



## Draft Genome Sequence of Norvancomycin-Producing Strain Amycolatopsis orientalis CPCC200066

## Xuan Lei, Fang Yuan, Yuanyuan Shi, Xingxing Li, Lifei Wang, Bin Hong

Key Laboratory of Biotechnology of Antibiotics of Ministry of Health, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

X.L. and F.Y. contributed equally to this work.

Amycolatopsis orientalis CPCC200066 is an actinomycete that can produce the glycopeptide antibiotic norvancomycin, which has significant inhibitory activity against Gram-positive cocci and bacilli. Here, we report the draft genome sequence of A. orientalis CPCC200066 and identified the genes involved in norvancomycin biosynthesis.

Received 2 April 2015 Accepted 7 April 2015 Published 14 May 2015

Citation Lei X, Yuan F, Shi Y, Li X, Wang L, Hong B. 2015. Draft genome sequence of norvancomycin-producing strain *Amycolatopsis orientalis* CPCC200066. Genome Announc 3(3):e00296-15. doi:10.1128/genomeA.00296-15.

**Copyright** © 2015 Lei et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Lifei Wang, lifeiwang2002@hotmail.com, or Bin Hong, binhong69@hotmail.com.

mycolatopsis orientalis CPCC200066 (B-37) was initially isolated from a soil sample in Guizhou Province, China, in 1959 (1) and was obtained by the China Pharmaceutical Culture Collection (CPCC). It produces the glycopeptide antibiotic norvancomycin, whose chemical structure is almost the same as that of vancomycin, except for an absent methyl group at the N terminus. Norvancomycin shows pharmacological properties and antibacterial activities similar to those of vancomycin, which exhibits significant inhibitory activity against Gram-positive cocci and bacilli, especially to methicillin-resistant Staphylococcus aureus (MRSA) and methicillin-resistant Staphylococcus epidermidis (MRSE). Due to its potential clinical application, norvancomycin was commercially developed by the North China Pharmaceutical Company and has been widely used in China to treat endocarditis, osteomyelitis, and other severe infections caused by S. aureus (including methicillin-resistant strains) (2) for >3 decades. Genome sequencing of A. orientalis CPCC200066 may provide information for discovering the biosynthetic gene cluster for norvancomycin. Here, we present a draft genome sequence of A. orientalis CPCC200066.

The strain was grown at 28°C in tryptic soy broth (TSB) liquid medium (2% tryptone, 0.5% NaCl, 0.25% glucose, 0.25%  $K_2$ HPO<sub>4</sub> [pH 7.2]), and genomic DNA was extracted using the DNA extraction kit (TANBead, China). Sequencing was performed using an Illumina HiSeq 2000 platform at the Beijing Genomics Institute (BGI) (Shenzhen, China), resulting in 14,002,362 reads with 39-fold average coverage. Short reads were assembled by SOAP*denovo* 2.04 (3), and Glimmer 3.02 was used to predict protein-coding sequences (CDSs) (4). The draft genome sequence was annotated based on the KEGG, COG, SwissProt, NR, GO, and PHI databases.

The draft genome sequence of *A. orientalis* CPCC200066 is 9,438,289 bp, with a G+C content of 68.85% distributed over 59 scaffolds containing 88 contigs. The  $N_{50}$  length of the scaffolds was 402,393 bp, and that of the contigs was 246,913 bp. A total of 8,233 CDSs were predicted, with an 86.36% coding density and a 990-bp

average length. For these CDSs, >300 proteins were identified as being involved in secondary metabolism, and this will help us to explore novel secondary metabolites in A. orientalis CPCC200066. The gene cluster responsible for norvancomycin biosynthesis was assigned on scaffold 20 and consists of 30 genes, which shows a high similarity with reported gene clusters of the glycopeptide antibiotics chloroeremomycin (5, 6) and vancomycin (7, 8). Consistent with the chemical structure difference of lacking one vancosamine compared to chloroeremomycin, the norvancomycin biosynthetic gene cluster contained only two glycosyltransferase genes compared with three in the chloroeremomycin cluster. Interestingly, one N-methylase gene was identified in the norvancomycin biosynthetic gene cluster as in the vancomycin biosynthetic clusters, while the main vancomycin analogue of CPCC200066 is absent of an N-methyl group. The function of the N-methylase gene in the norvancomycin biosynthetic gene cluster and the biosynthetic mechanism of norvancomycin in CPCC200066 warrant further investigation. The genome sequence of CPCC200066 will aid in further studies for designing strategies for the construction of strains with enhanced norvancomycin production and the discovery of new natural products by uncovering cryptic metabolic pathways.

Nucleotide sequence accession numbers. The whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession no. JXRD00000000. The version described in this paper is the first version, JXRD01000000.

## ACKNOWLEDGMENTS

This work was supported by the National Mega-Project for Innovative Drugs (grant 2014ZX09201001-004-001), the National Natural Science Foundation of China (grants 31170042, 81302677, 30973668, and 81402836), the Beijing Natural Science Foundation (grant 5102032), and the Fundamental Research Funds for the Central Universities (grants 2012N09 and 3332013088).

We thank the BGI (Shenzhen, China) for technical assistance.

## REFERENCES

- Li Q, Song AL, Liu JR, Wang XY. 1962. Actinomycetes Van23-vancomycin producing strain, p 85–91. In Tong C, Zhang WS (ed), Studies on antibiotic I. Shanghai Scientific & Technical Publishers, Shanghai, China. (In Chinese.)
- Wu XJ, Zhang J, Yu JC, Cao GY, Shi YG, Zhang YY, Wang MG. 2012. Establishment of norvancomycin fluorescence polarization immunoassay for therapeutic drug monitoring. J Antibiot (Tokyo) 65:35–39. http:// dx.doi.org/10.1038/ja.2011.89.
- Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J. 2010. *De novo* assembly of human genomes with massively parallel short read sequencing. Genome Res 20:265–272. http://dx.doi.org/10.1101/gr.097261.109.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23: 673–679. http://dx.doi.org/10.1093/bioinformatics/btm009.
- Van Wageningen AM, Kirkpatrick PN, Williams DH, Harris BR, Kershaw JK, Lennard NJ, Jones M, Jones SJ, Solenberg PJ. 1998. Sequencing and analysis of genes involved in the biosynthesis of a vancomycin group antibiotic. Chem Biol 5:155–162. http://dx.doi.org/10.1016/S1074 -5521(98)90060-6.
- Hubbard BK, Walsh CT. 2003. Vancomycin assembly: nature's way. Angew Chem Int Ed Engl 42:730–765. http://dx.doi.org/10.1002/anie.200390202.
- Xu L, Huang H, Wei W, Zhong Y, Tang B, Yuan H, Zhu L, Huang W, Ge M, Yang S, Zheng H, Jiang W, Chen D, Zhao GP, Zhao W. 2014. Complete genome sequence and comparative genomic analyses of the vancomycin-producing *Amycolatopsis orientalis*. BMC Genomics 15:363. http://dx.doi.org/10.1186/1471-2164-15-363.
- Jeong H, Sim YM, Kim HJ, Lee DW, Lim SK, Lee SJ. 2013. Genome sequence of the vancomycin-producing *Amycolatopsis orientalis* subsp. *orientalis* strain KCTC 9412<sup>T</sup>. Genome Announc 1(3):e00408-13. http:// dx.doi.org/10.1128/genomeA.00408-13.