

Data and text mining

drexplorer: A tool to explore dose–response relationships and drug–drug interactions

Pan Tong¹, Kevin R. Coombes², Faye M. Johnson³, Lauren A. Byers³,
Lixia Diao¹, Diane D. Liu⁴, J. Jack Lee⁴, John V. Heymach³ and
Jing Wang^{1,*}

¹Departments of Bioinformatics and Computational Biology, Houston, TX 77030, ²Department of Biomedical Informatics, The Ohio State University, Columbus, OH 43210, ³Thoracic and Head and Neck Medical Oncology and ⁴Biostatistics, The University of Texas MD Anderson Cancer Center, Houston, TX 77030, USA

*To whom correspondence should be addressed.

Associate editor: Jonathan Wren

Received on October 3, 2014; revised on December 11, 2014; accepted on January 11, 2015

Abstract

Motivation: Nonlinear dose–response models are primary tools for estimating the potency [e.g. half-maximum inhibitory concentration (IC) known as IC50] of anti-cancer drugs. We present `drexplorer` software, which enables biologists to evaluate replicate reproducibility, detect outlier data points, fit different models, select the best model, estimate IC values at different percentiles and assess drug–drug interactions. `drexplorer` serves as a computation engine within the R environment and a graphical interface for users who do not have programming backgrounds.

Availability and implementation: The `drexplorer` R package is freely available from GitHub at <https://github.com/nickytong/drexplorer>. A graphical user interface is shipped with the package.

Contact: jingwang@mdanderson.org

Supplementary information: [Supplementary data](#) are available at *Bioinformatics* online.

1 Introduction

Understanding the dose–response relationship of therapeutic compounds is a major focus of clinical oncology. Large-scale drug screening in cancer cell lines has demonstrated that different genomic characteristics (mutations) confer distinct drug sensitivity, supporting patient stratification into treatment regimens on the basis of genomic markers (Barretina *et al.*, 2012; Yang *et al.*, 2013). Cell lines resistant to single-drug treatment may respond to synergistic drug combinations (Lehár, *et al.*, 2009). As cell viability assays become widely used, it is important to provide easily accessible tools to analyze drug profiling data.

We developed `drexplorer` software in the R environment (R Core Team, 2014) to encompass several aspects of dose–response analysis: assess reproducibility of replicated experiments, detect outlier data points, fit different models, identify the best model, estimate inhibitory concentration (IC) values and evaluate drug–drug interactions.

2 Algorithms

A typical drug screening experiment exposes cells to a range of concentrations (*doses*) of a drug and quantifies cell viability (*response*). To account for experimental variations, multiple measurements are obtained for each dose. We denote the *j*th repeat of the *i*th dose by x_{ij} and the corresponding response by y_{ij} . For anti-cancer drug screening, response y_{ij} is further scaled by the mean response of the controls (i.e. under $x_{ij} = 0$) and is called relative viability.

2.1 Outlier detection

Outlier values in response y_{ij} will affect dose–response curve fitting and IC estimation. We implement the Newman test (Newman, 1939) to detect outliers, defining the test statistic as $q = w/s$, where w is the maximum difference among the response vector $(y_{i1}, \dots, y_{im_i})$ at the *i*th dose and s is the standard deviation of the response in untreated controls. Once calculated, the test statistic can be compared

with the threshold value from the null distribution under the specified type I error ($\alpha = 0.01$ or 0.05). We apply this test to identify outlier data points at each dose separately. Figure 1a shows outlier detection using the ryegrass data (Ritz and Streibig, 2005).

2.2 Assessing reproducibility

When replicate experiments have been performed, it is necessary to assess the quality of the reproducibility. We use the concordance correlation coefficient (CCC) to capture both the location shift and the scale shift between replicates (Lawrence and Lin, 1989). CCC ranges between -1 and 1 , where $CCC = 1$ means perfect agreement. Figure 1c illustrates the reproducibility assessment for two technical replicates run on different dates.

2.3 The dose–response model

The dose–response model can be expressed as a nonlinear function:

$$y_{ij} = f(x_{ij}; \theta) + \varepsilon_{ij}, \quad i = 1, \dots, n; j = 1, \dots, m_i,$$

where θ is the vector of unknown parameters, ε_{ij} is the measurement error for the response y_{ij} , n is the total number of doses administered and m_i is the number of repeats under the i th dose. The error term ε_{ij} is assumed to be identically independently distributed as $N(0, \sigma^2)$. In the case of the sigmoid Emax model, $f(\cdot)$ has the following form:

$$y_{ij} = E_0 + \frac{E_{\max} x_{ij}^b}{ED50^b + x_{ij}^b}.$$

Here, $\theta = (E_0, E_{\max}, ED50, b)$ is the vector of parameters to be estimated.

The dose–response models in `drexplorer` include all the models in the `drc` (Ritz and Streibig, 2005) and `DoseFinding`

(Bornkamp, *et al.*, 2010) packages and can be easily used through a uniform interface.

Parameters in the dose–response models can be estimated by minimizing the sum of the squared residuals (SSR):

$$SSR = \sum_i \sum_j \{y_{ij} - f(x_{ij}; \theta)\}^2.$$

After fitting the model, we estimate the IC at percentile τ by solving x in equation $\tau = f(x; \hat{\theta})$, where $\hat{\theta}$ is the vector of the fitted parameters.

Different models can be fit, so we identify the best model as that with minimum residual standard error, calculated as SSR divided by degrees of freedom. Approaches for parametric dose–response models may use the Akaike information criterion or Bayesian information criterion.

2.4 The Interaction Index for Drug Combinations

The interaction index (IAI) is useful to quantify the degree of interaction in drug combinations. For a 2-drug combination, IAI can be defined as

$$IAI = \frac{d_1}{D_{y,1}} + \frac{d_2}{D_{y,2}} \begin{cases} < 1, \text{ synergistic} \\ = 1, \text{ additive} \\ > 1, \text{ antagonistic} \end{cases}.$$

Here, $D_{y,1}$ and $D_{y,2}$ are the doses needed to induce response y for drugs 1 and 2 when administered alone; d_1 and d_2 are the doses in the mixture that produce the same response y . We adopt the source code used by Lee, *et al.* (2007) and Lee and Kong (2009) to estimate the IAI and 95% confidence interval (see Fig. 1b using UMSCC22B data; Lee and Kong, 2009). Graphical user interfaces (GUIs) help users without advanced programming skills to analyze dose–response data (Fig. 1d and e).

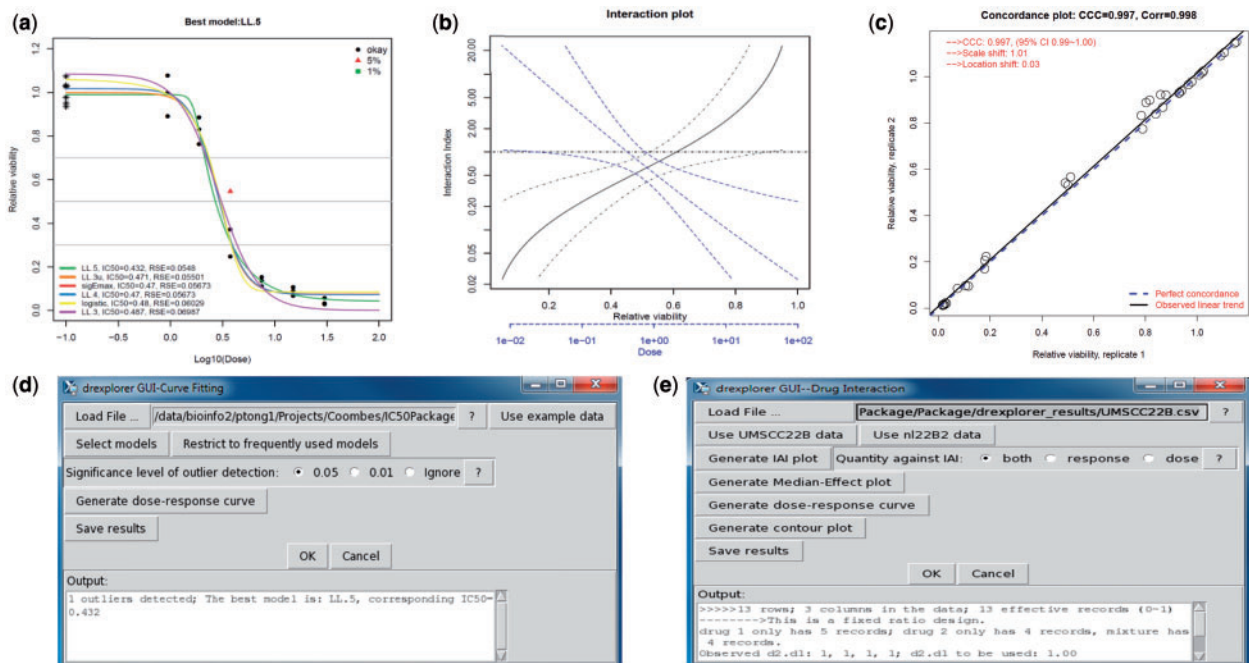


Fig. 1. Example of `drexplorer` output and GUI. (a) Outlier data at type I error = 0.01 and 0.05 are, respectively, indicated by the green square and red triangle. Various dose–response curves are fitted; RSEs are indicated; title indicates model selected by RSE. (b) Interaction index estimated at different doses (blue line) and relative viability (black line); 95% confidence interval also shown. (c) Reproducibility assessment using concordance. Responses from two replicates are highly concordant showing good reproducibility. (d) GUI screenshot for dose–response estimation. (e) GUI screenshot for interaction index estimation.

3 Conclusion

DreXplorer is a versatile R package encompassing several aspects of dose–response and drug–drug interaction analysis. The GUI enables biologists without programming skills to analyze their data.

Acknowledgements

This project was partially supported by the NCI/NIH through the Lung SPORC (P50 CA070907) and Cancer Center Support Grant (CA016672), and by the Mary K. Chapman Foundation.

Conflict of Interest: none declared.

References

- Barretina, J. *et al.* (2012) The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. *Nature*, **483**, 603–607.
- Bornkamp, B. *et al.* (2010) DoseFinding: planning and analyzing dose finding experiments, R package version 0.4-1. <http://www.icesi.edu.co/CRAN/web/packages/DoseFinding/DoseFinding.pdf>.
- Lawrence, I. and Lin, K. (1989) A concordance correlation coefficient to evaluate reproducibility. *Biometrics*, 255–268.
- Lee, J.J. and Kong, M. (2009) Confidence intervals of interaction index for assessing multiple drug interaction. *Stat. Biopharm. Res.*, **1**, 4–17.
- Lee, J.J. *et al.* (2007) Interaction index and different methods for determining drug interaction in combination therapy. *J. Biopharm. Stat.*, **17**, 461–480.
- Lehár, J. *et al.* (2009) Synergistic drug combinations tend to improve therapeutically relevant selectivity. *Nat. Biotechnol.*, **27**, 659–666.
- Newman, D. (1939) The distribution of range in samples from a normal population, expressed in terms of an independent estimate of standard deviation. *Biometrika*, **31**, 20–30.
- R Core Team. (2014) *R: A Language and Environment for Statistical Computing*. The R Foundation for Statistical Computing, Vienna, Austria. <http://www.R-project.org>.
- Ritz, C. and Streibig, J.C. (2005) Bioassay analysis using R. *J. Stat. Softw.*, **12**, 1–22.
- Yang, W. *et al.* (2013) Genomics of drug sensitivity in cancer (GDSC): a resource for therapeutic biomarker discovery in cancer cells. *Nucleic Acids Res.*, **41**, D955–D961.