tied to the publication reference), the submitter (because their data would be more accessible), and the scientific community as a whole (because of the increased database content and utility).

GXD accepts electronic data submissions for the types of data it collects; see the “Send us your data” tab at the bottom of the GXD Home Page for instructions (http://www.informatics.jax.org/mgihome/GXD/GEN/gxd_submission_guidelines.shtml). Other developmental databases, such as FlyBase, GEISHA, ZFIN and Xenbase, are welcoming electronic data submissions as well. The value of all these community resources is proportional to the amount and the quality of data they contain, and it is time for the research community to realize the potential of electronic data submissions.

FUTURE DIRECTIONS

GXD will continue to acquire data from the literature and electronic data submissions and to improve its search and display capabilities. Later this year, we plan to add interactive matrix views of expression data. Tissue-by-developmental stage matrices will provide high-level overviews of the spatio-temporal expression patterns of genes. Tissue-by-gene matrices will enable a comparison of expression patterns. Both types of matrices can be expanded (and collapsed) along the tissue axis, based on the hierarchical organization of the anatomy. Thus, these matrix views will provide users with intuitive high-level summaries of expression results from where they can drill down to more detail. GXD and the larger MGI resource are also planning to implement additional links to other developmental organism databases. In the shorter term, gene-based links will enable users to look up and compare expression data (and other types of data) for orthologous genes. As described above, we are also working on establishing cross-references between anatomical ontologies to enable, in the longer term, an anatomy-based comparison of developmental gene expression patterns.

USER SUPPORT AND OUTREACH

User Support and GXD curatorial staff actively seek to provide presentations, demonstrations and training sessions on GXD and the larger MGI resource at many meetings. Upon request user support personnel also provide on-site visits and training workshops, as well as remote interactive sessions. GXD is continually looking for ways to improve the user experience and, thus, direct user interactions, surveys and collaborations

provide valuable feedback on the project from biologists, biomedical researchers, ontology developers and computational biologists who utilize our data. Further, the GXD Advisory Board provides critical input and guidance for the project.

DATABASE ACCESS AND CONTACT INFORMATION

The GXD home page can be accessed directly at <http://www.informatics.jax.org/expression.shtml>; via the MGI home page (<http://www.informatics.jax.org/>) by following the topic “Gene Expression Database (GXG)”; or from any other page within MGI by clicking the “Expression” tab of the navigation bar. For web-based access to GXG and MGI, we recommend Firefox, Chrome, or Safari. Online help is available via the FAQs on the GXG home page and by clicking on the question marks in the upper right corner of most GXG pages. User Support personnel can be contacted via email to mgi-help@jax.org or via the “Contact Us” link in the navigation bar. This article focuses on web-based access to the GXG database proper. Additional computational tools to explore GXG’s expression data are available, such as the GXG BioMart (accessible via the GXG home page) and MouseMine (www.mousemine.org). MouseMine, in particular, offers advanced iterative search and filtering capabilities.

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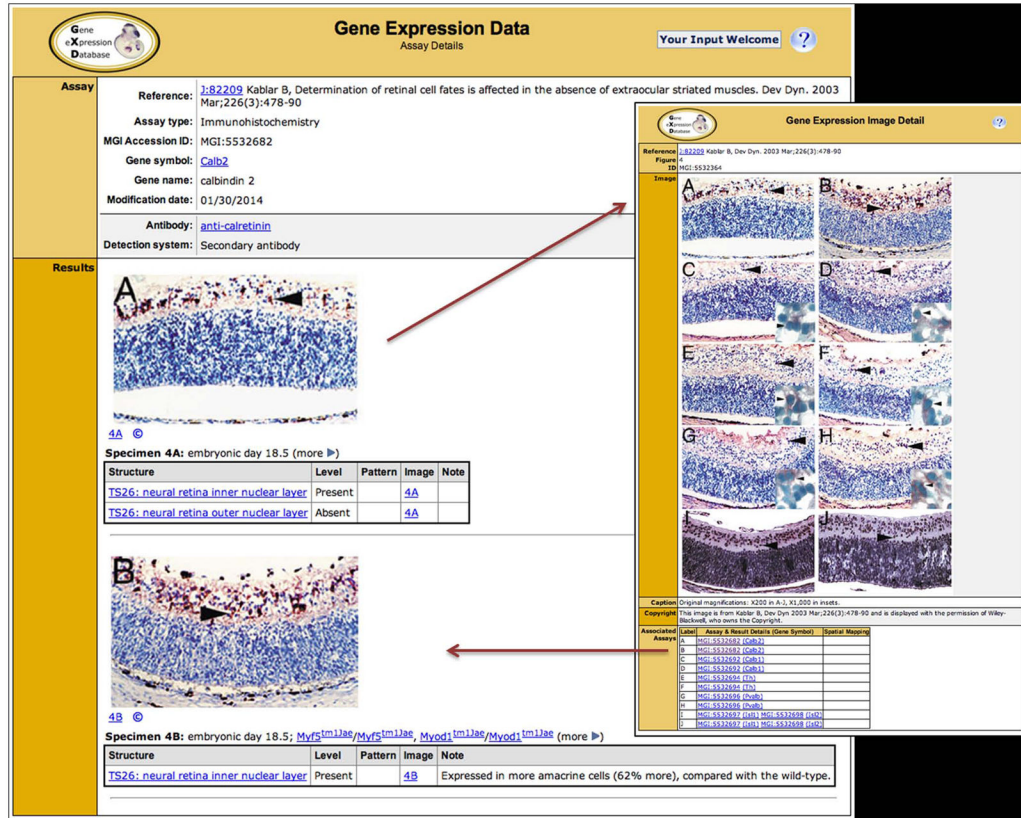


Figure 1. Left: GXD assay details pages provide detailed annotations of experimental parameters and expression results. Shown is an example of an Assay Detail page for an immunohistochemistry experiment illustrating the detailed content of expression results in GXD. The Assay section reports the reference from which the data were derived, the assay type, the gene analyzed, and the antibody used, with links to more details about the antibody, reference, and gene. The Results section reports the tissue (Theiler stage and anatomical structure) analyzed, as well as the level and pattern of expression, as described by the authors. Images of the original expression data are displayed together with the corresponding annotations whenever possible. Major specimen details such as the age and mutant alleles are always displayed on this page. Other information, such as genetic background, sex, and specimen preparation method, is accessible via the ‘more’ toggle. Right: Image detail pages are accessed by clicking on the image panes or pane labels shown in the assay details record. They show the entire figure, as published, providing the scientific context. As shown in the Associated Assays table of this example, several genes were studied in the same or similar tissue sections; the assay IDs and gene symbols link to the corresponding annotations for each image pane. While the image detail pages provide visual context, it is the detailed and standardized text annotations shown on the GXD assay detail pages (left) that make the expression data, including image data, accessible to searching.

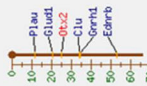
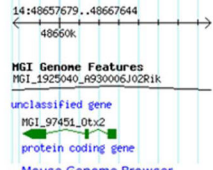

Symbol Name ID	Otx2 orthodenticle homolog 2 MGI:97451																
Synonyms	E130306E05Rik																
Feature Type	protein coding gene																
Genetic Map	Chromosome 14 25.36 cM Detailed Genetic Map ± 1 cM Mapping data(9) 																
Sequence Map	Chr14:48657679-48667644 bp, - strand From VEGA annotation of GRCm38 <input type="text" value="9966 bp"/> ± <input type="text" value="0"/> kb flank VEGA Genome Browser Ensembl Genome Browser UCSC Browser NCBI Map Viewer 																
Vertebrate homology	HomoloGene:11026 Vertebrate Homology Class 1 human; 1 mouse; 1 rat; 1 chimpanzee; 1 cattle; 1 dog; 1 chicken; 1 zebrafish Gene Tree: Otx2																
Human homologs	Human Homolog OTX2, orthodenticle homeobox 2 NCBI Gene ID 5015 nextProt AC NX_P32243 Human Synonyms CPHD6, MCOP55 Human Chr (Location) 14q22.3; chr14:57267425-57277194 (-) <i>GRCh37.p13</i> Disease Associations (2) Diseases Associated with Human OTX2																
Alleles and phenotypes	All alleles(47) : Targeted(33) Gene trapped(14) Mice homozygous for a knock-out allele exhibit embryonic lethality during organogenesis due to abnormal gastrulation and embryonic patterning in the brain and heart.																
Gene Ontology (GO) classifications	All GO classifications: (52 annotations) Process anatomical structure development, anterior/posterior pattern specification, ... Component cytoplasm, growth cone, ... Function DNA binding, protein binding, ... External Resources: FuncBase																
Expression	 Literature Summary: (404 records) Data Summary: Results (1370) Tissues (410) Images (466) Theiler Stages: 2 , 3 , 4 , 5 , 7 , 8 , 9 , 10 , 11 , 12 , 13 , 14 , 15 , 16 , 17 , 18 , 19 , 20 , 21 , 22 , 23 , 24 , 25 , 26 , 28 <table border="1"> <thead> <tr> <th>Assay Type</th> <th>Results</th> </tr> </thead> <tbody> <tr> <td>Immunohistochemistry</td> <td>55</td> </tr> <tr> <td>RNA in situ</td> <td>1116</td> </tr> <tr> <td>In situ reporter (knock in)</td> <td>81</td> </tr> <tr> <td>Northern blot</td> <td>1</td> </tr> <tr> <td>Western blot</td> <td>9</td> </tr> <tr> <td>RT-PCR</td> <td>105</td> </tr> <tr> <td>RNase protection</td> <td>3</td> </tr> </tbody> </table> cDNA source data(25) External Resources: Allen Institute GENSAT GEO ArrayExpress	Assay Type	Results	Immunohistochemistry	55	RNA in situ	1116	In situ reporter (knock in)	81	Northern blot	1	Western blot	9	RT-PCR	105	RNase protection	3
Assay Type	Results																
Immunohistochemistry	55																
RNA in situ	1116																
In situ reporter (knock in)	81																
Northern blot	1																
Western blot	9																
RT-PCR	105																
RNase protection	3																
Molecular reagents	All reagents																
Other database links	VEGA cDNA source data(25) External Resources: Allen Institute GENSAT GEO ArrayExpress																

Figure 2. Gene Detail pages summarize, and provide access to, all the information about a given gene in MGI, together with extensive links to external resources. The upper portion of the Otx2 gene detail page is shown. The expression section (expanded) indicates the types and amount of expression information available for the gene and provides links to the corresponding summary pages. Links to databases that store mouse expression data not available in GXD are provided as well: the Allen Institute (Lein et al., 2007), GENSAT (Heintz, 2004), GEO (Barret et al., 2013) and ArrayExpress (Petryszak et al., 2014).

Gene Expression Data

Standard Search | Differential Expression Search

Search Reset

Genes Find expression data for...

One gene e.g., *Shh* or *kit* oncogene
Genes with similar nomenclature e.g., *Hoxa**

OR

A set of genes defined by

- Function e.g., *cell-cell signalling*
- Phenotype e.g., *obese*
- Disease e.g., *DiGeorge Syndrome*

transcription factor binding - Function

Anatomical structure or stage Find assay results where expression is detected in not detected in either

Anatomical Structures: AND / OR Developmental Stages (dpc):

diencephalon

Use Theiler Stages Use Ages (dpc)

TS 15 (9.0-10.25 dpc)
TS 16 (9.5-10.75 dpc)
TS 17 (10.0-11.25 dpc)
TS 18 (10.5-11.25 dpc)
TS 19 (11.0-12.25 dpc)
TS 20 (11.5-13.0 dpc)
TS 21 (12.5-14.0 dpc)

Mutant / wild type Find expression data in ...

Specimens mutated in gene:
 Wild type specimens only
 All specimens

Assay types Find expression data in any assay type

Immunohistochemistry RNA in situ
 In situ reporter (knock in) RNase protection
 Northern blot RT-PCR
 Nuclease S1 Western blot

Search Reset

Figure 3.

The Gene Expression Data Query Form features two search tabs: Standard and Differential Expression. The Standard Search, shown here, enables queries for expression data using one or more parameters. The Genes section allows users to find expression data for a specific gene or for a set of genes based on their function [as defined by Gene Ontology terms (Gene Ontology Consortium, 2010)], their association with mouse phenotypes [as defined by Mammalian Phenotype Ontology terms (Smith et al., 2012)], or their association with human diseases [as defined by Online Mendelian Inheritance in Man (OMIM) terms (Amberger et al., 2011)]. In the anatomical/stage section, one can search for expression data in specific anatomical structures and/or developmental stages, and one can specify whether (1) all results should be returned or only those where expression was (2) detected (i.e. present) or (3) not detected (i.e. absent). Anatomical searches combine word searching and hierarchical searching. For example, a search for expression in “diencephalon” would return expression annotations for all anatomical structures that have “diencephalon” as part of their name as well as for all their anatomical substructures such as “thalamus”. In the mutant/wild type section one can limit the searches to expression data from wild-type mice or search for gene expression in specific mutants. The Assay types section allows selection of expression data types. Auto-fill utilities help to find appropriate search terms. The illustrated search asks for ‘transcription factor binding’ genes ‘detected’ in the ‘diencephalon’ at ‘Theiler stages 17, 18, or 19’. The corresponding search results page is shown in Fig. 4. The Differential Expression Search (not shown) allows searching for genes that are expressed in

some anatomical structures but not others and/or at some developmental stages but not others.

Author Manuscript

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Gene Expression Data

Click to modify search

Results

You searched for: **Function: transcription factor binding** Filter expression by: Anatomical System Assay Type Detected? Theiler Stage Wild type? << first < prev 1 2 3 next > last >> 100 Showing results(s) 1 - 100 of 264

Genes annotated to **Function: transcription factor binding** includes subterms
Detected in diencephalon includes synonyms & substructures
 at developmental stage(s): **(TS:17 or TS:18 or TS:19)**

Genes (108) Assays (177) Assay results(264) Images (330)

Export: Text File Excel File

Gene	Result Details	Assay Type	Anatomical System	Age	Structure	Detected?	Images	Mutant Allele(s)	Reference
Arnt	data (MGI:4440508)	RNA in situ	nervous system	E11.5	TS19: diencephalon	Yes			1:81291 Altola MH, et al., J Histochem Cytochem. 2003 Jan;51(1):41-54
Arnt2	data (MGI:4440509)	RNA in situ	nervous system	E11.5	TS19: diencephalon	Yes			1:81291 Altola MH, et al., J Histochem Cytochem. 2003 Jan;51(1):41-54
Ascl1	data (MGI:4440509)	RNA in situ	nervous system	E11.5	TS19: diencephalon	Yes			1:156017 Yokoyama S, et al., Dev Cell. 2009 Dec;17(6):836-48
Ascl1	data (MGI:4440509)	RNA in situ	nervous system	E11.5	TS19: diencephalon	Yes	3D		1:14701 Guillemot F, et al., Mech Dev. 1993 Aug;42(3):171-85

Expression Images in MGI are copyrighted; click on an image for details about their use.

Gene	Assay Type	Result Details
Gbx2	Immunohistochemistry	2E
Nkx2-1	Immunohistochemistry	2E

Gene	Assay Type	Result Details
Mef2a	Immunohistochemistry	3B

Gene	Assay Type	Result Details
Nes	Immunohistochemistry	10C,G
Ptx2	Immunohistochemistry	10A,G

Figure 4.

GXD data summaries can be viewed at different levels of detail and interactively refined and sorted. Searches using the Gene Expression Data Query form return a page with four tabbed summaries for the assay results, assays, genes, and images that match the search parameters. The assay results tab (upper) is displayed by default. It lists the gene studied, the assay type used, the anatomical system, age and tissue examined, indicates whether expression was detected, provides a link to the corresponding images, lists the mutant alleles of the specimen (if applicable), and provides the reference from which the data were derived. Links in the Result Details and Images columns lead to detailed expression records, such as the one shown in Fig. 1. Arrows in column headers indicate that the column is sortable (one set is circled). The assay results tab (as well as the genes tab) allows for the export of results in text and spread sheet formats (buttons in table header). The images tab (lower) shows all the images that match the search criteria, together with the gene(s) examined in that image and the assay type used and provides a link to the corresponding part of the detailed expression record. The expression summaries can be refined by using the 'click to modify search' button or by employing the filter options provided on the summary page. The content of all four tabbed summaries will change accordingly.

Figure 5.

The Mouse Developmental Anatomy Browser allows users to search for anatomical terms, to explore the anatomical hierarchies and locate specific anatomical structures in context, and to retrieve the expression data associated with these structures and their substructures. The anatomy search is facilitated by an auto-fill utility. As soon as a term is selected from the pick list, all matching anatomical structures are displayed in the search column, together with the developmental stage range during which these structures are present in the embryo. The best match is listed first and selected by default. Other matching terms can be selected by clicking. Upon selection, the Anatomical Tree View and the Anatomical Term Detail section are updated and the selected anatomical structure is highlighted. Using the Tree View, users can explore the ontology further by expanding and collapsing branches. Clicking on a term in the tree view will select (and highlight) that term. The number of expression results associated with each term is listed; following that link will lead to an expression summary page similar to the one shown in Fig. 4. The initial tree view shows the abstract version of the anatomy ontology. Accordingly, the associated expression results will include the annotations for all developmental stages at which the selected anatomical structure is present. The developmental stage pick list in the Anatomical Term Detail section allows users to toggle between stage-independent terms and tree views and stage-specific terms and tree views. Stage-specific terms will link to the expression results for the anatomical structure at that specific stage.

Gene Expression Literature Search

Gene SymbolName: Examples: one gene e.g., Shh
a set of genes with similar nomenclature e.g., Hoxa*

Assay type: **ANY**
 In situ protein (section)
 In situ RNA (section)
 In situ protein (whole mount)
 In situ RNA (whole mount)

Age: (days post conception)
 E4 E4.5 E5 E5.5 E6 E6.5 E7

Author: Any Author(s) First Author Last Author

Journal: Examples: Proc Natl Acad Sci USA (See [GDS](#))
J Cell Mol Med

Year: Examples: 2008
1990-2004
-2007 (from the earliest reference through 2007)
2009 (from 2009 through the present)

Text: Examples: gastrulation, morphogenesis
"pattern formation"

In Title In Abstract

Gene Expression Literature Summary

You searched for:
Marker SymbolName: Hoxb4
Ages: 7.5 or 8 or 8.5 or 9 or 9.5 or 10 or 10.5 or 11 or 11.5 or 12 or 12.5 or 13 or 13.5 or 14 or 14.5 or 15 or 15.5
66 matching records from 66 references.

Summary by Age and Assay: Numbers in the table indicate the number of results matching the search criteria.

Age	E7.5	E8	E8.5	E9	E9.5	E10	E10.5	E11	E11.5	E12	E12.5	E13	E13.5	E14	E14.5	E15	E15.5
In situ protein (section)	1	3	2		3	1	2							1	1		1
In situ RNA (section)		1	1	5	1	6		8		10		2		6			2
In situ protein (whole mount)			1	2	2	3	1										
In situ RNA (whole mount)	1	6	1	12	1	5		5		5		2		1			2
Northern blot								1		2		1		3			1
Western blot														1			1
RT-PCR					1	3	1	1		1	1					2	2
cDNA clones			1														
RNase protection										1				1			

Summary by Gene and Reference: Number indicates the number of results matching the search criteria recorded for each reference.
* Indicates detailed expression data entries available

Hoxb4 homeobox B4 (Synonyms: Hox-2.6)

Results	Reference
1*	J:33159 Akasaka T, Kanno M, Balling R, Mieza MA, Taniguchi M, Koseki H, A role for mel-18, a Polycomb group-related vertebrate gene, during the anteroposterior specification of the axial skeleton. <i>Development</i> . 1996 May;122(5):1513-1522
6*	J:35305 Bogue CW, Lou LJ, Vasavada H, Wilson CM, Jacobs HC, Expression of Hoxb genes in the developing mouse foregut and lung. <i>Am J Respir Cell Mol Biol</i> . 1996 Aug;15(2):163-71
1*	J:34198 Boulet AM, Capecci MR, Targeted disruption of hoxc-4 causes esophageal defects and vertebral transformations. <i>Dev Biol</i> . 1996 Jul 10;177(1):232-49
4*	J:83257 Brend T, Gilthorpe J, Summerbell D, Rigby PW, Multiple levels of transcriptional and post-transcriptional regulation are required to define the domain of Hoxb4 expression. <i>Development</i> . 2003 Jun;130(12):2717-28
3*	J:23135 Burke AC, Nelson CE, Morgan BA, Tabin C, Hox genes and the evolution of vertebrate axial morphology. <i>Development</i> . 1995 Feb;121(2):333-46
3	J:119560 Choi MY, Romer AI, Hu M, Lepourcelet M, Mechoor A, Yesilaltay A, Krieger M, Gray PA, Shivasani RA, A dynamic expression survey identifies transcription factors relevant in mouse digestive tract development. <i>Development</i> . 2006 Oct;133(20):4119-29
2*	J:14757 Detmer K, Lawrence HJ, Largman C, Expression of class I homeobox genes in fetal and adult murine skin. <i>J Invest Dermatol</i> . 1993 Oct;101(4):517-22
4	J:193244 Di Meglio T, Kratochwil CF, Vilain N, Loche A, Vitobello A, Yonehara K, Hrycaj SM, Roska B, Peters AH, Eichmann A, Wellik D, Ducret S, Rijli FM, Ezh2 orchestrates topographic migration and connectivity of mouse precerebellar neurons. <i>Science</i> . 2013 Jan 11;339(6116):204-7
1*	J:153498 Diez-Roux G, Banfi S, Sultan M, Geffers L, Anand S, Rozado D, Magen A, Canidio E, Pagani M, Peluso I, Lin-Marq N, Koch M, Billo M, Cantello I, Verde R, De Masi C, Bianchi SA, Cicchini J, Perroud E, Mehmeti S, Dagand E, Schrimmer S, Numbberger A, Schmidt K, Zwingmann C, Brieske N, Springer C, Hernandez AM, Herzog S, Grabbe F, Sieverding C, Fischer B, Schrader K, Brockmeyer M, Dettmer S, Helbig C, Alunni V, Battaini MA, Mura C, Hennrichsen CN, Garcia-Lopez R, Echevarria D, Puelles E, et al., A high-resolution anatomical atlas of the transcriptome in the mouse embryo. <i>PLoS Biol</i> . 2011;9(1):e1000582
2	J:50343 Ding J, Yang L, Yan YT, Chen A, Desai N, Wynshaw-Boris A, Shen MM, Crip to is required for correct orientation of the anterior-posterior axis in the mouse embryo [see comments]. <i>Nature</i> . 1998 Oct 15;395(6703):702-7
1*	J:41551 Dupe V, Ghyselinck NB, Wendling O, Chambon P, Mark M, Key roles of retinoic acid receptors alpha and beta in the patterning of the caudal hindbrain, pharyngeal arches and otocyst in the mouse. <i>Development</i> . 1999 Nov;126(23):5051-6

Gene Expression Literature Detail

Symbol Name: **Hoxb4**
homeobox B4
ID: MGI:96185

Reference: **J:35305** Bogue CW, Lou LJ, Vasavada H, Wilson CM, Jacobs HC, Expression of Hoxb genes in the developing mouse foregut and lung. *Am J Respir Cell Mol Biol*. 1996 Aug;15(2):163-71

Detailed expression data for these assays: 20 results

* Indicates gene expression was analyzed but not necessarily detected.

Age	E9.5	E10.5	E11.5	E12.5	E14.5
In situ protein (section)					
In situ RNA (section)					
In situ protein (whole mount)					
In situ RNA (whole mount)					
In situ reporter (knock in)					
Northern blot					
Western blot					
RT-PCR					
cDNA clones					
RNase protection					
Nuclease S1					
Primer Extension					

Figure 6. Querying the embryonic mouse expression literature. The Gene Expression Literature Search (top left) allows querying of the embryonic expression literature for genes and ages analyzed and expression assay types used, as well as querying for bibliographic information or specific words in the title or abstract. A portion of the summary return for the query formulated in the figure is displayed at right. The table at the top, “Index Results by Age and Assay”, shows the number of matching records grouped by the age of the specimen and the assay type used. The lower portion of the page, “Index Results by Gene and Reference”, lists the citations for the references where the matching index results were reported, as well as the number of matching results contained therein. Entries marked with an * indicate they have been annotated in detail in GXD. Links on this summary page access detail pages (lower left). These pages display the expression information about the gene contained in the reference and provide links to gene and reference detail pages, as well as to the detailed expression data from the paper if they have already been annotated in GXD.

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