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Glucokinase regulatory protein: complexity at the crossroads of triglyceride and glucose metabolism

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Abstract

Purpose of review—*Glucokinase regulator* (*GCKR*) encodes glucokinase regulatory protein (GKRP), a hepatocyte-specific inhibitor of the glucose-metabolizing enzyme glucokinase (GCK). Genome-wide association studies have identified a common coding variant within *GCKR* associated with multiple metabolic traits. This review focuses on recent insights into the critical role of GKRP in hepatic glucose metabolism that have stemmed from the study of human genetics. This knowledge has improved our understanding of glucose and lipid physiology and informed the development of targeted molecular therapeutics for diabetes.

Recent findings—Rare *GCKR* variants have effects on GKRP expression, localization, and activity. These variants are collectively associated with hypertriglyceridaemia but are not causal. Crystal structures of GKRP and the GCK–GKRP complex have been solved, providing greater insight into the molecular interactions between these proteins. Finally, small molecules have been identified that directly bind GKRP and reduce blood glucose levels in rodent models of diabetes.

Summary—*GCKR* variants across the allelic spectrum have effects on glucose and lipid homeostasis. Functional analysis has highlighted numerous molecular mechanisms for GKRP dysfunction. Hepatocyte-specific GCK activation via small molecule GKRP inhibition may be a new avenue for type 2 diabetes treatment, particularly considering evidence indicating GKRP lossof-function alone does not cause hypertriglyceridaemia.

Keywords

diabetes therapy; *glucokinase regulator*; glucokinase regulatory protein; glucose homeostasis; hypertriglyceridaemia

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INTRODUCTION

Glucokinase regulator (*GCKR*) and its cognate gene product glucokinase regulatory protein (GKRP) have been associated, both biologically and genetically, with several key metabolic pathways. In the liver, GKRP forms an inhibitory complex with glucokinase (GCK), the enzyme responsible for regulating the uptake and storage of dietary glucose [1,2]. Mechanistic studies have shown that disruption and reformation of this complex occurs in response to direct binding of glucose to GCK, and fructose 1-phosphate (F1P) (a by-product of dietary fructose metabolism) and fructose 6-phosphate (F6P, which accumulates during gluconeogenesis and glycogenolysis) to GKRP [3-5]. In this way, the GCK–GKRP complex acts as a metabolic 'switch' capable of initiating energy storage and release pathways in response to periods of feeding and fasting. Genome-wide association studies (GWAS) have identified multiple associations with the common coding variant p.P446L in GKRP, including type 2 diabetes (T2D) and an inverse modulation of fasting plasma glucose and triglyceride levels [6-8]. This variant affects GKRP function via a range of molecular mechanisms that ultimately result in GCK activation under conditions of normoglycaemia or hypoglycaemia [9,10]. As we move closer to an era of personalized medicine, our ability to deliver targeted and efficacious treatments for diabetes and other metabolic diseases will depend on combined insights from human genetics, molecular and structural biology, and whole-animal physiology. This review will focus on recent studies that have extended our understanding of the physiological impact of GKRP dysregulation on glucose and lipid homeostasis, and the ways in which we may be able to manipulate GKRP to provide new treatments for T2D.

GENETIC VARIATION IN GLUCOKINASE REGULATOR HAS BEEN IMPLICATED IN A WIDE RANGE OF METABOLIC TRAITS

Human genetics has played an important role in our understanding of lipid phenotypes. GWAS have identified more than 150 loci associated with lipid levels and other lipid-related traits, and the advent of next-generation sequencing has allowed for the identification of additional contributing rare and low-frequency variants [7,11-18]. The results of these studies have offered new insight into the molecular mechanisms that underpin lipid metabolism and have opened up new avenues towards the development of novel therapeutics.

The first human genetic evidence of a role for *GCKR* in lipid metabolism was the association of a more than 400 kb genomic region, encompassing *GCKR*, with plasma triglyceride levels in Europeans via GWAS [7]. A subsequent combination of imputation and fine-mapping highlighted the nonsynonymous single-nucleotide polymorphism (SNP) rs1260326 [c.1403 C>T, p.P446L, minor allele frequency (MAF) 0.34] as the likely causal variant [6,19]. Interestingly, this SNP was associated with an inverse modulation of fasting glucose and triglyceride levels, a finding that has since been replicated in other populations [8,13,18,20-23]. To date, SNP rs1260326 (or the intronic proxy SNP rs780094) has been associated with more than 25 metabolic traits, including T2D risk, fasting insulin, total cholesterol, and nonalcoholic fatty liver disease, as well as circulating levels of numerous

metabolites such as uric acid, C-reactive protein, creatinine, and albumin, indicating that it is a highly pleiotropic gene [14,24-39].

GLUCOKINASE REGULATORY PROTEIN HAS A CLEAR BIOLOGICAL ROLE IN GLUCOSE HOMEOSTASIS

The abovementioned insights are of particular interest in light of an already substantial body of evidence indicating a biological role for GKRP in glucose homeostasis. As its name suggests, GKRP was first identified as a protein that bound GCK and inhibited its activity in rodent hepatocytes [2]. GCK is a key regulator of glucose disposal and storage in both liver and pancreatic beta-cells, and responds to increases in circulating glucose concentration by initiating a signalling cascade that results in insulin secretion from the beta-cell and subsequent hepatic glucose uptake and storage [1]. The vast majority of human GKRP is expressed in the liver, suggesting that its primary effect on GCK activity occurs in this organ [9]. It is, however, also expressed at low levels in other tissues, although it does not appear to be appreciably expressed in beta-cells where GCK is clearly in excess [9,40].

The liver is responsible for the clearance and disposal of approximately 25–35% of dietary glucose, and responds to periods of hypoglycaemia by initiating gluconeogenic and glycogenolytic pathways designed to maintain blood glucose concentrations at homeostatic levels [41-43]. Formation of the GCK–GKRP inhibitory complex is disrupted by glucose, resulting in translocation of GCK from the hepatocyte nucleus to the cytoplasm where it can assist in glucose clearance [44-48]. A similar effect is also achieved in the presence of the phosphate ester F1P, an intermediary product in the conversion of dietary fructose [2-4]. Conversely, F6P, a by-product of gluconeogenesis and glycogenolysis, enhances the interaction between GCK and GKRP, thus promoting nuclear retention and inactivation of GCK during periods of hypoglycaemia [2,3]. This shuttling mechanism effectively creates a nuclear reserve of GCK capable of mobilizing rapidly in response to fluctuations in intrahepatic glucose levels (Fig. 1). Interestingly, GKRP also acts as a post-translational stabilizer of cellular GCK, as *Gckr*−/− mice display WT-like *Gck* transcript levels but reduced GCK protein expression and activity, as well as lower hepatic glycogen concentrations and a reduced ability to respond to an acute glucose load [47,49].

MOLECULAR MECHANISMS AND PHENOTYPIC IMPACT OF NATURALLY OCCURRING GLUCOKINASE REGULATOR MISSENSE VARIANTS

Functional analysis of the p.P446L GKRP variant provided us with the first mechanistic insights into the ways in which natural variation in *GCKR* may influence its cellular behaviour. A combination of *in vitro* and cell-based assays has shown that this variant has a reduced ability to sequester and inhibit GCK and a blunted response to F6P, both of which favour the generation of free and active cytoplasmic GCK [9,10]. Detailed biophysical experiments recently published by Zelent *et al.* [50^{\blacksquare}] suggest that the P > L substitution at this residue may have an overall effect on the structure of GKRP, resulting in destabilization of the GCK binding interface and altered phosphate ester binding. This explains the inverse correlation between fasting plasma glucose and triglycerides for this variant, as increased

Raimondo et al. Page 4

hepatic GCK activity would result in lower blood glucose levels and maintenance of energystoring triglyceride and glycogen synthesis pathways under conditions of normoglycaemia or hypoglycaemia (Fig. 2) [51].

Studies by both Johansen *et al.* and Rees *et al.* have recently extended the catalogue of naturally occurring *GCKR* variants, and provided further insight into the mechanistic basis to GKRP dysfunction across the allelic spectrum [14,52]. In one of the first studies to identify rare $(MAF < 0.01)$ variants of (potentially) large individual effect in a gene already implicated in a particular trait via GWAS (in this case, plasma lipid levels), Johansen *et al.* [14] performed targeted exome sequencing of *GCKR* in individuals with extreme lipid phenotypes. They identified an excess of rare *GCKR* variants in hypertriglyceridaemic cases (defined as having fasting plasma triglyceride levels $>95th$ percentile) relative to control individuals with normal lipid levels. Rees *et al.* [52] sequenced the exons of *GCKR* in 800 individuals from the ClinSeq cohort, who were recruited on the basis of an increased risk for coronary atherosclerosis, and identified a further 10 novel rare coding variants.

Functional studies have subsequently demonstrated a mutational mechanism for the vast majority of these variants, via a range of effects including cellular GKRP expression and localization, GCK interaction and inhibition, and phosphate ester binding [52,53■]. In one study, reclassification of variants based on empirical determination of pathogenicity increased the strength of the association with plasma triglycerides, total cholesterol, and LDL-Cholesterol, indicating that modulation of GKRP function has a demonstrable effect on clinical phenotype [52]. Interestingly, these empirical findings were repeatedly underpredicted by several widely used *in silico* prediction algorithms, emphasizing the need for robust analytical pipelines in the correct functional classification of novel variants $[14,52,53$].

GLUCOKINASE REGULATORY PROTEIN AS A TARGET FOR DIABETES THERAPY

Although rare loss-of-function *GCKR* variants are associated collectively with hypertriglyceridaemia, extended family studies have demonstrated that they generally do not co-segregate with triglyceride levels [53■]. This most likely reflects the complex heritability of lipid traits, to which rare functional *GCKR* alleles are contributory rather than deterministic, and the influence of additional genetic and environmental factors on the penetrance of lipid phenotypes [13,14,54]. It is also encouraging news in light of a recent report describing antidiabetic effects for two small molecule GKRP inhibitors in rodents $[55$ ^{\blacksquare}].

Lloyd *et al.* [55^{■■}] used a cell-free high-throughput screening approach to identify a lead molecule (AMG-1694) that activated hepatic GCK via direct binding to GKRP, and found it to be a robust nuclear-to-cytoplasmic translocator of GCK in Wistar and Zucker diabetic fatty (ZDF) rats and primary rat hepatocytes. The drug had a specific effect on blood glucose levels – without affecting insulin or triglycerides – over a 24-h period in diabetic (ZDF) rats and did not alter blood glucose levels in normoglycaemic (Wistar) rats. Further optimization of AMG-1694 resulted in a drug that displayed greater efficacy in mice

Raimondo et al. Page 5

(AMG-3969), with similar translocatory effects in diet-induced obesity (DIO), *ob/ob, db/db* and normoglycaemic C56BL/6 mice. Once again, GCK translocation was matched by a corresponding decrease in blood glucose levels only in diabetic (DIO, *ob/ob* and *db/db*) mice [55■■,56]. The precise mechanism of action remains uncertain; however, the blood glucose-lowering effects appear to be the result of increased hepatic carbohydrate oxidation $[55$ ^{\blacksquare}].

These results are of interest due to the potential for reduced risk of hypoglycaemia in humans, which has been a hallmark of clinical trials of glucokinase activators (GKAs) over the last 10 years. GKAs are a class of small molecules that increase the affinity of GCK for glucose by directly binding a pocket distal to its active site, thus lowering the set point for glucose-stimulated insulin secretion in the beta-cell [57,58]. In the liver, GKA binding causes GCK to dissociate from its inhibitory complex with GKRP [59]. A number of adverse side-effects to long-term GKA treatment have been reported, including hypoglycaemic episodes (due to improper insulin secretion), decreased efficacy over time, and increased hepatic triglycerides, the latter most likely due to persistent GCK activity in the liver [57,60-63].

Data suggest that increased hepatic GCK activity due to loss of GKRP function has negative long-term effects on whole-animal glucose and lipid homeostasis, particularly in the context of high fat and high sugar diets [47,61]. In humans, the p.P446L variant is associated with a modest decrease in plasma glucose but a proportionately larger increase in plasma triglycerides (8.76 mg/dl per allele in [13]), indicating that a satisfactory reduction in blood glucose levels via hepatic GCK activation may be outweighed by a correspondingly larger increase in triglyceride synthesis and storage [6,7,19,23]. The same variant is also associated with increased 2-h glucose – most likely reflecting a diminution of the nuclear GCK pool capable of being mobilized in response to a glucose challenge – making the long-term effects of chronic GCK activation on both glucose and triglycerides via GKRP inhibition difficult to predict [10,26]. As GKRP inhibitors do not increase the basal affinity of GCK for glucose, the effect on triglycerides at low glucose concentrations may be reduced relative to GKAs. Ultimately, these data emphasize that human clinical trials of AMG-1694 and AMG-3969 – or any other small molecule disruptors of the GCK–GKRP complex – will have to be closely monitored for adverse side-effects on both glucose and triglycerides, particularly in view of the fact that diabetes patients are already likely to have unfavourable lipid profiles that may be exacerbated by chronic hepatic GCK activation.

THE FUTURE FOR TARGETED DRUG DESIGN AND GLUCOKINASE REGULATORY PROTEIN

The study by Lloyd *et al.* [55■■] was the first to utilize crystal structure information to elucidate the precise mechanism of small molecule interference of GKRP. They crystallized the human GKRP protein bound to AMG-1694 and sorbitol 6-phosphate (an open-chain analogue of F6P), and used this information to model the effects of AMG-1694 binding on the GCK binding interface [55■■,64■■]. The very first two crystallographic studies of GKRP, however, were simultaneously published by Pautsch *et al.* and Beck and Miller 8

months previously. These two groups provided atomic-level information on human GKRP bound to F1P, and the human GCK–rat GKRP complex bound to F6P [65■■,66■■].

Cumulatively, these studies give a comprehensive picture of the structural changes that occur on the GKRP scaffold in response to GCK and phosphate ester binding. They demonstrate that GKRP is a trilobal protein, consisting of two sugar isomerase (SIS) homology domains capped by an alpha-helical 'lid'. The GCK binding interface is positioned opposite the lid and is mediated by a small number of polar contacts and multiple hydrophobic interactions [64■■,65■■]. It is also distinct from the F1P–F6P binding site, which is located in a deeply buried cavity where the lid meets the SIS domains [65■■, 66■■]. GCK binds to GKRP in a 'super-open' conformation in which its active site remains disordered and is released from GKRP in response to glucose via rearrangement into a βhairpin structure [65■■]. Phosphate ester binding appears to modulate the strength of the interaction between the lid and the second SIS domain such that GCK binding is favoured or disfavoured depending on whether F6P or F1P is bound [65■■]. This detailed molecular information verifies several years' worth of indirect evidence on the nature of the GKRP protein fold that had been deduced via a range of indirect methods, including homology modelling, mutagenesis, and biochemical and biophysical analyses [5,67-75].

Crystal structure information is of enormous significance in the context of current and future efforts to design targeted molecular inhibitors of GKRP, and has already been instrumental in structure-guided optimization of the initial screening hit AMG-1694 [56,76,77]. This molecule binds between the N-terminus of GKRP and the first SIS domain, revealing a novel binding pocket – distinct from the phosphate ester binding site – that clearly influences GKRP activity. This finding exemplifies the opportunities for novel, targeted therapeutics that can be inferred from structural details of previously unrecognized binding motifs, although the precise mechanism of GCK–GKRP disruption upon AMG-1694 binding remains unclear [55■■,78]. Interestingly, the p.P446 residue resides at the Cterminal end of a structural loop that interacts with GCK, and rare loss-of-function variants recently characterized by Johansen *et al.* and Rees *et al.* are distributed throughout the GKRP structure, the most severe of which are located proximal to the phosphate ester binding site [14,52,53■]. The extent to which small molecule inhibition of GKRP mimics the structural effects of these variants is yet to be elucidated, although they appear to have the same overall mechanistic effect via GCK–GKRP complex disruption.

CONCLUSION

Recent discoveries from fields as diverse as genetics, structural and cellular biology, and whole-animal physiology have underscored the importance of GKRP in hepatic glucose and triglyceride metabolism. The initial finding of multiple metabolic associations for the common coding variant p.P446L has now been extended to include collective associations for rare variants of larger individual effect with plasma lipids, and detailed functional analyses indicate that GKRP is highly sensitive to alteration in its activity via a range of molecular mechanisms [7,9,10,14,52,53■]. Rare *GCKR* variants, however, do not appear to be sufficient to cause hypertriglyceridaemia, giving hope for the suitability of small molecule GKRP inhibitors as an appropriate molecular therapy for hyperglycaemia and T2D

[53■,55■■]. However, the long-term effects of increased hepatic GCK activity on glucose and triglycerides remain to be elucidated. In addition to recent atomic-resolution structural information about GKRP and the GCK–GKRP interaction, existing human genetic data should also prove useful in monitoring the potential side-effects of novel small molecule GKRP modulators [55■■,64■■-66■■]. Such a multifaceted approach should improve our understanding of the extent to which GKRP dysfunction is deterministic of lipid phenotypes, and the ways in which we therefore may be able to intervene and modulate its activity in a clinical setting.

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REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- ■■ of outstanding interest
- 1. Gloyn, AL.; Odili, S.; Buettger, C., et al. Glucokinase and the regulation of blood sugar. In: Matschinsky, FM.; Magnuson, MA., editors. Glucokinase and glycaemic disease: from basics to novel therapeutics. Karger; Basel: 2004. p. 92-109.
- 2. Van Schaftingen E. A protein from rat liver confers to glucokinase the property of being antagonistically regulated by fructose 6-phosphate and fructose 1-phosphate. Eur J Biochem. 1989; 179:179–184. [PubMed: 2917560]
- 3. Detheux M, Vandercammen A, Van Schaftingen E. Effectors of the regulatory protein acting on liver glucokinase: a kinetic investigation. Eur J Biochem. 1991; 200:553–561. [PubMed: 1889418]
- 4. Vandercammen A, Detheux M, Van Schaftingen E. Binding of sorbitol 6-phosphate and of fructose 1-phosphate to the regulatory protein of liver glucokinase. Biochem J. 1992; 286(Pt 1):253–256. [PubMed: 1520277]
- 5. Veiga-da-Cunha M, Van Schaftingen E. Identification of fructose 6-phosphate- and fructose 1 phosphate-binding residues in the regulatory protein of glucokinase. J Biol Chem. 2002; 277:8466– 8473. [PubMed: 11756407]
- 6. Orho-Melander M, Melander O, Guiducci C, et al. Common missense variant in the glucokinase regulatory protein gene is associated with increased plasma triglyceride and C-reactive protein but lower fasting glucose concentrations. Diabetes. 2008; 57:3112–3121. [PubMed: 18678614]
- 7. Saxena R, Voight BF, Lyssenko V, et al. Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. Science. 2007; 316:1331–1336. [PubMed: 17463246]
- 8. Dupuis J, Langenberg C, Prokopenko I, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. Nat Genet. 2010; 42:105–116. [PubMed: 20081858]
- 9. Beer NL, Tribble ND, McCulloch LJ, et al. The P446L variant in GCKR associated with fasting plasma glucose and triglyceride levels exerts its effect through increased glucokinase activity in liver. Hum Mol Genet. 2009; 18:4081–4088. [PubMed: 19643913]
- 10. Rees MG, Wincovitch S, Schultz J, et al. Cellular characterisation of the GCKR P446L variant associated with type 2 diabetes risk. Diabetologia. 2012; 55:114–122. [PubMed: 22038520]

- 11. Kathiresan S, Melander O, Guiducci C, et al. Six new loci associated with blood low-density lipoprotein cholesterol, high-density lipoprotein cholesterol or triglycerides in humans. Nat Genet. 2008; 40:189–197. [PubMed: 18193044]
- 12. Aulchenko YS, Ripatti S, Lindqvist I, et al. Loci influencing lipid levels and coronary heart disease risk in 16 European population cohorts. Nat Genet. 2009; 41:47–55. [PubMed: 19060911]
- 13. Teslovich TM, Musunuru K, Smith AV, et al. Biological, clinical and population relevance of 95 loci for blood lipids. Nature. 2010; 466:707–713. [PubMed: 20686565]
- 14. Johansen CT, Wang J, Lanktree MB, et al. Excess of rare variants in genes identified by genomewide association study of hypertriglyceridemia. Nat Genet. 2010; 42:684–687. [PubMed: 20657596]
- 15. Lange LA, Hu Y, Zhang H, et al. Whole-exome sequencing identifies rare and low-frequency coding variants associated with LDL cholesterol. Am J Hum Genet. 2014; 94:233–245. [PubMed: 24507775]
- 16. Peloso GM, Auer PL, Bis JC, et al. Association of low-frequency and rare coding-sequence variants with blood lipids and coronary heart disease in 56 000 whites and blacks. Am J Hum Genet. 2014; 94:223–232. [PubMed: 24507774]
- 17. Kathiresan S, Willer CJ, Peloso GM, et al. Common variants at 30 loci contribute to polygenic dyslipidemia. Nat Genet. 2009; 41:56–65. [PubMed: 19060906]
- 18. Willer CJ, Sanna S, Jackson AU, et al. Newly identified loci that influence lipid concentrations and risk of coronary artery disease. Nat Genet. 2008; 40:161–169. [PubMed: 18193043]
- 19. Vaxillaire M, Cavalcanti-Proenca C, Dechaume A, et al. The common P446L polymorphism in GCKR inversely modulates fasting glucose and triglyceride levels and reduces type 2 diabetes risk in the DESIR prospective general French population. Diabetes. 2008; 57:2253–2257. [PubMed: 18556336]
- 20. Tam CH, Ma RC, So WY, et al. Interaction effect of genetic polymorphisms in glucokinase (GCK) and glucokinase regulatory protein (GCKR) on metabolic traits in healthy Chinese adults and adolescents. Diabetes. 2009; 58:765–769. [PubMed: 19073768]
- 21. Sparso T, Andersen G, Nielsen T, et al. The GCKR rs780094 polymorphism is associated with elevated fasting serum triacylglycerol, reduced fasting and OGTT-related insulinaemia, and reduced risk of type 2 diabetes. Diabetologia. 2008; 51:70–75. [PubMed: 18008060]
- 22. Li N, van der Sijde MR, Bakker SJ, et al. Pleiotropic effects of lipid genes on plasma glucose, HbA1c, and HOMA-IR levels. Diabetes. 2014; 63:3149–3158. [PubMed: 24722249]
- 23. Scott RA, Lagou V, Welch RP, et al. Large-scale association analyses identify new loci influencing glycemic traits and provide insight into the underlying biological pathways. Nat Genet. 2012; 44:991–1005. [PubMed: 22885924]
- 24. Kamatani Y, Matsuda K, Okada Y, et al. Genome-wide association study of hematological and biochemical traits in a Japanese population. Nat Genet. 2010; 42:210–215. [PubMed: 20139978]
- 25. Suhre K, Shin SY, Petersen AK, et al. Human metabolic individuality in biomedical and pharmaceutical research. Nature. 2011; 477:54–60. [PubMed: 21886157]
- 26. Saxena R, Hivert MF, Langenberg C, et al. Genetic variation in GIPR influences the glucose and insulin responses to an oral glucose challenge. Nat Genet. 2010; 42:142–148. [PubMed: 20081857]
- 27. Kettunen J, Tukiainen T, Sarin AP, et al. Genome-wide association study identifies multiple loci influencing human serum metabolite levels. Nat Genet. 2012; 44:269–276. [PubMed: 22286219]
- 28. Kolz M, Johnson T, Sanna S, et al. Meta-analysis of 28 141 individuals identifies common variants within five new loci that influence uric acid concentrations. PLoS Genet. 2009; 5:e1000504. [PubMed: 19503597]
- 29. Kottgen A, Pattaro C, Boger CA, et al. New loci associated with kidney function and chronic kidney disease. Nat Genet. 2010; 42:376–384. [PubMed: 20383146]
- 30. Ridker PM, Pare G, Parker A, et al. Loci related to metabolic-syndrome pathways including LEPR,HNF1A, IL6R, and GCKR associate with plasma C-reactive protein: the Women's Genome Health Study. Am J Hum Genet. 2008; 82:1185–1192. [PubMed: 18439548]
- 31. Chambers JC, Zhang W, Sehmi J, et al. Genome-wide association study identifies loci influencing concentrations of liver enzymes in plasma. Nat Genet. 2011; 43:1131–1138. [PubMed: 22001757]

- 32. Lemaitre RN, Tanaka T, Tang W, et al. Genetic loci associated with plasma phospholipid *n*-3 fatty acids: a meta-analysis of genome-wide association studies from the CHARGE Consortium. PLoS Genet. 2011; 7:e1002193. [PubMed: 21829377]
- 33. Speliotes EK, Yerges-Armstrong LM, Wu J, et al. Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. PLoS Genet. 2011; 7:e1001324. [PubMed: 21423719]
- 34. Gieger C, Radhakrishnan A, Cvejic A, et al. New gene functions in megakaryopoiesis and platelet formation. Nature. 2011; 480:201–208. [PubMed: 22139419]
- 35. Reiner AP, Beleza S, Franceschini N, et al. Genome-wide association and population genetic analysis of C-reactive protein in African American and Hispanic American women. Am J Hum Genet. 2012; 91:502–512. [PubMed: 22939635]
- 36. O'Seaghdha CM, Wu H, Yang Q, et al. Meta-analysis of genome-wide association studies identifies six new loci for serum calcium concentrations. PLoS Genet. 2013; 9:e1003796. [PubMed: 24068962]
- 37. Kraja AT, Chasman DI, North KE, et al. Pleiotropic genes for metabolic syndrome and inflammation. Mol Gen Metab. 2014; 112:317–338. [PubMed: 24981077]
- 38. Shin SY, Petersen AK, Wahl S, et al. Interrogating causal pathways linking genetic variants, small molecule metabolites, and circulating lipids. Genome Med. 2014; 6:25. [PubMed: 24678845]
- 39. Mahendran Y, Vangipurapu J, Cederberg H, et al. Association of ketone body levels with hyperglycemia and type 2 diabetes in 9398 Finnish men. Diabetes. 2013; 62:3618–3626. [PubMed: 23557707]
- 40. Nica AC, Ongen H, Irminger JC, et al. Cell-type, allelic, and genetic signatures in the human pancreatic beta cell transcriptome. Genome Res. 2013; 23:1554–1562. [PubMed: 23716500]
- 41. Ferrannini E, Bjorkman O, Reichard GA Jr, et al. The disposal of an oral glucose load in healthy subjects. A quantitative study. Diabetes. 1985; 34:580–588. [PubMed: 3891471]
- 42. Mari A, Wahren J, DeFronzo RA, Ferrannini E. Glucose absorption and production following oral glucose: comparison of compartmental and arteriovenous-difference methods. Metabolism. 1994; 43:1419–1425. [PubMed: 7968597]
- 43. Moore MC, Coate KC, Winnick JJ, et al. Regulation of hepatic glucose uptake and storage in vivo. Adv Nutr. 2012; 3:286–294. [PubMed: 22585902]
- 44. Toyoda Y, Miwa I, Kamiya M, et al. Evidence for glucokinase translocation by glucose in rat hepatocytes. Biochem Biophys Res Commun. 1994; 204:252–256. [PubMed: 7945367]
- 45. Toyoda Y, Miwa I, Kamiya M, et al. Tissue and subcellular distribution of glucokinase in rat liver and their changes during fasting–refeeding. Histochem Cell Biol. 1995; 103:31–38. [PubMed: 7736279]
- 46. Brown KS, Kalinowski SS, Megill JR, et al. Glucokinase regulatory protein may interact with glucokinase in the hepatocyte nucleus. Diabetes. 1997; 46:179–186. [PubMed: 9000692]
- 47. Farrelly D, Brown KS, Tieman A, et al. Mice mutant for glucokinase regulatory protein exhibit decreased liver glucokinase: a sequestration mechanism in metabolic regulation. Proc Natl Acad Sci USA. 1999; 96:14511–14516. [PubMed: 10588736]
- 48. Agius L, Peak M, Van Schaftingen E. The regulatory protein of glucokinase binds to the hepatocyte matrix, but, unlike glucokinase, does not translocate during substrate stimulation. Biochem J. 1995; 309(Pt 3):711–713. [PubMed: 7639682]
- 49. Grimsby J, Coffey JW, Dvorozniak MT, et al. Characterization of glucokinase regulatory proteindeficient mice. J Biol Chem. 2000; 275:7826–7831. [PubMed: 10713097]
- 50■. Zelent B, Raimondo A, Barrett A, et al. Analysis of the co-operative interaction between the allosterically regulated proteins GK and GKRP using tryptophan fluorescence. Biochem J. 2014; 459:551–564. [PubMed: 24568320] [Detailed biophysical characterization via tryptophan fluorescence of the p.P446L GKRP variant and the way in which it alters GCK–GKRP complex formation.]
- 51. Agius L, Peak M, Newgard CB, et al. Evidence for a role of glucose-induced translocation of glucokinase in the control of hepatic glycogen synthesis. J Biol Chem. 1996; 271:30479–30486. [PubMed: 8940014]

- 52. Rees MG, Ng D, Ruppert S, et al. Correlation of rare coding variants in the gene encoding human glucokinase regulatory protein with phenotypic, cellular, and kinetic outcomes. J Clin Invest. 2012; 122:205–217. [PubMed: 22182842]
- 53■. Rees MG, Raimondo A, Wang J, et al. Inheritance of rare functional GCKR variants and their contribution to triglyceride levels in families. Hum Mol Genet. 2014; 23:5570–5578. [PubMed: 24879641] [This article demonstrates that rare loss-of-function *GCKR* variants do not cosegregate with elevated plasma triglyceride levels in families.]
- 54. Johansen CT, Wang J, Lanktree MB, et al. An increased burden of common and rare lipidassociated risk alleles contributes to the phenotypic spectrum of hypertriglyceridemia. Arterioscler Thromb Vasc Biol. 2011; 31:1916–1926. [PubMed: 21597005]
- 55■■. Lloyd DJ, St Jean DJ Jr, Kurzeja RJ, et al. Antidiabetic effects of glucokinase regulatory protein small-molecule disruptors. Nature. 2013; 504:437–440. [PubMed: 24226772] [The first published report of small molecule GKRP inhibitors that reduce blood glucose levels in rodent models of diabetes, without short-term side effects on insulin or lipids.]
- 56. Ashton KS, Andrews KL, Bryan MC, et al. Small molecule disruptors of the glucokinaseglucokinase regulatory protein interaction. 1. Discovery of a novel tool compound for in vivo proof-of-concept. J Med Chem. 2014; 57:309–324. [PubMed: 24405172]
- 57. Matschinsky FM. Assessing the potential of glucokinase activators in diabetes therapy. Nat Rev Drug Discov. 2009; 8:399–416. [PubMed: 19373249]
- 58. Johnson D, Shepherd RM, Gill D, et al. Glucose-dependent modulation of insulin secretion and intracellular calcium ions by GKA50, a glucokinase activator. Diabetes. 2007; 56:1694–1702. [PubMed: 17360975]
- 59. Grimsby J, Sarabu R, Corbett WL, et al. Allosteric activators of glucokinase: potential role in diabetes therapy. Science. 2003; 301:370–373. [PubMed: 12869762]
- 60. Meininger GE, Scott R, Alba M, et al. Effects of MK-0941, a novel glucokinase activator, on glycemic control in insulin-treated patients with type 2 diabetes. Diabetes Care. 2011; 34:2560– 2566. [PubMed: 21994424]
- 61. De Ceuninck F, Kargar C, Ilic C, et al. Small molecule glucokinase activators disturb lipid homeostasis and induce fatty liver in rodents: a warning for therapeutic applications in humans. Br J Pharmacol. 2012; 168:339–353. [PubMed: 22925001]
- 62. Nissim I, Horyn O, Daikhin Y, et al. Effects of a glucokinase activator on hepatic intermediary metabolism: study with (13)C-isotopomer-based metabolomics. Biochem J. 2012; 444:537–551. [PubMed: 22448977]
- 63. Rees MG, Gloyn AL. Small molecular glucokinase activators: has another new antidiabetic therapeutic lost favour? Br J Pharmacol. 2013; 168:335–338. [PubMed: 22946641]
- 64■■. Choi JM, Seo MH, Kyeong HH, et al. Molecular basis for the role of glucokinase regulatory protein as the allosteric switch for glucokinase. Proc Natl Acad Sci USA. 2013; 110:10171– 10176. [PubMed: 23733961] [This article provides atomic-level resolution of the *Xenopus* GCK– GKRP complex bound to fructose 6-phosphate.]
- 65■■. Beck T, Miller BG. Structural basis for regulation of human glucokinase by glucokinase regulatory protein. Biochemistry. 2013; 52:6232–6239. [PubMed: 23957911] [This article provides atomic-level resolution of the mammalian GCK–GKRP complex bound to fructose 6 phosphate.]
- 66■■. Pautsch A, Stadler N, Lohle A, et al. Crystal structure of glucokinase regulatory protein. Biochemistry. 2013; 52:3523–3531. [PubMed: 23621087] [This article provides atomic-level resolution of the human GKRP protein bound to fructose 1-phosphate.]
- 67. Veiga-da-Cunha M, Courtois S, Michel A, et al. Amino acid conservation in animal glucokinases. Identification of residues implicated in the interaction with the regulatory protein. J Biol Chem. 1996; 271:6292–6297. [PubMed: 8626423]
- 68. Brocklehurst KJ, Davies RA, Agius L. Differences in regulatory properties between human and rat glucokinase regulatory protein. Biochem J. 2004; 378:693–697. [PubMed: 14627435]
- 69. Veiga-da-Cunha M, Sokolova T, Opperdoes F, Van Schaftingen E. Evolution of vertebrate glucokinase regulatory protein from a bacterial *N*-acetylmuramate 6-phosphate etherase. Biochem J. 2009; 423:323–332. [PubMed: 19671048]

- 70. Baltrusch S, Francini F, Lenzen S, Tiedge M. Interaction of glucokinase with the liver regulatory protein is conferred by leucine-asparagine motifs of the enzyme. Diabetes. 2005; 54:2829–2837. [PubMed: 16186382]
- 71. Baltrusch S, Lenzen S, Okar DA, et al. Characterization of glucokinase-binding protein epitopes by a phage-displayed peptide library. Identification of 6-phosphofructo-2-kinase/fructose-2,6 bisphosphatase as a novel interaction partner. J Biol Chem. 2001; 276:43915–43923. [PubMed: 11522786]
- 72. Anderka O, Boyken J, Aschenbach U, et al. Biophysical characterization of the interaction between hepatic glucokinase and its regulatory protein: impact of physiological and pharmacological effectors. J Biol Chem. 2008; 283:31333–31340. [PubMed: 18809676]
- 73. Vandercammen A, Van Schaftingen E. Competitive inhibition of liver glucokinase by its regulatory protein. Eur J Biochem. 1991; 200:545–551. [PubMed: 1889417]
- 74. Bourbonais FJ, Chen J, Huang C, et al. Modulation of glucokinase by glucose, small-molecule activator and glucokinase regulatory protein: steady-state kinetic and cell-based analysis. Biochem J. 2012; 441:881–887. [PubMed: 22044397]
- 75. Veiga-da-Cunha M, Xu LZ, Lee YH, et al. Effect of mutations on the sensitivity of human betacell glucokinase to liver regulatory protein. Diabetologia. 1996; 39:1173–1179. [PubMed: 8897004]
- 76. Nishimura N, Norman MH, Liu L, et al. Small molecule disruptors of the glucokinase-glucokinase regulatory protein interaction. 3. Structure-activity relationships within the aryl carbinol region of the *N*-arylsulfonamido-*N*′-arylpiperazine series. J Med Chem. 2014; 57:3094–3116. [PubMed: 24611879]
- 77. St Jean DJ Jr, Ashton KS, Bartberger MD, et al. Small molecule disruptors of the glucokinaseglucokinase regulatory protein interaction. 2. Leveraging structure-based drug design to identify analogues with improved pharmacokinetic profiles. J Med Chem. 2014; 57:325–338. [PubMed: 24405213]
- 78. Hong FT, Norman MH, Ashton KS, et al. Small molecule disruptors of the glucokinaseglucokinase regulatory protein interaction. 4. Exploration of a novel binding pocket. J Med Chem. 2014; 57:5949–5964. [PubMed: 25001129]

KEY POINTS

- **•** *GCKR* and its encoded protein GKRP are central to hepatic glucose and lipid metabolism via inhibition of GCK.
- **•** *GCKR* variants across the allelic spectrum have effects on glucose and lipid levels but are not deterministic for plasma triglycerides.
- **•** These variants act via a range of molecular mechanisms including protein expression, stability, localization, and GCK and phosphate ester binding.
- **•** GKRP may be an effective pharmacological target for hyperglycaemia; however, this may also negatively impact on plasma triglyceride levels.
- **•** Future efforts to design effective small molecular inhibitors of GKRP will be bolstered by resolution of the crystal structures of GKRP and the GCK–GKRP complex.

Raimondo et al. Page 13

FIGURE 1.

Model of GKRP regulation of hepatic glucose metabolism. In the fasting state (left), GCK is inhibited by GKRP and sequestered in the nucleus. The hepatocyte is active in producing glucose via glycogenolysis and gluconeogenesis, and exports glucose to the circulation for use by peripheral tissues. In the fed state (right), GCK is released from GKRP inhibition by glucose (binding to GCK) and F1P (binding to GKRP). Glucose phosphorylation leads to enhanced glycolytic flux and glucose disposal and storage. F1P, fructose 1-phosphate; GCK, glucokinase; GKRP, glucokinase regulatory protein.

FIGURE 2.

Model of the effects of P446L–GKRP on hepatic glucose metabolism. Normally (left), GCK is inhibited by wild-type (WT) GKRP and sequestered in the nucleus in the fasting state. F6P (small circle) enhances complex formation. GCK is released from WT-GKRP in response to glucose and moves to the cytoplasm where it initiates glucose storage pathways. The p.P446L variant (right) creates a GKRP protein with increased cytoplasmic localization and reduced affinity for GCK and F6P, resulting in decreased plasma glucose levels and maintenance of glycolysis and *de novo* lipogenesis. F6P, fructose 6-phosphate; GCK, glucokinase; GKRP, glucokinase regulatory protein.