www.jcbfm.com

BRIEF COMMUNICATION Hypertension fails to disrupt white matter integrity in young or aged Fisher (F44) Cyp1a1Ren2 transgenic rats

Philip R Holland^{1,2,7}, Mercede A Pannozzo¹, Mark E Bastin^{2,3}, Allison D McNeilly⁴, Karen J Ferguson², Sarah Caughey^{2,5}, Maurits A Jansen⁶, Gavin D Merrifield⁶, Ian Marshall³, John J Mullins⁵, Joanna M Wardlaw^{2,3}, Calum Sutherland⁴ and Karen Horsburgh^{1,2}

Hypertension is linked with an increased risk of white matter hyperintensities; however, recent findings have questioned this association. We examined whether hypertension and additional cerebrovascular risk factors impacted on white matter integrity in an inducible hypertensive rat. No white matter hyperintensities were observed on magnetic resonance imaging either alone or in conjunction with ageing and high-fat diet. Aged hypertensive rats that were fed a high-fat diet had moderately reduced fractional anisotropy in the corpus callosum with no overt pathological features. Herein we show that moderate hypertension alone or with additional risk factors has minimal impact on white matter integrity in this model.

Journal of Cerebral Blood Flow & Metabolism (2015) 35, 188–192; doi:10.1038/jcbfm.2014.201; published online 19 November 2014

Keywords: ageing; cerebrovascular disease; hypertension; MRI and white matter

INTRODUCTION

Hypertension has been associated with an increased risk of white matter (WM) hyperintensities (WMHs)¹ and is the second most important factor after ageing for the development of cerebrovascular disease, a known contributor to vascular dementia. Clinically, WM lesions (WMLs) are characterized on MRI as hyperintense regions on fluid-attenuated inversion recovery or T2-weighted magnetic resonance imaging (MRI), which are associated with disruption of the normal cellular architecture of myelinated fibers. However, increased tissue water mobility without altered fractional anisotropy (FA) is an early imaging feature, as shown by diffusion tensor (DT-) and magnetisation transfer (MT-) MRI that have been utilized to explore WM alterations in hypertension.² Hypertension is associated with decreased FA, increased mean diffusivity (MD), and reduced magnetization transfer ratio (MTR) in both WMLs and normal-appearing WM,^{2,4} commonly including periventricular WM and the corpus callosum (CC).³ However, hypertension may not be the primary precipitant of WMHs with other nonvascular factors involved, $^{\circ}$ which is further supported by the failure of antihypertensive treatment to prevent the progression of WMLs.^e

Animal models have been used to explore the impact of hypertension on the central nervous system, (see Bailey *et al*⁷ and Hainsworth *et al*⁸), but have yielded conflicting evidence regarding its role in central nervous system structural integrity. The spontaneously hypertensive rat-stroke prone (SHRSP) has demonstrated clear WMLs, whereas the spontaneously

hypertensive rat fails to recapitulate the damage observed in the SHRSP despite similar blood pressure (BP),⁹ with lesions confined to the caudate. This is in agreement with recent data indicating that SHRSP that survive into old age without developing stroke, fail to show gross WMLs.¹⁰

Given the discrepancies in the influence of hypertension between the two commonly utilized models, and the difficulty in exploring isolated hypertension in humans because of a multitude of comorbid risk factors including diet, diabetes, and genetic susceptibility,^{1,11} we aimed to explore the alterations in WM integrity using MRI and immunohistochemistry in an inducible hypertensive transgenic rat, in which the onset and duration of hypertension are controllable.¹² We have previously demonstrated that MRI is sensitive to subtle WM alterations in a mouse model of modest cerebral hypoperfusion,¹³ and as such we hypothesize that sustained hypertension would lead to microstructural alterations in central nervous system WM and further that ageing or additional cerebrovascular risk factors, such as high-fat intake, would exacerbate WM disruption.

MATERIALS AND METHODS

Animals

The Cyp1a1Ren2 transgenic rats on a Fisher-F44 background, expressing the mouse *Ren-2* gene in the liver, under the control of the cytochrome-P450 promoter (Cyp1a1) were bred in-house. Young rats were randomly divided into two groups (Supplementary Figure 1) and fed on a standard

E-mail: Karen.Horsburgh@ed.ac.uk

The authors thank funding from Age UK, Alzheimer's Research UK, and the Alzheimer Society. PRH, MB, KJF, SC, JMW and KH are members of The University of Edinburgh Centre for Cognitive Ageing and Cognitive Epidemiology (CCACE), part of the cross council Lifelong Health and Wellbeing Initiative (G0700704/84698). Funding from the BBSRC, EPSRC, ESRC and MRC is gratefully acknowledged. Part of the work was funded by a CCACE imaging pilot award and an award from the ARUK East of Scotland Dementia Network for which we are extremely grateful. Magnetic resonance imaging was carried out using the facilities of Edinburgh Preclinical Imaging, College of Medicine and Veterinary Medicine, University of Edinburgh.

Received 17 July 2014; revised 13 October 2014; accepted 20 October 2014; published online 19 November 2014

¹Centre for Neuroregeneration, University of Edinburgh, Edinburgh, UK; ²Centre for Cognitive Ageing and Cognitive Epidemiology, University of Edinburgh, Edinburgh, UK; ³Scottish Imaging Network, A Platform for Scientific Collaboration (SINAPSE), Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, UK; ⁴Diabetic and Cardiovascular Medicine, University of Dundee, Dundee, UK; ⁵Centre for Cardiovascular Science, University of Edinburgh, Edinburgh, UK; ⁶Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, UK; ⁶Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, UK and ⁷Present address: Department of Basic and Clinical Neuroscience, Institute of Psychiatry, Psychology and Neuroscience, King's College London, London, UK. Correspondence: Professor K Horsburgh, Centre for Neuroregeneration, University of Edinburgh, Chancellor's Building, 49 Little France Crescent, Edinburgh EH16 4SB, UK.

White matter in an inducible hypertensive rat PR Holland *et al*

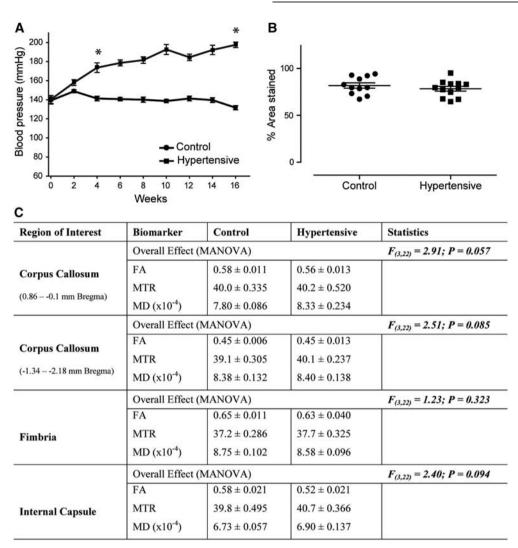


Figure 1. Physiological and magnetic resonance imaging findings in young rats. Addition of 0.15% I3C in normal rat chow results in a gradual increase in blood pressure in Cyp1a1Ren2 rats that is significantly different from control littermates from weeks 4 to 5 reaching > 160 mm Hg systolic and remaining significantly increased for the duration of the study (**A**). There were no significant alterations detected in any white matter region between control and hypertensive rats utilizing pathological (**B**, myelin basic protein immunohistochemistry) or MRI (**C**) techniques, suggesting minimal impact of hypertension alone on white matter integrity in young rats. FA, fractional anisotropy; MD, mean diffusivity; MRI, magnetic resonance imaging; MTR, magnetization transfer ratio.

diet (control; n = 12; 15 ± 1.2 weeks; range 14 to 17 weeks) or standard diet containing 0.15% indole-3-carbinol (I3C; hypertensive; n = 14;16±1.1 weeks; range 14 to 17 weeks) resulting in increased reninangiotensin cascade and elevated BP. Aged rats were again randomly assigned into four groups (Supplementary Figure 1) with groups 1 (n = 9; 69 ± 1.9 weeks; range 66 to 71 weeks) and group 2 (n = 9; 69 ± 2.1 weeks; range 66 to 70.5 weeks) receiving standard diet, and group 3 (n = 11; 67 ± 3.6 weeks; range 61 to 70.5 weeks), and group 4 (n = 11; 66 ± 2 weeks; range 62 to 67) receiving standard diet containing 0.15% I3C. In addition, groups 2 and 4 were switched to a high-fat diet containing no I3C (control and fat) or 0.15% I3C (hypertensive and fat), respectively, for the final 4 weeks (45% Atwater fuel energy FAT; Special Diets Services, Essex, UK). In addition, three aged animals were culled because of ill health, one each from groups 1, 3, and 4. All experiments were conducted under the UK Home Office Animals (Scientific Procedures) Act 1986, in agreement with local ethical and veterinary approval (Biomedical Research Resources, University of Edinburgh), and the ARRIVE guidelines.

Magnetic Resonance Imaging

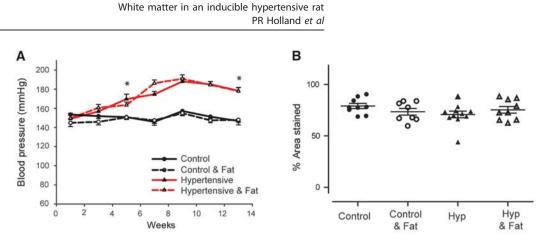
Rats were undefined and placed in an MRI holder (Rapid Biomedical, GmbH, Würzburg, Germany). Rectal temperature and respiration were

controlled to ensure normal physiological parameters. T₂, DT-, and MT-MRI data were collected using an Agilent 7T preclinical scanner (Agilent Technologies, Yarnton, UK); with a 72-mm volume coil and a two-channel phased-array rat brain coil (Rapid Biomedical). Diffusion tensor imaging was conducted using an axial fast spin–echo sequence and FA and MD were computed, MT-MRI was conducted using two spin–echo sequences (Supplementary Data). Mean diffusivity, FA, and MTR were measured in the CC, internal capsule, and fimbria.

Immunohistochemistry

After MRI, rats were perfused under deep isoflurane anesthesia with heparinized saline followed by 4% parafomaldehyde, their brains removed and postfixed before paraffin embedding. Appropriate $6 \mu m$ sections were cut corresponding to the regions assessed for MRI (0.86 to -0.1 mm and -1.34 to -2.18 mm bregma) as these areas have previously demonstrated vulnerability in spontaneously hypertensive rat and SHRSP strains. Immunostaining was conducted using standard methods after pretreatment to remove paraffin and for amyloid precursor protein antigen retrieval with microwaving in 10 mM citrate buffer (pH 6). Antibodies were selected to visualize the different cellular components of WM fiber bundles. Myelin integrity was assessed using anti-myelin basic protein (MBP) (1:5,000; Millipore, Watford, UK). Axonal damage was assessed using

189



С

190

Region of Interest	Biomarker	Control	Control & Fat	Hypertensive	Hypertensive & Fat	Statistics
	Overall Effect (MANOVA)					$F_{(9,98)} = 2.01; P = 0.045$
Corpus Callosum (0.86 – -0.1 mm Bregma)	FA	0.63 ± 0.010	0.60 ± 0.006	0.61 ± 0.008	0.59 ± 0.011	$F_{(3,36)} = 3.00; P = 0.043$ *
	MTR	44.3 ± 0.259	44.9 ± 0.260	43.9 ± 0.331	43.5 ± 0.542	$F_{(3,36)} = 0.90; P = 0.067$
	MD (x10 ⁻⁴)	7.24 ± 0.111	7.16 ± 0.135	7.24 ± 0.102	7.44 ± 0.164	$F_{(3,36)} = 2.61; P = 0.450$
Corpus Callosum (-1.34 – -2.18 mm Bregma)	Overall Effect (MANOVA)					$F_{(9,98)} = 0.66; P = 0.741$
	FA	0.51 ± 0.011	0.50 ± 0.012	0.49 ± 0.008	0.51 ± 0.011	
	MTR	43.5 ± 0.187	43.8 ± 0.185	43.7 ± 0.416	43.4 ± 0.286	
	MD (x10 ⁻⁴)	7.68 ± 0.126	7.43 ± 0.130	7.69 ± 0.153	7.57 ± 0.109	
Fimbria	Overall Effect (MANOVA)					$F_{(9,98)} = 0.53; P = 0.849$
	FA	0.68 ± 0.013	0.63 ± 0.055	0.71 ± 0.014	0.69 ± 0.012	
	MTR	42.1 ± 0.423	41.0 ± 1.180	42.4 ± 0.314	42.0 ± 0.358	
	MD (x10 ⁻⁴)	7.67 ± 0.160	7.77 ± 0.123	7.81 ± 0.116	7.77 ± 0.095	
Internal Capsule	Overall Effect (MANOVA)					$F_{(9,98)} = 0.66; P = 0.741$
	FA	0.60 ± 0.019	0.63 ± 0.019	0.62 ± 0.017	0.64 ± 0.020	
	MTR	42.6 ± 0.491	42.8 ± 0.305	42.8 ± 0.398	42.6 ± 0.319	
	MD (x10 ⁻⁴)	6.43 ± 0.090	6.46 ± 0.092	6.47 ± 0.045	6.52 ± 0.029	

Figure 2. Physiological and magnetic resonance imaging findings in aged rats. Addition of 0.15% I3C to the diet of aged Cyp1a1Ren2 rats results in a gradual increase in blood pressure that is significantly different from control littermates from weeks 4 to 5, reaching > 160 mm Hg systolic and remaining significantly increased for the duration of the study (**A**). Addition of high-fat to a subgroup of control or hypertensive (Hyp) rats had no significant impact on blood pressure. There was no significant alteration in white matter integrity as assessed by myelin basic protein immunohistochemistry (**B**); however, MRI techniques identified a significant overall difference when comparing MD, FA and MTR in the corpus callosum only at the boundary of the lateral ventricle (**C**). Follow-up ANOVA demonstrated a significant overall alteration in FA between control and hypertensive with high-fat rats only, with no significant change in MD or MTR values. There was no significant overall alteration in the remaining white matter regions (which precluded follow-up univariate analysis of MRI metrics to avoid the potential of type I errors), suggesting minimal impact of hypertension alone on white matter integrity in aged rats. ANOVA, analysis of variance; FA, fractional anisotropy; MD, mean diffusivity; MRI, magnetic resonance imaging; MTR, magnetization transfer ratio. *Significant change in an individual MRI metric (P < 0.05).

anti-amyloid precursor protein (1:1,000; Millipore). Sections were incubated with primary antibody overnight at 4°C after blocking then biotinylated secondary antibodies (1:100) and a solution of streptavidinbiotin-peroxidase complex. Peroxidase activity was localized using 3,3'diaminobenzadine tetrahydrochloride (Vector, Peterborough, UK). The percentage area stained of MBP was assessed in the CC above the lateral ventricle using Image J (v1.43, NIH, Bethesda, MD, USA). Bilateral images were taken at x200 magnification and the CC manually delineated by an observer blinded to the intervention. Four predefined regions-of-interest were then randomly placed over the images and the percentage area of immunopositive MBP calculated as a measure of myelin density. To assess axonal integrity the CC in all animals were carefully screened for the presence of intense amyloid precursor protein immunoreactivity in swollen or bulbous axons.

Statistical Analysis

Blood pressure data are presented as the average 2-weekly values across the different groups. Statistical comparisons were conducted using *t*-tests at varying time points (day 0, 4 to 5 weeks post induction, and at the conclusion of the experiment) and significance value were adjusted to 0.01 to correct for multiple comparisons. Magnetic resonance imaging data are presented as the average value of bilateral regions-of-interest for each subject. Statistical comparisons of MD, FA, and MTR values in each region measured between groups were carried out using a multivariate analysis of variance to correct for the use of multicollinearity (overall effect; Figures 1 and 2). Only when an appropriate significant overall effect was detected to protect against type I errors were analysis of variances conducted on individual MRI metrics to identify specific regional alterations in MD, FA, and MTR ($P \leq 0.05$). Percentage area stained for MBP was assessed by *t*-test for the young cohort and by analysis of variance for the aged cohort. All results are expressed as mean ± s.e. and statistical analysis was conducted using SPSS 19.0; SPSS Inc, Chicago, USA.

RESULTS

Impact of Hypertension in Young Rats

Addition of 0.15% I3C to young Cyp1a1Ren2 transgenic rats resulted in a gradual increase in BP which was significantly different from control littermates from week 4 to 5 onward ($t_{(18)} = 5.80$, P < 0.01; Figure 1A), when hypertensive rats had BP's greater than 160 mm Hg systolic. After 16 weeks I3C fed rats had

191

significantly higher systolic BPs of 197 \pm 2.8 compared with 131 \pm 2.2 mm Hg (t_{(18)} = 18.44, P < 0.01) for controls.

There were no overt WMH's on T₂-weighted MRI (Supplementary Figure 2) in young rats, and overall analysis failed to detect any significant microstructural alterations in all regions studied (Figure 1C). In agreement with the absence of WM alterations on MRI there was no overt axonal pathology in the CC and no significant change in myelin density ($t_{(21)} = 0.88$, P = 0.39; Figure 1B).

Impact of Hypertension in Aged Rats

Addition of 0.15% I3C to aged Cyp1a1Ren2 transgenic rats (groups 3 to 4) resulted in a gradual increase in BP that was significantly different from control littermates (groups 1 to 2) from weeks 4 to 5 onward ($t_{(18)}$ = 3.02, P < 0.01; Figure 2A), when hypertensive rats had BPs > 160 mm Hg systolic. After 14 weeks I3C fed rats had significantly higher systolic BPs of 178±3.6 mm Hg compared with 146±4.1 mm Hg ($t_{(15)}$ = 5.85, P < 0.01) for controls. Addition of high-fat diet to a subset of control (group 2) and hypertensive (group 4) aged rats, 4 weeks before imaging had no significant effect on BP (Figure 2A).

There were no overt WMLs on T_2 -weighted MRI in aged rats (Supplementary Figure 2), although overall analysis identified significant microstructural alterations between the four groups, in the CC ($F_{(9,98)} = 2.01$; P < 0.05; Figure 2C) at the boundary of the lateral ventricle (0.86 to -0.1 mm bregma), but not the fimbria $(F_{(9,98)} = 0.53; P > 0.05)$ or internal capsule $(F_{(9,98)} = 0.66; P > 0.05)$. Follow-up univariate comparisons in the CC showed that FA was significantly decreased ($F_{(3,36)} = 3.00$, P < 0.05); however, MD $(F_{(3,36)} = 0.90, P > 0.05)$ and MTR $(F_{(3,36)} = 2.61; P > 0.05)$ were unchanged. This specific alteration in FA was largely because of the differences between hypertensive rats fed a high-fat diet (group 4) compared with controls fed a normal diet (group 1). Analysis of axonal integrity identified no evidence of axonal disruption (Supplementary Figure 3) or significant change in myelin integrity ($F_{(3,33)} = 1.29$, P = 0.30; Figure 2B and Supplementary Figure 4). Correlation analysis using the Pearson's r statistics identified no significant correlation between CC FA values and end-stage systolic BP (r = 0.199, n = 40, and P = 0.22), or MBP density (r = 0.132, n = 37, and P = 0.44) with no additional effect of fat.

DISCUSSION

White matter lesions are commonly seen on MRI in older subjects and their presence is associated with age and cerebrovascular risk factors, including hypertension.¹ Herein we demonstrate that hypertension alone or in conjunction with further cerebrovascular risk factors, including ageing and high-fat intake does not result in gross WMLs as evidenced by the lack of WMHs on T2-weighted MRI. Recently alternate approaches including DT- and MT-MRI have demonstrated sensitivity for subtle WM disruption.¹³ In agreement we detected a significant subtle decrease in FA in the CC dorsal to the lateral ventricle; however, this was only evident in aged hypertensive rats fed a high-fat diet, suggestive of a summation effect. Despite the significant increase in BP there was no significant correlation between these indices and the observed alterations in FA. Our data are in agreement with others who recently identified no overt disruption in the WM of SHRSP rats, who survived stroke free up to ten months¹⁰ or required the addition of chronic hypoperfusion to elicit WMLs.¹⁴ Taken together the data suggest that hypertension alone for the duration tested here is insufficient to drive WM alterations and we further show that in aged rats (~21months) the combination of ageing and late onset hypertension does not impact on WM integrity in this model. As myelin damage may be a relatively late feature of WM damage secondary to vascular risk factors (as

suggested by pathologic study), it may be that we did not expose the rats to hypertension or high-fat for a sufficient length of time.

Although the use of a relatively novel model is beneficial to allow controllable hypertensive induction, it is a limitation of the study that the model has not been widely used to study central nervous system pathologic assessment after hypertension. To this end we have previously explored cerebrovascular alterations in young Cyp1a1Ren2 transgenic rats with varying durations of hypertension¹⁵ and identified thalamic cerebrovascular alterations. Thus the absence of overt WM alterations in hypertensive animals may indicate a regional vulnerability within thalamic regions. Alternatively, damage to myelin may occur late as suggested by human studies where post-mortem findings focus on loss of myelin, but much earlier changes detectable in vivo highlight change in water content as an early feature. The combination of cerebrovascular risk factors (ageing and high-fat diet) and the potential development of malignant hypertension in this transgenic line restricted our studies to 4 months duration. We cannot rule out that prolonged hypertension for >4 months or a more rapid increase in BP at onset may have an impact on white matter integrity; however, the gradual onset of BP elevation in this rat model is in keeping with the human condition.

In conclusion both young and aged hypertensive rats show no overt WM disruption raising further questions regarding the impact of hypertension on the development of WMHs. White matter structural integrity was only modestly altered in the combined presence of hypertension, ageing, and high-fat diet suggesting a cumulative effect of multiple risk factors.

DISCLOSURE/CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- 1 Raz N, Yang Y, Dahle CL, Land S. Volume of white matter hyperintensities in healthy adults: contribution of age, vascular risk factors, and inflammation-related genetic variants. *Biochim Biophys Acta* 2012; **1822**: 361–369.
- 2 Maclullich AM, Ferguson KJ, Reid LM, Deary IJ, Starr JM, Seckl JR et al. Higher systolic blood pressure is associated with increased water diffusivity in normalappearing white matter. Stroke 2009; 40: 3869–3871.
- 3 Salat DH, Williams VJ, Leritz EC, Schnyer DM, Rudolph JL, Lipsitz LA et al. Interindividual variation in blood pressure is associated with regional white matter integrity in generally healthy older adults. *Neuroimage* 2012; 59: 181–192.
- 4 Burgmans S, van Boxtel MP, Gronenschild EH, Vuurman EF, Hofman P, Uylings HB *et al.* Multiple indicators of age-related differences in cerebral white matter and the modifying effects of hypertension. *Neuroimage* 2010; **49**: 2083–2093.
- 5 Wardlaw JM, Allerhand M, Doubal FN, Valdes Hernandez M, Morris Z, Gow AJ et al. Vascular risk factors, large-artery atheroma, and brain white matter hyperintensities. *Neurology* 2014; 82: 1331–1338.
- 6 Weber R, Weimar C, Blatchford J, Hermansson K, Wanke I, Moller-Hartmann C et al. Telmisartan on top of antihypertensive treatment does not prevent progression of cerebral white matter lesions in the prevention regimen for effectively avoiding second strokes (PRoFESS) MRI substudy. *Stroke* 2012; **43**: 2336–2342.
- 7 Bailey EL, Smith C, Sudlow CL, Wardlaw JM. Is the spontaneously hypertensive stroke prone rat a pertinent model of sub cortical ischemic stroke? A systematic review. *Int J Stroke* 2011; 6: 434–444.
- 8 Hainsworth AH, Markus HS. Do in vivo experimental models reflect human cerebral small vessel disease? A systematic review. J Cereb Blood Flow Metab 2008; 28: 1877–1891.
- 9 Lin JX, Tomimoto H, Akiguchi I, Wakita H, Shibasaki H, Horie R. White matter lesions and alteration of vascular cell composition in the brain of spontaneously hypertensive rats. *Neuroreport* 2001; **12**: 1835–1839.
- 10 Brittain JF, McCabe C, Khatun H, Kaushal N, Bridges LR, Holmes WM et al. An MRIhistological study of white matter in stroke-free SHRSP. J Cereb Blood Flow Metab 2013; 33: 760–763.
- 11 Lopez LM, Bastin ME, Maniega SM, Penke L, Davies G, Christoforou A *et al.* A genome-wide search for genetic influences and biological pathways related to the brain's white matter integrity. *Neurobiol Aging* 2012; **33**: 1847.e1–14.

- 12 Kantachuvesiri S, Fleming S, Peters J, Peters B, Brooker G, Lammie AG et al. Controlled hypertension, a transgenic toggle switch reveals differential mechanisms underlying vascular disease. J Biol Chem 2001; 276: 36727–36733.
- 13 Holland PR, Bastin ME, Jansen MA, Merrifield GD, Coltman RB, Scott F *et al.* MRI is a sensitive marker of subtle white matter pathology in hypoperfused mice. *Neurobiol Aging* 2011; **32**: 2325, e1–2325, e6.
- 14 Jalal FY, Yang Y, Thompson J, Lopez AC, Rosenberg GA. Myelin loss associated with neuroinflammation in hypertensive rats. *Stroke* 2012; **43**: 1115–1122.
- 15 Pannozzo MA, Holland PR, Scullion G, Talbot R, Mullins JJ, Horsburgh K. Controlled hypertension induces cerebrovascular and gene alterations in Cyp1a1-Ren2 transgenic rats. J Am Soc Hypertens 2013; **7**: 411–419.

Supplementary Information accompanies the paper on the Journal of Cerebral Blood Flow & Metabolism website (http://www.nature. com/jcbfm)