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## EGFR: The Paradigm of an Oncogene-Driven Lung Cancer

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### Abstract

Somatic, activating mutations in Epidermal Growth Factor Receptor (*EGFR*) identify a significant minority of patients with non-small cell lung cancer (NSCLC). While these mutations are associated with an ~70% response rate to some EGFR tyrosine kinase inhibitors (gefitinib, erlotinib, and afatinib), patients develop resistance (i.e. “acquired resistance”) after a median of 9–12 months. In patients with clinical acquired resistance, repeat biopsy of tumors has identified a number of relevant mechanisms of resistance, but by far the most frequent event is the acquisition of EGFR T790M, a mutation in the “gatekeeper” residue that confers resistance to gefitinib, erlotinib, and afatinib. This emphasizes the critical dependence upon EGFR signaling for some tumors, a property that has been exploited therapeutically. Dual EGFR blockade using afatinib and cetuximab led to a 29% radiographic response rate. More recently, drugs which target EGFR T790M (e.g. rociletinib, AZD9291, and others) have entered clinical trials, with impressive results observed in phase 1 clinical trials. The development of these newer drugs, with efficacy after resistance to first line EGFR TKI, has led to exploration of these strategies in multiple disease settings: at resistance, in the first line, and adjuvant treatment of those with completely resected early stage disease who would otherwise die of recurrent/metastatic disease. This example of translational research that identifies mechanisms of resistance to first generation drugs, and then targets those mechanisms yielding clinical benefit is a paradigm for how targeted therapies can be developed.

### Introduction

Drugs targeting the epidermal growth factor receptor (EGFR) began initial development in the late 1990s and were hypothesized to be effective since a variety of epithelial malignancies, including non-small cell lung cancer (especially squamous cell lung cancer (1)) overexpressed EGFR protein. The earliest molecules to reach the clinic were cetuximab, gefitinib, and erlotinib. While the anti-EGFR antibody cetuximab did not show significant clinical activity (as a single-agent or in combination with chemotherapy), in large clinical trials both gefitinib and erlotinib had single-agent activity, with response rates <10% (2–5).

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#### Disclosure of Potential Conflicts of Interest

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This modest single agent activity led to initial regulatory approvals for both erlotinib and gefitinib in patients with previously treated advanced non-small cell lung cancer (Table 1). The US FDA approval for erlotinib was based on improvement in overall survival, compared to placebo, for an unselected group of patients with previously treated lung cancer (5). Gefitinib received an accelerated approval based upon response rate in single-arm trials of pretreated patients (2, 3), contingent upon the results of subsequent randomized trials. After a randomized trial comparing gefitinib and best supportive care to best supportive care alone in patients with previously treated lung cancer (analogous to (5) with erlotinib) failed to show an improvement in overall survival (6), in 2005, the US label for gefitinib was changed to effectively withdraw its approval.

Despite the low frequency of overall response rates, these clinical trials provided an opportunity to observe dramatic radiographic and clinical responses in a small proportion of patients treated with erlotinib or gefitinib. Initial trials noted higher response rates for patients from Asia, those who were never smokers, and those patients with adenocarcinoma histology (2, 7). Molecular analysis of tumors from patients with radiographic responses led to the identification of somatic activating mutations in the *EGFR* gene that were present more frequently in patients with response to erlotinib or gefitinib (8–10). These seminal papers identified the activating characteristics of these mutations and their association with response to erlotinib and gefitinib.

While the data regarding *EGFR* mutations and their association with response was clear, *EGFR* mutations occurred in just 10–20% of patients and in the early development of EGFR TKIs, some investigators explored the role of other predictive biomarkers, including *EGFR* copy number (not frank amplification but rather increased copy number). The IPASS trial was the single trial which best clarified the predictive nature of *EGFR* mutations (11). This trial randomized patients with clinical factors predictive of response to EGFR TKI (East Asian patients, never smokers, adenocarcinoma) to either gefitinib or paclitaxel and carboplatin. In the ensuing biomarker analysis, despite analyzing EGFR IHC, *EGFR* copy number, and clinical factors, the best predictor of response was *EGFR* mutations and any predictive effect of IHC or *EGFR* copy number was driven by their association with *EGFR* mutation. This led to the European approval of gefitinib as first-line treatment of *EGFR* mutant lung adenocarcinoma. Arguments have been made that a similar approval would be appropriate in the United States as well (12).

Subsequent to IPASS, multiple randomized phase 3 trials explored the use of erlotinib, gefitinib, and alectinib (an irreversible kinase inhibitor that blocks both EGFR and HER2) as first line treatment, in comparison with conventional platinum-based chemotherapy doublets (13–16). In each of these trials, which prospectively enrolled only patients with *EGFR* mutant lung cancer, the EGFR TKI improved the progression-free survival as compared to chemotherapy. Taken together these trials made clear that *EGFR* mutations were the predictor of choice and that EGFR TKI were the standard of care first line treatment.

The initial translational work to identify somatic activating *EGFR* mutations and their association with responsiveness to EGFR TKI, followed by rigorous randomized clinical trials have established that *EGFR* mutant lung cancer is a distinct clinical entity. These

discoveries have led to a new paradigm in how we understand lung cancer, how we identify new drug targets, and how we treat all types of lung cancer.

## Differences Among EGFR Mutations

With the initial discovery of *EGFR* mutations, the frequency of individual sensitizing *EGFR* mutations (exon 19 deletion L858R, G719X, etc.) was typically described but no additional significance was related to these individual genotypes. The two most common *EGFR* mutations, *EGFR* exon 19 deletion and L858R, typically represent the vast majority of patients identified. However, individual uncommon mutations, including G719, L861, and rarer mutations, are identified. The clinical relevance of these mutations is not clear and, many randomized trials have excluded such patients. More recently, larger retrospective series (17, 18) have been reported which suggest that many of these mutations are associated with good response rates to EGFR TKI (~50%) and there is a clear need for online resources that allow collation of results for uncommon mutations (such as My Cancer Genome [19]).

While some preliminary data with erlotinib and gefitinib (20, 21) suggested that the two most common *EGFR* genotypes (*EGFR* exon 19 deletion and *EGFR* L858R) may predict different outcomes, the dramatic sensitivities of all EGFR mutations to EGFR TKI were the focus. More recently, additional data have explored the difference between *EGFR* exon 19 deletion and *EGFR* L858R, the two most common EGFR mutation genotypes. In a combined analysis of randomized afatinib versus chemotherapy clinical trials (Lux Lung 3 and 6), investigators found that patients with *EGFR* exon 19 deletion randomized to initial chemotherapy had shorter overall survival compared to those patients allocated to afatinib (22). In contrast, patients with *EGFR* L858R who had been assigned to chemotherapy had a similar overall survival to those patients with initially assigned to afatinib, suggesting that patients with *EGFR* L858R had less of a benefit with afatinib than those with exon 19 deletion. More recently, circulating tumor DNA data from the EURTAC trial (a randomized trial of erlotinib vs chemotherapy) demonstrated significant difference in outcome based upon *EGFR* genotype, with a poorer outcome for patients with *EGFR* L858R (23). These data have re-emphasized the notion that there may be a differential effect of EGFR tyrosine kinase inhibitors for the two most common genotypes of *EGFR* mutation. While initial data supported this distinction based on findings with erlotinib and gefitinib, these new data extend these findings to afatinib.

## Mechanisms of Acquired Resistance to EGFR Tyrosine Kinase Inhibitors

After initial response to EGFR tyrosine kinase inhibitors, patients typically develop progression of disease after 9–12 months. Understanding how resistance develops in such patients remains a key question. Multiple pre-clinical and clinical approaches have been used to understand mechanisms of resistance to tyrosine kinase inhibitors with a broad list of pathways implicated (Table 2). Initial focused sequencing analysis of biopsy specimens from patients with acquired resistance looking for secondary mutations (built upon the identification of gatekeeper mutations in the BCR-ABL fusion oncogene in patients who had become resistant to imatinib) led to the identification of EGFR T790M as a secondary mutation in EGFR that was associated with acquired resistance (24, 25). In the laboratory,

investigators have developed cell lines and in vivo tumor xenografts that are resistant to EGFR TKIs. Analysis of such cell lines has helped to identify such findings as *MET* amplification (26, 27), AXL overexpression (28), and epithelial to mesenchymal transition (29). Similarly, analysis of genetically engineered mouse models of *EGFR* mutant cancers (mice with inducible expression of various mutant *EGFRs* that develop lung cancers which mimic the clinical responsiveness of human tumors to EGFR TKI) has been used, with resulting observations including the upregulation of the gene for PDL-1 (30). These data implicate development of an immunosuppressive environment in tumors with resistance to EGFR TKI. Immunotherapies currently in development may play a role in the treatment of EGFR mutant lung cancers (31).

In multiple biopsy series with analysis of a number of the previously reported mechanisms of resistance for the frequency of these events (though there has been little comprehensive analysis of samples for all reported mechanisms of resistance), it has become apparent that the most frequently identified mechanism of acquired resistance is the secondary mutation in EGFR T790M, occurring in >60% of tumors (32–34). This secondary mutation in EGFR is thought to alter kinase ATP affinity above that of gefitinib or erlotinib (35). The secondary mutations in EGFR emphasize the continued dependence upon EGFR signaling which is required for patients with *EGFR* mutant lung cancer. Without a complete understanding of mechanisms of acquired resistance to EGFR TKI and prior to development of drugs that target EGFR T790M, investigators explored a broad array of single-agent and combination therapies in the setting of acquired resistance to erlotinib, gefitinib, and afatinib with generally dismal results (reviewed in Yu et al (36)).

While understanding the molecular mechanisms of acquired resistance to EGFR TKIs has been important in developing new therapies, some clinical resistance occurs in individual sites (such as isolated central nervous system [CNS] lesions as well as visceral sites). A variety of approaches have been developed to deal with resistance to EGFR TKIs clinically, prior to development of new therapies. Primarily, these strategies revolve around the use of local therapies. Two groups have reported interesting retrospective series which suggest that local therapies (radiation or surgery) for CNS progression or extra-CNS progression may delay the time until an additional change in therapy is required (37, 38).

## Targeting Acquired Resistance to Erlotinib and Gefitinib with Dual Blockade of EGFR

The high frequency of EGFR T790M mutation in the acquired resistance setting (and other, less common secondary mutations such as T854A and D761Y), emphasized that continued signaling through EGFR was critical to survival of *EGFR* mutant lung cancer cells (34, 39, 40). This observation led to the hypothesis that dual EGFR blockade using an EGFR tyrosine kinase coupled with an antibody to EGFR (i.e. cetuximab) could sufficiently dampen EGFR signaling (even in the context of EGFR T790M) thus leading to cancer cell apoptosis (41). While an initial trial of erlotinib combined with cetuximab showed only modest activity and no RECIST-defined partial responses, there was significantly greater activity with the combination of afatinib and cetuximab (42, 43). After an initial dose escalation which defined the tolerability of full doses of afatinib and cetuximab in

combination, a broader study of efficacy was reported (42). In this study of both patients with and without EGFR T790M, but with clinically proven resistance to erlotinib or gefitinib, there was a response rate of 29% and a median progression-free survival of 5 months. The response rate understates the evidence of tumor shrinkage in the majority of patients treated with this combination therapy. Enthusiasm for the efficacy of this combination has led to a clinical trial that compares the efficacy and tolerability of the combination of afatinib and cetuximab to afatinib alone as initial therapy for *EGFR* mutant lung cancers.

## EGFR T790M-Directed Therapies

As the primary mechanism of acquired resistance, EGFR T790M was a clear target for drug development to address this important medical need. Moreover, a drug which preferentially targeted mutant EGFR (T790M along with the activating mutations) would likely reduce WT EGFR related adverse events often observed (including rash and diarrhea). There was a sea change in drug development for patients with acquired resistance to first generation EGFR TKI with the first described compound directed at EGFR T790M, WZ4002 (44). This drug, identified by screening an irreversible kinase inhibitor library for drugs that bound EGFR T790M, was 100 fold less potent against WT EGFR and 30–100 fold more potent against EGFR T790M. This compound binds irreversibly to mutant *EGFR* at the C797 residue. While this compound was not taken forward into clinical development, multiple other drugs with similar characteristics have begun early phase clinical trials (see Table 1). The first reported clinical data came with rociletinib (CO-1686, Clovis Oncology) (45) with clear evidence of single-agent clinical activity in patients with acquired resistance to erlotinib or gefitinib (46). Subsequently, AZD9291 (AstraZeneca) (47) entered phase 1 clinical trials in patients with *EGFR* mutant lung cancer and acquired resistance to erlotinib, gefitinib, or afatinib (48). Both of these drugs have been explored in single-arm studies with relatively large numbers of patients. A number of other compounds have more recently entered clinical trials with early clinical data anticipated in 2015 (Table 1).

AZD9291 and rociletinib, the EGFR T790M-directed drugs with the most data, have reported similar high response rates > 50%, but they appear to have unique patterns of adverse events which may distinguish some members of this class of drugs. Rociletinib has an 22% rate of observed grade 3 hyperglycemia and grade 3 QTc prolongation has been reported in 7%, but the worst rash reported was grade 1 (46). AZD9291 has been reported to have grade 3 rash and diarrhea in 2–5% (48). In early reports, there were no grade 3 events of QTc prolongation or hyperglycemia with AZD9291. These differences in adverse events may allow for preferential combination therapies with other classes of drugs.

While AZD9291 and rociletinib have shown impressive response rates as single-agent kinase inhibitors, it is likely that clinically meaningful drug resistance will occur for this class of drugs after a relatively short time, emphasizing the importance of beginning to understand mechanisms of resistance to these drugs. Potential mechanisms of resistance to EGFR T790M-specific kinase inhibitors have been identified in pre-clinical work. In the initial presentation of the WZ4002 compound, Zhou and colleagues identified the C797 binding site as a potential source of resistance and went on to show that a C797S mutation

would lead to a 100 fold increase in IC50 for cell lines with this mutation (44). Similarly, development of a cell line resistant to WZ4002 implicated amplification of the *MAPK1* gene and upregulation of MAPK signaling as mechanisms of acquired resistance to EGFR TKI treatment (49). Importantly, treatment with a MEK inhibitor restored EGFR-TKI sensitivity in their models. Combining the EGFR T790M specific drug with a MEK inhibitor significantly delayed the emergence of resistance *in vitro*, suggesting one of many rational combinations that will be explored as development of T790M specific drugs moves forward. In addition, in a cell line that was developed to be resistant to rociletinib, elevated levels of phosphorylated AKT were observed (50). Moreover, addition of an AKT inhibitor made the cell line sensitive to rociletinib. The impressive clinical activity observed with 3<sup>rd</sup>-generation, mutant specific EGFR TKI has led to a rapid move to compare these drugs to erlotinib, gefitinib, and afatinib, in the first line treatment of patients with *EGFR* mutant NSCLC.

### Can EGFR TKIs Be Used to Cure Some *EGFR* Mutant Lung Cancers?

Since the identification of *EGFR* mutations and their association with response to EGFR TKI in 2004, there have been intermittent calls for a targeted trial of EGFR TKI in the adjuvant setting for patients with early stage *EGFR* mutant lung cancer to build upon the palliative results seen in the advanced stage setting. While retrospective data support the use of EGFR TKI in the adjuvant setting, with likely improved disease-free and overall survival, only modest prospective randomized data exist (51). During the same time period, investigators have demonstrated that imatinib prolongs overall survival when given as adjuvant therapy for patients with resected gastrointestinal stromal tumors (GIST) (52). GIST investigators have gone on to show that 3 years of adjuvant imatinib is better than 1 year of adjuvant imatinib (53). With an annual incidence of 4-6000 cases per year in the US, these trials were completed in relatively short order.

Three clinical trials have been reported where the prospective study of adjuvant EGFR TKI was explored in patients with *EGFR* mutant lung cancer, all with significant methodological limitations. The BR.19 trial was a placebo-controlled trial of adjuvant gefitinib for patients with completely resected lung cancers that was stopped early after a trial of gefitinib in unselected patients with advanced lung cancer failed to improve overall survival (54). While an unselected patient population was included in this trial, *EGFR* mutation testing was done on available specimens and just 15 patients had *EGFR* mutant lung cancer, prohibiting meaningful analysis of the results. Similarly, the RADIANT study explored the value of adjuvant erlotinib in patients with completely resected lung cancer who had over-expression or increased gene copy number of *EGFR* (55). Subset analysis of the small proportion of patients with *EGFR* mutant lung cancer showed a disease-free survival benefit (that, due to hierarchical testing, was not deemed statistically significant), but was underpowered to detect a survival advantage. Finally, a randomized trial of adjuvant gefitinib after all patients with stage IIIA *EGFR* mutant lung cancer were treated with pemetrexed and carboplatin showed improved disease-free survival, and a trend toward improved OS, but, with just 60 patients, the study was underpowered to show statistically significant overall survival differences (56). Fortunately, there are multiple ongoing randomized studies that have enrolled patients with *EGFR* mutant lung cancer to treatment with erlotinib or gefitinib

(Table 3). In North America, the most prominent trial is the NCI-sponsored ALCHEMIST EGFR trial that seeks to enroll 410 people with resected, Stage IB–III, *EGFR* mutant lung cancers previously treated with standard chemotherapy to either erlotinib or placebo (57). The investigators are seeking to show a Hazard Ratio of 0.67 for overall survival, its primary endpoint. Similar trials are enrolling patients in Asia.

While there are ongoing trials of erlotinib or gefitinib in the adjuvant setting and retrospective data suggest that this strategy will improve overall survival, there are questions about whether these studies use a duration of therapy which is likely to show benefit and whether the optimal drug is being used. As described above, in GIST, there is evidence that 3 years of imatinib is better than 1 year. Similarly, in women with estrogen receptor expressing breast cancers, the use of adjuvant hormonal therapy is now recommended for 10 years for some patients. The early impressive clinical data for patients with the EGFR inhibitors which target EGFR T790M (i.e. rociletinib and AZD9291) in conjunction with a suggestion of better tolerability (lower rates of grade 3 rash, grade 3 diarrhea), would seem to make them better candidates for testing in the adjuvant setting. With these caveats, it is likely that optimization of duration of therapy and identification of the right drug can be accomplished and that the use of EGFR TKI may cure some patients with early stage resected *EGFR* mutant lung cancers who might otherwise die from recurrent or metastatic disease.

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## References

1. Gandara DR, Hammerman PS, Sos ML, Lara PN Jr, Hirsch FR. Squamous cell lung cancer: from tumor genomics to cancer therapeutics. *Clin Cancer Res.* 2015; 21:xxx–xxx.
2. Fukuoka M, Yano S, Giaccone G, Tamura T, Nakagawa K, Douillard JY, et al. Multi-institutional randomized phase II trial of gefitinib for previously treated patients with advanced non-small-cell lung cancer (The IDEAL 1 Trial). *J Clin Oncol.* 2003; 21:2237–46. Erratum in *J Clin Oncol* 2004, 22, 4863. [PubMed: 12748244]
3. Kris MG, Natale RB, Herbst RS, Lynch TJ Jr, Prager D, Belani CP, et al. Efficacy of gefitinib, an inhibitor of the epidermal growth factor receptor tyrosine kinase, in symptomatic patients with non-small cell lung cancer: a randomized trial. *JAMA.* 2003; 290:2149–58. [PubMed: 14570950]
4. Pirker R, Szczesna A, von Pawel J, Krzakowski M, Ramlau R, Park K, et al. FLEX: A randomized, multicenter, phase III study of cetuximab in combination with cisplatin/vinorelbine (CV) versus CV alone in the first-line treatment of patients with advanced non-small cell lung cancer (NSCLC). *J Clin Oncol.* 2008; 26(15 Suppl 3)
5. Shepherd FA, Pereira JR, Ciuleanu T, Tan EH, Hirsh V, Thongprasert S, et al. Erlotinib in previously treated non-small-cell lung cancer. *N Engl J Med.* 2005; 353:123–32. [PubMed: 16014882]
6. Thatcher N, Chang A, Parikh P, Rodrigues Pereira J, Ciuleanu T, von Pawel J, et al. Gefitinib plus best supportive care in previously treated patients with refractory advanced non-small-cell lung cancer: results from a randomised, placebo-controlled, multicentre study (Iressa Survival Evaluation in Lung Cancer). *Lancet.* 2005; 366:1527–37. [PubMed: 16257339]

7. Miller VA, Kris WG, Shah N, Patel J, Azzoli C, Gomez J, et al. Bronchioloalveolar pathologic subtype and smoking history predict sensitivity to gefitinib in advanced non-small-cell lung cancer. *J Clin Oncol*. 2004; 22:1103–9. [PubMed: 15020612]
8. Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med*. 2004; 350:2129–39. [PubMed: 15118073]
9. Paez JG, Janne PA, Lee JC, Tracy S, Greulich H, Gabriel S, et al. EGFR mutations in lung cancer: Correlation with clinical response to gefitinib therapy. *Science*. 2004; 304:1497–500. [PubMed: 15118125]
10. Pao W, Miller V, Zakowski M, Doherty J, Politi K, Sarkaria I, et al. EGF receptor gene mutations are common in lung cancers from “never smokers” and are associated with sensitivity of tumors to gefitinib and erlotinib. *Proc Natl Acad Sci U S A*. 2004; 101:13306–11. [PubMed: 15329413]
11. Mok TS, Wu YL, Thongprasert S, Yang CH, Chu DT, Saijo N, et al. Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. *N Engl J Med*. 2009; 361:947–57. [PubMed: 19692680]
12. Burotto M, Manasanch EE, Wilkerson J, Fojo T. Gefitinib and erlotinib in metastatic non-small cell lung cancer: a meta-analysis of toxicity and efficacy of randomized clinical trials. *Oncologist*. 2015 Mar 20. Epub ahead of print.
13. Maemondo M, Inoue A, Kobayashi K, Sugawara S, Oizumi S, Isobe H, et al. Gefitinib or chemotherapy for non-small-cell lung cancer with mutated EGFR. *N Engl J Med*. 2010; 362:2380–8. [PubMed: 20573926]
14. Mitsudomi T, Morita S, Yatabe Y, Negoro S, Okamoto I, Tsurutani J, et al. Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung cancer harbouring mutations of the epidermal growth factor receptor (WJTOG3405): an open label, randomised phase 3 trial. *Lancet Oncol*. 2010; 11:121–8. [PubMed: 20022809]
15. Rosell R, Carcereny E, Gervais R, Vergnenegre A, Massuti B, Felip E, et al. Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): a multicentre, open-label, randomised phase 3 trial. *Lancet Oncol*. 2012; 13:239–46. [PubMed: 22285168]
16. Sequist LV, Yang JCH, Yamamoto N, O’Byrne K, Hirsh V, Mok T, et al. Phase III study of afatinib or cisplatin plus pemetrexed in patients with metastatic lung adenocarcinoma with EGFR mutations. *J Clin Oncol*. 2013; 31:3327–34. [PubMed: 23816960]
17. Watanabe S, Minegishi Y, Yoshizawa H, Maemondo M, Inoue A, Sugawara S, et al. Effectiveness of gefitinib against non-small-cell lung cancer with the uncommon EGFR mutations G719X and L861Q. *J Thorac Oncol*. 2014; 9:189–94. [PubMed: 24419415]
18. Wu JY, Yu CJ, Chang YC, Yang CH, Shih JY, Yang PC. Effectiveness of tyrosine kinase inhibitors on “uncommon” epidermal growth factor receptor mutations of unknown clinical significance in non-small cell lung cancer. *Clin Cancer Res*. 2011; 17:3812–21. [PubMed: 21531810]
19. My Cancer Genome [database on the Internet]. Nashville (TN): Vanderbilt-Ingram Cancer Center; c2010–2015. [cited 2015 Mar 26]. Available from: <http://www.mycancergenome.org/>
20. Jackman DM, Yeap BY, Sequist LV, Lindeman N, Holmes AJ, Joshi VA, et al. Exon 19 deletion mutations of epidermal growth factor receptor are associated with prolonged survival in non-small cell lung cancer patients treated with gefitinib or erlotinib. *Clin Cancer Res*. 2006; 12:3908–14. [PubMed: 16818686]
21. Riely GJ, Pao W, Pham DK, Li AR, Rizvi N, Venkatraman ES, et al. Clinical course of patients with non-small cell lung cancer and epidermal growth factor receptor exon 19 and exon 21 mutations treated with gefitinib or erlotinib. *Clin Cancer Res*. 2006; 12:839–44. [PubMed: 16467097]
22. Yang JCH, Wu YL, Schuler M, Sebastian M, Popat S, Yamamoto N, et al. Afatinib versus cisplatin-based chemotherapy for EGFR mutation-positive lung adenocarcinoma (LUX-Lung 3 and LUX-Lung 6): analysis of overall survival data from two randomised, phase 3 trials. *Lancet Oncol*. 2015; 16:141–51. [PubMed: 25589191]
23. Karachaliou N, Mayo C, Queralt C. Association of EGFR L858R mutation in circulating free DNA with survival in the EURTAC Trial. *JAMA Oncol*. 2015 Feb 26. Epub ahead of print.



24. Kobayashi S, Boggon TJ, Dayaram T, Janne PA, Kocher O, Meyerson M, et al. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. *N Engl J Med*. 2005; 352:786–92. [PubMed: 15728811]
25. Pao W, Miller VA, Politi KA, Riely GJ, Somwar R, Zakowski MF, et al. Acquired resistance of lung adenocarcinomas to gefitinib or erlotinib is associated with a second mutation in the EGFR kinase domain. *PLoS Med*. 2005; 2:225–35.
26. Bean J, Brennan C, Shih JY, Riely G, Viale A, Wang L, et al. MET amplification occurs with or without T790M mutations in EGFR mutant lung tumors with acquired resistance to gefitinib or erlotinib. *Mol Cancer Ther*. 2007; 6:3333s–4s.
27. Engelman JA, Zejnullahu K, Mitsudomi T, Song YC, Hyland C, Park JO, et al. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science*. 2007; 316:1039–43. [PubMed: 17463250]
28. Zhang ZF, Lee JC, Lin LP, Olivas V, Au V, LaFramboise T, et al. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat Genet*. 2012; 44:852–60. [PubMed: 22751098]
29. Suda K, Tomizawa K, Fujii M, Murakami H, Osada H, Maehara Y, et al. Epithelial to mesenchymal transition in an epidermal growth factor receptor-mutant lung cancer cell line with acquired resistance to erlotinib. *J Thorac Oncol*. 2011; 6:1152–61. [PubMed: 21597390]
30. Akbay EA, Koyama S, Carretero J, Altabef A, Tchaicha JH, Christensen CL, et al. Activation of the PD-1 pathway contributes to immune escape in EGFR-driven lung tumors. *Cancer Discov*. 2013; 3:1355–63. [PubMed: 24078774]
31. Soria J-C, Marabelle A, Brahmer JR, Gettinger S. Immune checkpoint modulation for non-small cell lung cancer. *Clin Cancer Res*. 2015; 21:xxx–xxx.
32. Kim Y, Ko J, Cui Z, Abolhoda A, Ahn JS, Ou SH, et al. The EGFR T790M mutation in acquired resistance to an irreversible second-generation EGFR inhibitor. *Mol Cancer Ther*. 2012; 11:784–91. [PubMed: 22228822]
33. Sequist LV, Waltman BA, Dias-Santagata D, Digumarthy S, Turke AB, Fidias P, et al. Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. *Sci Transl Med*. 2011; 3:75ra26.
34. Yu HA, Arcila ME, Rekhtman N, Sima CS, Zakowski MF, Pao W, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clin Cancer Res*. 2013; 19:2240–7. [PubMed: 23470965]
35. Yun CH, Mengwasser KE, Toms AV, Woo MS, Greulich H, Wong KK, et al. The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. *Proc Natl Acad Sci U S A*. 2008; 105:2070–5. [PubMed: 18227510]
36. Yu HA, Riely GJ, Lovly CM. Therapeutic strategies utilized in the setting of acquired resistance to EGFR tyrosine kinase inhibitors. *Clin Cancer Res*. 2014; 20:5898–907. [PubMed: 25303979]
37. Weickhardt AJ, Scheier B, Burke JM, Gan G, Lu X, Bunn PA Jr, et al. Local ablative therapy of oligoprogressive disease prolongs disease control by tyrosine kinase inhibitors in oncogene-addicted non-small-cell lung cancer. *J Thorac Oncol*. 2012; 7:1807–14. [PubMed: 23154552]
38. Yu HA, Sima CS, Huang J, Solomon SB, Rimner A, Paik P, et al. Local therapy with continued EGFR tyrosine kinase inhibitor therapy as a treatment strategy in EGFR-mutant advanced lung cancers that have developed acquired resistance to EGFR tyrosine kinase inhibitors. *J Thorac Oncol*. 2013; 8:346–51. [PubMed: 23407558]
39. Balak MN, Gong YX, Riely GJ, Somwar R, Li AR, Zakowski MF, et al. Novel D761Y and common secondary T790M mutations in epidermal growth factor receptor - Mutant lung adenocarcinomas with acquired resistance to kinase inhibitors. *Clin Cancer Res*. 2006; 12:6494–501. [PubMed: 17085664]
40. Bean J, Riely GJ, Balak M, Marks JL, Ladanyi M, Miller VA, et al. Acquired resistance to epidermal growth factor receptor kinase inhibitors associated with a novel T854A mutation in a patient with EGFR-mutant lung adenocarcinoma. *Clin Cancer Res*. 2008; 14:7519–25. [PubMed: 19010870]

41. Regales L, Gong Y, Shen R, de Stanchina E, Vivanco I, Goel A, et al. Dual targeting of EGFR can overcome a major drug resistance mutation in mouse models of EGFR mutant lung cancer. *J Clin Invest*. 2009; 119:3000–10. [PubMed: 19759520]
42. Janjigian YY, Smit EF, Groen HJM, Horn L, Gettinger S, Camidge DR, et al. Dual inhibition of EGFR with afatinib and cetuximab in kinase inhibitor-resistant EGFR-mutant lung cancer with and without T790M mutations. *Cancer Discov*. 2014; 4:1036–45. [PubMed: 25074459]
43. Janjigian YY, Azzoli CG, Krug LM, Pereira LK, Rizvi NA, Pietanza MC, et al. Phase I/II trial of cetuximab and erlotinib in patients with lung adenocarcinoma and acquired resistance to erlotinib. *Clin Cancer Res*. 2011; 17:2521–7. [PubMed: 21248303]
44. Zhou WJ, Ercan D, Chen L, Yun CH, Li DN, Capelletti M, et al. Novel mutant-selective EGFR kinase inhibitors against EGFR T790M. *Nature*. 2009; 462:1070–4. [PubMed: 20033049]
45. Sequist LV, Soria J-C, Gadgeel SM, Wakelee HA, Camidge DR, Varga A, et al. First-in-human evaluation of CO-1686, an irreversible, selective, and potent tyrosine kinase inhibitor of EGFR T790M. *J Clin Oncol*. 2013; 31(suppl):abstr 2524.
46. Sequist LV, Soria J-C, Gadgeel SM, Wakelee HA, Camidge DR, Varga A, et al. First-in-human evaluation of CO-1686, an irreversible, highly selective tyrosine kinase inhibitor of mutations of EGFR (activating and T790M). *J Clin Oncol*. 2014; 32(suppl):5s. abstr 8010.
47. Cross DAE, Ashton SE, Ghiorghiu S, Eberlein C, Nebhan CA, Spitzler PJ, et al. AZD9291, an irreversible EGFR TKI, overcomes T790M-mediated resistance to EGFR inhibitors in lung cancer. *Cancer Discov*. 2014; 4:1046–61. [PubMed: 24893891]
48. Janne PA, Ramalingam SS, Yang JC-H, Ahn M-J, Kim D-W, Kim S-W, et al. Clinical activity of the mutant-selective EGFR inhibitor AZD9291 in patients (pts) with EGFR inhibitor-resistant non-small cell lung cancer (NSCLC). *J Clin Oncol*. 2014; 32(suppl):5s. abstr 8009.
49. Ercan D, Xu CX, Yanagita M, Monast CS, Pratilas CA, Montero J, et al. Reactivation of ERK Signaling Causes Resistance to EGFR Kinase Inhibitors. *Cancer Discov*. 2012; 2:934–47. [PubMed: 22961667]
50. Walter AO, Sjin RT, Haringsma HJ, Ohashi K, Sun J, Lee K, et al. Discovery of a mutant-selective covalent inhibitor of EGFR that overcomes T790M-mediated resistance in NSCLC. *Cancer Discov*. 2013; 3:1404–15. [PubMed: 24065731]
51. D'Angelo SP, Janjigian YY, Ahye N, Riely GJ, Chaft JE, Sima CS, et al. Distinct clinical course of EGFR-mutant resected lung cancers: results of testing of 1118 surgical specimens and effects of adjuvant gefitinib and erlotinib. *J Thorac Oncol*. 2012; 7:1815–22. [PubMed: 23154553]
52. Dematteo RP, Ballman KV, Antonescu CR, Maki RG, Pisters PW, Demetri GD, et al. Adjuvant imatinib mesylate after resection of localised, primary gastrointestinal stromal tumour: a randomised, double-blind, placebo-controlled trial. *Lancet*. 2009; 373:1097–104. [PubMed: 19303137]
53. Joensuu H, Eriksson M, Sundby Hall K, Hartmann JT, Pink D, Schutte J, et al. One vs three years of adjuvant imatinib for operable gastrointestinal stromal tumor: a randomized trial. *JAMA*. 2012; 307:1265–72. [PubMed: 22453568]
54. Goss GD, O'Callaghan C, Lorimer I, Tsao MS, Masters GA, Jett J, et al. Gefitinib versus placebo in completely resected non-small-cell lung cancer: results of the NCIC CTG BR19 study. *J Clin Oncol*. 2013; 31:3320–6. [PubMed: 23980091]
55. Kelly K, Altorki NK, Eberhardt WEE, O'Brien MER, Spigel DR, Crino L, et al. A randomized, double-blind phase 3 trial of adjuvant erlotinib (E) versus placebo (P) following complete tumor resection with or without adjuvant chemotherapy in patients (pts) with stage IB–IIIA EGFR positive (IHC/FISH) non-small cell lung cancer (NSCLC): RADIANT results. *J Clin Oncol*. 2014; 32(suppl):5s. abstr 7501.
56. Li N, Ou W, Ye X, Sun HB, Zhang L, Fang Q, et al. Pemetrexed-carboplatin adjuvant chemotherapy with or without gefitinib in resected stage IIIA-N2 non-small cell lung cancer harbouring EGFR mutations: a randomized, phase II study. *Ann Surg Oncol*. 2014; 21:2091–6. [PubMed: 24585406]
57. Gerber DE, Oxnard GR, Govindan R. ALCHEMIST: Bringing genomic discovery and targeted therapies to early-stage lung cancer. *Clin Pharmacol Ther*. 2015 Feb 11. Epub ahead of print.

58. Takezawa K, Pirazzoli V, Arcila ME, Nebhan CA, Song XL, de Stanchina E, et al. HER2 amplification: a potential mechanism of acquired resistance to EGFR inhibition in EGFR-mutant lung cancers that lack the second-site EGFR(T790M) mutation. *Cancer Discov.* 2012; 2:922–33. [PubMed: 22956644]
59. Chung JH, Rho JK, Xu X, Lee JS, Yoon HI, Lee CT, et al. Clinical and molecular evidences of epithelial to mesenchymal transition in acquired resistance to EGFR-TKIs. *Lung Cancer.* 2011; 73:176–82. [PubMed: 21168239]
60. Pirazzoli V, Nebhan C, Song XL, Wurtz A, Walther Z, Cai GP, et al. Acquired resistance of EGFR-mutant lung adenocarcinomas to afatinib plus cetuximab is associated with activation of mTORC1. *Cell Rep.* 2014; 7:999–1008. [PubMed: 24813888]
61. Zakowski MF, Ladanyi M, Kris MG. Memorial Sloan-Kettering Cancer Center Lung Cancer OncoGenome Group. EGFR mutations in small-cell lung cancers in patients who have never smoked. *N Engl J Med.* 2006; 355:213–5. [PubMed: 16837691]
62. Pietanza MC, Byers LA, Minna JD, Rudin CM. Small cell lung cancer: will recent progress lead to improved outcomes? *Clin Cancer Res.* 2015; 21:xxx–xxx.
63. Ohashi K, Sequist LV, Arcila ME, Moran T, Chmielecki J, Lin YL, et al. Lung cancers with acquired resistance to EGFR inhibitors occasionally harbor BRAF gene mutations but lack mutations in KRAS, NRAS, or MEK1. *Proc Natl Acad Sci U S A.* 2012; 109:E2127–E33. [PubMed: 22773810]
64. Postel-Vinay S, Ashworth A. AXL and acquired resistance to EGFR inhibitors. *Nat Genet.* 2012; 44:835–6. [PubMed: 22836088]
65. Ohashi K, Sequist LV, Arcila ME, Lovly CM, Chen X, Rudin CM, et al. Characteristics of Lung Cancers Harboring NRAS Mutations. *Clin Cancer Res.* 2013; 19:2584–91. [PubMed: 23515407]
66. Suda K, Mizuuchi H, Sato K, Takemoto T, Iwasaki T, Mitsudomi T. The insulin-like growth factor 1 receptor causes acquired resistance to erlotinib in lung cancer cells with the wild-type epidermal growth factor receptor. *Int J Cancer.* 2014; 135:1002–6. [PubMed: 24458568]
67. Cortot AB, Repellin CE, Shimamura T, Capelletti M, Zejnullahu K, Ercan D, et al. Resistance to irreversible EGF receptor tyrosine kinase inhibitors through a multistep mechanism involving the IGF1R pathway. *Cancer Res.* 2013; 73:834–43. [PubMed: 23172312]
68. Ware KE, Marshall ME, Heasley LR, Marek L, Hinz TK, Hercule P, et al. Rapidly acquired resistance to EGFR tyrosine kinase inhibitors in NSCLC cell lines through de-repression of FGFR2 and FGFR3 expression. *PLoS One.* 2010; 5:e14117. [PubMed: 21152424]
69. Ware KE, Hinz TK, Kleczko E, Singleton KR, Marek LA, Helfrich BA, et al. A mechanism of resistance to gefitinib mediated by cellular reprogramming and the acquisition of an FGF2-FGFR1 autocrine growth loop. *Oncogenesis.* 2013; 2:e39. [PubMed: 23552882]

**Table 1**

Representative EGFR TKIs currently in use or development

First generation (target WT EGFR)	Second generation (irreversible inhibitors of EGFR and HER2)	Third generation (EGFR mutant-specific, irreversible inhibitors)
Erlotinib *	Neratinib	Rociletinib (Clovis)
Gefitinib	Afatinib *	AZD9291 (AstraZeneca)
Icotinib	Dacomitinib	HM61713 (Hanmi) EGF816 (Novartis) ASP8273 (Astellas)

\* FDA-approved in the US for treatment of lung cancer

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**Table 2**

A partial list of mechanisms implicated in acquired resistance to EGFR TKI

<b>Secondary Mutations</b>	<b>Activation of alternate pathways</b>	<b>Other pathways</b>
<i>EGFR</i> T790M (24, 25)	<i>HER2</i> amplification (58)	Activation of PD-1 (30)
<i>EGFR</i> D761Y (39)	<i>MET</i> amplification (26, 27)	Epithelial to mesenchymal transition (29, 33, 59)
<i>EGFR</i> T854A (40)	mTORC1 (60)	Small Cell Transformation (61, 62)
<i>BRAF</i> V600E (63)	AXL (64)	
<i>NRAS</i> mutation (65)	IGF1-R (66, 67)	
	FGFR activation (68, 69)	

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**Table 3**

## Ongoing adjuvant EGFR TKI trials

<b>Trial</b>	<b>EGFR TKI</b>	<b>Comparison Arm</b>	<b>Population</b>
NCT0140214	erlotinib x 2 years	cisplatin/vinorelbine for 4 cycles	Stage III, EGFR mutations in exon 19 or 21
NCT01405079	gefitinib x 2 years	cisplatin/vinorelbine for 4 cycles	Stage II–III, EGFR exon 19 deletion or L858R
NCT01683175	erlotinib x 2 years	cisplatin/vinorelbine for 4 cycles	Stage III, EGFR exon 19 deletion or L858R
NCT02125240	icotinib x 2 years	placebo	Stage II–III, EGFR exon 19 deletion or L858R, treated with 4 cycles of platinum-based chemotherapy
NCT01746251	afatinib x 2 years	afatinib x 3 months	Stage I–III, EGFR mutation, prior chemotherapy allowed
NCT01929200	icotinib x 2 years	icotinib x 1 year	Stage II–III, EGFR mutation in exon 19 or 21
NCT01996098	chemotherapy followed by icotinib x 6 or 12 months	chemotherapy alone	Stage II–III, EGFR mutation in exon 19 or 21
NCT02194738	erlotinib x 2 years	placebo	Stage IB–III, EGFR mutation
NCT02264210	icotinib x 12 months	observation	Stage IB, EGFR mutation in exon 19 or 21
WJOG6410L	gefitinib x 2 years	cisplatin/vinorelbine	Stage II–III, EGFR exon 19 deletion or L858R