

# Epithelial–Mesenchymal Interactions in Fibrosis and Repair

## Transforming Growth Factor- $\beta$ Activation by Epithelial Cells and Fibroblasts

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### Abstract

Transforming growth factor- $\beta$  (TGF- $\beta$ ) plays a central role in driving tissue fibrosis. TGF- $\beta$  is secreted in a latent form, held latent by noncovalent association of the active cytokine with a peptide derived from cleavage of the N-terminal domain of the same gene product, and needs to be activated extracellularly to exert any of its diverse biological effects. We have shown that two of the three mammalian isoforms of TGF- $\beta$ , TGF- $\beta_1$  and TGF- $\beta_3$ , depend on interactions with cell surface integrins for activation. We found that the integrin  $\alpha v \beta_6$  is highly induced on injured alveolar epithelial cells, potently induces TGF- $\beta$  activation, and is critical for the development of pulmonary fibrosis and acute lung injury. However, although TGF- $\beta$  drives fibrosis in virtually every anatomic site,  $\alpha v \beta_6$ -mediated TGF- $\beta$  activation is much more restricted. For example,  $\alpha v \beta_6$  is not induced on injured hepatocytes and plays little or no role in cirrhosis induced by repetitive hepatocyte injury. Fibroblasts are highly contractile cells that express multiple integrins closely related to  $\alpha v \beta_6$ , which share the promiscuous

$\alpha v$  subunit, so we reasoned that perhaps one or more of these  $\alpha v$  integrins on fibroblasts might substitute for  $\alpha v \beta_6$  and activate the TGF- $\beta$  required to drive liver fibrosis. Indeed, deletion of the  $\alpha v$  subunit from activated fibroblasts protected mice from carbon tetrachloride-induced liver fibrosis. Importantly, these same mice were protected from bleomycin-induced pulmonary fibrosis and renal fibrosis caused by unilateral ureteral obstruction, despite the presence of epithelial  $\alpha v \beta_6$  in these mice. These results suggest that the generation and maintenance of sufficient quantities of active TGF- $\beta$  to cause tissue fibrosis in multiple organs probably depends on at least two sources—TGF- $\beta$  activation by injured epithelial cells that drives fibroblast expansion and activation and an amplification step that involves TGF- $\beta$  activation by an  $\alpha v$  integrin on activated fibroblasts. These results suggest that intervening at either of these steps could be useful for the treatment of fibrotic diseases.

**Keywords:** integrin; transforming growth factor- $\beta$ ; pulmonary fibrosis

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Transforming growth factor- $\beta$  has been long recognized to play a central role in tissue repair. Much work has focused on the well-characterized increase in TGF- $\beta$  production at sites of injury, with the presumption that TGF- $\beta$  production would be the critical axis for regulating this process. However, although each of the three mammalian TGF- $\beta$  isoforms can produce potent biological effects at picomolar concentrations, most organs of healthy mammals constitutively contain stored extracellular TGF- $\beta$  at concentrations several orders of magnitude

higher. Work over the past 2 decades has helped to explain this apparent paradox. All of the TGF- $\beta$  isoforms are secreted in a latent form that is sequestered in the extracellular matrix and on the surface of some cells (1). As a result, most of the regulation of TGF- $\beta$  function occurs at the level of activation of these stored latent complexes.

The first evidence implicating members of the integrin family in activation of latent TGF- $\beta$  came from observations of mice we generated lacking the epithelial integrin,  $\alpha v \beta_6$  (2). These mice develop normally and

are born at the appropriate mendelian frequency but develop exaggerated inflammation in the skin and lungs in response to normally trivial injuries (3). Despite exaggerated inflammatory responses, these mice are dramatically protected from bleomycin- (2) and radiation-induced pulmonary fibrosis (4). Because inactivation of one of the three mammalian TGF- $\beta$  isoforms, TGF- $\beta_1$ , leads to profound multiorgan inflammation, and TGF- $\beta$  has long been known to be a central mediator of tissue fibrosis, these findings raised the possibility

that TGF- $\beta$  could be acting downstream of the  $\alpha v \beta_6$  integrin. Together with John Munger, we showed that this integrin binds to the linear tripeptide, arginine-glycine-aspartic acid, present within an amino terminal fragment of TGF- $\beta_1$  (and TGF- $\beta_3$ ) (2, 5) called the latency-associated peptide and through a pathway that requires actin polymerization, actin-myosin contraction and the generation of traction force alter the conformation of stored extracellular TGF- $\beta$  to activate it (6). The recently solved crystal structure of latent TGF- $\beta_1$  together with cryo-electron microscopy of latent TGF- $\beta$  together with the secreted ectodomains of the  $\alpha v \beta_6$  integrin beautifully demonstrate how the latency-associated peptide prevents binding of TGF- $\beta$  to its receptors and how physical force transmitted through the integrin unfolds the latent complex to release free TGF- $\beta$  (7).

$\alpha v \beta_6$  is expressed at low levels on alveolar epithelial cells and plays an important role in the basal low-level activation of TGF- $\beta$  that is needed to suppress the activity of alveolar macrophages and maintain normal alveolar homeostasis (8). However, in response to epithelial injury, epithelial cells are induced to contract their subcortical actin-myosin cytoskeletons, exerting increased amounts of deforming physical force on latent TGF- $\beta$ , thereby increased the amount of activated TGF- $\beta$ . Because TGF- $\beta$  itself is the most potent inducer of  $\alpha v \beta_6$  expression, this process triggers a feed-forward loop of increasing integrin expression and further increases in TGF- $\beta$  activation (9, 10). As pathologic extracellular matrix accumulates in the fibrosing lung, the lung becomes stiffer, making it easier for any degree of epithelial cell contraction to activate more TGF- $\beta$ , further increasing  $\alpha v \beta_6$  expression. One consequence of this feed-forward loop is that  $\alpha v \beta_6$  protein expression is dramatically up-regulated in epithelial cells overlying regions of lung fibrosis (11), an effect we have consistently seen in tissue samples from more than 50 patients with a variety of fibrotic lung diseases. Based on this observation, multiple groups have developed methods to image  $\alpha v \beta_6$  expression *in vivo*, which could allow noninvasive evaluation of pulmonary fibrosis and potentially identify patients who are likely to be especially responsive

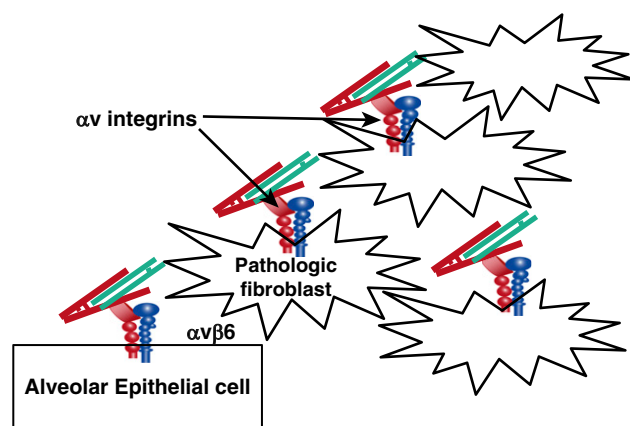
to inhibition of  $\alpha v \beta_6$  or other steps in this pathway (12).

We generated potent blocking antibodies that inhibit the function of  $\alpha v \beta_6$  in multiple species, and we and others have used these to demonstrate the potential therapeutic usefulness of targeting this pathway in murine models of bleomycin and radiation-induced fibrosis (4, 11), in renal fibrosis induced by unilateral ureteral obstruction or a genetic model of Alport syndrome (13), and in biliary fibrosis induced by ligation of the common bile duct (14).  $\beta_6$  knockout mice and our blocking antibodies have also allowed us to identify an important role for this integrin in alveolar flooding in multiple models of acute lung injury (15). A humanized version of one of these antibodies is currently in early phase 2 clinical trials for treatment of pulmonary fibrosis.

However, it is clear that the  $\alpha v \beta_6$  integrin is not the only important activator of TGF- $\beta$  that contributes to tissue fibrosis. For example, liver fibrosis in response to hepatocyte injury can be inhibited by blocking TGF- $\beta$  but proceeds normally in mice lacking the  $\beta_6$  subunit (16). We therefore sought to identify other relevant mechanisms of pathologic TGF- $\beta$  activation, using carbon tetrachloride-induced liver fibrosis as a model. *In vitro* work from Boris Hinz's laboratory showed that fibroblasts have the capacity to use integrins to activate TGF- $\beta$  (17), but this effect is clearly not dependent on  $\alpha v \beta_6$ , because  $\alpha v \beta_6$  is never expressed on fibroblasts. After trying a number of mouse lines expressing cre recombinase under the

control of putative fibroblast targeting promoters and failing to observe efficient recombination in liver fibroblasts, we settled on a line originally designed to target pericytes that expressed cre under the control of the platelet-derived growth factor receptor (PDGFR)- $\beta$  promoter. We chose this line because resting hepatic stellate cells, the major source for collagen-producing liver fibroblasts, closely resemble pericytes in other organs and because PDGFR- $\beta$  is highly expressed on activated fibroblasts. Although PDGFR $\beta$  expression is not restricted to fibroblasts, this line resulted in very efficient recombination in activated stellate cells in fibrotic livers. Based on evidence from our laboratory and others that multiple integrins that share the  $\alpha v$  subunit can activate TGF- $\beta$  *in vitro*, we deleted this whole family of integrins in activated fibroblasts by crossing the PDGFR $\beta$ -cre allele into mice designed for conditional deletion of  $\alpha v$  ( $\alpha v$  f/f mice).

$\alpha v$  f/f  $\times$  PDGFR $\beta$ -cre mice were significantly protected from CCl<sub>4</sub>-induced liver fibrosis (16). We then sought to determine which  $\alpha v$ -containing integrins are expressed on activated liver fibroblasts and found that these cells express moderate amounts of  $\alpha v \beta_1$ ,  $\alpha v \beta_3$ , and  $\alpha v \beta_5$ ; minimal amounts of  $\alpha v \beta_8$ ; and no  $\alpha v \beta_6$ . Mice globally lacking  $\alpha v \beta_3$ ,  $\alpha v \beta_5$ , or  $\alpha v \beta_6$  or mice conditionally lacking  $\alpha v \beta_8$  on activated fibroblasts all had normal fibrotic responses to CCl<sub>4</sub>. Unfortunately, because the  $\beta_1$  subunit is present in 12 different integrins, and deletion of  $\beta_1$  with PDGFR- $\beta$  results in perinatal mortality, we could not use mutant mice to directly examine the



**Figure 1.** Model of how distinct  $\alpha v$  integrins expressed on epithelial cells and pathologic fibroblasts each contribute to fibroblast activation and pathologic tissue fibrosis.

role of  $\alpha v \beta_1$  in this process. These results suggest that either there is redundancy among fibroblast  $\alpha v$  integrins in driving liver fibrosis or that the major integrin responsible for this effect is  $\alpha v \beta_1$ .

Although mice lacking the  $\alpha v \beta_6$  integrin are protected from pulmonary and renal fibrosis, fibrosis in those organs is also characterized by accumulation of contractile fibroblasts. Because pathologic fibrosis requires a sustained and substantial increase in active TGF- $\beta$ , we reasoned that loss of either  $\alpha v \beta_6$ -mediated activation by epithelial cells (as shown) or of  $\alpha v$  integrin-mediated TGF- $\beta$  activation by fibroblasts might protect against lung or kidney fibrosis. We therefore evaluated the efficiency of PDGFR- $\beta$ -mediated recombination on activated fibroblasts in the lung and kidney and found it to be equally effective to what we observed in the liver.  $\alpha v f/f \times$  PDGFR- $\beta$ -cre mice were also protected against bleomycin-induced

pulmonary fibrosis and unilateral ureteral obstruction-induced renal fibrosis.

Finally, to determine whether fibroblast  $\alpha v$  integrins could be reasonable therapeutic targets for fibrotic diseases, we examined the effects of a broadly active small molecule inhibitor of  $\alpha v$  integrins, CWHM-12, administered therapeutically beginning either on Day 21 after the start of CCl<sub>4</sub> administration or on Day 14 after treatment with intratracheal bleomycin. In both cases we found similar reductions in fibrosis to what we observed in  $\alpha v f/f \times$  PDGFR- $\beta$ -cre mice.

The combined results of our studies with inhibitors and knockouts of  $\alpha v \beta_6$  on epithelial cells and  $\alpha v$  integrins on fibroblasts support a model in which epithelial injury and ongoing epithelial cell dysfunction leads to up-regulation of the  $\alpha v \beta_6$  integrin and persistent TGF- $\beta$  activation (Figure 1). TGF- $\beta$  activation on the surface of epithelial cells drives

differentiation of adjacent cells into collagen-producing pathologic fibroblasts. TGF- $\beta$  activation by other  $\alpha v$  integrins on these fibroblasts provides an important amplification loop, leading to further expansion of the population of pathologic fibroblasts. Because there are potent homeostatic pathways to bring this system back into a healthy balance, pathologic fibrosis requires ongoing high-level TGF- $\beta$  activity, which explains why inhibition of each of these complementary pathways can substantially inhibit pathologic pulmonary fibrosis. These results provide encouragement that inhibition of either epithelial TGF- $\beta$  activation or fibroblast TGF- $\beta$  activation could provide significant therapeutic benefit and would ultimately provide a rationale for combining these approaches. ■

**Author disclosures** are available with the text of this article at [www.atsjournals.org](http://www.atsjournals.org).

## References

- Munger JS, Harpel JG, Gleizes PE, Mazziari R, Nunes I, Rifkin DB. Latent transforming growth factor-beta: structural features and mechanisms of activation. *Kidney Int* 1997;51:1376-1382.
- Munger JS, Huang X, Kawakatsu H, Griffiths MJ, Dalton SL, Wu J, Pittet JF, Kaminski N, Garat C, Matthay MA, et al. The integrin alpha v beta 6 binds and activates latent TGF beta 1: a mechanism for regulating pulmonary inflammation and fibrosis. *Cell* 1999;96:319-328.
- Huang XZ, Wu JF, Cass D, Erle DJ, Corry D, Young SG, Faresse RV Jr, Sheppard D. Inactivation of the integrin beta 6 subunit gene reveals a role of epithelial integrins in regulating inflammation in the lung and skin. *J Cell Biol* 1996;133:921-928.
- Puthawala K, Hadjiangelis N, Jacoby SC, Bayongan E, Zhao Z, Yang Z, Devitt ML, Horan GS, Weinreb PH, Lukashev ME, et al. Inhibition of integrin alpha(v)beta6, an activator of latent transforming growth factor-beta, prevents radiation-induced lung fibrosis. *Am J Respir Crit Care Med* 2008;177:82-90.
- Annes JP, Rifkin DB, Munger JS. The integrin alpha v beta 6 binds and activates latent TGF beta 3. *FEBS Lett* 2002;511:65-68.
- Giacomini MM, Travis MA, Kudo M, Sheppard D. Epithelial cells utilize cortical actin/myosin to activate latent TGF- $\beta$  through integrin  $\alpha(v)\beta(6)$ -dependent physical force. *Exp Cell Res* 2012;318:716-722.
- Shi M, Zhu J, Wang R, Chen X, Mi L, Walz T, Springer TA. Latent TGF- $\beta$  structure and activation. *Nature* 2011;474:343-349.
- Morris DG, Huang X, Kaminski N, Wang Y, Shapiro SD, Dolganov G, Glick A, Sheppard D. Loss of integrin alpha(v)beta6-mediated TGF-beta activation causes Mmp12-dependent emphysema. *Nature* 2003;422:169-173.
- Wang A, Yokosaki Y, Ferrando R, Balmes J, Sheppard D. Differential regulation of airway epithelial integrins by growth factors. *Am J Respir Cell Mol Biol* 1996;15:664-672.
- Sheppard D, Cohen DS, Wang A, Busk M. Transforming growth factor beta differentially regulates expression of integrin subunits in guinea pig airway epithelial cells. *J Biol Chem* 1992;267:17409-17414.
- Horan GS, Wood S, Ona V, Li DJ, Lukashev ME, Weinreb PH, Simon KJ, Hahm K, Allaire NE, Rinaldi NJ, et al. Partial inhibition of integrin alpha(v)beta6 prevents pulmonary fibrosis without exacerbating inflammation. *Am J Respir Crit Care Med* 2008;177:56-65.
- John AE, Lockett JC, Tatler AL, Awais RO, Desai A, Habgood A, Ludbrook S, Blanchard AD, Perkins AC, Jenkins RG, et al. Preclinical SPECT/CT imaging of  $\alpha v \beta_6$  integrins for molecular stratification of idiopathic pulmonary fibrosis. *J Nucl Med* 2013;54:2146-2152.
- Hahm K, Lukashev ME, Luo Y, Yang WJ, Dolinski BM, Weinreb PH, Simon KJ, Chun Wang L, Leone DR, Lobb RR, et al. Alpha v beta 6 integrin regulates renal fibrosis and inflammation in Alport mouse. *Am J Pathol* 2007;170:110-125.
- Wang B, Dolinski BM, Kikuchi N, Leone DR, Peters MG, Weinreb PH, Violette SM, Bissell DM. Role of alpha v beta 6 integrin in acute biliary fibrosis. *Hepatology* 2007;46:1404-1412.
- Pittet JF, Griffiths MJ, Geiser T, Kaminski N, Dalton SL, Huang X, Brown LA, Gotwals PJ, Koteliensky VE, Matthay MA, et al. TGF-beta is a critical mediator of acute lung injury. *J Clin Invest* 2001;107:1537-1544.
- Henderson NC, Arnold TD, Katamura Y, Giacomini MM, Rodriguez JD, McCarty JH, Pellicoro A, Raschperger E, Betsholtz C, Ruminski PG, et al. Targeting of  $\alpha v$  integrin identifies a core molecular pathway that regulates fibrosis in several organs. *Nat Med* 2013;19:1617-1624.
- Wipff PJ, Rifkin DB, Meister JJ, Hinz B. Myofibroblast contraction activates latent TGF-beta1 from the extracellular matrix. *J Cell Biol* 2007;179:1311-1323.